

Immunological Response During HAART and Determinants of Current CD4⁺ T-Cell Count Among HIV/AIDS Patients Attending University of Gondar Referral Hospital, Northwest Ethiopia

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Purpose: After the initiation of highly active antiretroviral therapy (HAART), successful HAART is characterized by an increase in the CD4⁺ count. Several factors affect the CD4⁺ T-cell count. This study aimed to assess the immunological response during HAART and determinants of the current CD4⁺ T-cell count among HIV/AIDS patients on HAART.

Patients and Methods: A hospital-based cross-sectional study was conducted from February 1 to April 1, 2017. A total of 423 HIV/AIDS patients on HAART were enrolled using simple random sampling. Descriptive statistics, and bivariate and multiple regression analyses were conducted. Variables with p -value <0.2 in the bivariate analysis were entered in the multiple regression models. p -Values <0.05 and 95% confidence intervals were used to identify determinants of the current CD4⁺ T-cell count.

Results: The mean CD4⁺ T-cell count gradually increased until 8 years on HAART but declined thereafter. An increased current CD4⁺ T-cell count was observed among patients with an initial regimen of pediatric d4T-3TC-NVP [$\beta=185.5$, 95% CI (8.8, 362.2)] ($p=0.040$), with increased baseline CD4⁺ T-cell count [$\beta=0.468$, 95% CI (0.342, 0.594)] ($p<0.0001$), and with long duration on HAART [$\beta=18.0$, 95% CI (9.9, 26.1)] ($p<0.0001$), whereas a decreased level of current CD4⁺ T-cell count was observed among males [$\beta=-72.7$, 95% CI (-114.5, -30.9)] ($p<0.0001$) and those with poor baseline adherence [$\beta=-108.9$, 95% CI (-210.9, -7.0)] ($p=0.036$) and viral load >1000 copies [$\beta=-189.2$, 95% CI (-243.5, -134.9)] ($p<0.0001$).

Conclusion: The trend in immunological response was not increased linearly throughout the HAART duration. Sex, type of initial regimen, baseline adherence, baseline CD4⁺ count, viral load, and duration on HAART were independent determinants of current CD4⁺ count. These determinants could be addressed by regular monitoring of HIV patients on HAART, and special attention should be paid to male patients.

Keywords: immunological responses, trends, current CD4⁺ count, HIV/AIDS, HAART

Introduction

Highly active antiretroviral therapy (HAART) helps to reduce the morbidity and mortality rates associated with human immunodeficiency virus (HIV) infection by decreasing the viral ribonucleic acid (RNA) concentration below detection levels and allowing immune restoration.^{1,2} The CD4⁺ T-cell count has been used to assess the urgency of HAART initiation and the need for opportunistic infection prophylaxis, and to provide information regarding the magnitude of immune reconstitution.³ Different

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studies have shown that the immunological response (measured by CD4⁺ T-cell count) increases as the duration on HAART increases.^{4–6} After the initiation of HAART, a successful HAART is characterized by a quicker response during the first 3 months, which gradually increases as the duration on HAART increases.^{7,8}

The CD4⁺ T-cell count and HIV viral load are routinely used prognostic tests for patients on HAART. Studies have shown that these two prognostic tests have an inverse relationship. After the initiation of HAART drugs, the viral load increases.^{9,10} In addition to viral load, the baseline CD4⁺ T-cell count also determines the immunological decrease in CD4⁺ cell response. There are controversial findings related to the correlation between the baseline CD4⁺ T-cell count and the current CD4⁺ T-cell count. Some studies show a positive correlation,^{11,12} whereas other studies indicate a negative correlation.^{13–15} Furthermore, sex,^{13,16,17} baseline adherence,^{18,19} duration on HAART,^{4,5} advanced WHO clinical stage, and treatment interruption also determine the level of current CD4⁺ T-cell count.¹²

Trends in the immunological response and the factors contributing to the current CD4⁺ T-cell count vary from one setting to another. Studies on the trends in immunological response and determinants of current CD4⁺ T-cell count are scarce in Ethiopia. Most available studies in Ethiopia are focused on immunological failure, which is defined as the decline in CD4 cells by 50% from their peak value, or persistently <100 cells/mm³, or a fall in CD4 counts below the baseline count.^{11–13,20–22} To the investigators' knowledge, no studies have been conducted on the trends in immunological response and determinants of current CD4⁺ T-cell count using more advanced regression analysis in this study area. Furthermore, most of the previous studies on immunological response in Ethiopia did not control for the confounding effect of viral load, as viral load testing was only introduced in Ethiopia after the end of 2016. Therefore, this study aimed to assess the trends in immunological response and determinants of the current CD4⁺ T-cell count using a more advanced regression analysis and controlling the confounding effects of viral load. Up-to-date information on immunological response and determinants of the current CD4⁺ T-cell count is important for understanding the immunological response to treatment outcome, regimen change, and patient management. This may, in turn, provide valuable inputs to achieve the ambitious 90–90–90 plan to end the AIDS epidemic by 2030.²³

