

miRNAs: A Promising Target in the Chemoresistance of Bladder Cancer

This article was published in the following Dove Press journal:
OncoTargets and Therapy

Zhonglin Cai^{1,*}
Fa Zhang^{2,*}
Wei jie Chen^{3,*}
Jianzhong Zhang¹
Hongjun Li¹

¹Department of Urology, Peking Union Medical College Hospital, Peking Union Medical College, Chinese Academy of Medical Sciences, Beijing, People's Republic of China; ²Department of Urology, First Hospital of Lanzhou University, Lanzhou, Gansu, People's Republic of China; ³Department of Urology, Shanghai Municipal Hospital of Traditional Chinese Medicine, Shanghai Traditional Chinese Medicine University, Shanghai, People's Republic of China

*These authors contributed equally to this work

Abstract: Chemotherapy is an important cancer treatment method. Tumor chemotherapy resistance is one of the main factors leading to tumor progression. Like other malignancies, bladder cancer, especially muscle-invasive bladder cancer, is prone to chemotherapy resistance. Additionally, only approximately 50% of muscle-invasive bladder cancer responds to cisplatin-based chemotherapy. miRNAs are a class of small, endogenous, noncoding RNAs that regulate gene expression at the posttranscriptional level, which results in the inhibition of translation or the degradation of mRNA. In the study of miRNAs and cancer, including gastric cancer, prostate cancer, liver cancer, and colorectal cancer, it has been found that miRNAs can regulate the expression of genes related to tumor resistance, thereby promoting the progression of tumors. In bladder cancer, miRNAs are also closely related to chemotherapy resistance, suggesting that miRNAs can be a new therapeutic target for the chemotherapy resistance of bladder cancer. Therefore, understanding the mechanisms of miRNAs in the chemotherapy resistance of bladder cancer is an important foundation for restoring the chemotherapy sensitivity of bladder cancer and improving the efficacy of chemotherapy and patient survival. In this article, we review the role of miRNAs in the development of chemotherapy-resistant bladder cancer and the various resistance mechanisms that involve apoptosis, the cell cycle, epithelial-mesenchymal transition (EMT), and cancer stem cells (CSCs).

Keywords: miRNAs, chemoresistant, bladder cancer, biomarkers, targeted therapy

Background

Bladder cancer (BCa) is the ninth most common cancer in the world, and the incidence is higher in men than in women.^{1,2} Of the cases of initially diagnosed BCa, 70% are non-muscle-invasive bladder cancer (NMIBC), and approximately 30% are muscle-invasive bladder cancer (MIBC).³ Although the incidence of MIBC is lower than that of NMIBC, MIBC has a worse prognosis and has become a great challenge for urologists. The standard treatment for patients with MIBC is radical cystectomy. However, despite aggressive treatment, the five-year survival rate for patients with advanced BCa is only 20–40%.⁴ To improve unsatisfactory treatment efficacy, cisplatin-based combination chemotherapies, such as methotrexate, vinblastine, doxorubicin and cisplatin (MVAC) and gemcitabine and cisplatin (GC),⁵ have become important adjuvant therapies for MIBC and have been used since the late 1980s; however, the median progression time of the GC regimen is only 6 months, and the regimen has no effect on overall survival after radical cystectomy in high-risk patients.⁵ Furthermore, only 50% of patients with MIBC respond to cisplatin-based chemotherapy.⁶ In addition, chemotherapy has failed in a large proportion of patients due to the gradual occurrence of chemoresistance,

Correspondence: Hongjun Li
Department of Urology, Peking Union Medical College Hospital, Peking Union Medical College, Chinese Academy of Medical Sciences, No. 1 Shuaifuyuan, Dongcheng District, Beijing 100730, People's Republic of China
Tel +86 139 0117 1724
Email lihongjun@pumch.cn

which leads to the relapse and progression of tumors. Therefore, overcoming multidrug resistance and exploring a novel safe and effective treatment strategy is urgently needed for BCa, especially MIBC.

Recent research has indicated that posttranscriptional regulatory mechanisms play a crucial role in various tumor biological properties, including chemotherapy resistance, and the most important molecules involved include miRNAs and human antigen R.^{7,8} miRNAs are a class of endogenous small noncoding RNAs that are approximately 18–25 nt in length and were first discovered in 1993.⁹ Since their discovery, an increasing number of miRNAs have been identified. miRNAs recognize and bind to the 3'-untranslated region (3'-UTR) of target mRNAs with complete or incomplete complementary pairs and cause the degradation of the target mRNA or the inhibition of translation, thereby negatively regulating the expression of cancer-related molecules.¹⁰ Studies have revealed that nearly one-third of human genes, including those involved in tumor development, angiogenesis, invasion, metastasis and drug resistance, can be regulated by miRNAs. Therefore, miRNAs act as oncogenes or tumor suppressor genes, depending on their complex regulatory mechanisms.¹¹ In recent years, tumor-related studies have shown that miRNAs are involved in the biological properties of BCa, including chemoresistance. Based on their important roles in BCa, miRNAs have been widely studied as therapeutic targets for BCa treatment. This paper will review the role of miRNAs in the chemoresistance of BCa and explore the related targeted therapeutic strategies.

Expression Patterns of Chemoresistance-Related miRNAs in BCa

Dysregulated expression of miRNAs in tumors involves pathophysiological processes.^{12,13} With the development of microarray analysis methods, real-time polymerase chain reaction (RT-PCR) and bioinformatic techniques, numerous cancer-related miRNAs have been discovered, most of which are closely associated with chemoresistance in many tumors, including BCa.^{14–17}

The 5637 cell line is the most multichemosensitive cell line of the BCa cell lines (5637, T24, EJ, H-bc and Bui87), and H-bc is the most resistant cell line. An RNA-seq-based miRomic analysis of the 5637 and H-bc cell lines showed that 83 miRNAs were differentially expressed by at least twofold (37 were more highly expressed and 45 were less highly expressed in the 5637 cells than in the H-bc cells).¹⁸

Microarray analysis of gemcitabine-resistant and parental cells revealed 66 differentially expressed miRNAs, including 41 miRNAs and 25 unidentified human miRPlus sequences in the miRBase database.¹⁹ Among these differentially expressed miRNAs, miR-1290 and miR-138 showed increased expression levels in gemcitabine-resistant cells, while let-7b and let-7i exhibited decreased expression.^{18,19} Transfection of pre-miR-138 and pre-miR-1290 into parental cells attenuated gemcitabine-induced cell death, while transfection of pre-miR-let-7b and pre-miR-let-7i into the resistant cells augmented cell death.^{18,19} These results demonstrated the role of these miRNAs in the gemcitabine resistance of BCa and implied the potential role of targeted therapy. Several studies have shown that the expression of multiple chemoresistance-related miRNAs in BCa tissues and cell lines was either upregulated or downregulated. As found by Li et al²⁰ the expression of miR-34a was frequently decreased in MIBC tissues and cell lines (5637, HT1376, J82, T24), while its upregulation promoted the sensitivity of BCa cells to cisplatin. Bu et al²¹ also found that the expression of miR-101 was downregulated in the BCa-resistant cell line T24/CDDP, while overexpression of miR-101 significantly enhanced cisplatin-induced apoptosis. Therefore, the differential expression of miRNAs in BCa confers the significance of miRNAs as oncogenes or tumor suppressor genes.

miRNAs and BCa Chemoresistance

Although chemotherapy plays an irreplaceable role in the treatment of advanced BCa, there are still numerous patients who are unable to tolerate it, and the emergence of drug resistance greatly limits the long-term curative effect of chemotherapy. Recent research has indicated that miRNA-mediated posttranscriptional regulation plays an important role in chemoresistance and can affect drug efficacy by regulating the expression of multiple drug-resistance-related proteins. We hypothesize that the effect of targeting miRNAs is stronger than the effect of targeting individual drug-associated proteins.

