

Targeted modulation of cell differentiation in distinct regions of the gastrointestinal tract via oral administration of differently PEG-PEI functionalized mesoporous silica nanoparticles

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Abstract: Targeted delivery of drugs is required to efficiently treat intestinal diseases such as colon cancer and inflammation. Nanoparticles could overcome challenges in oral administration caused by drug degradation at low pH and poor permeability through mucus layers, and offer targeted delivery to diseased cells in order to avoid adverse effects. Here, we demonstrate that functionalization of mesoporous silica nanoparticles (MSNs) by polymeric surface grafts facilitates transport through the mucosal barrier and enhances cellular internalization. MSNs functionalized with poly(ethylene glycol) (PEG), poly(ethylene imine) (PEI), and the targeting ligand folic acid in different combinations are internalized by epithelial cells in vitro and in vivo after oral gavage. Functionalized MSNs loaded with γ -secretase inhibitors of the Notch pathway, a key regulator of intestinal progenitor cells, colon cancer, and inflammation, demonstrated enhanced intestinal goblet cell differentiation as compared to free drug. Drug-loaded MSNs thus remained intact in vivo, further confirmed by exposure to simulated gastric and intestinal fluids in vitro. Drug targeting and efficacy in different parts of the intestine could be tuned by MSN surface modifications, with PEI coating exhibiting higher affinity for the small intestine and PEI-PEG coating for the colon. The data highlight the potential of nanomedicines for targeted delivery to distinct regions of the tissue for strict therapeutic control.

Keywords: intestinal targeting, PEG-PEI copolymer, Notch inhibition

Introduction

Targeting specific biological pathways provides an opportunity to devise more specific and more effective approaches in cancer and stem cell therapy. In conjunction with improved molecular diagnostics, this is the foundation of personalized medicine.^{1,2} However, given the widespread and complex functions of biological pathways, increased specificity and tissue- and cell-targeted delivery of modulators are necessary. Nanotechnology-based drug delivery systems provide a solution to improve therapeutic efficacy with reduced side effects. Chemical design of nanoparticles offers control over bioavailability and biodistribution, but we still need to address the form and function relationships for any given administration.^{3,4}

The Notch signaling pathway is the key regulator of stem cells in development and tissue homeostasis, and is deregulated in inflammatory intestinal disease and colon cancer.^{5,6} Clinical studies inhibiting Notch are focused on several types of cancers by mainly two approaches: use of antibodies against receptors and ligands, and

γ -secretase inhibitors, which inhibit proteolytic processing and activation of the Notch receptor. However, Notch is essential for tissue homeostasis including that of the intestine and immune system. Further, while Notch is an oncogene in most biological systems, in the skin and vasculature, as well as in squamous epithelia, it acts as a tumor suppressor. Given these diverse actions, it is imperative that Notch modulation is spatially targeted.⁷ With novel drug delivery platforms, we are likely to see successful development toward the clinical use of Notch modulators.

Oral delivery is the most common method for drug administration because of its simplicity, noninvasive nature, and patient compliance. However, the major hurdles with oral delivery of many drugs are poor stability in the gastric environment, low aqueous solubility, as well as inadequate penetration through mucosal barriers resulting in poor oral bioavailability.⁸ Nanoparticulate delivery systems offer great promise and advantages in the administration of drugs also via the oral route because they are able to carry sufficient amounts of drugs that could be released at specific sites, at a specific pH value, be resistant toward digestive enzymes, and control the release of encapsulated or associated drug.^{9,10} Due to these advantages, nanoparticle formulation approaches have proven to be very useful for drug delivery applications, including oral formulations.^{11–13} Moreover, employing nanomedical concepts such as cellular targeting and intracellular drug release in oral delivery further offers the possibility of efficient and specific delivery to cells within the intestinal epithelium and thereby to specific regions of the gastrointestinal (GI) tract, offering more efficacious treatment possibilities for intestinal diseases such as colon cancer and inflammatory bowel diseases, not attainable via systemic drug delivery.^{9,10}

In oral formulations, amorphous silicon dioxide (SiO₂) or silica, has been used as a pharmaceutical excipient for >50 years and is classified by the US Food and Drug Administration as generally regarded as safe.¹⁴ Amorphous silica is well known to be stable at low pH and dissolves rapidly at higher pH.¹⁵ Among the silica materials explored for drug delivery purposes, mesoporous silica nanoparticles (MSNs) offer several attractive features for drug delivery. These include a uniform and tunable pore size, high loading capacity of guest molecules, easily modifiable surface properties, and ability of the nanosized porous channels to render the crystalline state of a drug amorphous, thereby increasing its apparent solubility.¹⁶ All these features allow for better control of drug loading and increased solubility, and should lead to efficient

protection of drug molecules from the harsh conditions of the GI tract. This may lead to better oral bioavailability, a property that has also been exploited to some degree for bulk mesoporous silica.^{17,18} The nanoparticle containing the drug load could further be expected to penetrate the mucosal barrier, be taken up by the intestinal epithelial cells, and either release the drug intracellularly or further aid the drug permeability across the epithelial layers, thus offering some advantages not obtainable via traditional oral formulations.

In this study, we utilize the high positive charge density of poly(ethylene imine) (PEI) surface coating on MSNs for facilitating binding to the negatively charged cell surfaces, leading to cellular internalization. Pure PEI has been shown to result in toxic side effects directly related to the molecular weight of branched PEI, which would restrict its application for therapeutic drug delivery.¹⁹ However, derivatization or “charge capping” of PEI with other functional groups^{20,21} as well as when used as a constituent together with silica in the construction of hybrid materials²² has indicated that these drawbacks of pure PEI can be restrained. Poly(ethylene glycol) (PEG) is the most commonly used nonionic hydrophilic polymer in biomedical drug delivery design because it has been shown to reduce uptake by the reticuloendothelial system, provide good dispersibility in aqueous solvents, and diminish association with proteins.^{23,24} PEG has a low intrinsic toxicity that renders it very suitable for biomedical applications, and it is also considered to be a generally regarded as safe material (GRAS).²⁵ PEG has been reported as mucoadhesive agent, providing a net neutral surface, which aids to avoid binding with intestinal mucin networks and enhances penetration of nanoparticles through the mucin barrier.^{26,27} Hence, PEGylated nanoparticles have shown a higher ability for the interaction with the intestinal mucosa than the stomach.²⁸ Nevertheless, the type of surface conformation of PEG chains, molecular weight, grafting density, and even coupling chemistry may have a significant influence on the biological behavior of PEGylated nanoparticles. Thus, different conformations of the PEG chains could either facilitate the penetration of the PEGylated particles through the mucus layer or increase the residence time of the adhered fraction of particles in the mucosa.²⁸ In our approach, PEGylation was achieved by attaching PEG either directly to the particle surface or via a grafted PEI layer.²⁹ Further, folic acid (FA) was conjugated to the MSNs as a potential affinity ligand.

Our earlier reports have demonstrated targeted delivery of γ -secretase inhibitors, efficient Notch inhibitors,

to breast tumors with consequent reduction in tumor growth, through intravenous administration of MSNs.^{30,31} We also evaluated the effect on small intestinal stem cell differentiation by oral delivery of γ -secretase inhibitor-loaded FA-PEI-MSNs and demonstrated enhanced cell fate switching as compared to free drug.³⁰ However, the effect of different surface functionalizations, the interaction of the MSNs with the intestinal epithelium, and the effect in different GI regions were not evaluated. In this study, we took advantage of the same model system to evaluate the potential of a series of functionalized MSNs as drug carriers for cell-targeted drug delivery through the oral route, with focus on the effect of the different combinations of PEI, PEG, and FA surface modifications on intestinal targeting. We demonstrate enhanced intestinal cell targeting and therapeutic efficacy with the combination of PEG and PEI, and further show that different functionalizations target different regions of the intestinal tract. Our results should serve as a demonstration of the need for careful evaluation of the relationship between functionalization, site specificity/targetability, and therapeutic efficacy of nanomedicines through different administration routes.