Patients and Methods

Study Area and Period

The study was conducted at the University of Gondar Referral Hospital (UOGRH) from February 1 to April 1, 2017. The hospital is located 747 km from the capital city of Ethiopia, Addis Ababa. According to the recent administration, the town has 12 sub-cities, which consist of 21 kebeles (the smallest administrative unit in Ethiopia). According to the 2007 central statistical agency report, Gondar town has an estimated population of more than 206,987.²⁴ The town has one teaching referral hospital, eight health centers, and 15 private clinics. At the time of the study, there were a total of 13,753 HIV patients; of these, 5389 patients were on HAART.

We followed the methods of Ayele et al for the identification of eligible study participants, data collection, specimen collection, and analysis.²⁵

Study Design and Study Population

A hospital-based cross-sectional study design supported by a retrospective record review was conducted among adult HIV/AIDS patients on HAART.

Inclusion and Exclusion Criteria

All adult HIV patients on HAART whose age was greater than 18 years old, had a baseline CD4⁺ T-cell count, and had at least 6 months' follow-up on HAART were included in the study. Those patients who had been seriously sick and were unable to respond and give blood specimens, or who had incomplete secondary data records, especially for laboratory and clinical data such as baseline CD4⁺ T-cell count, were excluded from the study.

Operational Definitions

In the current study, “baseline data” refers to the data before the initiation of HAART and “current CD4⁺ T-cell count” refers to the CD4⁺ T-cell count during the data collection time. Adherence was calculated using $\frac{\text{No of dose of HAART taken}}{\text{No of prescribed doses of HAART}} \times 100\%$. We considered good adherence if >95%, fair adherence if 85–95%, and poor adherence if <85% of doses were taken.^{26,27}

Study Variables

In this study, the level of current CD4⁺ T-cell count was considered as a dependent variable and treated as a continuous variable. The independent variables included age, marital status, sex, residence, religion, education,

occupation, WHO clinical stage, HIV/AIDS co-infection, baseline CD4⁺ T-cell count, regimen type, duration on HAART, viral load, and adherence.

Sample Size Determination and Sampling Techniques

A single population proportion formula ($n = (Z\alpha/2)^2 p(1-p)/d^2$) was used to estimate the sample size. The following assumptions were considered when calculating the sample size: margin of error $d=0.05$, $Z\alpha/2=1.96$, P =population proportion (estimated prevalence)=0.5, and 95% confidence interval. After adding a 10% non-response rate, the final sample size was 423 participants. A systematic random sampling technique was used to select study participants. There was an average of 20 HIV/AIDS patients per day under follow-up in the antiretroviral therapy (ART) clinic

who gave a blood sample for viral load and CD4⁺ T-cell count concurrently. During the 3-month data collection period, 1320 HIV/AIDS patients were expected to visit the hospital for viral load and CD4⁺ T-cell count follow-up. The sampling interval (K) value was calculated by dividing the total number of HIV/AIDS patients during our study period by the sample size ($1320/423=3.12=3$). To determine the first participant, the lottery method was used among the first three patients and every third interval participant was selected (Figure 1).

Data Collection and Laboratory Methods

Sociodemographic characteristics, including age, gender, occupation, residence, marital status, religion, and educational status, were collected from each study participant using a structured questionnaire developed from the