Regarding BCa, the first identified drug-resistance-related miRNAs belonged to the miR-200 family. In 2009, Adam et al²² showed that the stable expression of miR-200 in mesenchymal UMUC3 cells increased E-cadherin levels and sensitivity to EGFR blockers (cetuximab) and decreased the expression of ZEB1, ZEB2, and ERFFI-1 and cell migration. Since then, studies have focused on the role of miRNAs in the chemoresistance of BCa. For instance, miR-21 promoted cell proliferation and increased cell resistance to doxorubicin treatment in the BCa cell line T24.²³ miR-203 was associated

with cancer progression and poor prognosis of BCa patients who received cisplatin-based adjuvant chemotherapy.²⁴ The restoration of miR-203 expression enhanced the sensitivity of BCa cells to cisplatin by promoting cell apoptosis by targeting Bcl-w and survivin. miR-218 increased the sensitivity of BCa to cisplatin by targeting Glut1 (glucose transporter isoform 1).²⁵ miR-193a-3p has been found to be more highly expressed in resistant cell lines (H-bc and UM-UC-3) than in chemosensitive cell lines (5637), and miR-193a-3p was shown to not only mediate BCa chemoresistance at the cellular level but was promote paclitaxel resistance by inhibiting the expression of SRSF2 and LOXL4 in nude mouse BCa tumor xenografts.²⁶ Similarly, it has been shown that other BCa chemoresistance-related miRNAs include miR-196a-5p, miR-203, miR-22-3p, miR-143, miR-193a-3b, miR-222, miR-27a, miR-145, miR-294, and miR-193b-3p (Table 1 and Figure 1).²⁷⁻³⁸ Based on these intracellular and preclinical studies, clinical research on miRNA-targeted therapeutic strategies will be the focus of cancer treatment. We believe that gene-targeted therapy will open a new chapter in cancer therapy.

miRNA-Mediated Pathways in the Chemoresistance of BCa

In terms of mechanisms, accumulating studies have revealed that miRNAs regulate the chemoresistance of BCa through a variety of signaling pathways. Current research indicates that the signaling pathways involving miRNAs are related to the following aspects:

Regulation of Cell Apoptosis

Apoptosis is one of the most important mechanisms by which multicellular organisms respond to environmental changes and maintain tissue homeostasis.³⁹⁻⁴¹ Recent studies have shown that miRNAs play a key role in regulating apoptosis and its relationship with chemoresistance in BCa. miRNAs mediate chemoresistance in BCa by regulating the expression of members of the Bcl-2 family. Zhan et al²⁴ showed that overexpression of miR-203 enhanced cisplatin sensitization by promoting apoptosis by directly targeting Bcl-w and survivin. Survivin is a key member of the inhibitor of apoptosis protein (IAP) family and exerts its antiapoptotic function by blocking caspase activity in a complex with the X-linked inhibitor of apoptosis protein (XIAP).⁴² In addition, studies have demonstrated that miR-133b regulates the proliferation and apoptosis of BCa cells (T24) by targeting Akt and

Bcl-w.⁴³ In BCa cells, Bcl-2 is regulated by miR-21, miR-192, miR-221, miR-9, miR-675, miR-29c, and Mcl-1 and is regulated by miR-192 and miR-29c.⁴⁴⁻⁴⁹ These miRNAs mediate the proliferation and apoptosis of BCa cells by regulating the expression of members of the Bcl-2 family. We speculate that the above miRNAs also play an important role in the chemotherapy resistance of BCa.

The PTEN/PI3K/Akt/mTOR signaling pathway plays a key role in the progression of BCa. The roles of Akt in cells are diverse, but all of the roles lead to antiapoptosis or cell proliferation. It is well known that the PI3K/Akt oncogenic signaling pathway is activated in response to various growth factors and extracellular matrix (ECM) proteins.^{50,51} PTEN has been reported to act as a dual-specific phosphatase, which, on the one hand, regulates cell growth, apoptosis, invasion and differentiation by negatively regulating the PI3K/Akt signaling pathway. On the other hand, PTEN downregulates the level of the lipid second messenger phosphoinositide-3,4,5-triphosphate (PIP3) via dephosphorylation and subsequently inhibits Akt phosphorylation.⁵² Studies have revealed that miRNAs mediate chemoresistance in BCa by regulating the PTEN/PI3K/Akt/mTOR signaling pathway. miR-21 can target PTEN and promote the proliferation and chemotherapy resistance (doxorubicin) of BCa cells (T24) through the PI3K-Akt pathway.⁵³ miR-222 activates the Akt/mTOR pathway and directly inhibits cisplatin-induced autophagy in BCa cells by directly targeting the protein phosphatase 2A subunit B (PPP2R2A).⁵⁴ In summary, a variety of miRNAs, including miR-21, miR-130b-3p, miR-495, miR-19a, miR-222, and the miR-130 family, have been found to target PTEN.⁵³⁻⁵⁸ Interestingly, miR-218 can indirectly regulate the expression of PTEN by targeting BMI-1, which plays a carcinogenic role and is related to tumor progression, inhibition of cell apoptosis and so on.⁵⁹ The insulin-like growth factor-1 receptor (IGF-1R) and its ligand play an important role in regulating cell proliferation and apoptosis. The combination of IGF-1R and its ligand triggers the downstream PI3K/Akt signaling pathway (Figure 2). Wang et al showed that miR-143 can target IGF-1R and promote the chemosensitivity of BCa cells (5637) to gemcitabine.²⁹

Regulation of the Cell Cycle

With advancements in our understanding of the basic mechanisms of tumor-related processes, cell cycle physiology, and apoptosis mechanisms, it is becoming increasingly apparent that the cell cycle plays a key role in chemosensitivity,

Table 1 Drug Resistance Related miRNAs in Bladder Cancer

miRNA	Expression	Cell Lines	Corresponding Drugs	Targets	Effects	First Author and Year, Refs
miR-193a-3p	down	5637, T24, EJ, H-bc, Biu87	Pirarubicin, Paclitaxel, Adriamycin, and Epirubicin Hydrochloride	SRSF2, PLAU, HIC2	MiR-193a-3p promotes both growth and paclitaxel chemoresistance.	Ly 2014 ¹⁸
miR-1290, miR-138, let-7i	up down	RT4, RT112, CUB11, TCCSUP, UM-UC-3, J82	gemcitabine	mucin-4	miRNAs 1290, 138, let-7i, and let-7b in imparting resistance to gemcitabine in UCB cell lines in part through the modulation of mucin-4.	Kozim 2013 ¹⁹
miR-34a	down	5637, HT1376, J82, T24 and 5 MIBC tissues	cisplatin	CD44	Increased miR-34a expression significantly sensitized MIBC cells to cisplatin.	Li 2014 ²⁰
miR-101	down	T24/CDDP	cisplatin	COX-2	Enforced expression of miR-101 enhances cisplatin sensitivity in human bladder cancer cells.	Bu 2014 ²¹
miR-200	up	epithelial cell lines	EGFR-blocking agents	ZEB1, ZEB2, ERRFI-1	Expression of miR-200 is sufficient to restore EGFR dependency.	Adam 2009 ²²
miR-21	up	T24	doxorubicin	PTEN	MiR-21 could modulate chemosensitivity of T24 cells to doxorubicin.	Tao 2011 ²³
miR-203	down	5637, T24 and 108 patients tissue	cisplatin	Bcl-w, Survivin	MiR-203 overexpression can enhance cisplatin sensitization.	Zhang 2015 ²⁴
miR-218	up	T24, EJ	cisplatin	Glut1	MiR-218 increases the sensitivity of bladder cancer to cisplatin by targeting Glut1.	Li 2017 ²⁵
miR-193a-3p	down	5637, T24, Biu87	Pirarubicin, Paclitaxel, Adriamycin, Cisplatin, Epirubicin Hydrochloride	SRSF2, LOXL4	MiR-193a-3p promotes both the growth and chemoresistance of the BCa cell derived tumor xenografts in nude mice.	Deng 2014 ²⁶
miR-196a-5p	up	5637	cisplatin/gemcitabine	p27	The reduced expression of miR-196a-5p enforced sensitivity to the growth inhibition effects of cisplatin/gemcitabine.	Pan 2016 ²⁷
miR-22-3p	up	H-bc	pirarubicin, paclitaxel; Adriamycin; hydrochloride; hydroxycamptothecin; cisplatin	NET1	MiR-22-3p promotes BCa chemoresistance by targeting NET1.	Xiao 2018 ²⁸