Experimental

Synthesis, functionalization, and bioconjugation of MSNs

A detailed description of the synthesis, the surface functionalization, and conjugation procedures can be found in the “[Supplementary materials](#)” section. Briefly, MSNs were synthesized according to previously published protocols.^{22,32,33} Surfactant-extracted MSNs were functionalized with PEI by surface hyperbranching polymerization of PEI as previously reported to yield PEI-MSNs.³⁴ Both plain MSNs and PEI-MSNs were further functionalized with PEG utilizing the primary amine groups of either the aminosilane (for plain MSNs) or terminal ends of PEI (for PEI-MSNs). Particles (50 mg) were dispersed in chloroform and were allowed to react with 12.5 mg hexamethylene diisocyanate (HMDI)-activated methoxy-PEG (mPEG)^{29,35} (Supplementary materials) in the presence of 5 μ L N,N-diisopropylethylamine (DIPEA) overnight at 60°C under reflux. After the reaction, the particles were washed with ethanol and vacuum-dried to yield PEG-MSNs and PEG-PEI-MSNs. To obtain FA-conjugated MSNs, we followed the protocol previously described.³⁶ To obtain FA-PEG-MSNs and FA-PEG-PEI-MSNs, maleimide (mal)-PEG-OH was reacted with HMDI to activate the –OH group of mal-PEG (similarly as mPEG activation) and further conjugated with folic acid-thiol.

Particle characterization methods

The particles were characterized according to previously reported methods.^{22,32,33} A detailed description can be found in the “Supplementary materials” section.

Cellular uptake studies by fluorescence-assisted cell sorting and confocal microscopy

Cellular uptake studies by fluorescence-assisted cell sorting and confocal microscopy were performed as previously described.^{32,33} A detailed description can be found in the “Supplementary materials” section.

Particle stability under simulated GI tract conditions

Stock suspensions of empty and DAPT (N-[N-(3,5-difluorophenacetyl)-L-alanyl]-S-phenylglycine t-butyl ester) loaded MSNs (PEG-PEI-MSNs and DAPT/PEG-PEI-MSNs) were prepared in 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid (HEPES) buffer (pH 7.2) at a concentration of 4 mg/mL. The untreated baseline samples (0 time point) were collected directly on transmission electron microscopy (TEM) grids and allowed to dry overnight. For the GI stability experiment, Spectra/Por® Float-A-Lyzer® G2 dialysis membrane of fixed 1 mL volume was used to transfer particles from HEPES to simulated gastric fluid (SGF), and further to simulated intestinal fluid (SIF). The dialysis membranes were washed and preconditioned according to the manufacturer’s guidelines. Additionally, the membranes were preincubated with SGF [2.0 g of sodium chloride and 3.2 g of purified pepsin (derived from porcine gastric mucosa; product number 77161; CAS Number 9001-75-6), Sigma; Sigma-Aldrich, St Louis, MO, USA] in 7.0 mL of hydrochloric acid and water up to 1,000 mL for an hour before use. The membranes were filled with 1 mL of PEG-PEI-MSN or DAPT/PEG-PEI-MSN suspension, and the outer chamber was filled with 9 mL of SGF. The buffer volume was selected according to the extracellular fluid volume in mice, which is approximately 5–8 mL (according to body weight).³⁷ The samples were kept at 37°C with agitation for 2 hours in the SGF. After that, a small amount of samples were placed on a TEM grid and allowed to dry. The SGF in the outer chamber was collected for high-performance liquid chromatography (HPLC) analysis. Subsequently, the samples were moved to the chamber containing 9 mL of SIF (6.8 g of monobasic potassium phosphate in 250 mL of water and then 77 mL of 0.2 N sodium hydroxide and 500 mL of water; 10.0 g of pancreatin [from porcine pancreas, Sigma] is added, and the

resulting solution is adjusted to a pH of 6.8 ± 0.1 and finally diluted to 1,000 mL) and transferred back to 37°C with agitation for 6 hours. After the end of incubation, the samples were collected on a TEM grid for electron microscopy. The remaining sample in the membrane was collected for HPLC analysis of DAPT. The TEM analysis was done using JEM-1400 Plus Transmission Electron Microscope (JEOL Ltd., Tokyo, Japan), and the HPLC was done using an HPLC system (Agilent, Santa Clara, CA, USA).

In vivo oral delivery and sample collection

All animal experiments were approved and conducted in accordance with the institutional animal care policies of the University of Turku and Åbo Akademi University (ÅAU) (Turku, Finland) under the permits ESAVI/1197/04.10.07/2013 similarly as was reported in our previous study.³⁰ For in vivo oral drug delivery studies, 18 FVB/N adult male mice were randomly allocated to six groups and subjected to gastric gavage once a day for 3 consecutive days, with group 1: dimethyl sulfoxide-vehicle solution, control MSN particles; group 2: FA-PEI-MSNs; group 3: FA-PEG-PEI-MSNs 100 mg/kg; group 4: free DAPT (10 mg/kg), and DAPT-loaded MSN particles; group 5: DAPT/FA-PEI-MSNs; and group 6: DAPT/FA-PEG-PEI-MSNs 100 mg/kg, 10 wt%. Total volume of administered substances was 0.5 mL per day. Stool samples (three mice per treatment) were collected directly from the animals, weighed, and dried overnight on the heating plate, and the water content (% hydration) was determined by the loss of weight upon drying. Goblet cell numbers were analyzed by regular periodic acid–Schiff staining and light microscopy by manual counting and by using ImageJ software, (US National Institutes of Health, Bethesda, MN, USA) as well as by measuring MUC2 mRNA content by reverse transcription polymerase chain reaction. Primers used are MUC2 forward primer 5'-CTT CTG TGC CAC CCT CGT-3' and MUC2 reverse primer 5'-TTC GGG ATC TGG CTT CTT T-3', and for normalization, β -actin forward primer 5'-TGG CTC CTA GCA CCA TGA AGA-3' and β -actin reverse primer 5'-GTG GAC AGT GAG GCC AGG AT-3'.

For in vivo uptake studies with fluorescein isothiocyanate (FITC)-labeled MSNs, eight C57BL/6 adult male mice were separated randomly into four groups and subjected to gastric gavage with different nanoparticulate mixture and control HEPES solution. Group 1 received PEI-MSNs, group 2: PEG-PEI-MSNs, group 3: FA-PEG-MSNs, and group 4: HEPES. MSNs at 8 mg/mL were dispersed in sterile HEPES, and 0.5 mL of the solution was introduced to each animal

by gastric gavage for 4 consecutive days. To increase the absorption rate in intestine, mice were fasted 10 hours before and 1 hour after gavage. At the end of all treatments, 1 hour after the last gavage, animals were killed by cervical dislocation and weighed. The whole GI tract was removed, divided into stomach, small intestine corresponding to duodenum, jejunum, and ileum, cecum, and proximal and distal colon (Figure S1). The sections were opened, and two-third of the section was collected in histology cassette and one-third of the section was snap frozen in liquid nitrogen. For the confocal microscopy, tissue sample were fixed in 4% paraformaldehyde (PFA) (in phosphate-buffered saline, pH 7.4) overnight and embedded in cryomold using optimum cutting temperature compound. The tissue samples were sectioned at 6 μm thickness by cryostat (CM3050 S, Leica, Nussloch, Germany), and the samples were imaged using Leica TCS SP5 confocal microscope (488 nm excitation). The frozen sections were analyzed by fluorescence spectroscopy and inductively coupled plasma mass spectrometry to determine the amount of internalized MSNs. All the experiments were done in low light to avoid possible loss for fluorescence.

