

# Diagnostic Role of Dysregulated Circular RNA hsa\_circ\_0036722 in Laryngeal Squamous Cell Carcinoma

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Yang Guo  
Qiang Huang  
Juan Zheng  
Chi-Yao Hsueh  
Xiaohui Yuan  
Yu Heng  
Liang Zhou

Department of Otorhinolaryngology  
Head and Neck Surgery, Shanghai Key  
Clinical Disciplines of  
Otorhinolaryngology, Eye & ENT  
Hospital of Fudan University, Shanghai,  
People's Republic of China

**Purpose:** Dysregulated circular RNAs (circRNAs) have been shown to play important roles in various cancers, and could serve as diagnostic biomarkers. However, research focusing on the roles of the circRNAs in laryngeal squamous cell carcinoma (LSCC) is limited. This research aimed to explore the expressions of hsa\_circ\_0036722 in LSCCs and its diagnostic significance.

**Materials and Methods:** The expression levels of the circular RNA, hsa\_circ\_0036722, and its parental gene, RHCG, in 41 pairs of LSCC tissues and paired adjacent normal tissues were validated with quantitative real-time polymerase chain reaction (qRT-PCR). The diagnostic values of hsa\_circ\_0036722 alone and combined with RHCG in LSCC were evaluated using receiver operating characteristic (ROC) curves. Bioinformatics analysis predicted likely cross-talk between hsa\_circ\_0036722 and RHCG. Then, qRT-PCR and luciferase reporter assay were used to examine the effect of hsa\_circ\_0036722 on miR-1248 and miR-1248 on RHCG expression. CCK-8 assays were conducted to investigate their effects on LSCC cell line.

**Results:** Hsa\_circ\_0036722 and RHCG were downregulated in LSCC tissues ( $P < 0.0001$ ). The expression level of hsa\_circ\_0036722 was significantly correlated with the differentiation level of LSCC ( $P = 0.018$ ). The area under the ROC curve of hsa\_circ\_0036722 was 0.838, which reached 0.859 when hsa\_circ\_0036722 was combined with RHCG as a biomarker. Mechanistically, hsa\_circ\_0036722 could directly sponge miR-1248 to antagonize its inhibitory effect on RHCG. And downregulation of hsa\_circ\_0036722 could promote the proliferation of LSCC cell line through upregulating miR-1248.

**Conclusion:** Our results indicated that hsa\_circ\_0036722 was downregulated in LSCC, which regulate the function of RHCG in LSCC via inhibiting miR-1248, and it could serve as a potential diagnostic marker for LSCC.

**Keywords:** circular RNA, diagnostic significance, laryngeal squamous cell carcinoma, biomarker

## Introduction

As the sixth most prevalent cancer worldwide, head and neck cancer has become a growing burden for the public.<sup>1,2</sup> Laryngeal cancer is the third most common head and neck cancer, and the vast majority is laryngeal squamous cell carcinoma (LSCC).<sup>3</sup> Larynx carcinogenesis is influenced by both environmental and genetic factors. The most extensively established environmental risk factors for LSCC are smoking and excessive alcohol consumption.<sup>4</sup> Various microRNAs (miRNAs) and long noncoding RNAs (lncRNAs) have been shown to be involved in the

Correspondence: Liang Zhou  
Department of Otorhinolaryngology  
Head and Neck Surgery, Shanghai Key  
Clinical Disciplines of  
Otorhinolaryngology, Eye & ENT Hospital  
of Fudan University, No. 83, Fenyang  
Road, Xuhui District, Shanghai 200031,  
People's Republic of China  
Fax +86 21-64377134  
Email zhoulent@126.com

carcinogenesis and development of LSCC.<sup>2,5-8</sup> Apart from miRNAs and lncRNAs, another type of non-coding RNA, called circular RNAs (circRNAs), has been demonstrated to play critical roles in LSCC.<sup>9</sup>

CircRNAs are covalently closed loops lacking 5' caps and 3' polyadenylation tails.<sup>10</sup> Initially believed to be byproducts of splicing errors, circRNAs have now been shown to participate in different pathophysiological conditions, especially in various cancers.<sup>11-15</sup> Generally, circRNAs function as “miRNA sponges” that bind to target miRNAs, suppressing their inhibiting effects on downstream mRNAs<sup>16</sup> and forming various competing endogenous RNA (ceRNA) networks. In addition, some circRNAs can interact with RNA-binding proteins to regulate different physiological processes, such as apoptosis and the cell cycle.<sup>12,17</sup> Some circRNAs can modulate the transcription, splicing and even translation of some genes.<sup>10,18</sup> The most relevant functions of circRNAs for translational medicine could be their potential as biomarkers for diverse diseases.<sup>19</sup>

The distinctive closed loop structure, which is resistant to RNA exonucleases, makes circRNAs more stable than their cognate linear mRNAs.<sup>20</sup> The characteristics of abundance and cell-, tissue-, developmental stage-, and disease-specificity of circRNA makes them uniquely advantageous as biomarkers for facilitating the diagnosis and prognosis estimation of diseases.<sup>10,21</sup> For example, hsa\_circ\_0001649 has been proposed as a potential biomarker for hepatocellular carcinoma<sup>22</sup> and hsa\_circ\_0067934 may serve as biomarker for esophageal squamous cell carcinoma (ESCC).<sup>23</sup> As far as potential circRNA biomarkers for head and neck cancer, most researches have focused on oral squamous cell carcinoma (OSCC);<sup>24-26</sup> the diagnostic and prognostic roles of circRNAs in LSCC are still largely unknown.

In this study, we focused on hsa\_circ\_0036722, a circRNA that was suggested in microarray study to be dysregulated in LSCC.<sup>9</sup> It is located at chr15:90,019,985-90,030,216 and its sample gene is Rh family C glycoprotein (RHCG), a confirmed tumor suppressor gene in various cancers.<sup>27,28</sup> First, we validated downregulated expression of hsa\_circ\_0036722 in LSCC tissues using quantitative real-time polymerase chain reaction (qRT-PCR). Second, we explored the relationships between hsa\_circ\_0036722 expression and clinicopathological characteristics in LSCC patients as well as its diagnostic value. Then, the potential LSCC-associated ceRNA network containing hsa\_circ\_0036722 and RHCG was predicted. Finally, the anti-tumor effect of

hsa\_circ\_0036722 through sponging miR-1248 to regulate RHCG in LSCC was validated.