miR-143	down	T24,5637 and 20 pairs tissues	gemcitabine	IGF-IR	Overexpression of miR-143 inhibited cell proliferation and promoted chemosensitivity of bladder cancer 5637 cells to gemcitabine.	Wang 2017 ²⁹
miR-193a-3p	down	5637	pirarubicin,epirubicin	HOXC9	MiR-193a-3p promotes the multi-chemoresistance of bladder cancer by targeting the HOXC9 gene.	Lv 2015 ³⁰
miR-193a-3p	down	5637,T24,EJ,H-bc, Bju88	Pirarubicin, Paclitaxel,Adriamycin, and Epirubicin Hydrochloride	PSENI	The miR-193a-3p regulated PSENI gene suppresses the multi-chemoresistance of bladder cancer.	Deng 2014 ³¹
miR-193a-3p	down	5637,T24,EJ,H-bc, Bju89	Pirarubicin, Paclitaxel,Adriamycin, and Epirubicin Hydrochloride	ING5	The miR-193a-3p-regulated ING5 gene activates the DNA damage response pathway and inhibits multi-chemoresistance in bladder cancer.	Li 2014 ³²
miR-222	down	T24,5637	cisplatin	PPP2R2A/Akt/mTOR axis	MiR-222 induces resistance of bladder cancer cells to cisplatin.	Zeng 2016 ³³
miR-27a	down	cisplatin-resistant cell lines (EJ-R, D4-R,and G7-R)	cisplatin	SLC7A11	Overexpression of miRNA-27a reduces levels of SLC7A11 and intracellular glutathione, and resensitizes resistant cells to cisplatin.	Drayton 2014 ³⁴
miR-27a	down	89 patients tissue,cell lines	cisplatin,Adriamycin,paclitaxel	RUNX-1	MiR-27a modulates sensitivity of chemotherapy through directly inhibiting RUNX-1 expression.	Deng 2015 ³⁵
miR-145	up	T24,5673	gemcitabine	NA	lncRNA-LET/NF90/miR-145 in UBC cells to increase CSC populations and promote chemoresistance.	Zhuang 2017 ³⁶
miR-294	up	J82, HT1376, T24, SW780	cisplatin	NA	MiR 294 suppression could promote the sensitivity of T24 cells to cisplatin.	Li 2016 ³⁷
miR-193b-3p	up	NTUB1	cisplatin	ETS1,Cyclin D1	MiR-193b-3p expression upon CDDP treatment	Lin 2016 ³⁸
miR-34a	down	27 patients tissue	cisplatin	Cdk6,SIRT-1	Increased miR-34a expression levels correlated with increased chemosensitivity.	Vinall 2011 ⁶¹

Abbreviations: COX-2, cyclooxygenase-2; PTEN, Phosphatase and tension homolog deleted on chromosome 10; Glut1, Glucose transporter isoform 1; NET1, neuroepithelial cell transforming 1; Cdk6, Cyclin dependent kinase 6; SIRT-1, sirtuin-1; IGF-IR, Insulin-like growth factor 1 receptor; HOXC9, homeobox C9; LOXL4, lysyl oxidase-like 4; PSENI, presenilin 1; ING5, inhibitor of growth 5;RUNX1, Runx-related transcription factor 1; PPP2R2A, protein phosphatase 2A subunit B.

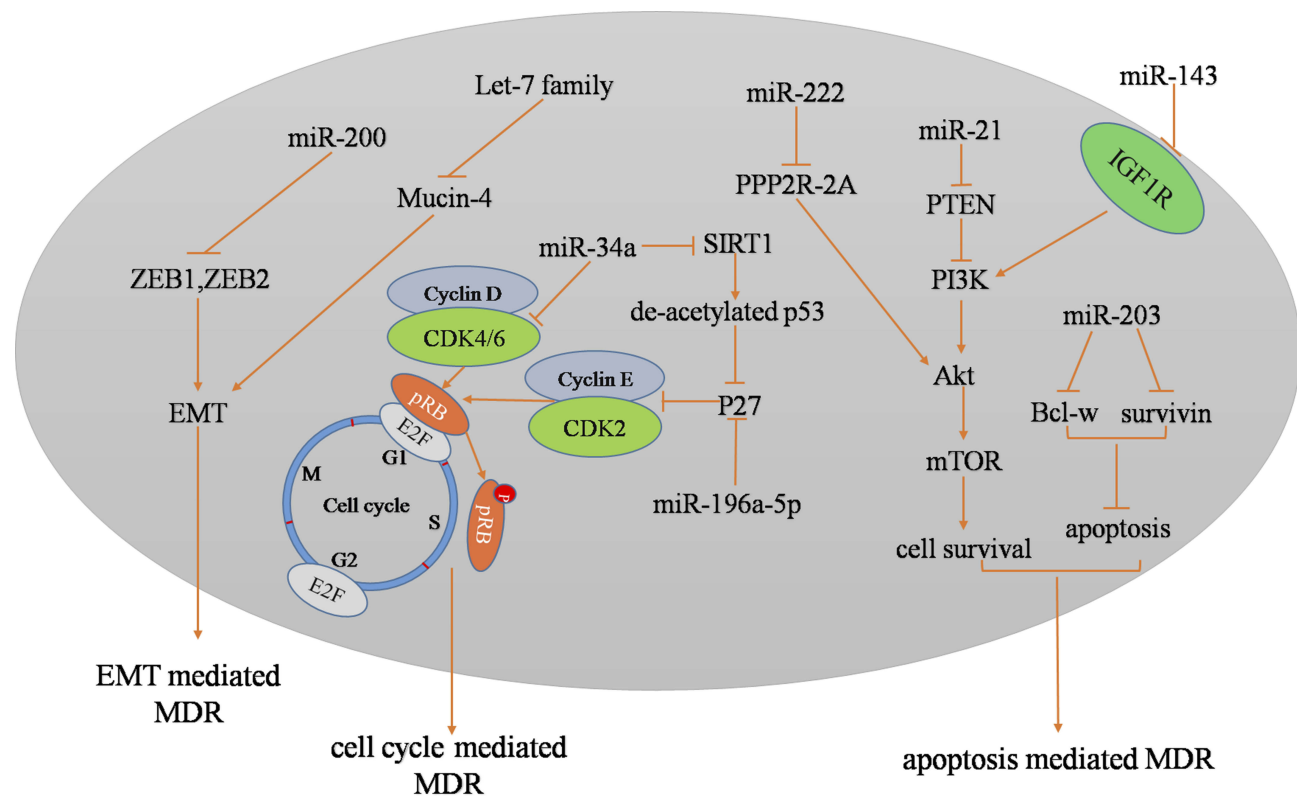


Figure 1 In BCa, these miRNAs are associated with chemoresistance.

especially sensitivity to combined chemotherapy.⁶⁰ Recent studies have shown that miRNAs participate in the processes related to cell cycle-related chemoresistance in BCa.

Increased miR-34a expression levels have been found to promote the chemosensitivity of BCa cell lines to cisplatin treatment by targeting CDK6 and SIRT-1.⁶¹ However, the inhibition of CDK6 and SIRT-1 was not as effective as the inhibition of pre-miR-34a in mediating chemosensitization. We speculate that this difference lies in the fact that miRNAs can simultaneously regulate multiple target genes, indicating that targeting these miRNAs may be more effective than targeting a single chemoresistance-related gene. UCA1, a class of lncRNAs, activates the transcription factor CREB, which leads to the expression of miR-196a-5p by binding with its promoter. miR-196a-5p is involved in the inhibition of apoptosis by UCA1 that is induced by cisplatin/gemcitabine by targeting p27.²⁷ These data suggest that the mechanisms of miRNA-mediated chemosensitivity involve a complex network system, that the novel lncRNA/miRNA/target gene axis plays an important role in tumor therapy and that any component of the axis can serve as a tumor treatment target. In addition, HOX gene family member-HOXC9 as an oncogene

and a novel miR-193a-3p target has also been found to exert a promoting effect of miR-193a-3p in BCa chemoresistance. HOXC9 has been reported to bind to and activate the expression of a large number of genes involved in the DNA damage response, such as TP53 and E2F6.⁶² Therefore, the miR-193a-3p/HOXC9/DNA damage response axis plays a key role in the chemoresistance of BCa. We found that the role of miRNAs depends on complex network axes and that an increasing number of upstream regulatory genes as well as downstream targets are involved. We hypothesize that the therapeutic effect of targeting upstream genes is superior to that of targeting downstream genes; however, complex network regulation axes still require further investigation. Studies have demonstrated that miRNAs mediate cell cycle-regulated chemoresistance by targeting CDK and p27. We also summarized that these miRNAs, including miR-29c, miR-124, miR-449a, miR-320c, miR-106a, miR-20b, and miR-195, can target CDK.^{63–69} In addition, miR-221 and miR-192 can target p27.⁷⁰ The roles of most miRNAs in the chemoresistance of BCa have not been studied, although these miRNAs can regulate the biological characteristics of BCa, which is one of the directions to be explored in the future.

