

HER2-Low Breast Cancer: Current Landscape and Future Prospects

Yelena Shirman, Shlomit Lubovsky, Ayelet Shai

Division of Oncology, Rambam Health Care Campus, Haifa, Israel

Correspondence: Ayelet Shai, Division of Oncology, Rambam Health Care Campus, HaAliya HaShniya St 8, PO Box 9602, Haifa, 3109601, Israel, Tel +972-47776400, Email a_shai@rambam.health.gov.il

Abstract: More than 50% of breast cancers are currently defined as “Human epidermal growth factor receptor 2 (HER2) low breast cancer (BC)”, with HER2 immunohistochemistry (IHC) scores of +1 or +2 with a negative fluorescence in situ hybridization (FISH) test. In most studies that compared the clinical and biological characteristics of HER2-low BC with HER2-negative BC, HER2-low was not associated with unique clinical and molecular characteristics, and it seems that the importance of HER2 in these tumors is being a docking site for the antibody portion of antibody drug conjugates (ADCs). Current pathological methods may underestimate the proportion of BCs that express low levels of HER2 due to analytical limitations and tumor heterogeneity. In this review we summarize and contextualize the most recent literature on HER2-low breast cancers, including clinical and translational studies. We also review the challenges of assessing low HER2 expression in BC and discuss the current and future therapeutic landscape for these tumors.

Keywords: HER2-low, ERBB2 low, breast cancer, HER2 targeted therapy, trastuzumab, trastuzumab-deruxtecan, T-DXd

Introduction

HER2 belongs to the epidermal growth factor (EGF) tyrosine kinase receptor family. HER2 is unique for both its functional characteristics as an orphan receptor that heterodimerizes with other tyrosine kinase receptors (TKIs) and its high oncogenic potential.¹ The HER2 gene is amplified in 15–20% of breast cancers, resulting in markedly increased HER2 protein content on the cell surface and enhanced signal transduction through HER2 heterodimers.² These tumors were associated with a higher risk of relapse and shorter survival before anti-HER2 treatments were available,^{3,4} however, the development of trastuzumab, pertuzumab and other HER2 targeting agents revolutionized the treatment and improved the prognosis of patients with advanced^{5,6} and early^{7–9} HER2-positive breast cancer.

HER2 positivity is defined by a circumferential, complete, and intense membrane immunohistochemistry (IHC) staining in >10% of the tumor, defined as IHC +3, or a weak to moderate complete membrane staining in >10% of tumor cells (IHC +2) with a positive ISH test.¹⁰ Trials designed to test the benefit of trastuzumab and of trastuzumab-emtansine in breast cancers with lower HER2 expression failed to show the benefit of these HER2 targeting agents,^{11,12} and these tumors were defined as “HER2-negative”. However, post-hoc analyses of the seminal adjuvant trastuzumab trials did not find an association between the degree of HER2 amplification or chromosome 17 polysomy and benefit from trastuzumab.^{13,14} In addition, analyses of the NSABP B-13 and the NCCTG N9831 trials suggested that some patients with tumors that were defined as HER2-positive by local pathology and HER2-negative by central pathology derived benefit from trastuzumab,^{13,15} raising questions about the impact of heterogeneity and analytical aspects of HER2 testing.

The success of clinical trials testing treatment with the HER2-directed antibody–drug conjugate T-DXd in breast cancers with a lower expression – IHC+1 and IHC+2 with a negative fluorescence in situ hybridization (FISH) test,^{16,17} previously considered “HER2-negative”, created a new terminology for these tumors, and they are now referred to as “HER2-low”. However, it remains unclear whether HER2 oncogenic signal transduction plays a role in the progression of HER2-low breast cancer, and if the mechanism underlying the anti-tumor effects of trastuzumab-deruxtecan in HER2-low cancers involves disruption of HER2 signaling or merely better cytotoxic drug delivery.¹⁸

In this review we will focus on 4 main topics: the molecular biology of HER2 in breast cancer and its relevance to HER2-low cancers, the clinical and prognostic significance of HER2-low status in local and metastatic breast cancers treated with standard regimens, the current and future landscape of HER2 targeted therapies for these tumors and the challenges of pathologic assessment of HER2-low BC.

Molecular Biology of HER2 in Breast Cancer

Mechanisms of HER2 Overexpression in Breast Cancer

To date, several molecular mechanisms of HER2 overexpression in breast cancer have been described, of these, amplification of the HER2 gene (also called *erbB-2* and *neu*) is the most common mechanism.¹⁹ Breast cancers can have up to 25–50 copies of the HER2 gene and up to 40–100-fold increases in HER2 protein expression, resulting in up to 2 million receptors expressed at the tumor cell surface.²⁰

Furthermore, it has been shown that transcriptional upregulation results in HER2 mRNA levels of 4- to 8-fold in breast cancer cells without HER2 gene amplification and of 64–128 fold in cells with HER2 gene amplification.²¹ Interplay between various promoters, promoter 1²² and promoter 2,²³ enhancers and transcription factors such as ESX,²⁴ TFAP2,^{25,26} YY1,²⁷ EGR2²⁸ is involved in HER2 transcription, resulting in overexpression.

Epigenetic mechanisms also contribute to transcriptional regulation of HER2. Histone modifications such as acquisition of H3K4me3 and H3K9ac increase *ErbB2* transcription independently from gene amplification.²⁹ It was shown that DNA hypomethylation of the HER2 gene body enhancer enables binding of the transcription factor TFAP2C, resulting in increased transcription.³⁰ Furthermore, in vitro studies have shown that disruption of ubiquitination of HER2 by chaperone-interacting protein (CHIP)^{30,31} results in reduced degradation.³² A recent study reported that HER2 neddylation, the process of removing the ubiquitin-like protein (NEDD8) from HER2, is a post-translational modification that results in reduced degradation and stabilized HER2 expression.³³

Finally, anti-cancer therapy may affect the expression of the HER2 receptor. In ER+/HER2- breast cancer cells following endocrine treatment, defects in MutL mismatch repair complex (*MLH1/3*, *PMS1/2*) result in interference with lysosomal protein trafficking. Studies suggested that HER2 levels increase with MutL loss even before endocrine therapy, uncovering another mechanism of HER2 overexpression.^{34,35} Radiation therapy can induce HER2 expression through radiation-induced NF- κ B activity. As NF- κ B activation through the PI3K/Akt pathway is a major downstream event of HER2 overexpression, a loop-like HER2-NF- κ B-HER2 pathway in radiation-induced adaptive resistant breast cancer cells has been proposed.³⁶

Of note, close inspection of the literature disclosed that some of the mechanisms were described in HER2-low tumors as well, including gene amplification,¹⁹ HER2 gene body enhancer,³⁰ radiation therapy and endocrine therapy.^{35,36}

Molecular Biology of the Carcinogenic Effect of HER2

HER2, a member of the HER receptor family consisting of 4 cell surface receptor tyrosine kinases (EGFR/*erbB1*/HER1, *erbB2*/HER2, *erbB3*/HER3, *erbB4*/HER4),^{37–39} was found to be a key player in the pathogenesis of breast cancer in 1987.²³ Since then, an abundance of experimental evidence on the molecular mechanisms underlying HER2 tumorigenesis has accumulated.⁴⁰ Signal transduction via the HER receptors is initiated upon ligand-induced⁴¹ dimerization. HER2 does not bind a ligand and is activated by heterodimerization with other HER family members. This results in transphosphorylation of their intracellular domains,⁴² which in turn activates signaling pathways, principally the PI3K/Akt, mitogen-activated protein kinase (MAPK), protein kinase C (PKC), RAS/RAF/MEK. These events lead to deregulation of the cell cycle through upregulation of various cyclins such as cyclin D1, E and CDK6, and degradation of cell cycle inhibitors such as p27Kip1 and results in proliferation, survival, and differentiation.^{43–46}

Additionally, crosstalk with other signaling pathways further promotes the carcinogenic effect of HER2. About 50% of HER2-amplified tumors express the estrogen receptor. HER2/ER crosstalk, through ligand-independent phosphorylation of estrogen-receptor (ER), results in activation and consequent enhancement of ER activity at the DNA level, stimulating cell proliferation and mutagenesis. Moreover, phosphorylation of co-repressors results in disruption of regulation of ER transcriptional complexes in the nucleus.⁴⁷

Although HER2 overexpression alone, that is, independent from mutational activation, holds tumorigenic potential, breast carcinomas can harbor HER2-activating mutations that act as oncogenic drivers.⁴⁸ Interestingly, HER2 somatic mutations occur in only about 2.7% of breast cancer patients⁴⁹ and they more frequently occur in HER2-negative or HER2-low breast carcinomas.⁴⁸

Molecular Biology of HER2-Low Breast Cancer: What is the Significance of the HER2 Level of Expression?