Statistical analysis

The experiments were repeated a minimum of two to three times and represented as mean \pm standard error of the mean. One-way analysis of variance or Student's *t*-test (GraphPad Prism version 5.00 for Windows, GraphPad Software, La Jolla, CA, USA) was used to analyze the data: cell viability, stool hydration, goblet cell numbers, and MUC2 analysis experiments. The level of significance was set at probabilities of $*P < 0.05$, $**P < 0.01$, and $***P < 0.001$.

Results and discussion

Physicochemical characterization of functionalized MSNs

MSNs were synthesized as previously described adopting a co-condensation approach^{22,32,33} further modified by functionalization for cellular and oral delivery with PEI, PEG, and FA as described in the "Experimental" section, and summarized in Figure 1. The resulting particles and their physicochemical characteristics are summarized in Table 1. Dynamic light scattering demonstrated that particles modified with PEI were fully redispersible and colloidally stable in HEPES buffer (25 mM, pH 7.2) with a hydrodynamic size of 400–500 nm, whereas their counterparts without PEI either aggregated or sedimented, as indicated by their hydrodynamic size of $> 1,000$ nm (Table 1) due to their near-neutral net charge under the studied conditions. To determine that

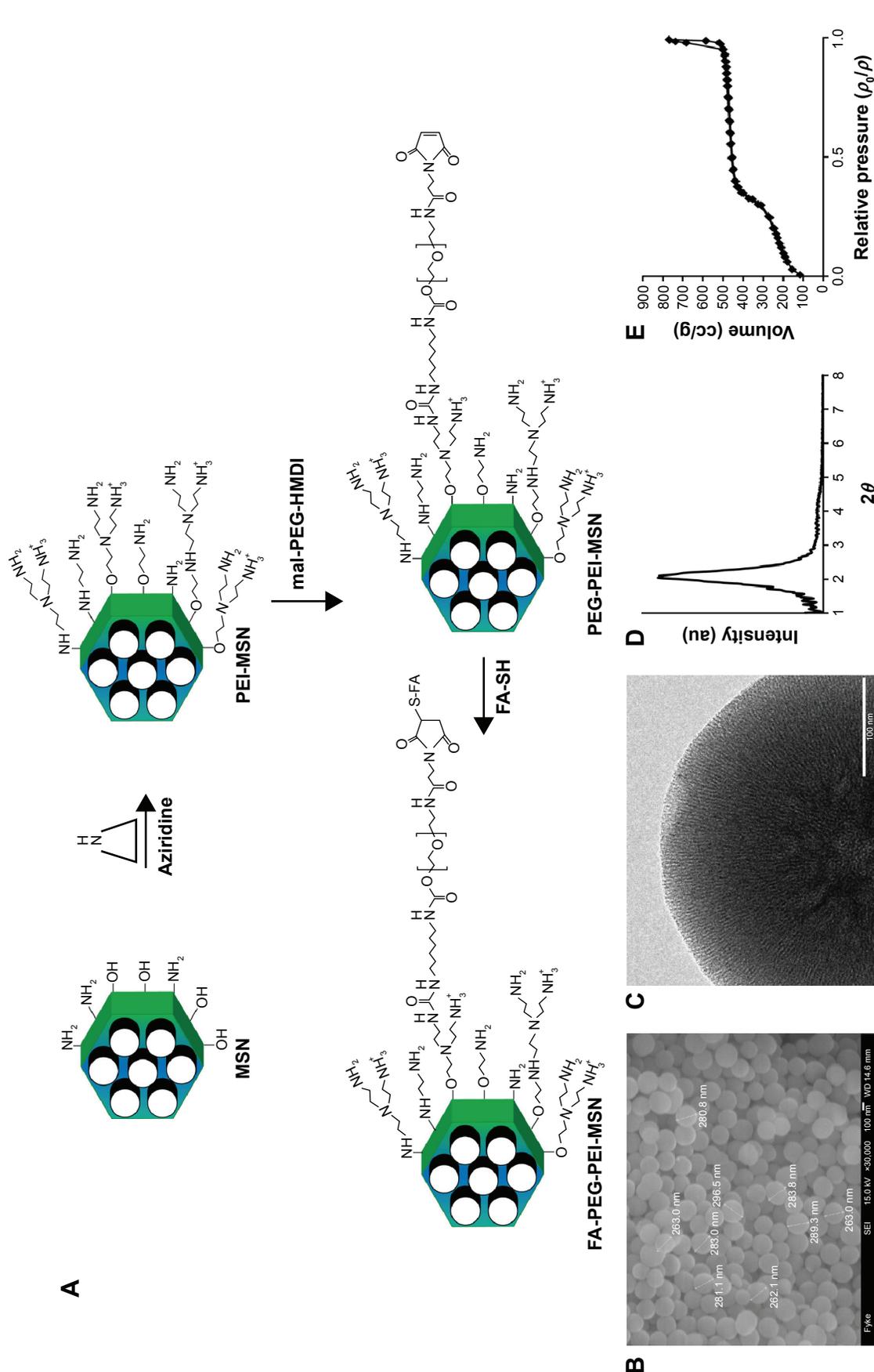


Figure 1 Synthesis, modification, and characterization of MSNs.

Notes: (A) Schematic representation for the synthesis and further modification of MSNs. (B) SEM image of MSNs shows spherical particles in the size range of 250–300 nm. (C) TEM image of MSNs shows radially aligned mesopores (scale bar = 100 nm). (D) Powder X-ray diffraction pattern of a typical MSN structure with radially arranged pores (lattice spacing = 4.34 nm). (E) N₂-sorption isotherm for MSN (S_{BET} = 905 m²/g, V_p = 0.79 cc/g, D_p = 3.5 nm) by nonlocal density functional theory.

Abbreviations: MSN, mesoporous silica nanoparticle; SEM, scanning electron microscopy; TEM, transmission electron microscopy; PEI, poly(ethylene imine); PEG, poly(ethylene glycol); FA, folic acid; mal, maleimide; au, arbitrary units; sBET, surface area calculated using BET theory; V_p, pore volume; D_p, pore diameter; SH, thiol.

Table 1 Physicochemical features of plain and functionalized MSNs

Type of particle	Hydrodynamic size (nm) in HEPES buffer (pH 7.2)		Hydrodynamic size (nm) in H ₂ O		Net surface charge/zeta potential (mV) in HEPES buffer (pH 7.2)	Total mass loss (wt%)	FA content (wt%)	Fluorescence intensity (au) at $\lambda=521$ nm in HEPES buffer (pH 7.2)
	Size (nm \pm SD)	PDI	Size (nm \pm SD)	PDI				
MSN	> 1,000	1	384 \pm 6.9	0.22	0.8	12	NA	944
PEI-MSN \pm SD	478 \pm 12	0.36	ND		52	28	NA	436
PEG-MSN	> 1,000	1	474 \pm 12	0.26	-2.1	15	NA	308
PEG-PEI-MSN	472 \pm 5.8	0.25	ND		10	35	NA	206
FA-MSN	> 1,000	1	424 \pm 6.7	0.24	-0.3	14	0.04	721
FA-PEI-MSN	410 \pm 1.9	0.21	ND		40	34	0.06	292
FA-PEG-MSN	> 1,000	1	382 \pm 30	0.19	-0.9	21	0.13	511
FA-PEG-PEI-MSN	496 \pm 3.0	0.26	ND		26	34	0.05	215

Abbreviations: MSN, mesoporous silica nanoparticle; HEPES, 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid; FA, folic acid; au, arbitrary units; PDI, polydispersity index; NA, not applicable; PEI, poly(ethylene imine); ND, not determined; PEG, poly(ethylene glycol); SD, standard deviation.

these particles were not agglomerated, but could still be fully redispersed in aqueous solvent, the hydrodynamic diameter was also determined in pure water, revealing particle sizes in the same range as those modified with PEI in HEPES buffer (400–500 nm; Table 1 and Figure S2A). All particle suspension measurements had a low polydispersity index indicating that the particles were indeed present as single particles in suspension (Table 1).