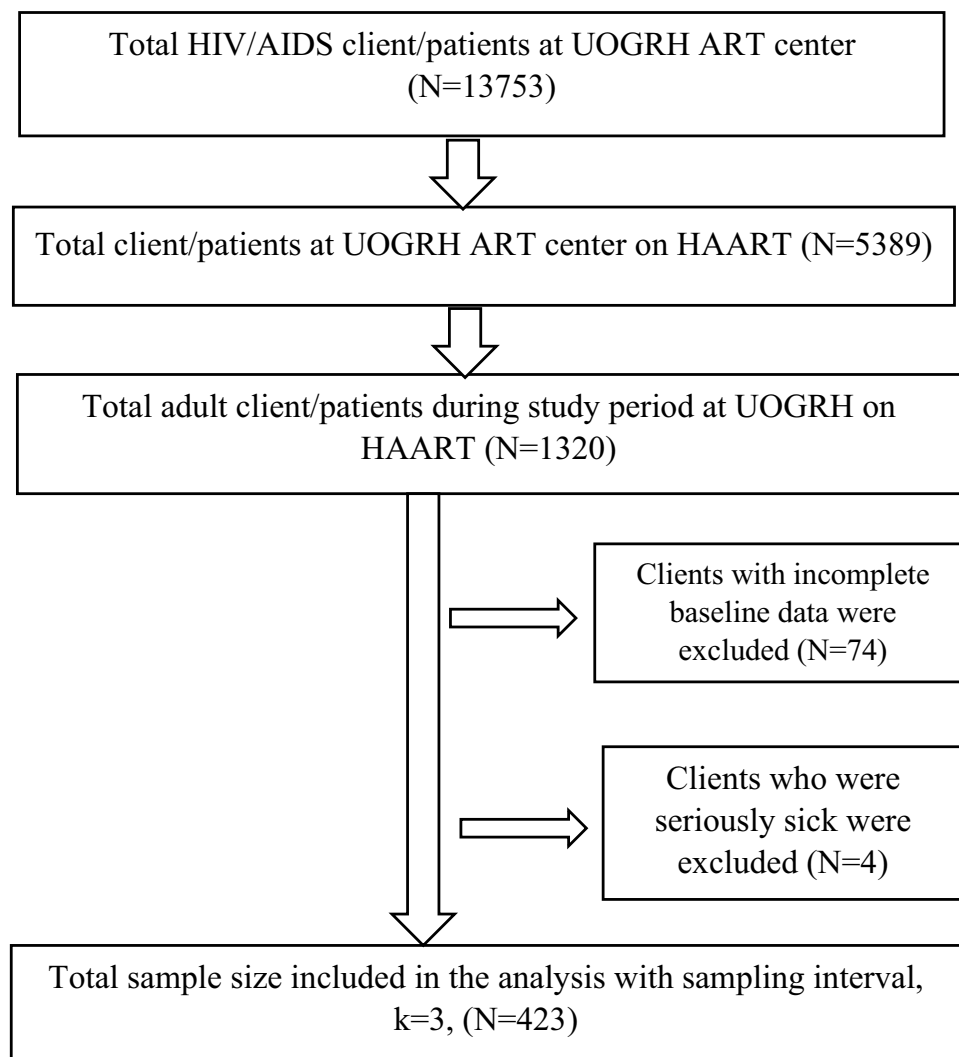


Figure 1 Schematic representation of the sampling procedure of HIV-positive adults on HAART at University of Gondar Referral Hospital from February to April 2017.

literature.^{21,28} Relevant clinical characteristics of participants (weight, WHO clinical stage, CD4⁺ T-cell count, HAART regimen, adherence with and duration on HAART, initial regimen, types of co-infections, adverse effects, and reason for drug switching) were retrieved from medical records by trained nurses using a data extraction checklist (S1 file).

For CD4⁺ T-cell count and viral load tests, 3–5 mL of whole blood was drawn from each participant using Vacutainer tubes, separately in two tubes containing ethylenediamine tetra-acetic acid (EDTA) following standard blood collection procedures. After centrifugation (3000 rpm for 20 minutes), plasma was separated and aliquots were prepared for viral load testing. Collected specimens were labeled and immediately transported to the Gondar Hospital laboratory. The CD4⁺ T-cell count specimens were stored at room temperature and processed within 24 hours. For viral load testing, plasma was separated within 5 hours and two aliquots of cryo-vials (with a capacity of 1 mL each) were prepared for transportation. The specimens were transported on dry ice and stored at –80°C until processing. Centrifugation, pipetting, and aliquoting were performed following standard protocol and laboratory biosafety precautions at both the collection and testing sites.

Quantification of absolute CD4⁺ T-cell count on whole blood specimens was performed using the BD FACSCalibur flow cytometry system (BD, CA, USA) following the standard operational procedures in the laboratory. The CD4⁺ T-cell count was carried out by adding 50 µL whole blood to a reagent tube containing 20 µL of monoclonal antibodies, followed by vortexing and incubation for 30 minutes under dark conditions. Plasma viral load was measured using a quantitative real-time PCR HIV-1 assay, with the COBAS[®] AmpliPrep instrument (Roche, Homburg, Germany). Plasma was prepared from 5 mL of blood by centrifugation at 3000 rpm for 20 minutes.

Data Analysis

The data were entered, cleaned, checked for completeness, and analyzed using SPSS version 21. Descriptive statistics, and bivariate and multiple regression analyses were conducted. Before applying the multiple regression models, all the assumptions were assessed. Variables with *p*-value <0.2 in the bivariate analysis were entered in the multiple regression model. *p*-Values <0.05 and 95% confidence intervals were used to determine the independent factors associated with the current CD4⁺ T-cell count.