Recent studies have sought to investigate the molecular biology of HER2-low breast cancer. In the past decade, “Molecular portraits” of human breast tumors have been developed through hierarchical clustering methods, in which genes are grouped based on similarity in their expression pattern.⁵⁰ The most widely accepted molecular portraits, known as intrinsic subtypes, ie luminal A, luminal B, HER2-enriched, and basal-like, provide insight on the molecular biology of breast tumors and have been translated into clinical assays that are now widely implemented in clinical practice.^{51,52} Schettini et al and Agostinetto et al studied the molecular characteristics of HER2-low breast cancer using the PAM50 assay, a qRT-PCR based, 50-gene molecular profile assay.⁵¹ Both have demonstrated that HER2-low breast cancers are a heterogeneous group of tumors comprising primarily of luminal A (50.8–56.9%) and luminal B (22.8–28.8%), with a minority being HER2-enriched (3.5–3.6%) and basal-like (13.3–17.7%).^{52,53} Zahng et al evaluated HER2-low genomic profile using MammaPrint testing, a microarray-based, 70-gene expression assay⁵⁴ and Blueprint testing, a microarray-based, 80-gene molecular subtyping assay⁵⁵ and showed similar distributions of these molecular subtypes.⁵⁶ Additional studies which used the PAM50-based Prosigna assay^{57,58} and IHC-based molecular subtype distribution⁵⁹ found that molecular subtype distribution of the HER2-low subgroup was comprised predominantly of luminal B tumors (58.9–76% %) and 20–28.6% of luminal A tumors. While these studies used different assays, they consistently found a high prevalence of luminal subtypes and a low prevalence of HER2-enriched and basal-like subtypes among HR+/HER2-low tumors. Along these lines, in the studies by both Schettini et al and Agostinetto et al, proliferation-related genes and tyrosine-kinase receptor genes were more frequently over-expressed in HER2-0 tumors compared to HER2-low tumors, whereas HER2-low tumors had higher expression of luminal-related genes.

When tested separately, hormone receptor (HR) negative/ HER2-low (HR-/HER2-low) tumors were mostly basal-like, with a slightly higher proportion being HER2-enriched compared to HR-/HER2-negative tumors.⁶⁰ Zahng et al used a 520-gene panel and did not find genes or signaling pathways that were differentially expressed between HR-/HER2-low and HR-/HER2-negative tumors. Denkert et al reported that, in 556 tumors sequenced in the GeparSepto trial, there was no difference in the frequency of PIK3CA and TP53 mutations between HER2-low and HER2-negative tumors when analyzed differentially by HR expression.⁶¹ Additionally, Schettini et al emphasized the lack of difference in gene expression between the two HER2 subgroups within the triple-negative (TN) breast tumors. Taken together, these findings suggest that HR status and luminal genes, and not HER2 expression, are the key determinants of the biology of HER2-negative and HER2-low breast cancers.

Several studies evaluated the mRNA levels of ERBB2 and found them to be directly proportional to HER2 protein expression, with HER2-low mRNA levels being closer to HER2-negative levels.^{53,58,60} Higher ERBB2 expression was observed in HR+/HER2-low tumors compared with HR-/HER2-low tumors, without corresponding enrichment of the HER2-enriched subtype. This finding was explained by the fact that the HER2-enriched phenotype is not defined solely by expression of *ERBB2*.^{53,60}

Zhang et al analyzed the genomic data of 523 breast cancers by next generation sequencing using a 520-gene panel. Pathway analysis demonstrated that HER2-low tumors had significantly more mutations involved in PI3K-Akt signaling than HER2-positive and HER2-negative breast tumors, and less mutations in checkpoint genes, Fanconi anemia, and p53 signaling and cell cycle pathway compared to HER2-negative breast tumors. However, when analyzing HR-positive and HR-negative tumors separately, they could not detect significant differences between HER2-low and HER2-negative tumors.⁵⁹

Finally, Denkert et al analyzed germline mutation data from 549 patients and reported higher rates of germline BRCA1/2 mutations or other breast cancer predisposition genes (26.8% vs 18.9%) in patients with HER2-negative breast cancers compared to HER2-low breast cancers.

To date, there are no published studies that assess if HER2 is an active oncogene in HER2-low breast cancers, and it is currently unknown if activation of HER2 signaling contributes to malignant transformation and progression of these tumors. It is possible that HER2 signaling is active in the small fraction of HER2-low tumors that are defined as “HER2 enriched” by gene expression analysis,⁶⁰ but the role of HER2 in the pathogenesis of most HER2-low breast cancers is unknown.

Clinical Significance of HER2-Low Status

HER2-low status is more frequent in HR+ than HR- breast cancer,⁶² and rates of HER2-low staining increase as levels of expression of HR increase.⁶³ HER2-low status was found to be associated with better prognostic pathological tumor characteristics compared with HER2-negative, both in HR-positive and HR-negative tumors.^{62,64,65} Among HR-negative tumors, HER2-low cancers are less often “basal like” when classified by the PAM50 gene array⁶⁰ compared to HER2-negative tumors, and in another study on HR-negative tumors, HER2-low status was more frequently seen in low-grade tumors (35% vs 18%) and in tumors with apocrine IHC markers (57% vs 36%) compared to HER2-negative tumors.⁶⁶ Thus, it seems that among cancers without HER2 amplification, low HER2 expression is found more often in tumors that have markers of better prognosis.

Multiple retrospective studies looked at the association of HER2-low status and prognosis among HER2 non-amplified cancers, with conflicting results. In localized HR-positive disease, a study looking at 23,000 patients from 6 Asian centers reported better disease-free survival (DFS) and overall survival (OS) for HER2-low compared to HER2-negative breast cancers.⁶⁷ Xu et al reported that, among 678 patients with ER positive, HER2 non-amplified tumors, HER2-low cancers had similar DFS as HER2-negative tumors in the initial 5 years after diagnosis, and better DFS from year 5 onwards.⁶⁴ A study from Japan with 2890 patients found that HER2-low status was not associated with 5-year DFS and OS.⁶³ Along these lines, a recent cohort study of 1,136,016 patients from the National Cancer Database in the US similarly found that patients with HER2-low breast cancers have a similar prognosis to those with HER2-negative breast cancer.⁶⁸ Similar results were reported in additional cohort studies in several countries.^{60,63,65,69,70} Mutai et al reported that in tumors with a high 21-gene recurrence score (RS), those with HER2-low status had better DFS rates than those with HER2-negative status, despite similar clinical-pathological features. Better DFS and OS were also reported in HER2-low node-negative BC patients who did not receive any systemic adjuvant treatment.⁷¹

Several retrospective studies did not find an association between HER2-low status and DFS or OS in localized HR-negative breast cancer.^{60,63,64,66} However, a nationwide study from Korea with a median follow-up of 12 years reported better breast cancer-specific survival in patients with HR-negative HER2-low breast cancer compared with HER2-negative HR-negative breast cancer,⁶⁵ and Tan et al also reported better OS for HR-negative HER2-low breast cancer patients.⁶⁷ Jacot et al reported that, in HR-negative breast tumors, HER2 +2, ISH negative tumors had worse DFS and OS compared to HER2-negative and HER2 +1, combined.⁶⁶

The effect of HER2-low status on the prognosis of patients undergoing neoadjuvant chemotherapy was reported in several studies. A pooled analysis of neoadjuvant trials from Germany reported that HER2-low status was associated with lower pathological complete response (pCR) rate in HR-positive⁶¹ but not HR-negative breast cancer. In this study, patients with HR-negative HER2-low breast cancer had better DFS and OS, and in an exploratory analysis this difference was seen only in patients without a pCR after neoadjuvant chemotherapy. Patients with HR-positive BC had similar outcomes regardless of HER2 status. A single-center retrospective study from Korea similarly reported that in patients with HR-negative tumors, but not in those with HR-positive tumors, HER2-low status was associated with higher DFS rates after neoadjuvant chemotherapy.⁷² In contrast, 4 retrospective studies found no association between HER2-low status and prognosis in HR-positive and HR-negative BC patients undergoing neoadjuvant chemotherapy.^{73–76}

Thus, although controversial, several studies suggest that HER2-low status may be associated with better prognosis compared with HER2-negative status in high-risk tumors like HR-negative tumors without a pCR and HR-positive tumors with a high 21-gene RS, or those who did not get any adjuvant treatment. This better prognosis is achieved without anti-HER2 therapy, suggesting that HER2-low status may be a marker rather than an oncogenic driver in these tumors.