The main difference in the characteristics of functionalized particles was observed in their zeta potential values, which indicates the net/effective surface charge of the particles at the studied conditions (Table 1). Non-coated MSN particles that displayed a net surface charge (zeta potential) of \sim 0 mV was shifted to \sim 50 mV upon PEI functionalization, indicative of successful grafting of PEI (Table 1 and Figure S2B). The total amount of organic functions for each particle type was estimated based on the mass loss up to 900°C from thermogravimetric analysis measurements (Table 1 and Figure S2C). Efficient removal of the surfactant template (CTAC) from the mesopores was confirmed by Fourier transform infrared spectroscopy (Figure S2D). FA content, determined with the aid of ultraviolet-visible spectrophotometry, showed that FA was successfully conjugated to the concerned particles (Table 1).

To be able to track the particle trafficking/distribution in vitro and in vivo, the starting particles (before functionalization) were fluorescently labeled in situ with fluorescein in the form of FITC. The fluorescence intensity of the particle suspensions in HEPES buffer at a particle concentration of 1 mg/mL is given in Table 1. The scanning and transmission electron microscopy images (Figure 1B and C) confirmed the mesoporous structure, spherical morphology, and the particle sizes, which were \sim 300 nm. Radially arranged mesopores can be discerned, with the radial arrangement of pores distinguishable in the center of the particle (Figure 1C; viewed from the top). The porous structure of the particle was further characterized by powder X-ray scattering and nitrogen sorption measurements (Figure 1D and E).

PEI-, PEG-, and FA-functionalized MSNs are nontoxic at therapeutic concentrations

The in vitro cytocompatibility of all produced MSNs was studied in colon cancer Caco-2 cells using the WST-1 assay, 48 hours after incubation with MSNs. All MSNs were non-toxic to Caco-2 cells up to 50 μ g/mL concentration (Figure S3). At 100 μ g/mL, most of the particles were still non-toxic, except for PEG-PEI-MSNs and FA-PEG-PEI-MSNs, which showed a 20%–30% decrease in cell viability

compared to the vehicle dimethyl sulfoxide. This could be explained by a much higher uptake of these MSNs by the cells (Figure S3). The results show that all the produced MSNs are biocompatible at concentrations as high as 50 $\mu\text{g}/\text{mL}$. Further in vitro studies were performed at the concentration of 10 $\mu\text{g}/\text{mL}$.

MSNs can be readily internalized by colon cancer cells

The effect of surface functionalization on the internalization by two different human colon cancer cell lines (HT-29 and Caco-2 cells) was assessed by flow cytometry. As the fluorescence intensity of fluorescein varies depending on the surrounding pH³⁸ as well as the particle functionalization regime^{22,32} (Table 1), we detected the percent of cells

which have internalized particles as a measure of uptake efficiency (Figure 2A). In Caco-2 cells, the cellular uptake was determined as a function of cell confluency. This is important as the intestinal epithelial tissue comprises a sheet of interconnected cells. Particles were more efficiently internalized in non-confluent cells as compared to confluent cells (Figure 2A), as expected, since more surface area is available for uptake in non-confluent cells. Both HT-29 and Caco-2 are folate receptor-positive cell lines, but FA modification only enhanced the uptake of PEGylated particles (PEG-MSNs) and did not add further benefit to PEG-PEI-MSNs at the studied concentrations (Figure 2A and B).³⁹ This might be because the inherent high uptake of PEI- and PEG-PEI-modified particles may have already reached the saturation levels,⁴⁰ and consequently, further modification

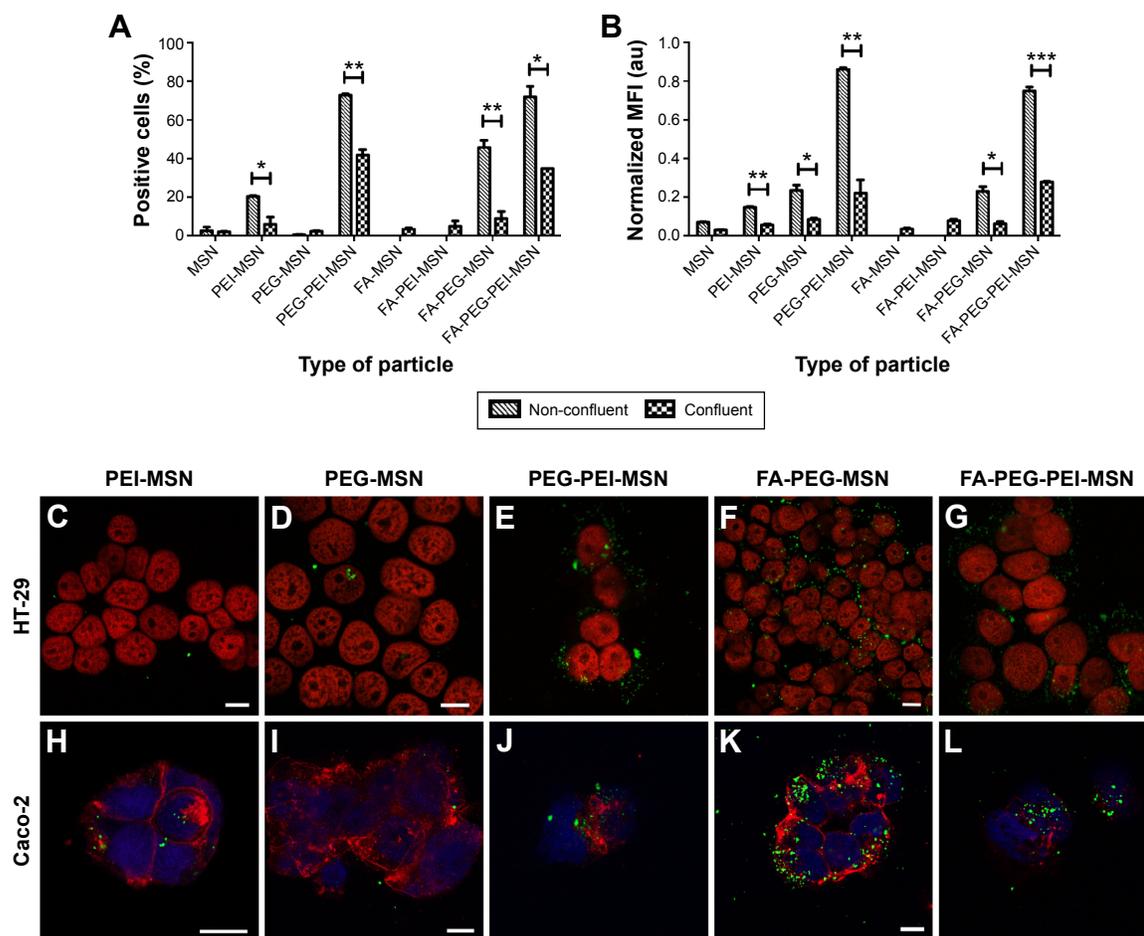


Figure 2 Effect of surface functionalization and confluency of cells on uptake of MSNs in Caco-2 and HT-29 colon cancer cells.

Notes: (A) Flow cytometric analysis of the uptake of functionalized and FITC-labeled MSNs in confluent and non-confluent Caco-2 cells after 12-hour incubation. (B) Normalized fluorescence intensity of FITC-labeled functionalized MSNs in confluent and non-confluent Caco-2 cells. Error bars represent SD ($n \geq 4$). (C–L) Confocal microscopy images are shown of HT-29 and Caco-2 cells incubated for 12 hours with PEI-MSNs, PEG-MSNs, PEG-PEI-MSNs, FA-PEG-MSNs, and FA-PEG-PEI-MSNs (green) as indicated for 12 hours. Nuclei were stained using propidium iodide and shown in red for HT-29 cells and in blue for Caco-2 cells. The Caco-2 cell membrane was stained with E-cadherin Cy 5.0 (shown in red). Scale bars = 10 μm . The level of significance was set at probabilities of * $P < 0.05$, ** $P < 0.01$, and *** $P < 0.001$.