## Materials and Methods

### Patients and Specimens

A total of 41 paired samples of LSCC and adjacent normal tissues were obtained from patients who underwent surgeries in Department of Otorhinolaryngology Head and Neck Surgery, Eye & ENT Hospital of Fudan University between 2018 and 2019. No patients received radiotherapy or chemotherapy before surgery and all LSCC tissues were confirmed by pathologists. Clinical and pathological characteristics of patients were recorded. This study was approved by the Institutional Ethics Committee of the Eye & ENT Hospital of Fudan University. Informed consent forms were signed by all patients before sample collection and the study was performed in accordance with the Declaration of Helsinki. All tissues were stored at -80 °C until use.

### RNA Extraction and qRT-PCR Analyses

TRIzol reagent (Invitrogen, Carlsbad, California, USA) was used to extract total RNA in tissues following manufacturer's instructions. Quantity and quality of RNA were examined with Nanodrop ND-2000 (Thermo Fisher Scientific, Inc). PrimeScript™ RT Master Mix (TaKaRa, Dalian, China) was used to perform reverse transcription and SYBR Green Premix Ex Taq™ II (TaKaRa, Dalian, China) was used to perform qRT-PCR according to the manufacturer's instructions. GAPDH was chosen as the internal standard control, and values of  $\Delta Ct$  or  $2^{-\Delta\Delta Ct}$  were calculated to represent the relative expression of RNAs. Higher values of  $\Delta Ct$  or  $2^{-\Delta\Delta Ct}$  represented relatively lower or higher expression levels, respectively. The primers used were as follows: forward primer of 5'-TCCAACCTTCTGTAGGGCTC-3' and reverse primer of 5'-GAAGCCTCCCAGTAGACC-3' for hsa\_circ\_0036722; forward primer of 5'-GGTTCCACTTCTTACAAGACCG-3' and reverse primer of 5'-GGGCTGACTTTACCCAGAACT-3' for RHCG; forward primer of 5'-CAATGACCCCTTCATTGACC-3' and reverse primer of 5'-TTGATTTTGGAGGGATCTCG-3' for GAPDH.

### Prediction of hsa\_circ\_0036722-miRNA-RHCG Network

According to the “miRNA sponge” theory, hsa\_circ\_0036722 might regulate the expression of its parental gene, RHCG, by modulating the miRNAs targeting

RHCG. To test this idea, we first predicted the potential miRNAs that could be bound by hsa\_circ\_0036722 using the Circular RNA Interactome (<https://circinteractome.nia.nih.gov/>) and RegRNA2.0 (<http://regrna2.mbc.nctu.edu.tw/>). The miRNAs targeting RHCG were forecasted with Targetscan (<http://www.targetscan.org>) and miRWalk 2.0 (<http://mirwalk.umm.uni-heidelberg.de/>). Finally, the miRNAs predicted to be involved in the hsa\_circ\_0036722-miRNA-RHCG network were selected from the intersection of the above results.

## Cell Culture and Transfection

AMC-HN-8 cells were cultured in RPMI-1640 (HyClone, Logan, UT) medium supplemented with 10% FBS (Gibco, Grand Island, NY), 1% streptomycin and penicillin, as well as 1% L-glutamine (Gibco, Grand Island, NY) at 37 °C in humidified air with 5% CO<sub>2</sub> as previously reported.<sup>7</sup> And siRNA targeting hsa\_circ\_0036722 and miR-1248 inhibitor as well as negative controls (NCs) were obtained from Genomeditech (Shanghai, China). The transfection was completed with Lipofectamine 2000 (Invitrogen, Carlsbad, CA, USA) according to the manufacturer's instructions.

## Cell Counting Kit-8 (CCK-8) Assay

CCK-8 assays were performed to determine cell proliferation abilities of various transfected cells. LSCC cell lines AMC-HN-8 transfected with indicated siRNAs, inhibitors or both were seeded at a concentration of  $1 \times 10^3$  cells/well into 96-well plates and cultured in 37 °C under a 5% CO<sub>2</sub> atmosphere. Cell proliferation was measured by CCK-8 reagent (CCK8, Dojindo, Osaka, Japan) following the instructions of manufacturer.

## Luciferase Reporter Assay

For hsa\_circ\_0036722-miR-1248 binding luciferase assays, the HEK-293T cells were co-transfected with miR-1248 mimics or NC combined with wild-type or mutated hsa\_circ\_0036722-luciferase reporter plasmids using Lipofectamine 2000; for the miR-1248-RHCG luciferase assay, the HEK-293T cells were co-transfected with miR-1248 mimics or NC combined with wild-type or mutated RHCG-luciferase reporter plasmids (Genomeditech, Shanghai) using Lipofectamine 2000. Luciferase activity was detected using Dual-Glo Luciferase Assay system (Promega, Madison, WI) and then Firefly luciferase activities were normalized to the Renilla luciferase activities according to the manufacturers' protocols.

## Statistical Analysis

GraphPad Prism 6.02 (GraphPad Software, La Jolla, CA, USA) and SPSS version 22.0 (SPSS Inc, Chicago, IL, USA) were used to perform the statistical analyses. The results of qRT-PCR were presented as mean  $\pm$  standard deviation (SD). Paired *t*-tests were used to compare the relative expression of hsa\_circ\_0036722 as well as RHCG in LSCC and adjacent normal tissues. The relationships between hsa\_circ\_0036722 expression levels and clinicopathological characteristics were assessed using *t*-test or one-way analysis of variance (ANOVA). The correlation between hsa\_circ\_0036722 and RHCG in LSCC patients was explored with Pearson correlation. The diagnostic values of hsa\_circ\_0036722 and RHCG were evaluated with ROC curves. *P* value <0.05 was considered statistically significant.