In patients with metastatic breast cancer, HER2-low status is more often found in HR-positive disease. When adjusted to other variables such as ER expression, visceral disease and age, patients with HER2-low metastatic BC were found to have a slightly better prognosis than those with HER2-negative status in a study involving more than 15,000 patients diagnosed between the years 2000–2015 (HR 0.95, CI 0.91–0.99; P = 0.02).⁷⁷ In a more recent series of patients treated with the CDK 4/6 inhibitor Palbociclib, HER2-low status was not associated with outcomes.⁷⁸ Others have also reported similar outcomes for HER2-low and HER2-negative metastatic BC patients.⁷⁹

Current Treatments and Ongoing Trials of HER2 Targeted Therapy for HER2-Low Breast Cancer

Trastuzumab deruxtecan (T-DXd) is a potent antibody drug conjugate (ADC) that has proved to be superior to trastuzumab emtansine in the 2nd line treatment of patients with HER2-positive MBC.⁸⁰ T-DXd is the 1st ADC to be approved for use in patients with HER2-low breast cancer, based on the Destiny breast -04 trial. This Phase III trial randomized 557 patients with metastatic breast cancer and centrally confirmed HER2-low (1+ or 2+ with negative FISH) expression to receive T-DXd vs standard chemotherapy (capecitabine, eribulin, gemcitabine, paclitaxel or nab-paclitaxel). All patients had received one or more prior lines of chemotherapy for metastatic breast cancer and those with HR-positive disease were endocrine refractory. At a median follow-up of 18.4 months, a 49% reduction in the risk of progressive cancer and a 36% reduction in the risk of death were observed in patients treated with T-DXd vs standard chemotherapy. Median progression-free survival was 10.1 months for the T-DXd-treated patients vs 5.4 months for those treated with standard chemotherapy. In the patients with HR-positive disease, median overall survival was 23.9 months vs 17.8 months, respectively. Median overall survival in the total study population was 23.4 months for T-DXd recipients vs 16.8 months for standard chemotherapy recipients—a significant gain of 6.6 months in median survival favoring the antibody-drug conjugate (ADC).¹⁶ In the neoadjuvant setting, t-DXd treatment achieved a high response rate of 75% when administered alone and 63% when administered with endocrine therapy in HR-positive, HER2-low localized breast cancer in the Phase II TRIO-US B12 TALENT trial.⁸¹

The activity of T-DXd is achieved by its potent payload and by the high ratio of payload molecules per ADC molecule.⁸² It is the 1st anti-HER2 agent that has shown activity in tumors that express a low amount of HER2. This has led the oncology and research community to realize that any amount of HER2 expression may be sufficient to elicit an anti-tumor response by potent ADC targeting HER2 and has further strengthened the understanding of the bystander effect, by which the chemotherapy portion of ADCs enters and kills nearby tumor cells—even those that have little or no HER2 expression. It is unclear, however, if the ability of the antibody portion of T-DXd to block HER2 signaling is part of its mechanism of action in HER2-low tumors, or if it merely serves to deliver the potent payload into HER2-expressing tumor cells.

Ongoing and pending trials in the HER2-low setting include approximately 17 recruiting studies as of a search done in January 2023. DESTINY08 is a Phase Ib trial assessing T-DXd with various combinations, including durvalumab in metastatic HER2-low breast cancer.⁸³ A Phase II trial is assessing the addition of pyrotinib, an oral inhibitor of the intracellular portion of HER2, to chemotherapy in the neoadjuvant treatment of HR-positive HER2-low breast cancer.⁸⁴

Several trials are testing novel agents in HER2-low tumors. A Phase I trial is assessing safety and efficacy of a novel anti-HER2 antibody drug conjugate MRG002 in MBC. This molecule is composed of a humanized anti-HER2 IgG1 monoclonal antibody conjugated to a microtubule disrupting agent, monomethyl auristatin E (MMAE).⁸⁵ Another Phase I study is assessing the combination of an enhancer of zeste homolog (EZH) ½ dual inhibitor, valemotostat, in combination with t-DXd.⁸⁶ Valemotostat targets epigenetic regulation by inhibiting both the EZH1 and EZH2 enzymes that act through histone methylation to regulate gene expression. In pre-clinical studies, reactivation of the silenced genes resulted in decreased proliferation of EZH2-expressing cancer cells.^{87,88}

The I-spy-P1.01 trial is evaluating the safety of a novel ADC, trastuzumab duocarmazine with weekly paclitaxel in several tumor types, including HER2-low metastatic breast cancer.⁸⁹ Trastuzumab duocarmazine consists of trastuzumab conjugated to a highly potent duocarmycin payload through maleimide attachment to interchain disulfides.⁹⁰ The Phase III TULIP trial compared T-duocarmazine with anti-HER2 therapy and chemotherapy in patients with metastatic HER2-

positive BC that progressed on 2 prior anti HER2 therapies, and resulted in improvement in progression-free survival for patients treated with T-duocarmazine.⁹¹

A pending single-arm Phase II trial will be looking at ARX788 in HER2-low MBC.⁹² ARX788 is a next-generation, site-specific anti-HER2 ADC that utilizes a unique non-natural amino acid-enabled conjugation technology and a non-cleavable amberstatin (AS269) drug-linker to generate a homogeneous ADC with a high drug-to-antibody ratio. The payload AS269 is conjugated by the synthetic amino acid para-acetylphenylalanine (pAF) to a humanized anti-HER2 mAb.⁸⁷ A recent Phase I trial demonstrated high stability and low serum exposure of pAF-AS269 resulting in low systemic toxicity.⁹³

Another Phase I trial is investigating the safety and activity of another novel ADC, BL-M07D1, in patients with HER2-amplified and HER2-low MBC. This ADC is based on trastuzumab, with a wild type Fc portion that is expected to elicit an immune response and a toxic payload.⁹⁴ The DecipHER trial is investigating a dendritic cell (DC) vaccine given with chemotherapy in several breast cancer subsets, including HER2-low BC. DCs have been in clinical use for three decades for boosting anti-tumor immunity. They present antigens to naïve T cells and polarize them into effector or tolerogenic subsets.⁹⁵ The DecipHER trial utilizes HER2 and HER3 primed dendritic cells and delivers 8 intra-tumoral injections in order to assess maximal tolerated dose and efficacy.