Abbreviations: MSN, mesoporous silica nanoparticle; FITC, fluorescein isothiocyanate; SD, standard deviation; PEI, poly(ethylene imine); PEG, poly(ethylene glycol); FA, folic acid; MFI, mean fluorescence intensity.

with FA has no advantage in these cases in terms of total uptake. A boosting effect of FA on cellular uptake was only, but clearly, detected in the case when FA was linked to PEGylated particles where PEG was conjugated to the MSNs directly, without an intermediate PEI layer; and for which the basal uptake was thus, was low. All particles functionalized with PEI were internalized more efficiently than their counter-particle without PEI, in accordance with our previous studies^{22,33} (Figure S4A and B), and we have previously shown that this type of particles were internalized by the clathrin-mediated endocytosis route.²² Particles without any polymeric coating or only modified with PEG showed low uptake probably associated with their almost neutral charge as well as tendency to aggregate at neutral conditions (Table 1).⁴¹ The combination of PEI and PEG showed the most efficient uptake (Figure 2A and B). The data was verified by normalizing the mean fluorescence intensity values against fluorescence intensity of these specific particles given in Table 1 (Figure 2B and S4B). Confocal microscopy further confirmed the cellular uptake (Figure 2C–L). Here, PEG-MSN particles appeared to be aggregated, in contrast to PEI-modified particles that exhibited a more dispersed intracellular pattern (Figure 2C–L). Taken together, these data showed superior performance of particles functionalized both with PEG and PEI in terms of maximizing the cellular internalization in the targeted cell lines.

Enhancement of Notch inhibition and goblet cell differentiation in the colon is facilitated by PEG-functionalized and inhibitor-loaded MSNs in vivo

To determine the therapeutic potential of MSNs as drug delivery carriers in vivo for targeted oral drug delivery to the intestinal epithelium, we tested MSNs loaded with the γ -secretase inhibitor DAPT that inhibits the activity of Notch (Notch inhibitory particle).³⁰ We have previously used DAPT-loaded FA-PEI-MSNs to successfully demonstrate their ability to control stem cell fate in the small intestine,³⁰ where inhibiting Notch with DAPT increases the number of goblet cells resulting in increased stool hydration. To test if oral delivery and drug efficacy in vivo could be further improved by functionalization of the particles also with PEG, DAPT-loaded or control (unloaded) FA-PEI-MSNs, and FA-PEG-PEI-MSNs, DAPT alone or the vehicle control was fed to mice by oral gavage for 3 consecutive days. The uptake and effects were tested both in the small intestine, and colon, which is the target tissue in most intestinal diseases. The readout of Notch-mediated cell fate switches

was monitored by the number of goblet cells using periodic acid–Schiff staining, MUC2⁴² (the predominant intestinal mucin) mRNA analysis, and stool hydration. As expected, DAPT alone increased the number of goblet cells in colon (Figure 3A) and in the small intestine (Figure 4A and B). Importantly, the number of goblet cells in the colon of mice given DAPT/FA-PEG-PEI-MSNs were significantly higher than DAPT alone or control particles (FA-PEG-PEI-MSNs; Figure 3A and C). DAPT/FA-PEG-PEI-MSNs were also the only Notch inhibitory particles able to increase the expression of MUC2 in the colon (Figure 3B). DAPT/FA-PEI-MSNs were not able to improve the colonic Notch inhibition. Interestingly, a similar trend of improvement and goblet cell hyperplasia with PEG-PEI-coating was seen in the small intestine, whereas DAPT-loaded MSNs with FA and PEI (but not PEG) showed a significantly better inhibition of Notch compared to DAPT alone (Figure 4A and B). This effect was seen both in crypts and villi but more prominently in the villi, which have a bigger cell compartment and more goblet cells (Figure 4). These results are supported by an increased level of stool hydration only in mice fed with DAPT alone or DAPT/FA-PEG-PEI-MSNs (Figure S5), which reflects the Notch inhibition-induced cell fate switch from decreased numbers of water-absorbing colonocytes to mucus-producing goblet cells.^{43,44} The discrepancy between small intestine and colon could be related to the fact that the small intestine, being higher up in the GI tract, had been exposed to the particles for slightly longer time than the colon, and that DAPT/FA-PEI-MSNs were more efficiently internalized by the intestinal cells, subsequently leading to a more efficient drug delivery (as shown in increased uptake in cells over time; Figure S4). It is also likely that differences in colon and small intestine mucus layers influence particle penetration, for instance, the pore size and charge of mucin molecules may vary considerably along the GI tract.⁴⁵ It has been shown that particles more easily penetrate the mucus in small intestine and hence there might be no beneficial effect of PEGylation, while this modification may be necessary in the colon which has much more goblet cells/area than the small intestine (Figures 3 and 4).^{46,47} Since the main intestinal disorders, colorectal cancer, and inflammatory bowel disease occur in the colon and not in the small intestine, the PEGylation of MSN should provide a useful tool for nanomedical drug development targeting the colon. In summary, MSNs in particular when functionalized with combination of PEG and PEI are efficient carriers for drug delivery and targeted Notch inhibition by oral administration, and significantly more efficient as compared to free drug administration, especially in the colon.