## Results

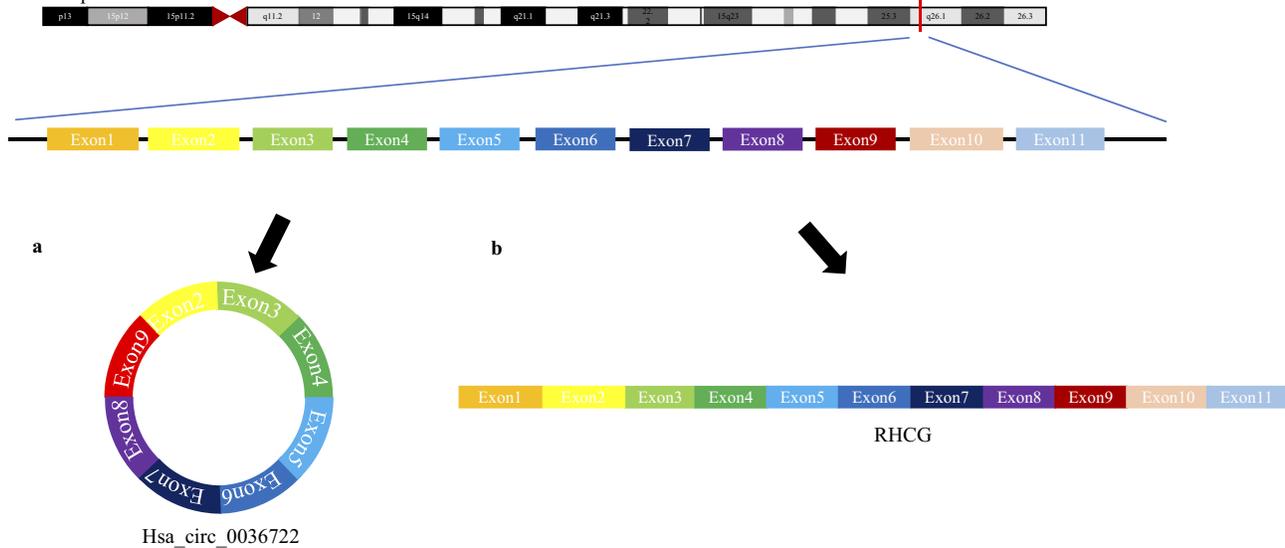
### Downregulated Expression of hsa\_circ\_0036722 in LSCC Tissues

Hsa\_circ\_0036722 consists of exons 2–9 and its spliced sequence length is 1127 bp (Figure 1), which is the longest among known exonic circRNAs generated from RHCG.<sup>29</sup> Divergent primers were used to detect the expression levels of hsa\_circ\_0036722 in 41 pairs of LSCC tissues and adjacent normal tissues. The products of qRT-PCR were sequenced and the results of Sanger sequencing experiment validated the back-splice junction of hsa\_circ\_0036722 (Figure 2A). In addition, hsa\_circ\_0036722 remain stable after RNase R treatment while its cognate linear mRNA RHCG were degraded (Figure 2B). And the results of the qRT-PCR assay indicated that the expression level of hsa\_circ\_0036722 in LSCC was significantly lower than in paired adjacent mucosa tissues ( $n = 41$ ,  $P < 0.0001$ ) (Figure 3A).

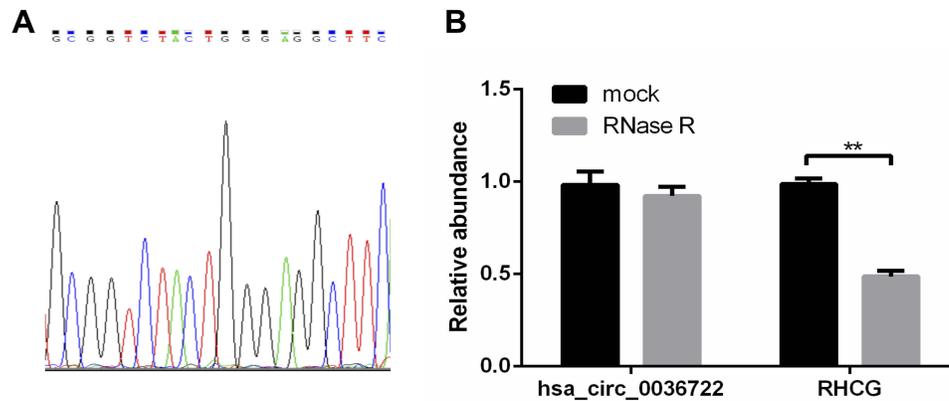
### Relationship Between hsa\_circ\_0036722 Expression Levels and Clinicopathological Factors

Relationship between the clinicopathological factors of enrolled patients and the expression levels of hsa\_circ\_0036722 was explored. Hsa\_circ\_0036722 expression levels were significantly associated with the pathological differentiation of LSCC ( $P = 0.018$ ) (Table 1). However, no other clinicopathological factors, including age, T stage, N stage or tumor sizes, were associated with hsa\_circ\_0036722 expression.

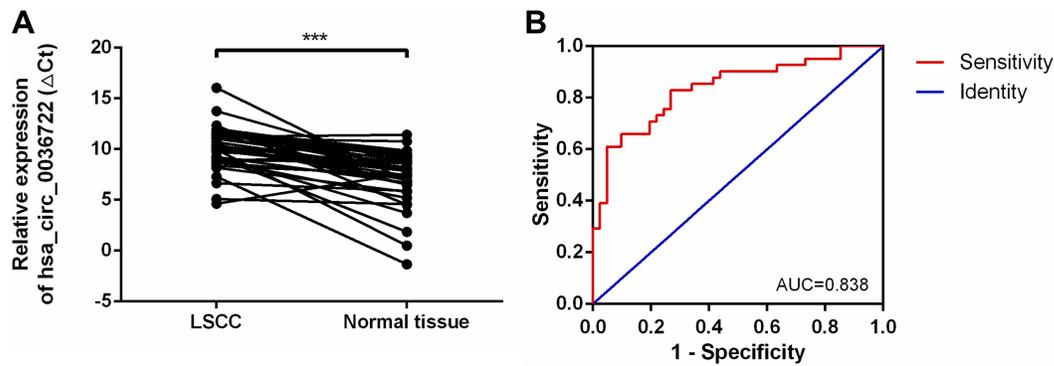
Chr15: q26.1



**Figure 1 (A)** Hsa\_circ\_0036722 is generated from chr15:90,019,985–90,030,216 through back-splicing and is composed of eight exons spanning from exon 2 to 9 of RHCg. **(B)** Canonical splicing encodes the corresponding linear mRNA, RHCg.