Table 1 Sociodemographic Characteristics of HIV/AIDS Patients on HAART in University of Gondar Referral Hospital, 2017

Variable		Frequency	%
Mean±SD age (years)	39±9.8		
Marital status	Single	97	22.9
	Married	209	49.4
	Divorced	81	19.1
	Widowed	36	8.5
Gender	Female	272	64.3
	Male	151	35.7
Residence	Urban	343	81.1
	Rural	80	18.9
Occupation	Farmer	51	12.1
	Merchant	58	13.7
	Student	17	4
	Government employee	43	10.2
	Daily laborer	62	14.7
	Housewife	101	23.9
	Private employee	66	15.6
	Other	25	5.9
Educational status	Illiterate	121	28.6
	Primary school	117	27.7
	Secondary school	142	33.6
	Tertiary	43	10.2
Religion	Orthodox Christian	386	91.3
	Muslim	30	7.1
	Protestant	7	1.7
	Total	423	100.0

Results

Sociodemographic Characteristics of Participants

A total of 423 HIV/AIDS participants who received HAART were enrolled in the study. Of these, 272 (64.3%) were female and 151 (35.7%) were male. The mean±SD age of the participants was 39±9.8 years (range 18–78 years). The mean weight of participants at the baseline and at the time of the study (current) was 50 kg and 56 kg, respectively. At the time of the study, almost half of the participants, 209 (49.4%), were married, 343 (81.1%) were urban inhabitants, and 386 (91.6%) were Orthodox Christians (Table 1).

Clinical Characteristics of Participants

The study patients were on HAART for a minimum of 6 months up to 12 years, with an average duration of 7(3 years. At baseline, most of the participants, 269

(63.6%), had WHO clinical stage III and IV disease, 267 (63.1%) had a CD4⁺ T-cell count <200 cells/mm³, 408 (96.5%) had good adherence, 156 (36.9%) were on a regimen of AZT+3TC+NVP, and 87 (20.6%) were on TDF+3TC+EFV. At the time of data collection, 420 (99.1%) had WHO clinical stage I and II disease, 44 (10.4%) had a CD4⁺ T-cell count <200 cells/mm³, 420 (99.3%) had good adherence, and 162 (38.2%) had switched, to either a first-line regimen, 150 (35.5%), or a second-line regimen, 12 (2.8%) (Table 2).

Trends in Immunological Response

The mean CD4⁺ T-cell count gradually increased from baseline to 8 years on HAART (from 184 to 493 cells/mm³). However, the mean CD4⁺ T-cell count slightly declined 8 years after HAART initiation (464 cells/mm³) (Figure 2).

Correlation Between Current Virological and Immunological Status

Using paired sample *t*-test analysis, baseline CD4⁺ T-cell count and CD4⁺ T-cell count at the time of data collection were positively correlated ($r=0.285$, $p<0.0001$) (Figure 3). Furthermore, a Pearson's correlation revealed that there was weak negative correlation between current CD4⁺ T-cell count and viral load ($r=-0.243$, $p<0.0001$) (Figure 4).

Bivariate Regression Analysis Results

Without controlling potential confounding, sex, occupation, type of initial HAART regimen, duration on HAART, baseline adherence, baseline CD4⁺ T-cell count and viral load, and duration on HAART were significantly associated with current CD4⁺ T-cell count ($p<0.05$). However, age, marital status, residence, educational status, baseline WHO stage, opportunistic infections, switching of drug regimens, current adherence, baseline weight, current weight, and type of second regimens were not significantly associated with the current CD4⁺ T-cell count ($p>0.05$). Variables with p -value <0.2 were included in the multiple regression models.

Multiple Regression Analysis Results

The multiple regression analysis showed that 32.8% of the variation in CD4⁺ T-cell count was explained by our model ($R^2=0.328$, $p<0.0001$). The assumption of a linear relationship between the continuous independent variables and current CD4⁺ T-cell count was met. Analysis of collinearity statistics showed that there were no variables with a variance inflation factor of greater than 10 and tolerance scores above

Table 2 Clinical Characteristics of HIV/AIDS Patients on HAART in University of Gondar Referral Hospital 2017