Pathologic Assessment of HER2 Breast Cancer

While T-DXd is approved for treatment of HER2-low breast cancer, a Phase II trial suggested that it is active in tumors that are defined as HER2-negative.⁹⁶ Since this ADC's mechanism of action involves binding to HER2, it is plausible that current immunohistochemistry does not accurately detect and quantify low levels of HER2 expression. Categorizing breast cancers as “HER2-low” carries inherent challenges,⁹⁷ and only a 26% concordance rate was found between 18 pathologists in designating breast tumors as HER2+1 or HER2-negative,⁹⁸ questioning the accuracy of the HER2-low diagnosis. An IHC score of +1 is defined as faint, barely perceptible membranous reactivity in >10% of tumor cells and an IHC score of HER2-negative is defined as no membrane staining or faint, barely perceptible membranous reactivity in <10% of cells,⁹⁹ thus distinguishing between these scores is understandably difficult. Discordance in scoring between the biopsy and the surgical specimen was seen in more than 20% of patients treated with neoadjuvant chemotherapy without anti-HER2 therapy,¹⁰⁰ and HER2-low IHC results change between primary tumors and metastatic relapses in more than 35% of patients.^{100,101}

There are numerous pitfalls associated with the available preanalytical, analytical, and post-analytical methods^{102,103} that may explain these discordant HER2 IHC results. Among these are tissue sample processing which might affect protein detection rate,¹⁰⁴ true HER2 intratumoral heterogeneity,¹⁰⁵ true heterogeneity between different metastatic sites in HER2 expression level,¹⁰⁶ as well as inter-observer variability,¹⁰⁷ leading to reduced sensitivity and reproducibility of HER2 testing. The American Society of Clinical Oncology (ASCO)/College of American Pathologists (CAP) has recently updated their guidelines for HER2 testing in breast cancer. The thresholds for HER2 +1 remain the same as in the previous guideline version; however, it is recommended that tumors be assessed at 40X magnification, using appropriate controls, and that a second pathologist reviews the slides when results are close to 0. Due to heterogeneity in HER2 levels, medical oncologists are encouraged to consider HER2-low results from any biopsy, not only the most recent, in treatment decisions.¹⁶

The changing landscape of anti-HER2 therapy has led investigators to search for more sensitive assays and novel diagnostic methods that could better quantify HER2 expression levels and refine the historically binary approach to HER2 classification. Boyars et al re-evaluated the scores of breast carcinomas with low HER2 expression using up to 400x total magnification, which resulted in re-classification of most HER2-negative cases to HER2 “very low” (incomplete faint/barely perceptible membranous staining in up to 10% of tumor cells) or HER2 +1.¹⁰⁸ A recent study reported the promising results of HER2 detection using a novel assay combining quantitative immunofluorescence and mass spectrometry,¹⁰⁹ which resulted in a lower threshold of detection and better quantification than IHC. Kennedy et al evaluated a targeted mass spectrometry-based assay for quantifying HER2 protein in formalin-fixed paraffin-embedded (FFPE) and frozen BC biopsies and reported higher detection rates for tumors that were initially classified by IHC as HER2-negative or -low, and improved concordance by normalizing to glyceraldehyde-3-phosphate dehydrogenase to account for tissue heterogeneity.¹¹⁰ Moreover, artificial intelligence and digital image analysis (DIA) offer the potential to

supplement conventional pathologic analysis and enhance the precision of HER2 testing. Recent studies have shown promising results, particularly in relation to HER2 heterogeneity in HER2-low breast cancer.^{111–115} Nonetheless, caution should be taken before implementing these technologies for routine diagnosis of HER2-low BC, as at least 1 study reported that they could under-estimate HER2 staining, especially in heterogenous HER2-low cases.¹¹⁶

Summary and Future Directions

When categorizing breast cancer by HER2 expression, “HER2-low” cancers make up the majority of cases.¹¹⁷ The promising results of Tdx-D in breast cancer patients with low HER2 expression have led to comprehensive research aiming to understand the clinical landscape of these tumors. Numerous trials have shown that HER2-low is associated with HR expression and other pathologic correlates of better prognosis, and that these tumors are associated with similar or better prognosis compared with HER2-negative tumors when treated with current therapies. Thus, to date, low expression of HER2 is clinically relevant only to patients who are candidates for treatment with Tdx-D in the metastatic setting. A trial that tested Tdx-D in the neoadjuvant setting in HER2-low cancers achieved impressive response rates,⁸¹ and thus detecting HER2-low status may become relevant to locally advanced BC as well. The clinical significance of HER2 expression in these tumors seems to be its function as a docking site for the antibody portion of this ADC, which allows targeted delivery of potent chemotherapy.

Additional potent agents targeting HER2 are currently tested as treatment for HER2-low BC, and assessing tumors for low HER2 levels is an important clinical issue. Pathologists should meticulously test tumors at high magnification and use appropriate controls¹⁶ so that patients will not be denied potentially beneficial therapy. As levels of HER2 expression change between different metastases when tested simultaneously,¹⁰⁶ clinicians should consider previous biopsy results documenting HER2-low BC in determining eligibility for Tdx-D therapy. Heterogeneity should also be considered when designing clinical trials for this patient population, and it might be appropriate to include patients based on any biopsy interpreted as “HER2-low”, and not only the most recent biopsy.

The association between HER2 level and benefit from Tdx-D in HER2-low tumors has not been explored; however, HER2 +3 tumors derive a greater benefit from Tdx-D, and benefit from other ADCs seems to be related to level of expression of their target.¹¹⁸ ADCs are expensive and toxic, and patient selection based on expected benefit is important. If HER2 targeting ADCs is tested in the neoadjuvant setting, aiming to replace standard chemotherapy, it would be essential to accurately define the patients who benefit and those who still need chemotherapy. Novel methods for HER2 quantification are in development,^{108–110,112,114} and their value as predictive markers for benefitting from HER2 targeting ADCs in HER2-low tumors should be tested. Trials testing HER2 targeting ADCs in combination with other targeted agents in patients with HER2-low BC are also warranted.¹⁰⁰

Conclusions

With the development of novel ADCs and other targeted agents, the treatment landscape for HER2-low breast cancer is expanding, and the importance of detecting low expression levels of HER2 is becoming increasingly relevant. Improvements in pathology review and novel laboratory methods are needed to make sure that patients are not spared effective therapy, to stratify patients according to the level of HER2 expression and test for associations between level of expression and response. Inclusion of patients with HER2-low breast cancer in clinical trials of targeted therapies will hopefully increase our treatment arsenal for these common tumors and allow further translational research on the role of HER2 and other molecular pathways in tumor progression and treatment response.

Author Contributions

All authors made a significant contribution to the work reported, whether that is in the conception, study design, execution, acquisition of data, analysis and interpretation, or in all these areas; took part in drafting, revising or critically reviewing the article; gave final approval of the version to be published; have agreed on the journal to which the article has been submitted; and agree to be accountable for all aspects of the work.

Funding

This research received no external funding.

Disclosure

AS reports receiving travel funds and honoraria from AstraZeneca. YS and SL report no conflicts of interest in this work.