**Figure 2 (A)** The back-splicing junction of hsa\_circ\_0036722 was validated with Sanger sequencing experiments. **(B)** The stabilities of hsa\_circ\_0036722 and its cognate linear mRNA RHCg treated with RNase R was assessed by qRT-PCR (\*\**P* < 0.01).



**Figure 3 (A)** Hsa\_circ\_0036722 was downregulated in LSCC tissues compared to paired adjacent normal tissues (n = 41, \*\*\**P* < 0.0001). Higher values of ΔCt represent relatively lower expression levels. **(B)** The diagnostic potential of hsa\_circ\_0036722 in LSCC. The area under the ROC curve (AUC) was 0.838 (95% CI: 0.750 to 0.925, *P* < 0.0001).

**Table 1** Correlation Between Clinicopathological Factors and hsa\_circ\_0036722 Expression Levels ( $\Delta$ Ct) in Laryngeal Squamous Cell Carcinoma Patients

| Characteristics  | Number of Patients | Mean $\pm$ SD    | P value |
|------------------|--------------------|------------------|---------|
| Age              |                    |                  |         |
| <60              | 13                 | 9.77 $\pm$ 1.91  | 0.4482  |
| $\geq$ 60        | 28                 | 10.31 $\pm$ 2.18 |         |
| Tumor sizes (cm) |                    |                  |         |
| <3               | 18                 | 10.05 $\pm$ 2.76 | 0.8073  |
| $\geq$ 3         | 23                 | 10.21 $\pm$ 1.43 |         |
| Differentiation  |                    |                  |         |
| Well             | 11                 | 8.83 $\pm$ 1.92  | 0.0180* |
| Moderate         | 26                 | 10.43 $\pm$ 1.75 |         |
| Poor             | 4                  | 11.89 $\pm$ 3.06 |         |
| T stage          |                    |                  |         |
| T1–2             | 18                 | 9.80 $\pm$ 2.35  | 0.2607  |
| T3–4             | 23                 | 10.41 $\pm$ 1.88 |         |
| N stage          |                    |                  |         |
| N0               | 25                 | 9.95 $\pm$ 1.78  | 0.4784  |
| N1–3             | 16                 | 10.44 $\pm$ 2.54 |         |

**Note:** \*Indicated statistical significance.

## Diagnostic Value of hsa\_circ\_0036722 in LSCC

Considering the significant relationship between the expression levels of hsa\_circ\_0036722 and the clinicopathological characteristics of LSCC patients, we performed receiver operating characteristic (ROC) curve analysis to investigate the diagnostic value of hsa\_circ\_0036722 expression. The area under the ROC curve (AUC) for hsa\_circ\_0036722 distinguishing LSCC from adjacent normal tissues was 0.838 (95% confidence interval (CI): 0.750 to 0.925;  $P < 0.0001$ ; **Figure 3B**), indicating its potential as a biomarker for LSCC.

## Downregulated Expression of RHCG and Its Diagnostic Value in LSCC Tissues

Usually, the possible functions of circRNAs are associated with their parental genes.<sup>30–32</sup> Thus, we hypothesized that the functions of RHCG could partly explain the probable functions of hsa\_circ\_0036722 in LSCC. The expression levels of RHCG in LSCC and paired adjacent normal tissues were explored by qRT-PCR. The results suggested that RHCG expression was significantly lower in LSCC than in adjacent normal tissues ( $n = 41$ ,  $***P < 0.0001$ ) (**Figure 4A**). The ROC curve was built to explore the

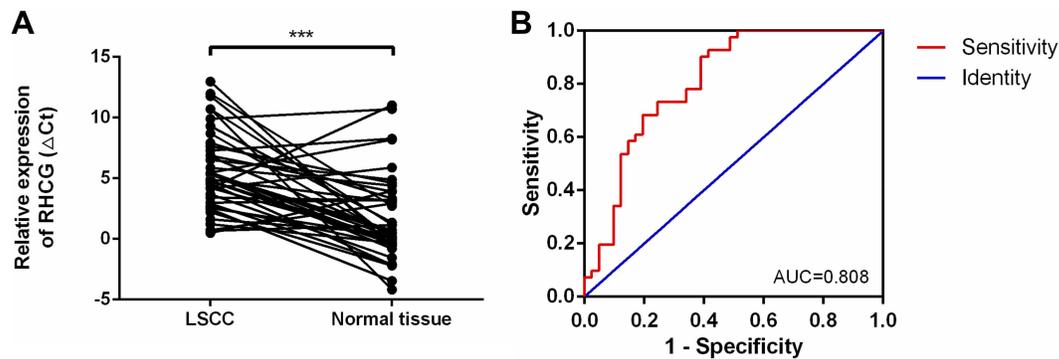
diagnostic value of RHCG; the AUC of RHCG for differentiating LSCC from paired normal tissues was 0.808 (95% CI: 0.713 to 0.904,  $P < 0.0001$ ) (**Figure 4B**).