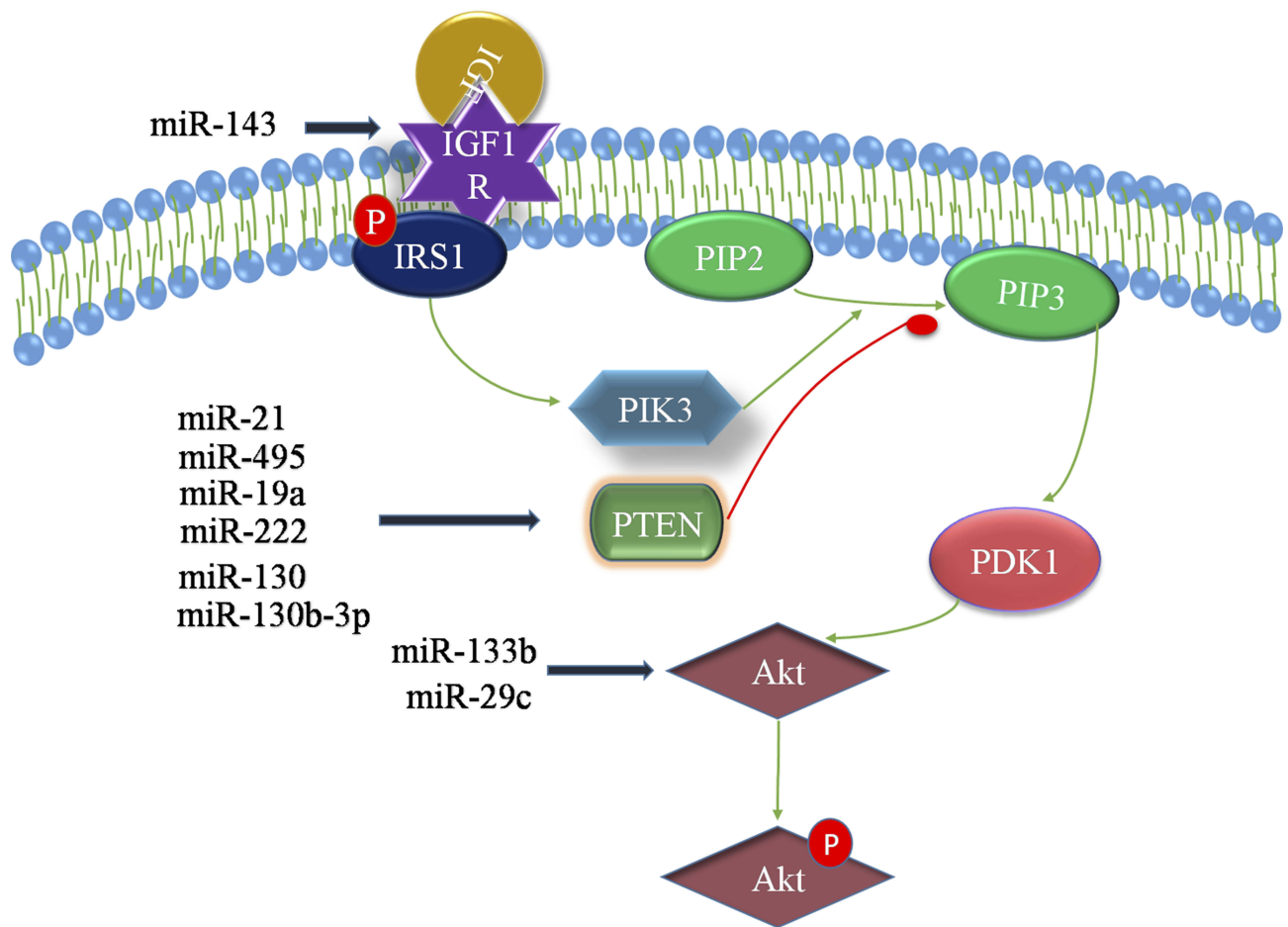


Figure 2 PI3K/PIP3 signaling activates Akt signaling via Akt/PDK-1 activation, which results in the downregulation of apoptosis. However, the conversion of PIP2 to PIP3 is reversed by PTEN. In addition, insulin-like growth factor-1 receptor (IGF-1R) and its ligand play an essential role in regulating cellular proliferation and apoptosis. The binding of the ligand to IGF-1R triggers various downstream signaling pathways, including the PI3K/Akt pathway, which is essential for cell survival. In BCa, these miRNAs regulate the PTEN/PI3K/Akt/mTOR signaling pathway.

Tumor Stem Cells

Stem cells are undifferentiated cells that have the ability to self-renew while producing differentiated tissues or organ-specific cells by asymmetric cell division. Knowledge of the importance of stem cells in normal tissue biology has led to the belief that cancer may also come from a pool of progenitor cells (the cancer stem cell (CSC) hypothesis). CSCs are a subpopulation of cancer cells responsible for tumor initiation, differentiation, recurrence, metastasis and drug resistance.^{71–73} CSCs can be isolated from a large number of tumor cells based on characteristic cell surface markers, such as CD44 and CD133. CD44 is also described as a marker for bladder CSCs that are resistant to therapeutic drugs.^{74,75} CD44 is targeted by miR-34a in MIBC cells after cisplatin treatment, and increased expression of CD44 could effectively reverse the effects of miR-34a on the proliferation, cloning potential and chemosensitivity of

MIBC cells.⁷⁶ The mechanism by which CSCs are involved in chemoresistance may be related to the fact that they produce drug-resistant daughter cells under the pressure of drug action. However, the exact mechanism remains unclear.

Epithelial-Mesenchymal Transition

Epithelial-mesenchymal transition (EMT) is a process that plays major roles in development and wound healing and is characterized by the loss of homotypic adhesion and cell polarity and increased invasion and migration. At the molecular level, EMT is characterized by the loss of E-cadherin and the increased expression of several transcriptional repressors of E-cadherin expression (ZEB1, ZEB2, Twist, Snail, and Slug).^{77,78} Studies have shown that EMT is involved in the resistance of a variety of cancer drug treatments^{79–82} and is regulated by multiple signaling pathways.^{83,84} In BCa, the

stable expression of miR-200 in mesenchymal UMUC3 cells increases E-cadherin levels; decreases the expression of ZEB1, ZEB2 and the tumor suppressor gene ERRFI-1 and cell migration; and increases EGFR blocker sensitivity.²² In addition, it has been shown that ERRFI-1 can be targeted by miR-200.²² The differential expression of the following miRNAs in BCa is associated with the EMT process: miR-433, miR-323a-3p, miR-22, miR-92, miR-96, miR-199a-5p, miR-301b, miR-613, miR-370-3p and miR-451.⁸⁵⁻⁹⁴ These miRNAs are associated with BCa cell proliferation, migration, invasion and EMT. For instance, miR-433 targets c-Met and CREB1 and inhibits EMT in BCa cells by modulating the c-Met/Akt/GSK-3 β /Snail signaling pathway.⁸⁵ miR-323a-3p regulates the EMT progression of BCa by targeting c-Met and SMAD3 and by negatively regulating their expression by modulating the c-Met/SMAD3/Snail pathway.⁸⁶ In addition, the overexpression of miR-613 enhanced the expression of the epithelial biomarker E-cadherin and inhibited the expression of mesenchymal biomarkers (vimentin, Snail and N-cadherin).⁹² Moreover, sphingosine kinase 1 (Sphk1), as an oncogene, promotes tumor cell survival by converting ceramide to

sphingosine and has been identified as a direct target gene of miR-613 in BCa cells.⁹² The recovery of Sphk1 partially reversed the inhibition of the proliferation, invasion and EMT of BCa cells induced by miR-613. Therefore, miR-613 exerts a tumor suppressive effect in BCa by targeting Sphk1. The Notch pathway has a negative regulatory effect on EMT, and DNA methylation in BCa regulates the high expression of mir-193a-3p, thus inhibiting the Notch pathway to promote EMT-induced multidrug resistance.^{18,95} Figure 3 shows the miRNAs associated with BCa EMT as well as their target genes and the EMT regulatory pathways.

Other Pathways

Generally, the chemoresistance of tumors involves complex networks. Mechanisms of multidrug resistance have been the subject of a great amount of research. In addition to the above-mentioned signaling pathways, miRNAs can also regulate chemoresistance through other novel signaling pathways.