References

1. Alroy I, Yarden Y. Biochemistry of HER2 oncogenesis in breast cancer. *Breast Dis.* 2000;11(1):31–48. doi:10.3233/BD-1999-11104
2. Rubin I, Yarden Y. The basic biology of HER2. *Ann Oncol.* 2001;12(SUPPL. 1). doi:10.1093/ANNONC/12.SUPPL_1.S3
3. Slamon DJ, Clark GM, Wong SG, Levin WJ, Ullrich A, McGuire WL. Human breast cancer: correlation of relapse and survival with amplification of the HER-2/neu oncogene. *Science.* 1987;235(4785):182–191. doi:10.1126/SCIENCE.3798106
4. Seshadri R, Firgaira FA, Horsfall DJ, McCaul K, Setlur V, Kitchen P. Clinical significance of HER-2/neu oncogene amplification in primary breast cancer. The South Australian Breast Cancer Study Group. *J Clin Oncol.* 1993;11(10):1936–1942. doi:10.1200/JCO.1993.11.10.1936
5. Slamon DJ, Leyland-Jones B, Shak S, et al. Use of chemotherapy plus a monoclonal antibody against HER2 for metastatic breast cancer that overexpresses HER2. *N Engl J Med.* 2001;344(11):783–792. doi:10.1056/NEJM200103153441101
6. Swain SM, Miles D, Kim SB, et al. Pertuzumab, trastuzumab, and docetaxel for HER2-positive metastatic breast cancer (CLEOPATRA): end-of-study results from a double-blind, randomised, placebo-controlled, phase 3 study. *Lancet Oncol.* 2020;21(4):519–530. doi:10.1016/S1470-2045(19)30863-0
7. Cameron D, Piccart-Gebhart MJ, Gelber RD, et al. 11 years' follow-up of trastuzumab after adjuvant chemotherapy in HER2-positive early breast cancer: final analysis of the HERceptin Adjuvant (HERA) trial. *Lancet.* 2017;389(10075):1195–1205. doi:10.1016/S0140-6736(16)32616-2
8. Bradley R, Braybrooke J, Gray R, et al. Trastuzumab for early-stage, HER2-positive breast cancer: a meta-analysis of 13 864 women in seven randomised trials. *Lancet Oncol.* 2021;22(8):1139–1150. doi:10.1016/S1470-2045(21)00288-6
9. Piccart M, Procter M, Fumagalli D, et al. Adjuvant pertuzumab and trastuzumab in early HER2-positive breast cancer in the APHINITY trial: 6 years' follow-up. *J Clin Oncol.* 2021;39(13):1448–1457. doi:10.1200/JCO.20.01204
10. Wolff AC, Elizabeth Hale Hammond M, Allison KH, et al. Human epidermal growth factor receptor 2 testing in breast cancer: American Society of Clinical Oncology/College of American Pathologists Clinical Practice Guideline Focused Update. *J Clin Oncol.* 2018;36(20):2105–2122. doi:10.1200/JCO.2018.77.8738
11. Fehrenbacher L, Cecchini RS, Geyer CE, et al. NSABP B-47/NRG oncology Phase III randomized trial comparing adjuvant chemotherapy with or without trastuzumab in high-risk invasive breast cancer negative for HER2 by FISH and with IHC 1+ or 2+. *J Clin Oncol.* 2020;38(5):444. doi:10.1200/JCO.19.01455
12. Jacot W, Cottu P, Berger F, et al. Actionability of HER2-amplified circulating tumor cells in HER2-negative metastatic breast cancer: the CirCe T-DM1 trial. *Breast Cancer Res.* 2019;21(1):1–9. doi:10.1186/S13058-019-1215-Z/FIGURES/3
13. Perez EA, Reinholz MM, Hillman DW, et al. HER2 and chromosome 17 effect on patient outcome in the N9831 adjuvant trastuzumab trial. *J Clin Oncol.* 2010;28(28):4307. doi:10.1200/JCO.2009.26.2154
14. Dowsett M, Procter M, McCaskill-Stevens W, et al. Disease-free survival according to degree of HER2 amplification for patients treated with adjuvant chemotherapy with or without 1 year of trastuzumab: the HERA trial. *J Clin Oncol.* 2009;27(18):2962–2969. doi:10.1200/JCO.2008.19.7939
15. Paik S, Kim C, Wolmark N. HER2 status and benefit from adjuvant trastuzumab in breast cancer. *N Engl J Med.* 2008;358(13):1409–1411. doi:10.1056/NEJM0801440
16. Modi S, Jacot W, Yamashita T, et al. Trastuzumab deruxtecan in previously treated HER2-low advanced breast cancer. *N Engl J Med.* 2022;387(1):9–20. doi:10.1056/NEJM0A2203690/SUPPL_FILE/NEJM0A2203690_DATA-SHARING.PDF
17. Modi S, Park H, Murthy RK, et al. Antitumor activity and safety of trastuzumab deruxtecan in patients with her2-low-expressing advanced breast cancer: results from a Phase Ib study. *J Clin Oncol.* 2020;38(17):1887–1896. doi:10.1200/JCO.19.02318
18. Tarantino P, Morganti S, Curigliano G. Biologic therapy for advanced breast cancer: recent advances and future directions. *Expert Opin Biol Ther.* 2020;20(9):1009–1024. doi:10.1080/14712598.2020.1752176
19. Venter DJ, Kumar S, Tuzi NL, Gullick WJ. Overexpression of the c-erbB-2 oncoprotein in human breast carcinomas: immunohistological assessment correlates with gene amplification. *Lancet.* 1987;2(8550):69–72. doi:10.1016/S0140-6736(87)92736-X
20. Kallioniemi OP, Kallioniemi A, Kurisu W, et al. ERBB2 amplification in breast cancer analyzed by fluorescence in situ hybridization. *Proc Natl Acad Sci U S A.* 1992;89(12):5321. doi:10.1073/PNAS.89.12.5321
21. Kraus MH, Popescu NC, Amsbaugh SC, King CR. Overexpression of the EGF receptor-related proto-oncogene erbB-2 in human mammary tumor cell lines by different molecular mechanisms. *EMBO J.* 1987;6(3):605–610. doi:10.1002/J.1460-2075.1987.TB04797.X
22. Nezu M, Sasaki H, Kuwahara Y, et al. Identification of a Novel Promoter and Exons of the c-ERBB-2 Gene. *Biochem Biophys Res Commun.* 1999;258(3):499–505. doi:10.1006/BBRC.1999.0634
23. Tal M, King CR, Kraus MH, Ullrich A, Schlessinger J, Givol D. Human HER2 (neu) promoter: evidence for multiple mechanisms for transcriptional initiation. *Mol Cell Biol.* 2003;23(7):2597–2601. doi:10.1128/MCB.7.7.2597-2601.1987
24. Chang CH, Scott GK, Kuo WL, et al. ESX: a structurally unique Ets overexpressed early during human breast tumorigenesis. *Oncogene.* 1997;14(13):1617–1622. doi:10.1038/SJ.ONC.1200978
25. Vernimmen D, Begon D, Salvador C, Gofflot S, Grootclaes M, Winkler R. Identification of HTF (HER2 transcription factor) as an AP-2 (activator protein-2) transcription factor and contribution of the HTF binding site to ERBB2 gene overexpression. *Biochem J.* 2003;370(Pt 1):323. doi:10.1042/BJ20021238
26. Delacroix L, Begon D, Chatel G, Jackers P, Winkler R. Distal ERBB2 promoter fragment displays specific transcriptional and nuclear binding activities in ERBB2 overexpressing breast cancer cells. *DNA Cell Biol.* 2005;24(9):582–594. doi:10.1089/DNA.2005.24.582