Variable		Frequency	%
Duration of HAART (years)	≤6	162	38.3
	>6	261	61.7
Mean±SD HAART duration (years)	7±3)		
Baseline WHO stage	I	57	13.5
	II	97	22.9
	III	214	50.6
	IV	55	13.0
WHO stage during data collection	I	11	2.6
	II	408	96.5
	III	4	0.9
Initial regimen	d4T+3TC+NVP	78	18.4
	d4T+3TC+EFV	29	6.9
	AZT+3TC+NVP	156	36.9
	AZT+3TC+EFV	30	7.1
	TDF+3TC+EFV	87	20.6
	TDF+3TC+NVP	31	7.3
	Pediatric (d4T+3TC+NVP)	7	1.7
	Pediatric 4C (AZT+3TC+NVP)	5	1.2
Switching	No	261	61.7
	Yes	162	38.3
Switching drug type	To 1st line drug	150	92.6
	To 2nd line drug	12	7.4
Second regimen	AZT+3TC+NVP	61	37.7
	AZT+3TC+EFV	25	15.4
	TDF+3TC+NVP	25	15.4
	TDF+3TC+EFV	39	24.1
	ABC+ddl+LPV/R	11	6.8
	TDF+ddl+IPV/R	1	0.6
Reasons for switching drug	Toxicity	109	67.3
	Pregnancy	7	4.3
	TB	18	11.1
	Clinical failure	1	0.6
	Immunological failure	12	7.4
	Default	6	3.7
	Age	9	5.6
ARV drug adherence at baseline	Good	408	96.5
	Fair	2	0.5
	Poor	13	3.1
ARV drug adherence during data collection	Good	420	99.3
	Poor	3	0.7

(Continued)

Table 2 (Continued).

Variable		Frequency	%
Baseline CD4 ⁺ T-cell count	≤199	267	63.1
	200–349	120	28.4
	350–499	27	6.4
	≥500	9	2.1
CD4 ⁺ T-cell count during data collection	≤199	44	10.4
	200–349	110	26.0
	350–499	114	27.0
	≥500	155	36.6
Viral load count (copies/mm ³)	Undetected	224	53.0
	0–19	84	19.9
	20–999	53	12.5
	≥1000	62	14.7

Abbreviations: ARV, antiretroviral; d4T, stavudine; 3TC, lamivudine; NVP, nevirapine; EFV, efavirenz; AZT, zidovudine; TDF, tenofovir disoproxil fumarate; ABC, abacavir; ddl, didanosine; LPV/R, lopinavir/ritonavir.

0.2. The Durbin–Watson statistic showed that the values of the residuals were independent (Durbin–Watson=1.81). The plot of standardized residuals versus standardized predicted values showed no obvious signs of funneling, suggesting that the assumption of homoscedasticity had been met. In addition, the normal P-P plot suggested that the assumption of values of the residuals being normally distributed may have been violated. However, the deviations from normality were not extreme and are not likely to have a significant impact on

our findings. Therefore, the results are still valid. Finally, Cook's distance values were all under 1, suggesting that individual cases were not improperly influencing the model. After applying the multiple regression models, occupation and type of second-line HAART regimens were not significantly associated with the current CD4⁺ T-cell count. The other variables (sex, type of initial HAART regimen, duration on HAART, baseline adherence, baseline CD4⁺ T-cell count, and viral load) remained significantly associated with current CD4⁺ T-cell count.

Keeping other independent variables constant, for each cell/mm³ of blood increment in the baseline CD4⁺ T-cell count, the current CD4⁺ T-cell count increased by 0.468 cells/mm³ of blood ($p<0.0001$). For every additional one-year stay on HAART, the current CD4⁺ T-cell count increased by 18.02 cells/mm³ of blood ($p<0.0001$). Males' current CD4⁺ T-cell count was lower than females', by 72.73 cells/mm³ of blood ($p<0.0001$). Similarly, the current CD4⁺ T-cell count of HIV patients who took the initial HAART regimen type pediatric d4T-3TC-NVP was higher by 185.5 cells/mm³ of blood compared to others ($p=0.040$). Furthermore, the current CD4⁺ T-cell count of HIV patients with poor adherence was less than those with good adherence by 108.93 cells/mm³ of blood ($p=0.036$). Lastly, current CD4⁺ T-cell count among HIV patients with ≥ 1000 copies/mm³ of viral load was lower than in those with an undetectable viral load by 189.17 copies/mm³ ($p<0.0001$) (Table 3).