Low expression of miR-101 induced cell survival and cisplatin resistance by negatively regulating COX-2 expression.²¹ In addition, COX-2 was also shown to be

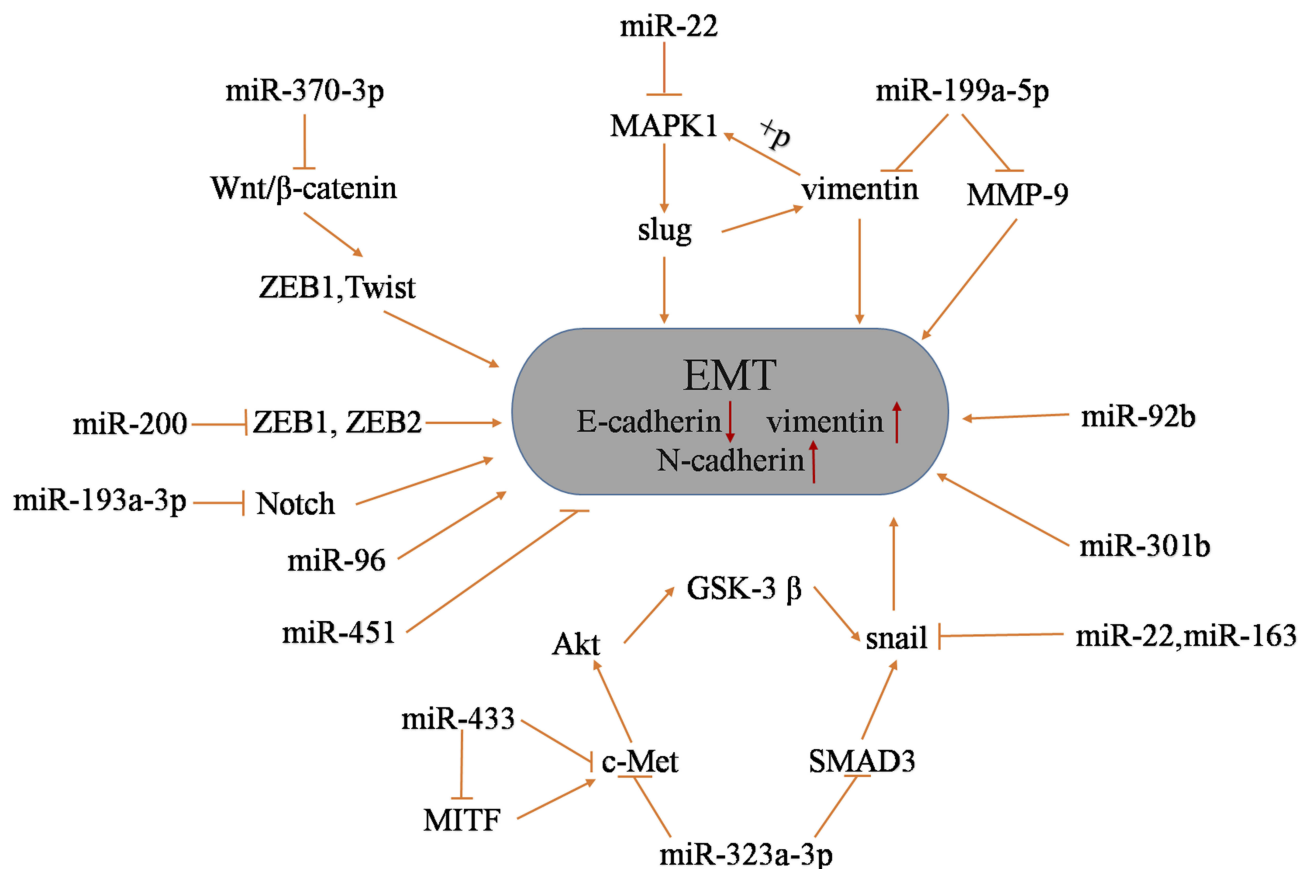


Figure 3 In BCa, these miRNAs regulate the EMT process.

a direct target of miR-101.²¹ COX-2 plays an important role in a variety of tumor drug resistances; therefore, strategies targeting the miR-101/COX-2 axis may be increasingly effective. Li et al found that miR-218 increased the sensitivity of BCa cells to cisplatin by targeting Glut1.²⁵ Glut1 is a key rate-limiting enzyme that controls glycolysis flux in cells and plays a crucial role in tumorigenesis and progression.^{96,97} Overexpression of Glut1 enhances glycolysis activity, increases cancer cell proliferation, promotes tumor invasion and metastasis and is associated with poor prognosis of various malignancies, including BCa.⁹⁸⁻¹⁰⁰ In addition, miR-22-3p has been shown to promote chemotherapy resistance in BCa by targeting NET1. NET1 is a member of the transmembrane 4 superfamily (TM4SF)¹⁰¹ and is a novel tumor-associated gene associated with many malignancies through multiple regulatory mechanisms.¹⁰²⁻¹⁰⁵ Finally, overexpression of miRNA-27a has been reported to be associated with reduced levels of SLC7A11 and the intracellular levels of glutathione and to cause resistant cells to become resensitized to cisplatin.³⁴

Notably, the tumor microenvironment, including the extracellular matrix and stromal cells, is closely related to the increase in drug resistance.¹⁰⁶ Stromal cells, such as macrophages and fibroblasts, can produce growth factors to change the characteristics of tumor cells and increase drug resistance.¹⁰⁶ Interactions between the tumor microenvironment and tumor, as well as the interior of the microenvironment, are mediated by important signal molecules. As an important signal molecule, miRNA levels are closely related to the tumor microenvironment. In BCa, it has been shown that tumor-related macrophages can promote EMT by increasing miR-30a levels,¹⁰⁷ and EMT is one of the important mechanisms of drug resistance in BCa. Although there is limited evidence of the relationship between the tumor microenvironment and miRNA levels in BCa, this is an important direction and deserves further study.

Conclusions

Tumor-related research has evolved from studying a single oncogene or tumor suppressor gene to developing the current network regulation axis model. Changes in various biological properties of tumors, including the development of chemoresistance, are closely related to the dysregulation of miRNAs. lncRNAs, circRNAs and RNA-binding proteins are involved in the regulation of miRNA expression. The lncRNA/miRNA/target gene axis, the circRNA/miRNA/target gene axis, the RBP/miRNA/target gene axis, etc., constitute a complex tumor regulatory network. miRNAs may be the

core factor, and the therapeutic effect of targeting miRNAs may be more effective than the effect of targeting a single oncogene. In tumors, miRNAs are involved in a variety of chemoresistance-related signaling pathways to regulate tumor resistance. Currently, miRNA-based therapeutic strategies include the use of agomir-miRNA, miR-101 mimic, and miRNA inhibitors. Extensive cell experiments have demonstrated the potential of these strategies to reverse chemoresistance in chemotherapy. miRNAs also have potential value in the treatment of chemotherapy resistance in BCa. In addition, the bladder is a smooth-muscle organ with an independent cavity that can be treated by perfusion administration, which greatly reduces targeting and safety problems. Therefore, miRNA-targeted therapy strategies to treat BCa chemotherapy resistance are a very promising treatment direction. Unfortunately, there are still many components of miRNA-targeted therapy strategies that need to be further elucidated. For example, how many miRNAs are associated with BCa resistance? Which miRNA is the most important? What duration of miRNA targeting is required to regulate drug resistance? Research on these issues will demonstrate the utility of miRNA-targeted therapeutic strategies and lay the foundation for further clinical trials.

Data Sharing Statement

Data sharing not applicable to this article as no datasets were generated or analysed during the current study.

Author Contributions

All authors contributed to data analysis, drafting or revising the article, gave final approval of the version to be published, and agree to be accountable for all aspects of the work.

Funding

This work is supported by the grant from National Natural Science Foundation of China (Grant No. 81671448).

Disclosure

The authors report no conflicts of interest in this work.

References

1. Ploeg M, Aben KK, Kiemeny LA. The present and future burden of urinary bladder cancer in the world. *World J Urol.* 2009;27(3):289-293. doi:10.1007/s00345-009-0383-3
2. Bo J, Yang G, Huo K, et al. microRNA-203 suppresses bladder cancer development by repressing bcl-w expression. *FEBS J.* 2011;278(5):786-792. doi:10.1111/j.1742-4658.2010.07997.x