## Relationship Between hsa\_circ\_0036722 and RHCG, and Diagnostic Value of the Combination of hsa\_circ\_0036722 and RHCG in LSCC

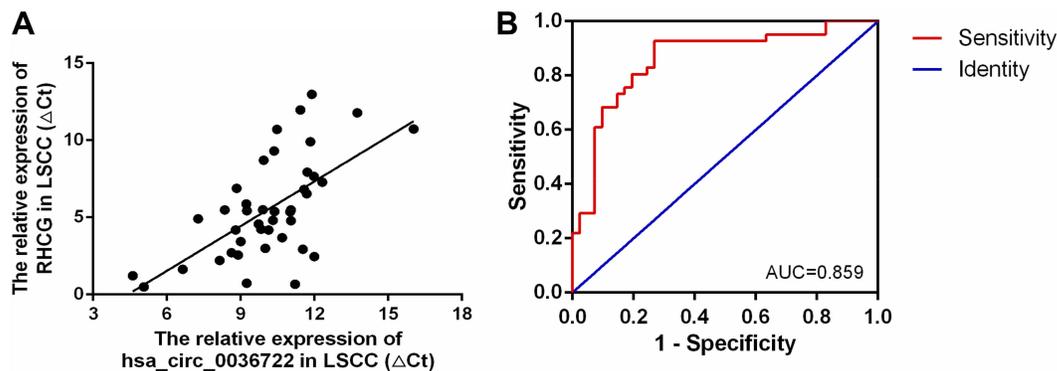
As both hsa\_circ\_0036722 and RHCG were downregulated in LSCC tissues and hsa\_circ\_0036722 was generated from sample gene of RHCG, it was of interest to see if there was a correlation between the expression levels of hsa\_circ\_0036722 and RHCG. Pearson correlation analysis indicated the expression levels of hsa\_circ\_0036722 and RHCG were positively correlated in LSCC tissues ( $r = 0.6220$ ,  $P < 0.001$ ) (**Figure 5A**). To explore the potential value of the combination of hsa\_circ\_0036722 and RHCG as a diagnostic biomarker, ROC curve analysis was conducted. The AUC of the combination reached 0.859 (95% CI: 0.775 to 0.943,  $P < 0.0001$ ), suggesting that the combination of the two markers was a better diagnostic biomarker for LSCC (**Figure 5B**).

## Potential ceRNA Network Involved in hsa\_circ\_0036722-RHCG Interactions

To explore the likely ceRNA regulatory network involving hsa\_circ\_0036722 and RHCG, the miRNAs that can bind to both hsa\_circ\_0036722 and RHCG were bioinformatically predicted. The Circular RNA Interactome indicated that 52 miRNAs could be targeted by hsa\_circ\_0036722, while RegRNA 2.0 found fewer candidate miRNAs. TargetsCan and miRWalk 2.0 showed that RHCG could be targeted by 1642 and 2313 miRNAs, respectively. MiR-1248 was the only miRNA predicted by all four website tools (**Figure 6A**). The results were represented with Venny 2.0 (<http://bioinfo.gp.cnb.csic.es/tools/venny/index.html>); sequence matching between the predicted circRNA-miRNA-mRNA interactions are shown in **Figure 6B**. Thus, the hsa\_circ\_0036722 might play a role in LSCC through the miR-1248/RHCG axis. We next investigated the clinical significance of miR-1248 in head and neck squamous cell carcinoma (HNSCC) using an integrated online bioinformatic tool (<http://kmplot.com/analysis/>) established by Lánczky et al.<sup>33</sup> Higher levels of miR-1248 predicted poorer prognosis of HNSCC patients, indicating that miR-1248 acted as oncogene in HNSCC (**Figure 6C**). According to the ceRNA theory, this was consistent with the tumor



**Figure 4 (A)** RHCG was downregulated in LSCC tissues compared to paired adjacent normal tissues ( $n = 41$ ,  $***P < 0.0001$ ). Higher values of  $\Delta Ct$  represented relatively lower expression levels. **(B)** The diagnostic potential of RHCG in LSCC. The area under the ROC curve (AUC) was 0.808 (95% CI: 0.713 to 0.904,  $P < 0.0001$ ).



**Figure 5 (A)** Expression levels of RHCG were significantly correlated with the expression levels of *hsa\_circ\_0036722* in LSCC tissues ( $r = 0.6220$ ,  $P < 0.001$ ). **(B)** The diagnostic potential of the combination of *hsa\_circ\_0036722* and RHCG in LSCC. The area under the ROC curve (AUC) was 0.859 (95% CI: 0.775 to 0.943,  $P < 0.0001$ ).

suppressor roles of *hsa\_circ\_0036722* and RHCG in LSCC considering their downregulation observed in this study.

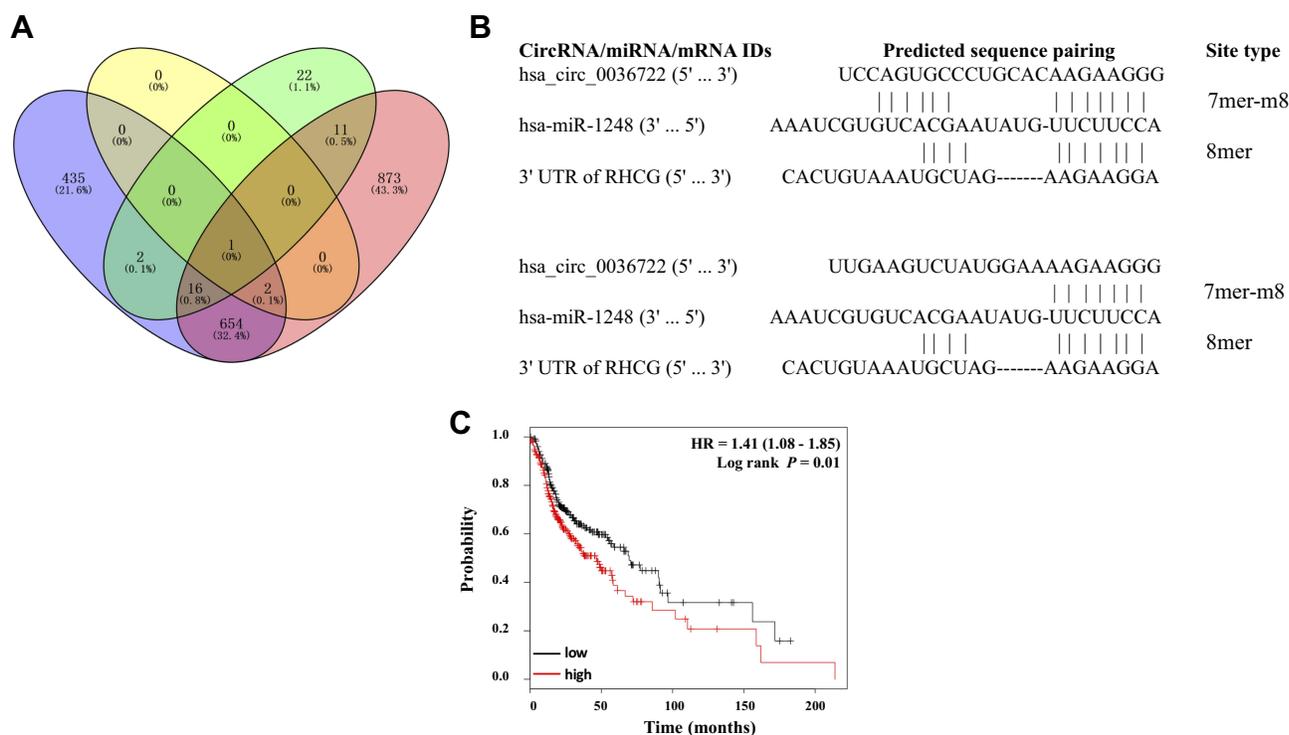
### Hsa\_circ\_0036722 Functions Through miR-1248/RHCG Axis in AMC-HN-8 LSCC Cells