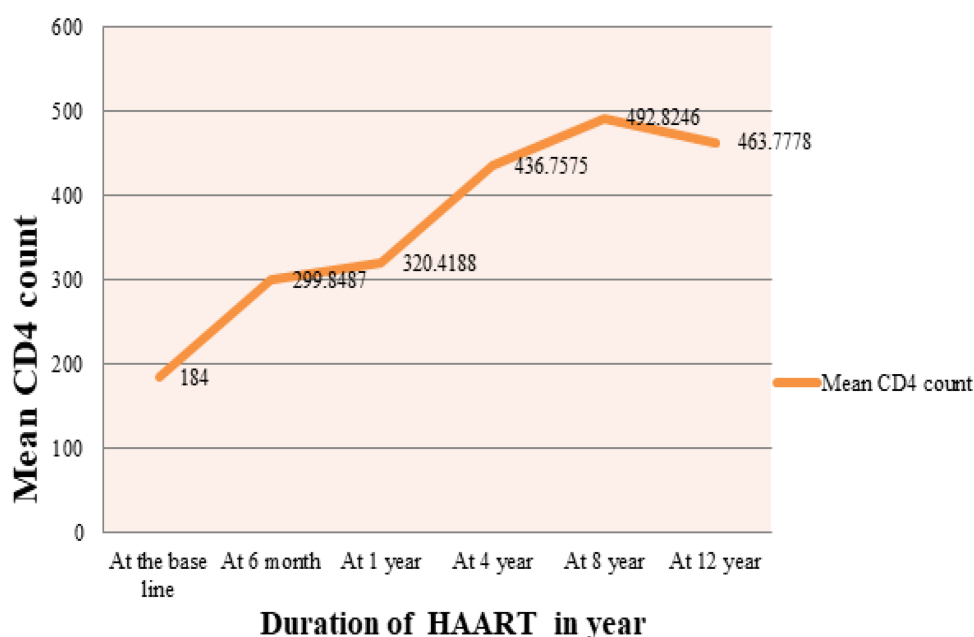


Figure 2 Trend in CD4⁺ T-cell count at baseline, 6 months, and 1, 4, 8, and 12 years in HIV/AIDS patients on HAART at the University of Gondar Referral Hospital 2017.

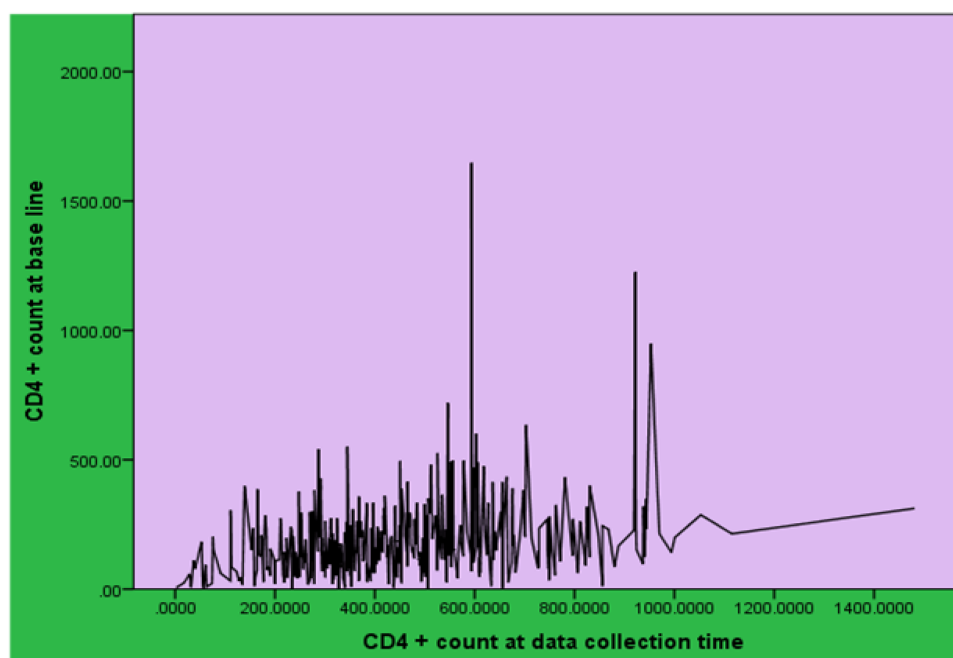


Figure 3 Correlation between baseline CD4⁺ T-cell count and current CD4⁺ T-cell count in HIV/AIDS patients on HAART at the University of Gondar Referral Hospital 2017.