3. Botteman MF, Pashos CL, Redaelli A, et al. The health economics of bladder cancer: a comprehensive review of the published literature. *Pharmacoeconomics*. 2003;21(18):1315–1330. doi:10.1007/BF03262330
4. Noguchi S, Mori T, Hoshino Y, et al. MicroRNA-143 functions as a tumor suppressor in human bladder cancer T24 cells. *Cancer Lett*. 2011;307(2):211–220. doi:10.1016/j.canlet.2011.04.005
5. Witjes JA, Compérat E, Cowan NC, et al. EAU guidelines on muscle-invasive and metastatic bladder cancer: summary of the 2013 guidelines. *Eur Urol*. 2014;65(4):778–792. doi:10.1016/j.eururo.2013.11.046
6. Herr HW, Dotan Z, Donat SM, et al. Defining optimal therapy for muscle invasive bladder cancer. *J Urol*. 2007;177(2):437–443. doi:10.1016/j.juro.2006.09.027
7. Srikantan S, Gorospe M. HuR function in disease. *Front Biosci (Landmark Ed)*. 2012;17:189–205. doi:10.2741/3921
8. von Roretz C, Di Marco S, Mazroui R, et al. Turnover of AU-rich-containing mRNAs during stress: a matter of survival. *Wiley Interdiscip Rev RNA*. 2011;2(3):336–347. doi:10.1002/wrna.55
9. Liang H, Gong F, Zhang S, et al. The origin, function, and diagnostic potential of extracellular microRNAs in human body fluids. *Wiley Interdiscip Rev RNA*. 2014;5:285–300. doi:10.1002/wrna.1208
10. Lytle JR, Yario TA, Steitz JA. Target mRNAs are repressed as efficiently by microRNA-binding sites in the 5' UTR as in the 3' UTR. *Proc Natl Acad Sci U S A*. 2007;104(23):9667–9672. doi:10.1073/pnas.0703820104
11. Song JH, Meltzer SJ. MicroRNAs in pathogenesis, diagnosis, and treatment of gastroesophageal cancers. *Gastroenterology*. 2012;143(1):35–47.e2. doi:10.1053/j.gastro.2012.05.003
12. Ambros V. The functions of animal microRNAs. *Nature*. 2004;431(7066):350–355. doi:10.1038/nature02871
13. Kloosterman WP, Plasterk RH. The diverse functions of microRNAs in animal development and disease. *Dev Cell*. 2006;11(4):441–450. doi:10.1016/j.devcel.2006.09.009
14. Fadejeva I, Olschewski H, Hrzenjak A, et al. MicroRNAs as regulators of cisplatin-resistance in non-small cell lung carcinomas. *Oncotarget*. 2017;8(70):115754–115773. doi:10.18632/oncotarget.v8i70
15. Yang W, Ma J, Zhou W, et al. Molecular mechanisms and therapeutic potential of miRNAs in drug resistance of gastric cancer. *Expert Opin Ther Targets*. 2017;21(11):1063–1075. doi:10.1080/14728222.2017.1389900
16. Li F, Mahato RI. MicroRNAs and drug resistance in prostate cancers. *Mol Pharm*. 2014;11(8):2539–2552. doi:10.1021/mp500099g
17. Tölle A, Ratert N, Jung K, et al. miRNA panels as biomarkers for bladder cancer. *Biomark Med*. 2014;8(5):733–746. doi:10.2217/bmm.14.26
18. Lv L, Deng H, Li Y, et al. The DNA methylation-regulated miR-193a-3p dictates the multi-chemoresistance of bladder cancer via repression of SRSF2/PLAU/HIC2 expression. *Cell Death Dis*. 2014;5:e1402. doi:10.1038/cddis.2014.367
19. Kozinn SI, Harty NJ, DeLong JM, et al. MicroRNA profile to predict gemcitabine resistance in bladder carcinoma cell lines. *Genes Cancer*. 2013;4(1–2):61–69. doi:10.1177/1947601913484495
20. Li H, Yu G, Shi R, et al. Cisplatin-induced epigenetic activation of miR-34a sensitizes bladder cancer cells to chemotherapy. *Mol Cancer*. 2014;13:8.
21. Bu Q, Fang Y, Cao Y, et al. Enforced expression of miR-101 enhances cisplatin sensitivity in human bladder cancer cells by modulating the cyclooxygenase-2 pathway. *Mol Med Rep*. 2014;10(4):2203–2209. doi:10.3892/mmr.2014.2455
22. Adam L, Zhong M, Choi W, et al. miR-200 expression regulates epithelial-to-mesenchymal transition in bladder cancer cells and reverses resistance to epidermal growth factor receptor therapy. *Clin Cancer Res*. 2009;15(16):5060–5072. doi:10.1158/1078-0432.CCR-08-2245
23. Tao J, Lu Q, Wu D, et al. microRNA-21 modulates cell proliferation and sensitivity to doxorubicin in bladder cancer cells. *Oncol Rep*. 2011;25(6):1721–1729. doi:10.3892/or.2011.1245
24. Zhang X, Zhang Y, Liu X, et al. MicroRNA-203 is a prognostic indicator in bladder cancer and enhances chemosensitivity to cisplatin via apoptosis by targeting Bcl-w and survivin. *PLoS ONE*. 2015;10(11):e0143441. doi:10.1371/journal.pone.0143441
25. Li P, Yang X, Cheng Y, et al. MicroRNA-218 increases the sensitivity of bladder cancer to cisplatin by targeting Glut1. *Cell Physiol Biochem*. 2017;41(3):921–932. doi:10.1159/000460505
26. Deng H, Lv L, Li Y, et al. miR-193a-3p regulates the multi-drug resistance of bladder cancer by targeting the LOXL4 gene and the oxidative stress pathway. *Mol Cancer*. 2014;13:234.
27. Pan J, Li X, Wu W, et al. Long non-coding RNA UCA1 promotes cisplatin/gemcitabine resistance through CREB modulating miR-196a-5p in bladder cancer cells. *Cancer Lett*. 2016;382(1):64–76. doi:10.1016/j.canlet.2016.08.015
28. Xiao J, Niu S, Zhu J, et al. miR-22-3p enhances multi-chemoresistance by targeting NET1 in bladder cancer cells. *Oncol Rep*. 2018;39(6):2731–2740. doi:10.3892/or.2018.6355
29. Wang H, Li Q, Niu X, et al. miR-143 inhibits bladder cancer cell proliferation and enhances their sensitivity to gemcitabine by repressing IGF-1R signaling. *Oncol Lett*. 2017;13(1):435–440. doi:10.3892/ol.2016.5388
30. Lv L, Li Y, Deng H, et al. MiR-193a-3p promotes the multi-chemoresistance of bladder cancer by targeting the HOXC9 gene. *Cancer Lett*. 2015;357(1):105–113. doi:10.1016/j.canlet.2014.11.002
31. Deng H, Lv L, Li Y, et al. The miR-193a-3p regulated PSEN1 gene suppresses the multi-chemoresistance of bladder cancer. *Biochim Biophys Acta*. 2015;1852(3):520–528. doi:10.1016/j.bbdis.2014.12.014
32. Li Y, Deng H, Lv L, et al. The miR-193a-3p-regulated ING5 gene activates the DNA damage response pathway and inhibits multi-chemoresistance in bladder cancer. *Oncotarget*. 2015;6(12):10195–10206. doi:10.18632/oncotarget.3555
33. Zeng LP, Hu ZM, Li K, et al. attenuates cisplatin-induced cell death by targeting the PPP2R2A/Akt/mTOR axis in bladder cancer cells. *J Cell Mol Med*. 2016;20(3):559–567. doi:10.1111/jcmm.12760
34. Drayton RM, Dudzic E, Peter S, et al. Reduced expression of miRNA-27a modulates cisplatin resistance in bladder cancer by targeting the cystine/glutamate exchanger SLC7A11. *Clin Cancer Res*. 2014;20(7):1990–2000. doi:10.1158/1078-0432.CCR-13-2805
35. Deng Y, Bai H, Hu H. rs11671784 G/A variation in miR-27a decreases chemo-sensitivity of bladder cancer by decreasing miR-27a and increasing the target RUNX-1 expression. *Biochem Biophys Res Commun*. 2015;458(2):321–327. doi:10.1016/j.bbrc.2015.01.109
36. Zhuang J, Shen L, Yang L, et al. TGFβ1 promotes gemcitabine resistance through regulating the LncRNA-LET/NF90/miR-145 signaling axis in bladder cancer. *Theranostics*. 2017;7(12):3053–3067. doi:10.7150/thno.19542
37. Li Y, Shan Z, Liu C, et al. MicroRNA-294 promotes cellular proliferation and motility through the PI3K/AKT and JAK/STAT pathways by upregulation of NRAS in bladder cancer. *Biochemistry (Mosc)*. 2017;82(4):474–482. doi:10.1134/S0006297917040095
38. Lin SR, Yeh HC, Wang WJ, et al. MiR-193b mediates CEBPD-induced cisplatin sensitization through targeting ETS1 and Cyclin D1 in human urothelial carcinoma cells. *J Cell Biochem*. 2017;118(6):1563–1573. doi:10.1002/jcb.25818
39. Danial NN, Korsmeyer SJ. Cell death: critical control points. *Cell*. 2004;116(2):205–219. doi:10.1016/S0092-8674(04)00046-7
40. Hanahan D, Weinberg RA. The hallmarks of cancer. *Cell*. 2000;100(1):57–70. doi:10.1016/S0092-8674(00)81683-9
41. Ashkenazi A. Directing cancer cells to self-destruct with pro-apoptotic receptor agonists. *Nat Rev Drug Discov*. 2008;7(12):1001–1012. doi:10.1038/nrd2637