After transfection of siRNA targeting *hsa\_circ\_0036722*, the expression levels of *hsa\_circ\_0036722* were downregulated as proved by qRT-PCR (Figure 7A). And the expression levels of miR-1248 and RHCG in AMC-HN-8 transfected with siRNAs targeting *hsa\_circ\_0036722* were studied. The qRT-PCR results indicated that miR-1248 was upregulated while RHCG was downregulated after *hsa\_circ\_0036722* was inhibited in AMC-HN-8 cells (Figure 7B and C). To investigate the role of miR-1248 in LSCC, AMC-HN-8 was transfected with miR-1248 inhibitors, and the transfection efficiency was examined by qRT-PCR (Figure 7D). In accordance with our prediction, the downregulation of miR-1248 significantly increased the expression of RHCG (Figure 7E). And co-

transfection of miR-1248 inhibitors could partly contradict with RHCG decrease induced by downregulated *hsa\_circ\_0036722* (Figure 7F). Furthermore, luciferase reporter assays were performed to confirm the relationship between miR-1248 and *hsa\_circ\_0036722*/RHCG, respectively. The data illustrated that miR-1248 upregulation significantly restrained the activity of *hsa\_circ\_0036722*-WT reporter and RHCG-WT reporter (Figure 7G and H).

### Ablation of *hsa\_circ\_0036722* Promote the Proliferation of AMC-HN-8 LSCC Cells Which Could Be Partly Reversed by the Inhibition of miR-1248

CCK-8 assay showed that the proliferation ability of AMC-HN-8 transfected with siRNAs targeting *hsa\_circ\_0036722* was enhanced compared to controls (Figure 8A). The proliferation ability of AMC-HN-8 transfected with miR-1248 inhibitors was significantly decreased compared to negative controls (Figure 8B). And it worth noting that co-transfection of miR-1248



**Figure 6 (A)** MiR-1248 was predicted to be able to bind to both hsa\_circ\_0036722 and RHCg. Green area and yellow area indicate predictions from Circular RNA Interactome and RegRNA 2.0, respectively, of which miRNAs could be potentially sponged by hsa\_circ\_0036722; blue area and red area represent the potential miRNAs targeting RHCg predicted by Targetscan and miRWalk 2.0, respectively. **(B)** Predicted sequences pairing in the hsa\_circ\_0036722/miR-1248/RHCg ceRNA network. Hsa\_circ\_0036722 contains two predicted binding sites for miR-1248 and RHCg had one predicted 8-mer type binding site for miR-1248. **(C)** Higher levels of miR-1248 predict poorer prognosis of head and neck squamous cell carcinoma (HNSCC) patients.

inhibitors in AMC-HN-8 transfected with siRNA targeting hsa\_circ\_0036722 could partly reverse enhanced proliferation ability induced by hsa\_circ\_0036722 downregulation (Figure 8C).