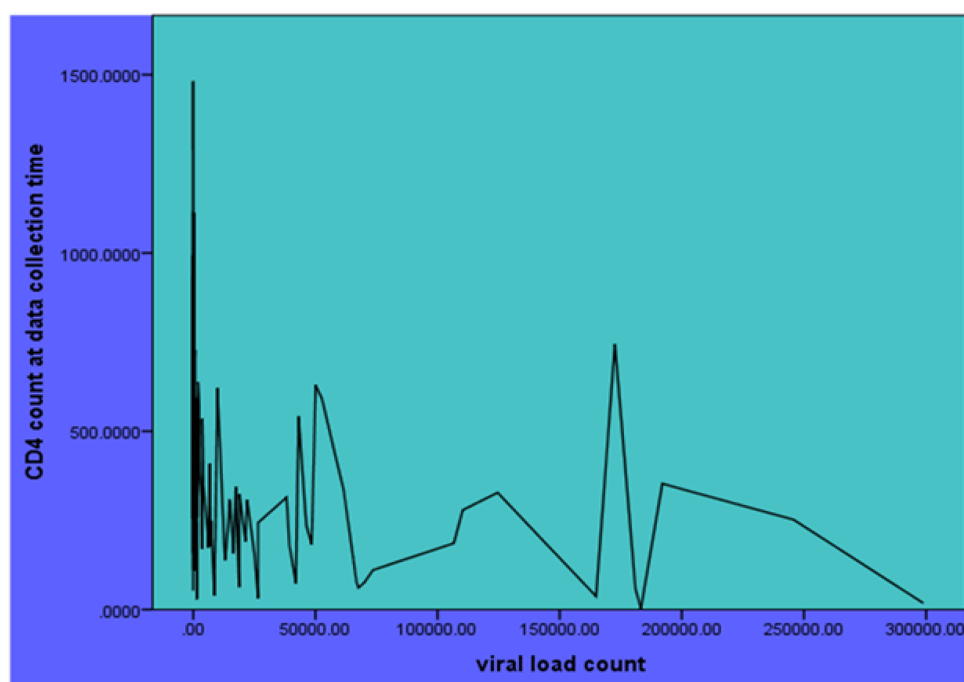


Figure 4 Correlation between current CD4⁺ T-cell count and viral load in HIV/AIDS patients on HAART at the University of Gondar Referral Hospital 2017.

Discussion

In this study, the trend in the immunological response comprised an increase until 8 years on HAART and then a slight decrease after 8 years. The trends in the immunological response observed in this study were similar to

other study findings in Ethiopia^{22,29} and Ghana.³⁰ The large increases in the CD4⁺ T-cell count until 8 years on HAART may be due to immune reconstitution and redistribution of CD4 T-cells that are sequestered from the lymphoid tissues into the circulating blood. The reduction

Table 3 Bivariate and Multiple Regression Analysis Results of HIV/AIDS Patients on HAART Attending University of Gondar Referral Hospital, Northwest Ethiopia, 2017

Variable	No (%)	Bivariate (Simple Regression Analysis)			Multiple Regression Analysis			
		Unstandardized β Coefficient	95% CI for β	p-Value	Unstandardized β Coefficient	Standardized β Coefficient	95% CI for β	p-Value
Sex								
Female	272 (64.3)	R			R			
Male	151 (35.7)	-66.2	-108.7, -23.7	0.002	-72.7	-0.2	-114.5, -30.9	0.001*
Occupation								
Farmer	51 (12.1)	4.9	-68.9, -78.7		33.2	0.05	-32.6, 99.1	0.321
Merchant	58 (13.7)	-29.3	-100.2, -41.6		-24.6	-0.04	-86.9, 37.7	0.439
Student	17 (4.0)	25.9	-85.6, 137.4		0.53	0.0001	-103.4, 104.5	0.992
Government employee	43 (10.2)	-82.4	-160.5, -4.3	0.039	-46.6	-0.07	-115.6, 22.4	0.185
Daily laborer	62 (14.7)	-16.0	-85.5, 53.9		19.8	0.033	-40.9, 80.6	0.521
Housewife	101 (23.9)	17.6	-43.4, 78.6		19.8	-0.009	-58.4, 49.7	0.874
Private employee	66 (15.6)	R			R			
Type of initial regimen								
d4T-3TC-NVP	78 (18.4)	20.276	-37.8, 78.3		-28.0	-0.05	-117.4, 61.4	0.538
d4T-3TC-EFV	29 (6.9)	108.6	23.9, 193.3	0.012	64.6	0.08	-40.6, 169.8	0.228
AZT-3TC-NVP	156 (36.9)	R			R			
AZT-3TC-EFV	30 (7.1)	107.5	24.0, 191.0	0.012	71.8	0.09	-12.2, 155.9	0.094
TDF-3TC-EFV	87 (20.6)	-9.4	-65.4, 46.6		-18.7	-0.04	-71.9, 34.4	0.489
TDF-3TC-NVP	31 (7.3)	-20.4	-102.8, 61.9		-21.5	-0.03	-100.8, 57.7	0.594
Pediatric d4T-3TC-NVP	7 (1.7)	77.6	-84.2, 239.4		185.5	0.11	8.8, 362.2	0.040*
AZT-3TC-NVP	5 (1.2)	-62.3	-252.6, 128.0		-135.2	-0.07	-324.6, 54.1	0.161
Baseline adherence								
Good	407 (96.2)	R			R			
Fair	2 (0.5)	-332.7	-630.9, -34.6	0.375	-117.8	-0.04	-378.7, 143.0	0.375
Poor	14 (3.3)	-80.2	-194.6, 34.1	0.036	-108.9	-0.09	-210.9, -6.9	0.036*
Baseline CD4 ⁺ T-cell count								
		0.404	0.273, 0.5	0.000	0.468	0.33	0.342, 0.594	0.00*
Viral load								
Undetectable	224 (53.0)	R			R			
1–19 copies/mm ³	84 (19.9)	-25.7	-76.703, 25.2		2.5	0.005	-44.2, 49.3	0.914
20–999 copies/mm ³	53 (12.5)	5.5	-56.1, 67.0		40.9	0.063	-16.2, 98.1	0.160
≥1000 copies/mm ³	62 (14.7)	-206.9	-264.3, -149.6	0.000	-189.2	-0.311	-243.5, -134.9	0.000*
Duration on HAART		14.0	7.3, 20.7	0.000	18.0	0.252	9.9, 26.1	0.000*