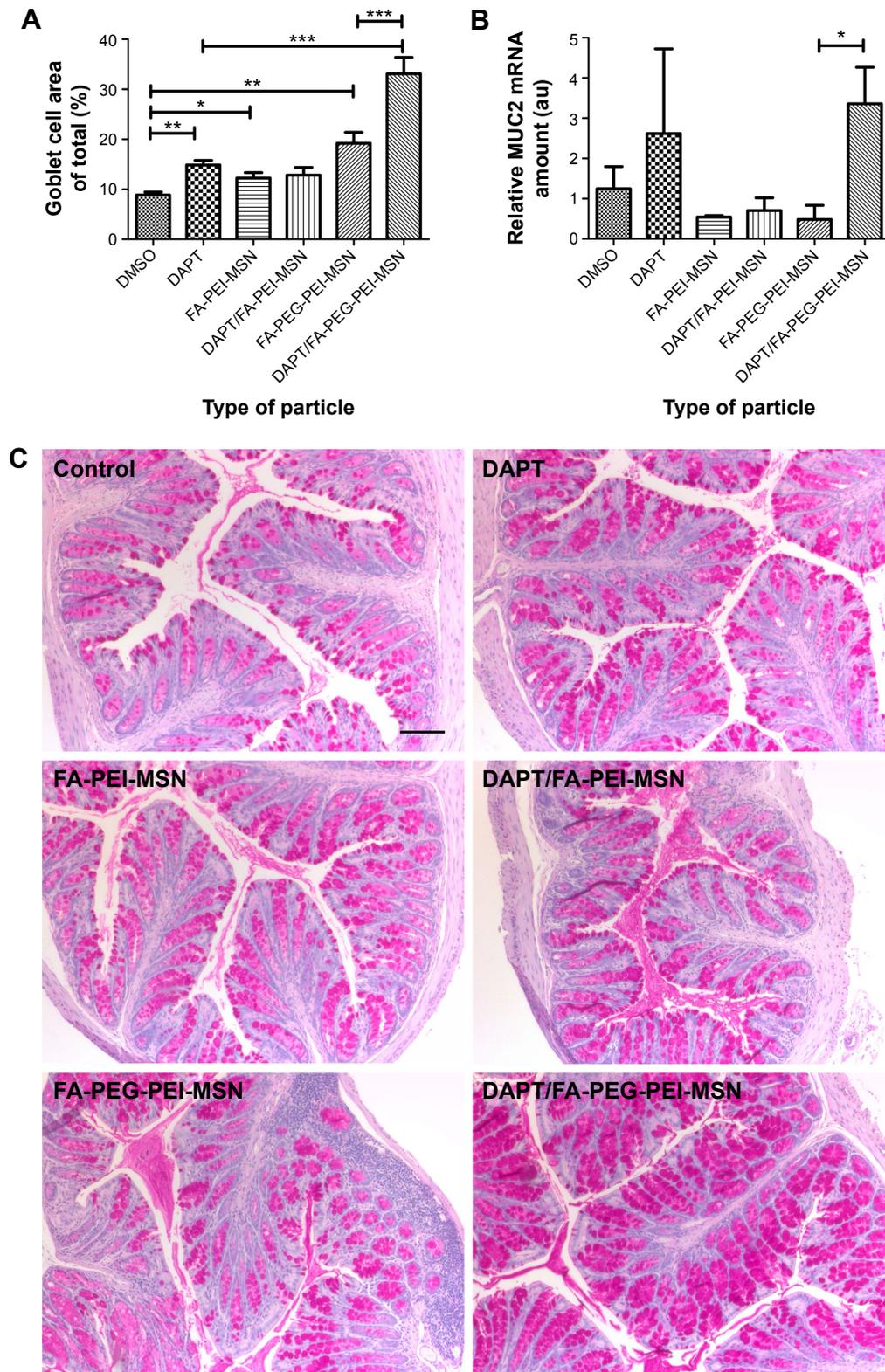


Figure 3 PEG-PEI-functionalized and Notch inhibitor DAPT-loaded MSNs induce goblet cell hyperplasia and increased levels of MUC2 in the colon.

Notes: Mice were fed vehicle control (HEPES), control particles (FA-PEI-MSNs, FA-PEG-PEI-MSNs), the Notch inhibitor DAPT, or DAPT-loaded MSNs (DAPT/FA-PEI-MSNs, DAPT/FA-PEG-PEI-MSNs) by oral gavage ($n=3$ mice per treatment). **(A and C)** The colon was excised, fixed in 4% paraformaldehyde, and processed for paraffin embedding. Sections of $6\ \mu\text{m}$ thickness were stained for goblet cells by PAS staining, and goblet cell area in colon was quantified using ImageJ ($n=3$ mice per treatment). **(B)** Colon samples were stored in RNA later, and MUC2 mRNA levels were determined by quantitative RT-PCR. Scale bar = $100\ \mu\text{m}$. Statistical analysis was done by Student's *t*-test: * $P<0.05$, ** $P<0.01$, and *** $P<0.001$.

Abbreviations: PEG, poly(ethylene glycol); PEI, poly(ethylene imine); MSN, mesoporous silica nanoparticle; HEPES, 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid; FA, folic acid; PAS, periodic acid-Schiff; RT-PCR, reverse transcription polymerase chain reaction; DMSO, dimethyl sulfoxide.

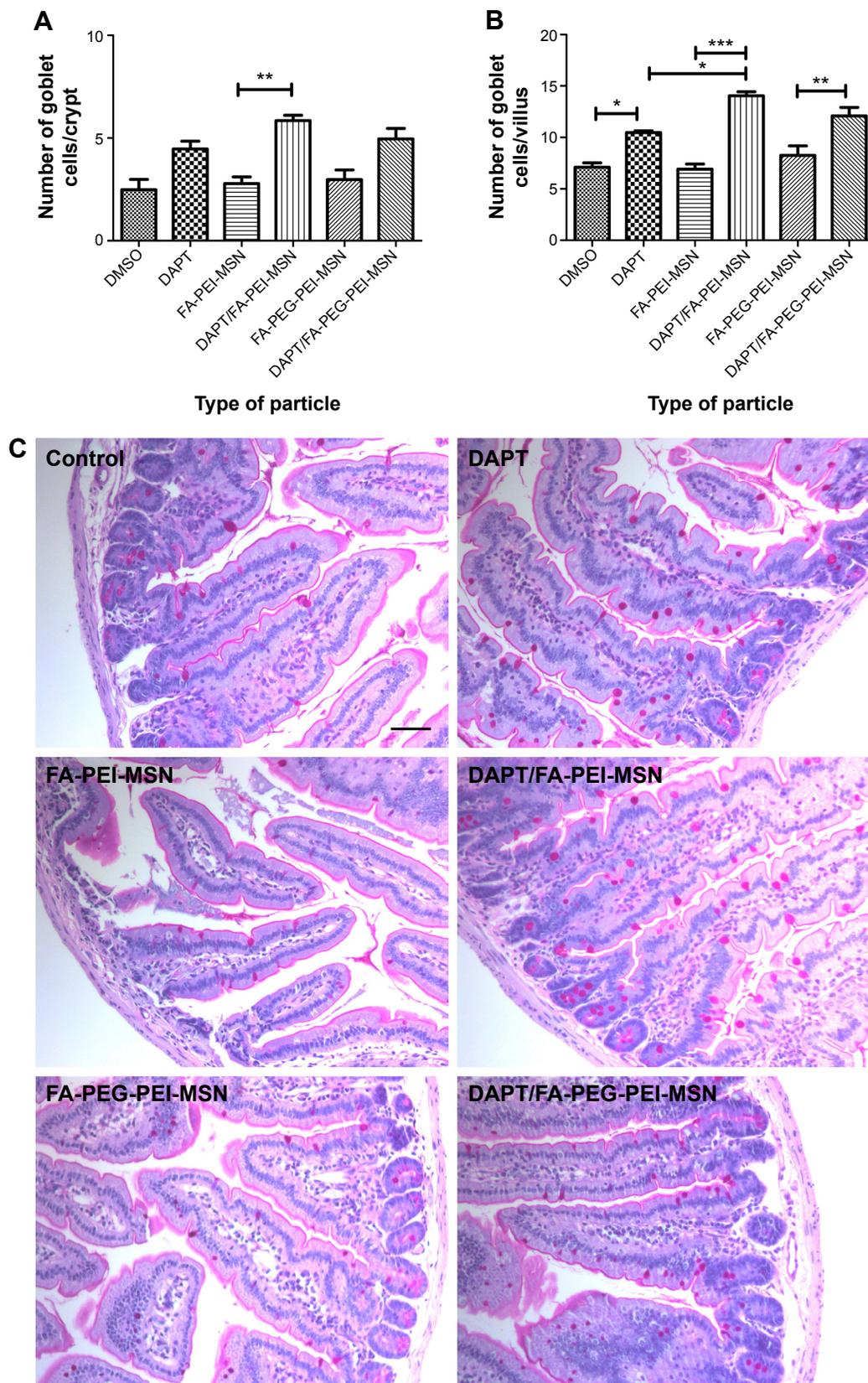


Figure 4 PEG-PEI- or PEI-functionalized and Notch inhibitor DAPT-loaded MSNs induce goblet cell hyperplasia in the small intestine.

Notes: Duodenum from mice fed with MSNs was collected, fixed in 4% paraformaldehyde, and processed for paraffin embedding. Sections of 6 μm thickness were stained for goblet cells by PAS staining, and goblet cells in intact crypts or villi were counted (**A** and **B**) and visualized in (**C**). Scale bar = 100 μm ; n=3 mice per treatment. Statistical analysis was done by Student's *t*-test: * $P < 0.05$, ** $P < 0.01$, and *** $P < 0.001$.

Abbreviations: PEG, poly(ethylene glycol); PEI, poly(ethylene imine); MSN, mesoporous silica nanoparticle; PAS, periodic acid-Schiff; DMSO, dimethyl sulfoxide; FA, folic acid.