27. Begon DY, Delacroix L, Vernimmen D, Jackers P, Winkler R, Yin Yang 1 cooperates with activator protein 2 to stimulate ERBB2 gene expression in mammary cancer cells. *J Biol Chem.* 2005;280(26):24428–24434. doi:10.1074/JBC.M503790200
28. Dillon RL, Brown ST, Ling C, Shioda T, Muller WJ. An EGR2/CITED1 transcription factor complex and the 14-3-3sigma tumor suppressor are involved in regulating ErbB2 expression in a transgenic-mouse model of human breast cancer. *Mol Cell Biol.* 2007;27(24):8648–8657. doi:10.1128/MCB.00866-07
29. Mungamuri SK, Murk W, Grumolato L, Bernstein E, Aaronson SA. Chromatin modifications sequentially enhance ErbB2 expression in ErbB2 positive breast cancers. *Cell Rep.* 2013;5(2):302. doi:10.1016/j.celrep.2013.09.009
30. Liu Q, Kulak MV, Borcherding N, et al. A novel HER2 gene body enhancer contributes to HER2 expression. *Oncogene.* 2018;37(5):687–694. doi:10.1038/ONC.2017.382
31. Connell P, Ballinger CA, Jiang J, et al. The co-chaperone CHIP regulates protein triage decisions mediated by heat-shock proteins. *Nat Cell Biol.* 2001;3(1):93–96. doi:10.1038/35050618
32. Ballinger CA, Connell P, Wu Y, et al. Identification of CHIP, a novel tetratricopeptide repeat-containing protein that interacts with heat shock proteins and negatively regulates chaperone functions. *Mol Cell Biol.* 1999;19(6):4535–4545. doi:10.1128/MCB.19.6.4535
33. Xu W, Marcu M, Yuan X, Mimnaugh E, Patterson C, Neckers L. Chaperone-dependent E3 ubiquitin ligase CHIP mediates a degradative pathway for c-ErbB2/Neu. *Proc Natl Acad Sci U S A.* 2002;99(20):12847–12852. doi:10.1073/PNAS.202365899
34. Xia X, Hu T, He X, et al. Neddylation of HER2 inhibits its protein degradation and promotes breast cancer progression. *Int J Biol Sci.* 2023;19(2):377. doi:10.7150/IJBS.75852
35. Punturi NB, Seker S, Devarakonda V, et al. Mismatch repair deficiency predicts response to HER2 blockade in HER2-negative breast cancer. *Nat Commun.* 2021;12(1):1–11. doi:10.1038/s41467-021-23271-0
36. Haricharan S, Punturi N, Singh P, et al. Loss of mutl disrupts CHK2-dependent cell-cycle control through CDK4/6 to promote intrinsic endocrine therapy resistance in primary breast cancer. *Cancer Discov.* 2017;7(10):1168–1183. doi:10.1158/2159-8290.CD-16-1179
37. Yamamoto T, Ikawa S, Akiyama T, et al. Similarity of protein encoded by the human c-erb-B-2 gene to epidermal growth factor receptor. *Nature.* 1986;319(6050):230–234. doi:10.1038/319230A0
38. Kraus MH, Issing W, Miki T, Popescu NP, Aaronson SA. Isolation and characterization of ERBB3, a third member of the ERBB/epidermal growth factor receptor family: evidence for overexpression in a subset of human mammary tumors. *Proc Natl Acad Sci U S A.* 1989;86(23):9193–9197. doi:10.1073/PNAS.86.23.9193
39. Plowman GD, Culouscou JM, Whitney GS, et al. Ligand-specific activation of HER4/p180erbB4, a fourth member of the epidermal growth factor receptor family. *Proc Natl Acad Sci U S A.* 1993;90(5):1746. doi:10.1073/PNAS.90.5.1746
40. Moasser MM. The oncogene HER2: its signaling and transforming functions and its role in human cancer pathogenesis. *Oncogene.* 2007;26(45):6469–6487. doi:10.1038/SJ.ONC.1210477
41. Beerli RR, Hynes NE. Epidermal growth factor-related peptides activate distinct subsets of ErbB receptors and differ in their biological activities. *J Biol Chem.* 1996;271(11):6071–6076. doi:10.1074/JBC.271.11.6071
42. Ushiro H, Cohens S. Identification of phosphotyrosine as a product of epidermal growth factor-activated protein kinase in A-431 cell membranes*. *J Biol Chem.* 1980;255(18):8363–8365. doi:10.1016/S0021-9258(18)43497-7
43. Ottenhoff-Kalff AF, Rijksen G, van Beurden EA, Hennipman A, Michels AA, Staal GE. Characterization of protein tyrosine kinases from human breast cancer: involvement of the c-src oncogene product. *Cancer Res.* 1992;52(17):4773–4778.
44. Reissig D, Clement J, Sanger J, Berndt A, Kosmehl H, Bohmer FD. Elevated activity and expression of Src-family kinases in human breast carcinoma tissue versus matched non-tumor tissue. *J Cancer Res Clin Oncol.* 2001;127(4):226–230. doi:10.1007/S004320000197/METRICS
45. Belsches-Jablonski AP, Biscardi JS, Peavy DR, Tice DA, Romney DA, Parsons SJ. Src family kinases and HER2 interactions in human breast cancer cell growth and survival. *Oncogene.* 2001;20(12):1465–1475. doi:10.1038/SJ.ONC.1204205
46. Timms JF, White SL, O’Hare MJ, Waterfield MD. Effects of ErbB-2 overexpression on mitogenic signalling and cell cycle progression in human breast luminal epithelial cells. *Oncogene.* 2002;21(43):6573–6586. doi:10.1038/SJ.ONC.1205847
47. Montemurro F, Di Cosimo S, Arpino G. Human epidermal growth factor receptor 2 (HER2)-positive and hormone receptor-positive breast cancer: new insights into molecular interactions and clinical implications. *Ann Oncol.* 2013;24(11):2715–2724. doi:10.1093/ANNONC/MDT287
48. Bose R, Kavuri SM, Searleman AC, et al. Activating HER2 mutations in HER2 gene amplification negative breast cancer. *Cancer Discov.* 2013;3(2):224–237. doi:10.1158/2159-8290.CD-12-0349
49. Petrelli F, Tomasello G, Barni S, Lonati V, Passalacqua R, Ghidini M. Clinical and pathological characterization of HER2 mutations in human breast cancer: a systematic review of the literature. *Breast Cancer Res Treat.* 2017;166(2):339–349. doi:10.1007/S10549-017-4419-X
50. Perou CM, Sorile T, Eisen MB, et al. Molecular portraits of human breast tumours. *Nature.* 2000;406(6797):747–752. doi:10.1038/35021093
51. Schettini F, Braso-Maristany F, Kuderer NM, Prat A. A perspective on the development and lack of interchangeability of the breast cancer intrinsic subtypes. *NPJ Breast Cancer.* 2022;8(1). doi:10.1038/S41523-022-00451-9
52. Bernard PS, Parker JS, Mullins M, et al. Supervised risk predictor of breast cancer based on intrinsic subtypes. *J Clin Oncol.* 2009;27(8):1160. doi:10.1200/JCO.2008.18.1370
53. Schettini F, Chic N, Braso-Maristany F, et al. Clinical, pathological, and PAM50 gene expression features of HER2-low breast cancer. *NPJ Breast Cancer.* 2021;7(1). doi:10.1038/S41523-020-00208-2
54. Cardoso F, Van’t Veer LJ, Bogaerts J, et al. 70-gene signature as an aid to treatment decisions in early-stage breast cancer. *N Engl J Med.* 2016;375(8):717–729. doi:10.1056/NEJMOA1602253
55. Krijgsman O, Roepman P, Zwart W, et al. A diagnostic gene profile for molecular subtyping of breast cancer associated with treatment response. *Breast Cancer Res Treat.* 2012;133(1):37–47. doi:10.1007/S10549-011-1683-Z
56. Zhang H, Katerji H, Turner BM, Audeh W, Hicks DG. HER2-low breast cancers: incidence, HER2 staining patterns, clinicopathologic features, MammaPrint and Blueprint genomic profiles. *Mod Pathol.* 2022;35(8):1075–1082. doi:10.1038/S41379-022-01019-5
57. Wallden B, Storhoff J, Nielsen T, et al. Development and verification of the PAM50-based Prosigna breast cancer gene signature assay. *BMC Med Genomics.* 2015;8(1). doi:10.1186/S12920-015-0129-6
58. Marchio C, Dell’Orto P, Annaratone L, et al. The dilemma of HER2 double-equivocal breast carcinomas: genomic profiling and implications for treatment. *Am J Surg Pathol.* 2018;42(9):1190–1200. doi:10.1097/PAS.0000000000001100