42. Ambrosini G, Adida C, Altieri DC. A novel anti-apoptosis gene, survivin, expressed in cancer and lymphoma. *Nat Med.* 1997;3(8):917–921. doi:10.1038/nm0897-917
43. Chen XN, Wang KF, Xu ZQ, et al. MiR-133b regulates bladder cancer cell proliferation and apoptosis by targeting Bcl-w and Akt1. *Cancer Cell Int.* 2014;14:70. doi:10.1186/s12935-014-0070-3
44. Zhou C, Ding J, Wu Y. Resveratrol induces apoptosis of bladder cancer cells via miR-21 regulation of the Akt/Bcl-2 signaling pathway. *Mol Med Rep.* 2014;9(4):1467–1473. doi:10.3892/mmr.2014.1950
45. Jin Y, Lu J, Wen J, et al. Regulation of growth of human bladder cancer by miR-192. *Tumour Biol.* 2015;36(5):3791–3797. doi:10.1007/s13277-014-3020-8
46. Fu B, Wang Y, Zhang X, et al. MiR-221-induced PUMA silencing mediates immune evasion of bladder cancer cells. *Int J Oncol.* 2015;46(3):1169–1180. doi:10.3892/ijo.2015.2837
47. Wang H, Zhang W, Zuo Y, et al. miR-9 promotes cell proliferation and inhibits apoptosis by targeting LASS2 in bladder cancer. *Tumour Biol.* 2015;36(12):9631–9640. doi:10.1007/s13277-015-3713-7
48. Liu C, Chen Z, Fang J, et al. H19-derived miR-675 contributes to bladder cancer cell proliferation by regulating p53 activation. *Tumour Biol.* 2016;37(1):263–270. doi:10.1007/s13277-015-3779-2
49. Xu XD, Wu XH, Fan YR, et al. Exosome-derived microRNA-29c induces apoptosis of BIU-87 cells by down regulating BCL-2 and MCL-1. *Asian Pac J Cancer Prev.* 2014;15(8):3471–3476. doi:10.7314/APJCP.2014.15.8.3471
50. Yuan TL, Cantley LC. PI3K pathway alterations in cancer: variations on a theme. *Oncogene.* 2008;27(41):5497–5510. doi:10.1038/onc.2008.245
51. Courtney KD, Corcoran RB, Engelman JA. The PI3K pathway as drug target in human cancer. *J Clin Oncol.* 2010;28(6):1075–1083. doi:10.1200/JCO.2009.25.3641
52. Askham JM, Platt F, Chambers PA, et al. AKT1 mutations in bladder cancer: identification of a novel oncogenic mutation that can co-operate with E17K. *Oncogene.* 2010;29(1):150–155. doi:10.1038/onc.2009.315
53. Lei M, Xie W, Sun E, et al. microRNA-21 regulates cell proliferation and migration and cross talk with PTEN and p53 in bladder cancer. *DNA Cell Biol.* 2015;34(10):626–632. doi:10.1089/dna.2015.2868
54. Calderaro J, Rebouissou S, de Koning L, et al. PI3K/AKT pathway activation in bladder carcinogenesis. *Int J Cancer.* 2014;134(8):1776–1784. doi:10.1002/ijc.28518
55. Tan M, Mu X, Liu Z, et al. microRNA-495 promotes bladder cancer cell growth and invasion by targeting phosphatase and tensin homolog. *Biochem Biophys Res Commun.* 2017;483(2):867–873. doi:10.1016/j.bbrc.2017.01.019
56. Feng Y, Liu J, Kang Y, et al. miR-19a acts as an oncogenic microRNA and is up-regulated in bladder cancer. *J Exp Clin Cancer Res.* 2014;A,33::67. doi:10.1186/s13046-014-0067-8
57. Lv M, Zhong Z, Chi H, et al. Genome-wide screen of miRNAs and targeting mRNAs reveals the negatively regulatory effect of miR-130b-3p on PTEN by PI3K and Integrin β 1 signaling pathways in bladder carcinoma. *Int J Mol Sci.* 2016;18(1). doi:10.3390/ijms18010078.
58. Egawa H, Jingushi K, Hirono T, et al. miR-130 family promotes cell migration and invasion in bladder cancer through FAK and Akt phosphorylation by regulating PTEN. *Sci Rep.* 2016;6:20574. doi:10.1038/srep20574
59. Cheng Y, Yang X, Deng X, et al. MicroRNA-218 inhibits bladder cancer cell proliferation, migration, and invasion by targeting BMI-1. *Tumour Biol.* 2015;36(10):8015–8023. doi:10.1007/s13277-015-3532-x
60. Cheng JD, Werness BA, Babb JS, Meropol NJ. Paradoxical correlations of cyclin-dependent kinase inhibitors p21waf1/cip1 and p27kip1 in metastatic colorectal carcinoma. *Clin Cancer Res.* 1999;5(5):1057–1062.
61. Vinall RL, Ripoll AZ, Wang S. MiR-34a chemosensitizes bladder cancer cells to cisplatin treatment regardless of p53-Rb pathway status. *Int J Cancer.* 2012;130(11):2526–2538. doi:10.1002/ijc.26256
62. Wang X, Choi JH, Ding J, et al. HOXC9 directly regulates distinct sets of genes to coordinate diverse cellular processes during neuronal differentiation. *BMC Genomics.* 2013;14:830. doi:10.1186/1471-2164-14-830
63. Zhao X, Li J, Huang S, et al. MiRNA-29c regulates cell growth and invasion by targeting CDK6 in bladder cancer. *Am J Transl Res.* 2015;7(8):1382–1389.
64. Zhang T, Wang J, Zhai X, et al. MiR-124 retards bladder cancer growth by directly targeting CDK4. *Acta Biochim Biophys Sin (Shanghai).* 2014;46(12):1072–1079. doi:10.1093/abbs/gmu105
65. Chen H, Lin YW, Mao YQ, et al. MicroRNA-449a acts as a tumor suppressor in human bladder cancer through the regulation of pocket proteins. *Cancer Lett.* 2012;320(1):40–47. doi:10.1016/j.canlet.2012.01.027
66. Wang X, Wu J, Lin Y, et al. MicroRNA-320c inhibits tumorous behaviors of bladder cancer by targeting Cyclin-dependent kinase 6. *J Exp Clin Cancer Res.* 2014;33:69.
67. Shin SS, Park SS, Hwang B, et al. MicroRNA-106a suppresses proliferation, migration, and invasion of bladder cancer cells by modulating MAPK signaling, cell cycle regulators, and Ets-1-mediated MMP-2 expression. *Oncol Rep.* 2016;36(4):2421–2429. doi:10.3892/or.2016.5015
68. Park SL, Cho TM, Won SY, et al. MicroRNA-20b inhibits the proliferation, migration and invasion of bladder cancer EJ cells via the targeting of cell cycle regulation and Sp-1-mediated MMP-2 expression. *Oncol Rep.* 2015;34(3):1605–1612. doi:10.3892/or.2015.4119
69. Lin Y, Wu J, Chen H, et al. Cyclin-dependent kinase 4 is a novel target in microRNA-195-mediated cell cycle arrest in bladder cancer cells. *FEBS Lett.* 2012;586(4):442–447. doi:10.1016/j.febslet.2012.01.027
70. Lu Q, Lu C, Zhou GP, et al. MicroRNA-221 silencing predisposed human bladder cancer cells to undergo apoptosis induced by TRAIL. *Urol Oncol.* 2010;28(6):635–641. doi:10.1016/j.urolonc.2009.06.005
71. Galluzzi L, Vitale I, Michels J, et al. Systems biology of cisplatin resistance: past, present and future. *Cell Death Dis.* 2014;5:e1257.
72. Magee JA, Piskounova E, Morrison SJ. Cancer stem cells: impact, heterogeneity, and uncertainty. *Cancer Cell.* 2012;21(3):283–296. doi:10.1016/j.ccr.2012.03.003
73. Valent P, Bonnet D, De Maria R, et al. Cancer stem cell definitions and terminology: the devil is in the details. *Nat Rev Cancer.* 2012;12(11):767–775. doi:10.1038/nrc3368
74. Patrawala L, Calhoun T, Schneider-Broussard R, et al. Highly purified CD44+ prostate cancer cells from xenograft human tumors are enriched in tumorigenic and metastatic progenitor cells. *Oncogene.* 2006;25(12):1696–1708. doi:10.1038/sj.onc.1209327
75. Patrawala L, Calhoun-Davis T, Schneider-Broussard R, et al. Hierarchical organization of prostate cancer cells in xenograft tumors: the CD44+ α 2 β 1+ cell population is enriched in tumor-initiating cells. *Cancer Res.* 2007;67(14):6796–6805. doi:10.1158/0008-5472.CAN-07-0490
76. Yu G, Yao W, Xiao W, et al. MicroRNA-34a functions as an anti-metastatic microRNA and suppresses angiogenesis in bladder cancer by directly targeting CD44. *J Exp Clin Cancer Res.* 2014;33:779. doi:10.1186/s13046-014-0115-4
77. Peinado H, Olmeda D, Cano A. Snail, Zeb and bHLH factors in tumour progression: an alliance against the epithelial phenotype? *Nat Rev Cancer.* 2007;7(6):415–428. doi:10.1038/nrc2131
78. Baumgart E, Cohen MS, Silva Neto B, et al. Identification and prognostic significance of an epithelial-mesenchymal transition expression profile in human bladder tumors. *Clin Cancer Res.* 2007;13(6):1685–1694. doi:10.1158/1078-0432.CCR-06-2330
79. Wang Z, Li Y, Kong D, et al. Acquisition of epithelial-mesenchymal transition phenotype of gemcitabine-resistant pancreatic cancer cells is linked with activation of the notch signaling pathway. *Cancer Res.* 2009;69(6):2400–2407. doi:10.1158/0008-5472.CAN-08-4312