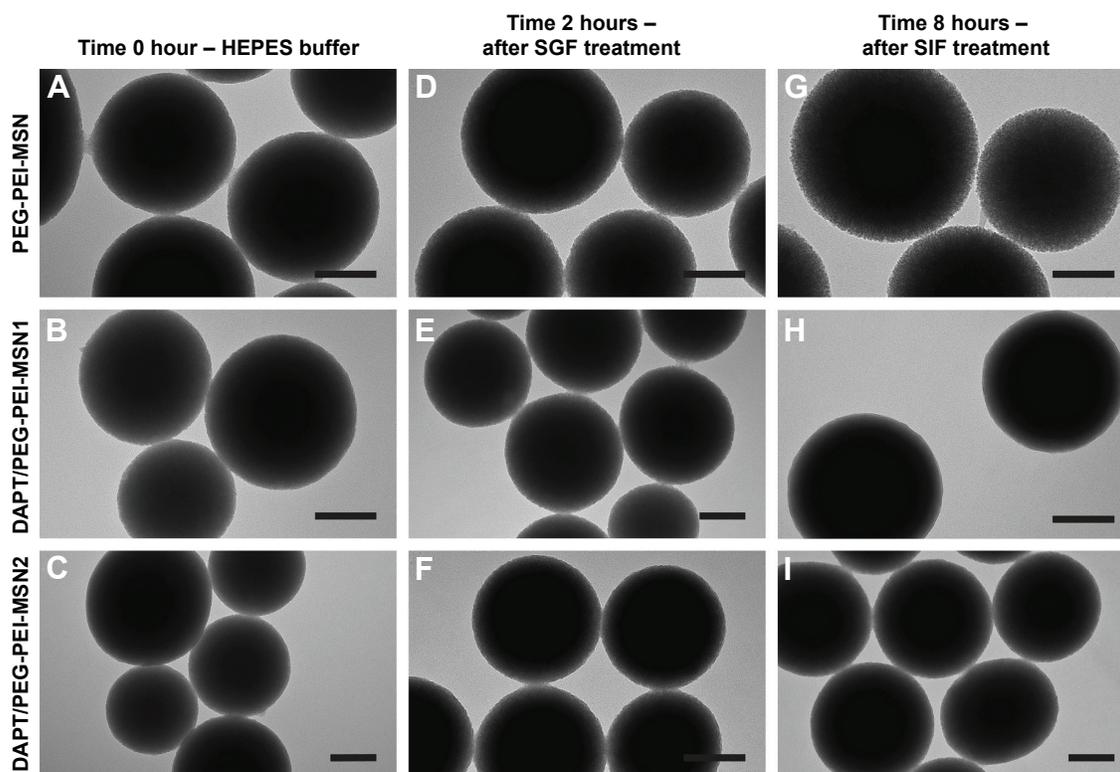


Figure 5 Functionalized and drug-loaded MSNs are stable in simulated gastric and intestinal fluids.

Notes: Transmission electron microscopy images of PEG-PEI-MSNs and DAPT/PEG-PEI-MSNs ($n=2$) (A–C) at time 0 hour in HEPES buffer, (D–F) after 2-hour incubation with SGF and (G–I) after further 6-hour incubation in SIF. Scale bar = 100 nm.

Abbreviations: MSN, mesoporous silica nanoparticle; PEG, poly(ethylene glycol); PEI, poly(ethylene imine); HEPES, 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid; SGF, simulated gastric fluid; SIF, simulated intestinal fluid.

Functionalized MSNs are stable in SGF and SIF

To evaluate whether PEG-PEI-functionalized MSNs are stable in the harsh conditions of GI tract and that they are able to protect the loaded drug/cargo, PEG-PEI-MSNs and DAPT/PEG-PEI-MSNs were exposed to SGF (containing the gastric enzyme pepsin and hydrochloric acid, pH 1.2) and SIF (containing, eg, the enzyme pancreatin, pH 6.8) whereafter the particle morphology was analyzed by TEM as described in the “Experimental” section. The TEM images show that the MSNs were intact and the structure was virtually unaltered both after the SGF (Figure 5D–F) and after the consequent SIF (Figure 5G–I) treatments compared to the untreated (starting) particles (Figure 5A–C). In order to analyze if MSNs can safely protect DAPT while passing the intestinal fluids, DAPT was eluted from MSNs and analyzed by chromatography before and after SGF and SIF incubation, which showed that DAPT is intact and has not been prematurely released nor degraded by exposure to SGF and SIF (Figure S6). Thus, functionalized MSNs can serve as an effective carrier to deliver cargo to the target site without losing its integrity even after passing through

the GI environment with low pH and presence of digestive enzymes.

Functionalized MSNs can be internalized by intestinal epithelial in vivo

To determine whether orally administered MSNs with different types of polymeric coatings were taken up by the intestinal epithelial cells in vivo, we performed daily oral gavage for 4 days of PEI-MSNs, FA-PEG-MSNs, and PEG-PEI-MSNs. The mice were healthy during the treatment and were sacrificed at the end of the experiment. The biodistribution and in vivo uptake of MSNs in the GI tract (Figure S1) were assessed in tissue cryo-sections and by confocal microscopy. While tissue sections of the control mice gavaged with vehicle (HEPES buffer) alone show only autofluorescence from the tissue (Figure 6A–D), tissue sections from the mice fed with particles showed brighter green particulate fluorescence from the ~300 nm FITC-labeled MSNs either attached to the mucosal barrier or internalized by epithelial cells (Figure 6E–P; colon not shown). This was evident in all studied GI sections but easily discernible in the stomach and the small intestinal villi. Mice given PEG-PEI-MSNs