59. Zhang G, Ren C, Li C, et al. Distinct clinical and somatic mutational features of breast tumors with high-, low-, or non-expressing human epidermal growth factor receptor 2 status. *BMC Med.* 2022;20(1):1–15. doi:10.1186/S12916-022-02346-9/FIGURES/5
60. Agostinetti E, Rediti M, Fimereli D, et al. Her2-low breast cancer: molecular characteristics and prognosis. *Cancers.* 2021;13(11):2824. doi:10.3390/CANCERS13112824/S1
61. Denkert C, Seither F, Schneeweiss A, et al. Clinical and molecular characteristics of HER2-low-positive breast cancer: pooled analysis of individual patient data from four prospective, neoadjuvant clinical trials. *Lancet Oncol.* 2021;22(8):1151–1161. doi:10.1016/S1470-2045(21)00301-6
62. Horisawa N, Adachi Y, Takatsuka D, et al. The frequency of low HER2 expression in breast cancer and a comparison of prognosis between patients with HER2-low and HER2-negative breast cancer by HR status. *Breast Cancer.* 2022;29(2):234–241. doi:10.1007/S12282-021-01303-3
63. Tarantino P, Jin Q, Tayob N, et al. Prognostic and biologic significance of ERBB2-low expression in early-stage breast cancer. *JAMA Oncol.* 2022;8(8):1177–1183. doi:10.1001/JAMAONCOL.2022.2286
64. Xu H, Han Y, Wu Y, et al. Clinicopathological characteristics and prognosis of HER2-low early-stage breast cancer: a single-institution experience. *Front Oncol.* 2022;12:906011. doi:10.3389/FONC.2022.906011/BIBTEX
65. Won HS, Ahn J, Kim Y, et al. Clinical significance of HER2-low expression in early breast cancer: a nationwide study from the Korean Breast Cancer Society. *Breast Cancer Res.* 2022;24(1):1–11. doi:10.1186/S13058-022-01519-X/TABLES/3
66. Jacot W, Maran-Gonzalez A, Massol O, et al. Prognostic value of HER2-low expression in non-metastatic triple-negative breast cancer and correlation with other biomarkers. *Cancers.* 2021;13(23). doi:10.3390/CANCERS13236059
67. Tan RSYC, Ong WS, Lee KH, et al. HER2 expression, copy number variation and survival outcomes in HER2-low non-metastatic breast cancer: an international multicentre cohort study and TCGA-METABRIC analysis. *BMC Med.* 2022;20(1):1–15. doi:10.1186/S12916-022-02284-6/FIGURES/6
68. Peiffer DS, Zhao F, Chen N, et al. Clinicopathologic characteristics and prognosis of ERBB2-low breast cancer among patients in the national cancer database. *JAMA Oncol.* 2023;9(4):500–510. doi:10.1001/JAMAONCOL.2022.7476
69. Chen M, Chen W, Liu D, et al. Prognostic values of clinical and molecular features in HER2 low-breast cancer with hormonal receptor overexpression: features of HER2-low breast cancer. *Breast Cancer.* 2022;29(5):844. doi:10.1007/S12282-022-01364-Y
70. Mutai R, Barkan T, Moore A, et al. Prognostic impact of HER2-low expression in hormone receptor positive early breast cancer. *Breast.* 2021;60:62. doi:10.1016/J.BREAST.2021.08.016
71. Almstedt K, Heimes AS, Kappenberg F, et al. Long-term prognostic significance of HER2-low and HER2-zero in node-negative breast cancer. *Eur J Cancer.* 2022;173:10–19. doi:10.1016/J.EJCA.2022.06.012
72. Kang S, Lee SH, Lee HJ, et al. Pathological complete response, long-term outcomes, and recurrence patterns in HER2-low versus HER2-zero breast cancer after neoadjuvant chemotherapy. *Eur J Cancer.* 2022;176:30–40. doi:10.1016/J.EJCA.2022.08.031
73. de Moura Leite L, Cesca MG, Tavares MC, et al. HER2-low status and response to neoadjuvant chemotherapy in HER2 negative early breast cancer. *Breast Cancer Res Treat.* 2021;190(1):155–163. doi:10.1007/S10549-021-06365-7
74. Shao Y, Yu Y, Luo Z, et al. Clinical, pathological complete response, and prognosis characteristics of HER2-low breast cancer in the neoadjuvant chemotherapy setting: a retrospective analysis. *Ann Surg Oncol.* 2022;29(13):8026–8034. doi:10.1245/S10434-022-12369-4
75. Zhou S, Liu T, Kuang X, et al. Comparison of clinicopathological characteristics and response to neoadjuvant chemotherapy between HER2-low and HER2-zero breast cancer. *Breast.* 2023;67:1. doi:10.1016/J.BREAST.2022.12.006
76. Domergue C, Martin E, Lemarié C, et al. Impact of HER2 status on pathological response after neoadjuvant chemotherapy in early triple-negative breast cancer. *Cancers.* 2022;14(10). doi:10.3390/CANCERS14102509
77. De Calbiac O, Lusque A, Mailliez A, et al. Comparison of management and outcomes in ERBB2-low vs ERBB2-zero metastatic breast cancer in France. *JAMA Netw Open.* 2022;5(9):e2231170. doi:10.1001/JAMANETWORKOPEN.2022.31170
78. Carlino F, Diana A, Ventriglia A, et al. HER2-low status does not affect survival outcomes of patients with Metastatic Breast Cancer (MBC) undergoing first-line treatment with endocrine therapy plus palbociclib: results of a multicenter, retrospective cohort study. *Cancers.* 2022;14(20). doi:10.3390/CANCERS14204981
79. Gampenrieder SP, Rinnerthaler G, Tinchon C, et al. Landscape of HER2-low metastatic breast cancer (MBC): results from the Austrian AGMT_MBC-Registry. *Breast Cancer Res.* 2021;23(1). doi:10.1186/S13058-021-01492-X
80. Cortés J, Kim SB, Chung WP, et al. Trastuzumab deruxtecan versus trastuzumab emtansine for breast cancer. *N Engl J Med.* 2022;386(12):1143–1154. doi:10.1056/NEJMOA2115022
81. Hurvitz SA, Wang LS, Chan D, et al. TRIO-US B-12 TALENT: Phase II neoadjuvant trial evaluating trastuzumab deruxtecan with or without anastrozole for HER2-low, HR+ early-stage breast cancer. *J Clin Oncol.* 2022;40(16_suppl):TPS623. doi:10.1200/JCO.2022.40.16_SUPPL.TPS623
82. Linehan AS, Fitzpatrick OM, Morris PG. Profile of trastuzumab deruxtecan in the management of patients with HER2-positive unresectable or metastatic breast cancer: an evidence-based review. *Breast Cancer.* 2021;13:151–159. doi:10.2147/BCTT.S245024
83. Andre F, Hamilton EP, Loi S, et al. Dose-finding and -expansion studies of trastuzumab deruxtecan in combination with other anti-cancer agents in patients (pts) with advanced/metastatic HER2+ (DESTINY-Breast07 [DB-07]) and HER2-low (DESTINY-Breast08 [DB-08]) breast cancer (BC). *J Clin Oncol.* 2022;40(16_suppl):3025. doi:10.1200/JCO.2022.40.16_SUPPL.3025
84. Phase II neoadjuvant pyrotinib combined with neoadjuvant chemotherapy in HER2-low-expressing and HR positive early or locally advanced breast cancer: a single-arm, non-randomized, single-center, open label trial - Full text view - ClinicalTrials.gov. Available from: <https://classic.clinicaltrials.gov/ct2/show/NCT05165225>. Accessed July 15, 2023.
85. Li H, Zhang X, Xu Z, et al. Preclinical evaluation of MRG002, a novel HER2-targeting antibody-drug conjugate with potent antitumor activity against HER2-positive solid tumors. *Antib Ther.* 2021;4(3):175–184. doi:10.1093/ABT/TBAB017
86. Phase 1b study of EZH1/2 inhibitor valemestostat in combination with trastuzumab deruxtecan in subjects with HER2 low/ultra-low/null metastatic breast cancer - Full text view - ClinicalTrials.gov. Available from: <https://classic.clinicaltrials.gov/ct2/show/NCT05633979>. Accessed July 18, 2023.
87. Shastri PN, Zhu J, Skidmore L, et al. Nonclinical development of next-generation site-specific HER2-targeting antibody-drug conjugate (ARX788) for breast cancer treatment. *Mol Cancer Ther.* 2020;19(9):1822–1832. doi:10.1158/1535-7163.MCT-19-0692
88. Daiichi Sankyo's EZH1/2 dual inhibitor valemestostat (DS-3201) receives SAKIGAKE designation for treatment of patients with relapsed/refractory peripheral T-cell lymphoma from Japan MHLW - Daiichi Sankyo US. Available from: <https://daiichisankyo.us/press-releases/-/article/daiichi-sankyo-s-ezh1-2-dual-inhibitor-valemestostat-ds-3201-receives-sakigake-designation-for-treatment-of-patients-with-relapsed-refractory-periphera>. Accessed July 15, 2023.