## Discussion

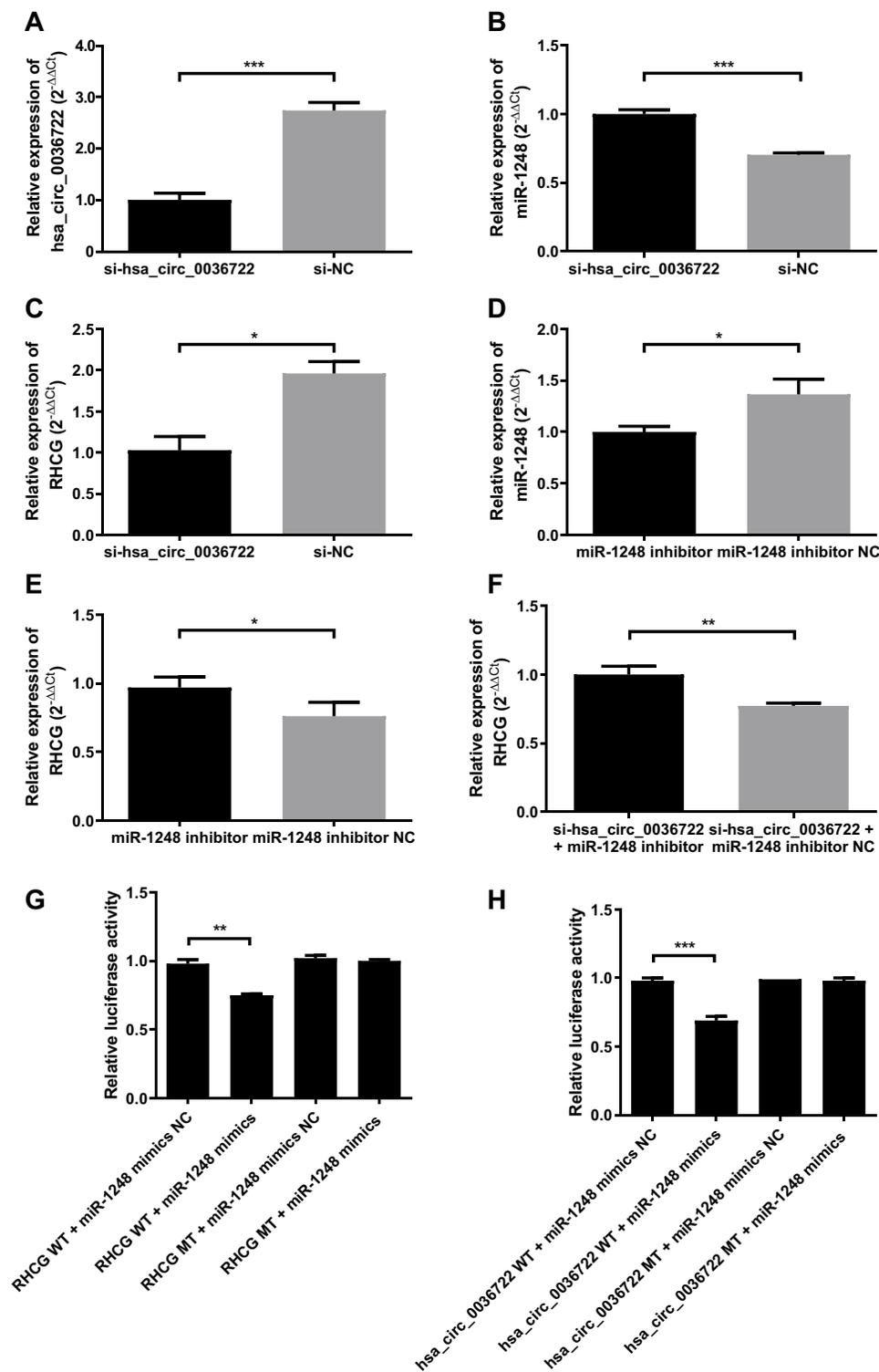
Despite tremendous research efforts and huge therapeutic advancements in LSCC, overall survival of this cancer has not been improved in the past years.<sup>34,35</sup> Considering the poor prognosis and negative impact on the quality of life of LSCC patients, researches into potential biomarkers might shed a light on the early detection and improvement of therapy for LSCC.

Although they were discovered about 40 years ago, the important roles played by circRNAs have only recently been appreciated.<sup>36,37</sup> Various circRNAs have now been validated as involved in different cancers.<sup>14,38-40</sup> Considering the stability and the spatio-temporal specificity of circRNAs, they are optimal biomarkers for cancer diagnostics.<sup>14,19,41</sup> The downregulated hsa\_circ\_0008309 and hsa\_circ\_001242 have been shown to be novel biomarkers and therapy targets in OSCC.<sup>25,26</sup> Similarly,

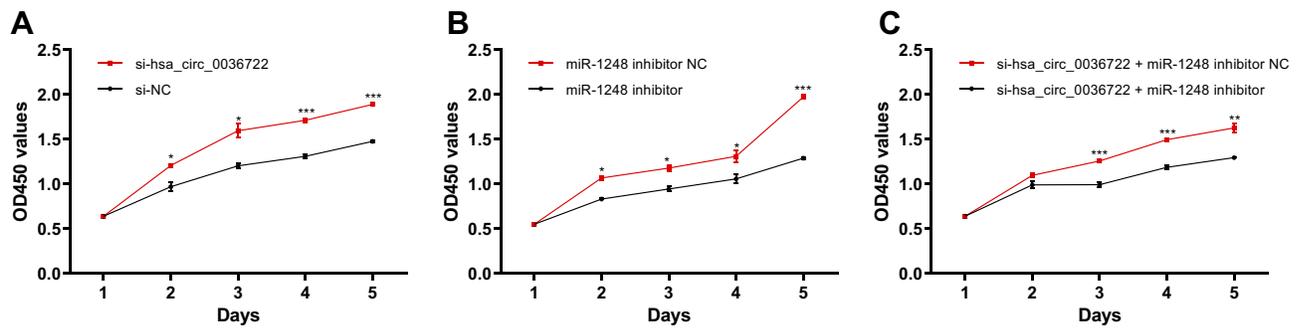
decreased expression of hsa\_circ\_0000190 and hsa\_circ\_002059 could serve as biomarkers for gastric cancer.<sup>42,43</sup> However, although a few studies have examined the dysregulation of circRNAs in LSCC,<sup>9,10</sup> research about circRNA biomarkers in LSCC remains rare.

Here, we first validated the downregulation of hsa\_circ\_0036722 in LSCC tissues compared with paired normal tissues. Examining the relationships between clinicopathological factors and the hsa\_circ\_0036722 expression levels suggested that as the differentiation levels became poorer, the expression levels of hsa\_circ\_0036722 also decreased. This suggested that hsa\_circ\_0036722 expression levels were negatively associated with LSCC differentiation. ROC curve analysis illustrated that hsa\_circ\_0036722 could serve as a diagnostic biomarker for LSCC with AUC of 0.838.

Many circRNAs were found to participate in cancer progression by affecting their parental genes.<sup>30,32,44-46</sup> We therefore wondered if the parental gene of hsa\_circ\_0036722 was dysregulated in LSCC. The qRT-PCR indicated RHCg was downregulated in LSCC. This was consistent with the role of RHCg as tumor suppressor



**Figure 7** (A) Hsa\_circ\_0036722 was downregulated in AMC-HN-8 cell line transfected with siRNA targeting its junction sequence ( $***P < 0.0001$ ). (B and C) MiR-1248 was upregulated ( $***P < 0.0001$ ) while RHCG ( $*P < 0.05$ ) was downregulated after hsa\_circ\_0036722 was downregulated in AMC-HN-8 cell line. (D) The expression of miR-1248 was decreased by miR-1248 inhibitors, and the transfection efficiency was examined by qRT-PCR ( $*P < 0.05$ ). (E) Downregulation of miR-1248 significantly increased the expression of RHCG ( $*P < 0.05$ ). (F) Co-transfection of miR-1248 inhibitors could partly reverse the RHCG decrease induced by downregulated hsa\_circ\_0036722 ( $**P < 0.01$ ). Higher values of  $2^{-\Delta\Delta C_t}$  represented relatively higher expression levels in this figure. (G and H) Luciferase reporter assay showed that ectopic expression of miR-1248 could suppress the activity of hsa\_circ\_0036722-WT ( $***P < 0.001$ ) and RHCG-WT ( $**P < 0.01$ ) reporters, respectively.