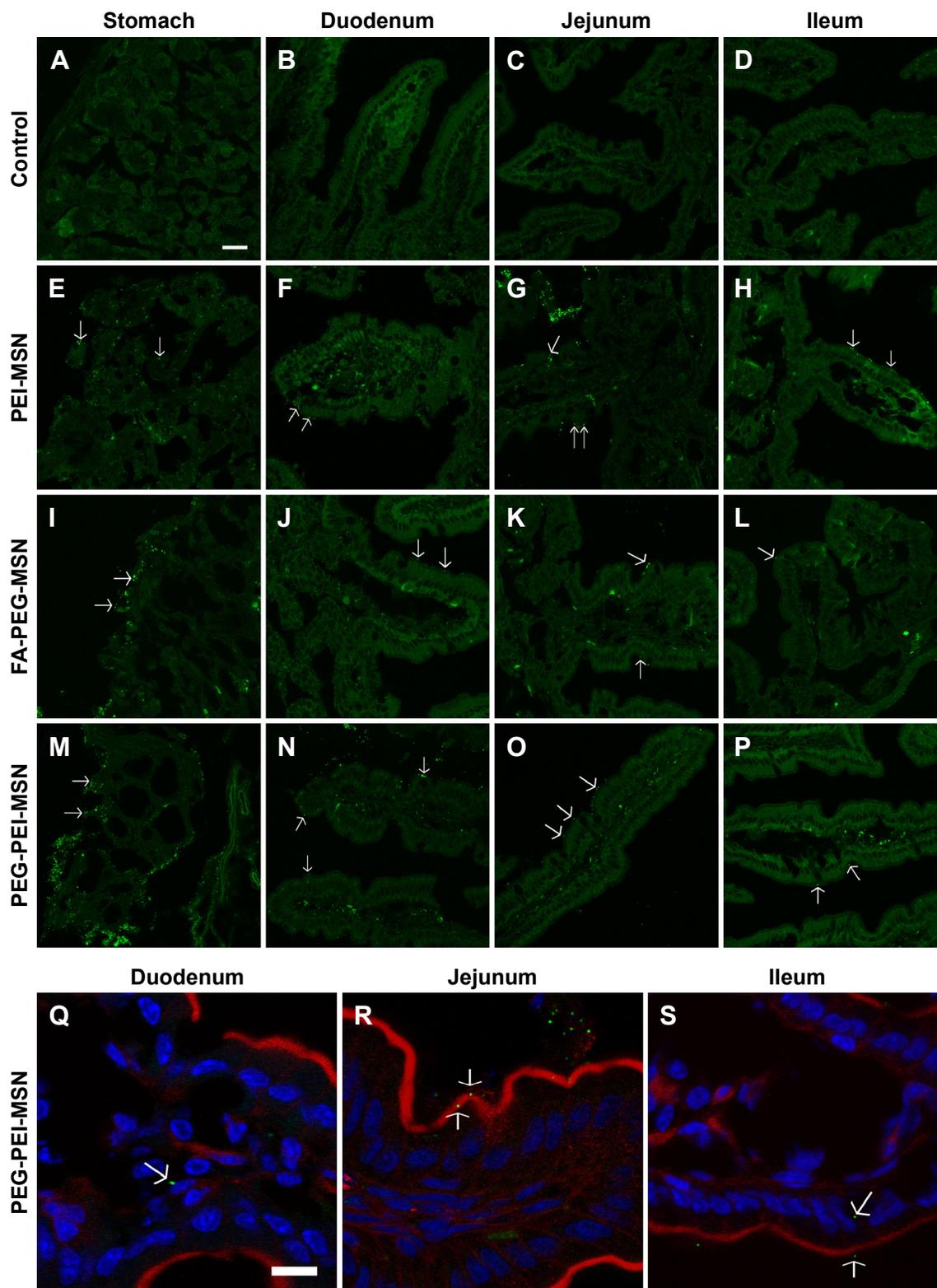


Figure 6 Functionalized MSNs are internalized to murine intestinal epithelia *in vivo* after oral administration.

Notes: (A–P) MSNs were fed to mice by oral gavage, and intestinal tissues were excised, fixed in 4% paraformaldehyde, embedded in OCT compound, sectioned, and imaged. Confocal laser scanning microscopy images of the tissue sections are shown from stomach, duodenum, jejunum, and ileum of mice fed with PEI-MSNs, FA-PEG-MSNs, and PEG-PEI-MSNs in HEPES. Control mice were fed with HEPES alone. Arrows indicate internalized nanoparticles. Scale bar = 25 μm . (Q–S) High-magnification confocal laser scanning microscopy show images of tissue sections from small intestine; duodenum, jejunum, and ileum of the mice fed with PEG-PEI-MSNs (green). Nuclei were stained with DAPI (blue) and F-actin with phalloidin (red) to indicate the brush border microvilli. Arrows indicate some of the nanoparticles particles taken up by the cells. Scale bar = 10 μm .

Abbreviations: MSN, mesoporous silica nanoparticle; OCT, optimum cutting temperature; PEI, poly(ethylene imine); FA, folic acid; PEG, poly(ethylene glycol); HEPES, 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid; DAPI, 4',6-diamidino-2-phenylindole.

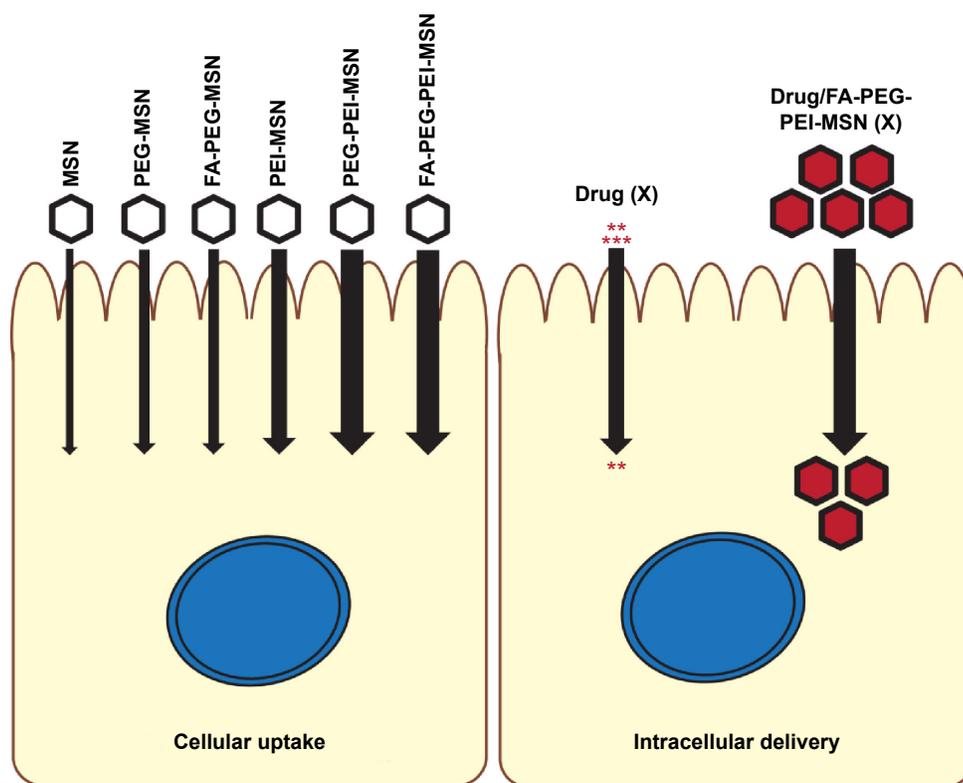


Figure 7 Graphical summary of findings.

Notes: The uptake of MSNs in intestinal cells, as determined by fluorescence-labeled MSNs, is dependent on their functionalization. The best uptake is seen by MSNs functionalized with PEI and PEG, where the addition of FA is not affecting the uptake significantly. The combination of PEI and PEG is more powerful than one of the modifications alone. Drugs are taken up more efficiently to intestinal cells if loaded into MSNs functionalized with (FA-)PEG-PEI compared to drug alone or single functionalization, suggesting that FA-PEG-PEI could be useful for targeting and drug loading to intestinal cells.

Abbreviations: MSN, mesoporous silica nanoparticle; PEI, poly(ethylene imine); PEG, poly(ethylene glycol); FA, folic acid.

appeared to have slightly more internalized particles in the epithelium, and this was further confirmed by staining for F-actin in tissue sections from mice given fluorescent MSNs to highlight epithelial apical microvilli toward the lumen and DRAQ5[®] to visualize nuclei (Figure 6Q–S). We further attempted to quantify the amount of fluorescent particles in dissected tissue sections; however, neither fluorescence spectroscopy of homogenized tissue samples (Figure S7) nor quantification of elemental silicon by inductively coupled plasma measurements (Figure S8) proved to be sensitive enough to detect changes in fluorescence or Si above the high baseline in control mice gavaged with vehicle (HEPES) alone due to autofluorescence and inherent Si content in the tissue. In summary, we have shown that PEI-, FA-PEG-, or PEG-PEI-functionalized MSNs can be taken up by intestinal epithelial cells *in vivo* after administration by oral gavage.

There are a few studies demonstrating increased drug bioavailability and/or permeability for drug-loaded MSN formulations after oral administration^{48–51} relying on the enhanced solubility that can be attained for poorly soluble drugs upon loading into a porous matrix with subsequent release into the GI tract.^{52,53} To our knowledge, however,

the present study is the first to demonstrate the potential of intestinal targeting of the MSN drug delivery carrier itself, with subsequent cellular uptake and intracellular release of a poorly soluble drug important for clinical use. Depending on the employed surface functionalization, the drug efficacy level varied in different parts of the GI tract, highlighting the potential of intestinal targeting imparted by nanomedicine.

Conclusion

We have developed a series of biocompatible MSNs with surface modifications suitable for oral drug delivery and intestinal targeting. The combination of PEI and PEG as particle surface modification is superior to enhance the internalization of MSNs to intestinal epithelial cells (Figure 7). Further, the carrier MSNs and loaded drug stay intact and functional after exposure to the harsh conditions of the stomach and the intestine. DAPT-loaded orally delivered FA-PEG-PEI-MSNs more efficiently decrease Notch signaling compared to free drug in the intestine, and the drug targeting can be further tuned by MSN surface modifications, since PEI coating exhibits higher affinity for the small intestine and PEG-PEI coating for the colon. These findings reveal

significant potential for the use of MSNs as targeted oral drug delivery carriers through epithelial barriers, provided the right surface functionalization design is employed.

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Disclosure

The authors report no conflicts of interest in this work.

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