89. ISPY-P1.01: evaluating the safety of weekly paclitaxel with trastuzumab duocarmazine (SYD985) in patients with metastatic cancer - Full text view - ClinicalTrials.gov. Available from: <https://classic.clinicaltrials.gov/ct2/show/NCT04602117>. Accessed July 15, 2023.
90. Banerji U, van Herpen CML, Saura C, et al. Trastuzumab duocarmazine in locally advanced and metastatic solid tumours and HER2-expressing breast cancer: a phase I dose-escalation and dose-expansion study. *Lancet Oncol.* 2019;20(8):1124–1135. doi:10.1016/S1470-2045(19)30328-6
91. SYD985 vs Physician's choice in participants with HER2-positive locally advanced or metastatic breast cancer - Full text view - ClinicalTrials.gov. Available from: <https://classic.clinicaltrials.gov/ct2/show/NCT03262935>. Accessed July 15, 2023.
92. ARX788 in breast cancer with low expression of HER2 - Full text view - ClinicalTrials.gov. Available from: <https://classic.clinicaltrials.gov/ct2/show/NCT05018676>. Accessed July 15, 2023.
93. Hurvitz SA, Park H, Frentzas S, et al. Safety and unique pharmacokinetic profile of ARX788, a site-specific ADC, in heavily pretreated patients with HER2-overexpressing solid tumors: results from two phase I clinical trials. *J Clin Oncol.* 2021;39(15_suppl):1038. doi:10.1200/JCO.2021.39.15_SUPPL.1038
94. A study of BL-M07D1 in patients with locally advanced or metastatic HER2 positive/low expression breast cancer and other solid tumors - Full text view - ClinicalTrials.gov. Available from: <https://classic.clinicaltrials.gov/ct2/show/NCT05461768>. Accessed July 15, 2023.
95. Calmeiro J, Carrascal MA, Tavares AR, et al. Dendritic cell vaccines for cancer immunotherapy: the role of human conventional type 1 dendritic cells. *Pharmaceutics.* 2020;12(2). doi:10.3390/PHARMACEUTICS12020158
96. 260P Antitumor activity of trastuzumab deruxtecan (T-DXd) in patients with metastatic breast cancer (mBC) and brain metastases (BMs) from DAISY trial - Annals of Oncology. Available from: [https://www.annalsofncology.org/article/S0923-7534\(22\)02150-0/fulltext](https://www.annalsofncology.org/article/S0923-7534(22)02150-0/fulltext). Accessed July 18, 2023.
97. Lu Y, Zhu S, Tong Y, et al. HER2-low status is not accurate in breast cancer core needle biopsy samples: an analysis of 5610 consecutive patients. *Cancers.* 2022;14(24):6200. doi:10.3390/CANCERS14246200/S1
98. Fernandez AI, Liu M, Bellizzi A, et al. Examination of Low ERBB2 Protein Expression in Breast Cancer Tissue. *JAMA Oncol.* 2022;8(4):1–4. doi:10.1001/JAMAONCOL.2021.7239
99. Wolff AC, Somerfield MR, Dowsett M, et al. Human epidermal growth factor receptor 2 testing in breast cancer: ASCO-College of American Pathologists Guideline Update. *J Clin Oncol.* 2023;JCO2202864. doi:10.1200/JCO.22.02864
100. Miglietta F, Griguolo G, Bottosso M, et al. ARTICLE evolution of HER2-low expression from primary to recurrent breast cancer. *NPJ Breast Cancer.* 2021;7(1). doi:10.1038/s41523-021-00343-4
101. Tarantino P, Gandini S, Nicolò E, et al. Evolution of low HER2 expression between early and advanced-stage breast cancer. *Eur J Cancer.* 2022;163:35–43. doi:10.1016/J.EJCA.2021.12.022
102. Sajjadi E, Venetis K, Ivanova M, Fusco N. Improving HER2 testing reproducibility in HER2-low breast cancer. *Cancer Drug Resist.* 2022;5(4):882. doi:10.20517/CDR.2022.29
103. Sajjadi E, Guerini-Rocco E, De Camilli E, et al. Pathological identification of HER2-low breast cancer: tips, tricks, and troubleshooting for the optimal test. *Front Mol Biosci.* 2023;10:1176309. doi:10.3389/fmolb.2023.1176309
104. Perez EA, Cortés J, Maria Gonzalez-Angulo A, Bartlett JMS. Laboratory-Clinic Interface HER2 testing: current status and future directions. *Cancer Treat Rev.* 2014;40(2):276–284. doi:10.1016/j.ctrv.2013.09.001
105. Allison KH, Dintzis SM, Schmidt RA. Frequency of HER2 heterogeneity by fluorescence in situ hybridization according to CAP expert panel recommendations: time for a new look at how to report heterogeneity. *Am J Clin Pathol.* 2011;136(6):864–871. doi:10.1309/AJCPXTZSKBRIP07W
106. Geukens T, De Schepper M, Richard F, et al. Intra-patient and inter-metastasis heterogeneity of HER2-low status in metastatic breast cancer. *Eur J Cancer.* 2023;188:152–160. doi:10.1016/j.ejca.2023.04.026
107. Lambein K, Van Bockstal M, Vandemaele L, et al. Distinguishing score 0 from score 1+ in HER2 immunohistochemistry-negative breast cancer: clinical and pathobiological relevance. *Am J Clin Pathol.* 2013;140(4):561–566. doi:10.1309/AJCP4A7KTAYHZSOE
108. Boyraz B, Ly A. Discerning subsets of breast cancer with very low and absent HER2 protein expression*. *Hum Pathol.* 2022;127:50–55. doi:10.1016/j.humpath.2022.05.019
109. Moutafi M, Robbins CJ, Yaghoobi V, et al. Quantitative measurement of HER2 expression to subclassify ERBB2 unamplified breast cancer. *Lab Invest.* 2022;102(10):1101–1108. doi:10.1038/S41374-022-00804-9
110. Kennedy JJ, Whiteaker JR, Kennedy LC, et al. Quantification of human epidermal growth factor receptor 2 by immunopeptide enrichment and targeted mass spectrometry in formalin-fixed paraffin-embedded and frozen breast cancer tissues. *Clin Chem.* 2021;67(7):1008–1018. doi:10.1093/clinchem/hvab047
111. Palm C, Connolly CE, Masser R, et al. Determining HER2 status by artificial intelligence: an investigation of primary, metastatic, and HER2 low breast tumors. *Diagnostics.* 2023;13(1). doi:10.3390/DIAGNOSTICS13010168/S1
112. Frey P, Mamilos A, Minin E, et al. AI-based HER2-low IHC scoring in breast cancer across multiple sites, clones, and scanners. *J Clin Oncol.* 2023;41(16_suppl):516. doi:10.1200/JCO.2023.41.16_SUPPL.516
113. Yue M, Zhang J, Wang X, et al. Can AI-assisted microscope facilitate breast HER2 interpretation? A multi-institutional ring study. *Virchows Arch.* 2021;479(3):443–449. doi:10.1007/S00428-021-03154-X
114. Wu S, Yue M, Zhang J, et al. The role of artificial intelligence in accurate interpretation of HER2 immunohistochemical scores 0 and 1+ in Breast Cancer. *Mod Pathol.* 2023;36(3):100054. doi:10.1016/J.MODPAT.2022.100054
115. Farahmand S, Fernandez AI, Ahmed FS, et al. Deep learning trained on hematoxylin and eosin tumor region of Interest predicts HER2 status and trastuzumab treatment response in HER2+ breast cancer. *Mod Pathol.* 2022;35(1):44–51. doi:10.1038/S41379-021-00911-W
116. Sode M, Thagaard J, Eriksen JO, Laenkholm AV. Digital image analysis and assisted reading of the HER2 score display reduced concordance: pitfalls in the categorisation of HER2-low breast cancer. *Histopathology.* 2023. doi:10.1111/HIS.14877
117. Tarantino P, Hamilton E, Tolaney SM, et al. HER2-low breast cancer: pathological and clinical landscape. *J Clin Oncol.* 2020;38(17):1951–1962. doi:10.1200/JCO.19.02488
118. Bardia A, Tolaney SM, Punie K, et al. Biomarker analyses in the phase III ASCENT study of sacituzumab govitecan versus chemotherapy in patients with metastatic triple-negative breast cancer. *Ann Oncol.* 2021;32(9):1148–1156. doi:10.1016/j.annonc.2021.06.002

Breast Cancer: Targets and Therapy

Dovepress

Publish your work in this journal

Breast Cancer - Targets and Therapy is an international, peer-reviewed open access journal focusing on breast cancer research, identification of therapeutic targets and the optimal use of preventative and integrated treatment interventions to achieve improved outcomes, enhanced survival and quality of life for the cancer patient. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Submit your manuscript here: <https://www.dovepress.com/breast-cancer—targets-and-therapy-journal>