**Figure 8** (A) CCK-8 assay showed that the proliferation ability of AMC-HN-8 transfected with siRNAs targeting *hsa\_circ\_0036722* was enhanced compared to controls (\* $P < 0.05$ ; \*\*\* $P < 0.001$ ). (B) The proliferation ability of AMC-HN-8 was inhibited by miR-1248 inhibitors compared to negative controls (\* $P < 0.05$ ; \*\*\* $P < 0.001$ ). (C) Inhibition of miR-1248 rescued the enhanced proliferation ability induced by *hsa\_circ\_0036722* downregulation (\*\* $P < 0.01$ ; \*\*\* $P < 0.001$ ).

gene in another subtype of HNSCCs, tongue squamous cell carcinoma.<sup>47</sup> Similarly, the expression of RHCG was decreased in ESCC and correlated with poorer differentiation, invasion, lymph node metastasis and poorer prognosis, and RHCG could inhibit the tumorigenicity and progression of ESCC in-vitro and in-vivo via suppressing MMP1 expression.<sup>27</sup> In addition, RHCG was found to be downregulated in cervical cancer and could induce apoptosis and suppress migration ability of cervical cancer cells by lowering TGF- $\beta$ 1 expression.<sup>28</sup>

It is worth noting that the decreased expression of RHCG in LSCC was positively correlated with the expression of *hsa\_circ\_0036722*. The ROC curve analysis suggested that RHCG could distinguish LSCC from adjacent normal tissues with an AUC value of 0.808. Though this value was lower than that of *hsa\_circ\_0036722* (AUC = 0.838), the addition of RHCG to *hsa\_circ\_0036722* improved the diagnostic value, with the AUC of the combination reaching 0.859. Thus, *hsa\_circ\_0036722* could serve as diagnostic biomarker for LSCC, and could do so more efficiently if combined with RHCG.

In diverse cancers, circRNAs act as transcriptional regulators, protein decoys and templates for translation.<sup>48</sup> In addition, most circRNAs function as “miRNA sponges” by targeting downstream miRNAs, which in turn affecting their target mRNAs.<sup>38</sup> In most cases, the cognate linear mRNAs from the parental genes that produce the circRNAs are regulated at the end of circRNA/miRNA/mRNA axes.<sup>30,44,46,49-51</sup> A typical example is the anti-oncogenic circ-ITCH. In lung cancer, circ-ITCH sponges miR-7 and miR-214 to increase the expression of its parental gene, ITCH, which suppresses the progression of lung cancer via inhibiting the Wnt/ $\beta$ -catenin pathway.<sup>30</sup> This regulation mechanism also

applies to colorectal cancer.<sup>44</sup> Considering the ceRNA network theory and the positive correlation between *hsa\_circ\_0036722* expression and RHCG expression, we hypothesized that *hsa\_circ\_0036722* might partly influence the course of LSCC via regulating its anti-oncogenic parental gene RHCG through shared miRNAs.

One miRNA, miR-1248, that could bind to both *hsa\_circ\_0036722* and RHCG was predicted by all four bioinformatics databases. A recent study suggested that miR-1248 played an oncogenic role in osteosarcoma by inhibiting apoptotic protein angiotensin II type 1 receptor (AGTR1) at the post-transcriptional level and was correlated with poorer survival and chemotherapy resistance.<sup>52</sup> Furthermore, higher expression of miR-1248 was associated with BRCA1/2 mutation-associated hereditary breast cancer.<sup>53</sup> To investigate the role of miR-1248 in LSCC, the relationship of miR-1248 expression with the prognosis of HNSCC patients was explored based on a web-tool that could analyze data from 523 HNSCC patients.<sup>33</sup> The result indicated that the prognosis of HNSCC patients with higher expression of miR-1248 was poorer. Thus, miR-1248 was also an oncogene in HNSCC. This was in line with the predicted tumor suppressor roles of the decreased *hsa\_circ\_0036722* and RHCG in LSCC.

Furthermore, we conducted CCK-8 assays that demonstrated the tumor-suppressor roles of *hsa\_circ\_0036722* and oncogenic roles of miR-1248 in LSCC cell line. Downregulation of *hsa\_circ\_0036722* could increase the expression of miR-1248, while RHCG could be upregulated by miR-1248 inhibition. What's more, luciferase reporter assays illustrated the direct interactions between miR-1248 and *hsa\_circ\_0036722* as well as RHCG. Thus, we concluded

that hsa\_circ\_0036722 sponges miR-1248, inhibiting the suppression effect of miR-1248 on its parental gene, RHCG, to function as tumor suppressor in LSCC.

## Conclusion

Our study revealed that hsa\_circ\_0036722 was downregulated in LSCC compared with adjacent normal tissues and its expression levels were significantly associated with the differentiation levels of LSCC. For the first time, we demonstrated the diagnostic value of hsa\_circ\_0036722 for LSCC. In addition, its expression was associated with the expression of its parental gene, RHCG. The diagnostic value of the combination of RHCG and hsa\_circ\_0036722 was higher than that of hsa\_circ\_0036722 alone. Further function experiments indicated that hsa\_circ\_0036722 inhibit LSCC proliferation through sponging miR-1248 to regulate RHCG. Our results suggested that downregulated hsa\_circ\_0036722 could serve as a potential diagnostic biomarker for LSCC.

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## Disclosure

The authors report no conflicts of interest in this work.

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