80. Kim HP, Han SW, Song SH, et al. Testican-1-mediated epithelial-mesenchymal transition signaling confers acquired resistance to lapatinib in HER2-positive gastric cancer. *Oncogene*. 2014;33(25):3334–3341. doi:10.1038/onc.2013.285
81. Du F, Wu X, Liu Y, et al. Acquisition of paclitaxel resistance via PI3K-dependent epithelial-mesenchymal transition in A2780 human ovarian cancer cells. *Oncol Rep*. 2013;30(3):1113–1118. doi:10.3892/or.2013.2567
82. Kajiyama H, Shibata K, Terauchi M, et al. Chemoresistance to paclitaxel induces epithelial-mesenchymal transition and enhances metastatic potential for epithelial ovarian carcinoma cells. *Int J Oncol*. 2007;31(2):277–283.
83. Horiguchi K, Shirakihara T, Nakano A, et al. Role of Ras signaling in the induction of snail by transforming growth factor-beta. *J Biol Chem*. 2009;284(1):245–253. doi:10.1074/jbc.M804777200
84. Levy L, Hill CS. Smad4 dependency defines two classes of transforming growth factor {beta} (TGF- β) target genes and distinguishes TGF- β -induced epithelial-mesenchymal transition from its antiproliferative and migratory responses. *Mol Cell Biol*. 2005;25(18):8108–8125. doi:10.1128/MCB.25.18.8108-8125.2005
85. Xu X, Zhu Y, Liang Z, et al. and CREB1 are involved in miR-433-mediated inhibition of the epithelial-mesenchymal transition in bladder cancer by regulating Akt/GSK-3 β /Snail signaling. *Cell Death Dis*. 2016;7:e2088. doi:10.1038/cddis.2015.274
86. Li J, Xu X, Meng S, et al. MET/SMAD3/SNAIL circuit mediated by miR-323a-3p is involved in regulating epithelial-mesenchymal transition progression in bladder cancer. *Cell Death Dis*. 2017;8(8):e3010. doi:10.1038/cddis.2017.331
87. Xu M, Li J, Wang X, et al. MiR-22 suppresses epithelial-mesenchymal transition in bladder cancer by inhibiting Snail and MAPK1/Slug/vimentin feedback loop. *Cell Death Dis*. 2018;9(2):209. doi:10.1038/s41419-017-0206-1
88. Huang J, Wang B, Hui K, et al. miR-92b targets DAB2IP to promote EMT in bladder cancer migration and invasion. *Oncol Rep*. 2016;36(3):1693–1701. doi:10.3892/or.2016.4940
89. He C, Zhang Q, Gu R, et al. miR-96 regulates migration and invasion of bladder cancer through epithelial-mesenchymal transition in response to transforming growth factor- β 1. *J Cell Biochem*. 2018. doi:10.1002/jcb.27172
90. Zhou M, Wang S, Hu L, et al. miR-199a-5p suppresses human bladder cancer cell metastasis by targeting CCR7. *BMC Urol*. 2016;16(1):64. doi:10.1186/s12894-016-0181-3
91. Yan L, Wang Y, Liang J, et al. MiR-301b promotes the proliferation, mobility, and epithelial-to-mesenchymal transition of bladder cancer cells by targeting EGR1. *Biochem Cell Biol*. 2017;95(5):571–577. doi:10.1139/bcb-2016-0232
92. Yu H, Duan P, Zhu H, et al. miR-613 inhibits bladder cancer proliferation and migration through targeting SphK1. *Am J Transl Res*. 2017;9(3):1213–1221.
93. Huang X, Zhu H, Gao Z, et al. Wnt7a activates canonical Wnt signaling, promotes bladder cancer cell invasion, and is suppressed by miR-370-3p. *J Biol Chem*. 2018;293(18):6693–6706. doi:10.1074/jbc.RA118.001689
94. Zeng T, Peng L, Chao C, et al. miR-451 inhibits invasion and proliferation of bladder cancer by regulating EMT. *Int J Clin Exp Pathol*. 2014;7(11):7653–7662.
95. Fang D, Kitamura H. Cancer stem cells and epithelial-mesenchymal transition in urothelial carcinoma: possible pathways and potential therapeutic approaches. *Int J Urol*. 2018;25(1):7–17. doi:10.1111/iju.2018.25.issue-1
96. Moreno-Sánchez R, Rodríguez-Enríquez S, Marín-Hernández A, et al. Energy metabolism in tumor cells. *FEBS J*. 2007;274(6):1393–1418. doi:10.1111/j.1742-4658.2007.05686.x
97. Koch A, Lang SA, Wild PJ, et al. Glucose transporter isoform 1 expression enhances metastasis of malignant melanoma cells. *Oncotarget*. 2015;6(32):32748–32760. doi:10.18632/oncotarget.v6i32
98. Amann T, Maegdefrau U, Hartmann A, et al. GLUT1 expression is increased in hepatocellular carcinoma and promotes tumorigenesis. *Am J Pathol*. 2009;174(4):1544–1552. doi:10.2353/ajpath.2009.080596
99. Shen YM, Arbman G, Olsson B, et al. Overexpression of GLUT1 in colorectal cancer is independently associated with poor prognosis. *Int J Biol Markers*. 2011;26(3):166–172. doi:10.5301/IJBM.2011.8550
100. Zhang W, Liu Y, Chen X, et al. Novel inhibitors of basal glucose transport as potential anticancer agents. *Bioorg Med Chem Lett*. 2010;20(7):2191–2194. doi:10.1016/j.bmcl.2010.02.027
101. Serru V, Dessen P, Boucheix C, et al. Sequence and expression of seven new tetraspans. *Biochim Biophys Acta*. 2000;1478(1):159–163. doi:10.1016/S0167-4838(00)00022-4
102. Ye K, Chang S, Li J, et al. A functional and protein-protein interaction analysis of neuroepithelial cell transforming gene 1 in hepatocellular carcinoma. *Tumour Biol*. 2014;35(11):11219–11227. doi:10.1007/s13277-014-2454-3
103. Zhang J, Wang J, Chen L, et al. Expression and function of NET-1 in human skin squamous cell carcinoma. *Arch Dermatol Res*. 2014;306(4):385–397. doi:10.1007/s00403-013-1423-9
104. Ecimovic P, Murray D, Doran P, et al. Propofol and bupivacaine in breast cancer cell function in vitro - role of the NET1 gene. *Anticancer Res*. 2014;34(3):1321–1331.
105. Fang L, Zhu J, Ma Y, et al. Neuroepithelial transforming gene 1 functions as a potential prognostic marker for patients with non-small cell lung cancer. *Mol Med Rep*. 2015;12(5):7439–7446. doi:10.3892/mmr.2015.4385
106. Assaraf YG, Brozovic A, Gonçalves AC, et al. The multi-factorial nature of clinical multidrug resistance in cancer. *Drug Resist Updat*. 2019;46:100645. doi:10.1016/j.drug.2019.100645
107. Zhang Q, Mao Z, Sun J. NF- κ B inhibitor, BAY11-7082, suppresses M2 tumor-associated macrophage induced EMT potential via miR-30a/NF- κ B/Snail signaling in bladder cancer cells. *Gene*. 2019;710:91–97. doi:10.1016/j.gene.2019.04.039

OncoTargets and Therapy

Dovepress

Publish your work in this journal

OncoTargets and Therapy is an international, peer-reviewed, open access journal focusing on the pathological basis of all cancers, potential targets for therapy and treatment protocols employed to improve the management of cancer patients. The journal also focuses on the impact of management programs and new therapeutic

agents and protocols on patient perspectives such as quality of life, adherence and satisfaction. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Submit your manuscript here: <https://www.dovepress.com/oncotargets-and-therapy-journal>