Note: *Significant ($p < 0.05$).

Abbreviation: R, reference category.

in CD4⁺ T-cell count after 8 years on HAART in our study may be due to two factors: 1) natural aging leads to a long-term decline in the immune system (CD4⁺ T-cell count through low-grade chronic immune activation); and 2) long-term HIV infection despite viral suppression from combined antiretroviral therapy (cART) may accelerate the body's natural aging trajectory, primarily

as a result of persistent immune activation and cART-induced pro-inflammatory effects leading to premature immunosenescence.³¹

In this study, there was a positive correlation between baseline and current CD4⁺ T-cell count. This result was in line with research conducted in southern India.^{32–34} The lower the CD4⁺ T-cell count at baseline, the less probable

it is that the patient will normalize and convalesce completely,^{35,37} and the greater the CD4⁺ T-cell count at baseline, the more possible it is for them to normalize and convalesce completely. For instance, many individuals who initiate treatment with a CD4⁺ T-cell count <350 cells/mm³ never attain a CD4⁺ T-cell count of >500 cells/mm³, even with up to 10 years on HAART.^{38,39} Some studies showed that HIV patients who started HAART with a lower CD4⁺ T-cell count have a shorter life expectancy than those starting at higher CD4⁺ T-cell counts.^{38–40} As expected, a negative correlation between the current CD4⁺ T-cell count and viral load was observed. This agrees with studies conducted in Rwanda and the USA.^{41–43}

The reconstitution of the immune system through viral suppression and increased CD4⁺ T-cell count is the goal of HAART. However, attaining the target immune response to HAART is affected by several factors. In the current study, sex was an independent predictor of the current CD4⁺ T-cell count. Compared with female HIV patients, male HIV patients on HAART showed a decreased current CD4⁺ count. This result is in line with study findings in Nigeria,¹⁷ Northern Ethiopia,¹³ and China,¹⁶ and in a multi-center study conducted in low-income countries.⁴⁴ The increased CD4⁺ T-cell count among females could be explained by the fact that females may have more access to expanded routine HIV testing in antenatal care during pregnancy and thus they may be linked to ART facilities early on. Females are more likely to be diagnosed with HIV infection earlier than males.^{45,46} Early diagnoses may lead to early initiation of HAART. This may, in turn, improve the immunological response.⁴⁷ On the other hand, health-seeking behavior of males is poor, which may result in lower rates of screening for HIV, lower acceptance of screening results, and less linkage to ART facilities after a HIV-positive diagnosis.^{48,49} A delay in the diagnosis of HIV and failure to initiate HAART early favor a poor response to HAART.⁵⁰

In the current study, poor baseline adherence to HAART was significantly associated with a decreased current CD4⁺ T-cell count. Study findings in resource-limited settings¹⁸ and a study conducted in Uganda and Zimbabwe¹⁹ showed that poor first-year adherence to HAART exposes the patient to a higher risk of immunological failure.

In addition to poor baseline adherence, baseline CD4⁺ T-cell count was found to be a significant predictor of current

CD4⁺ T-cell count. This finding is in line with previous studies conducted in Ethiopia, where HAART initiated at higher CD4⁺ T-cell count had positive effects on the immunological response.^{11,12,20} Furthermore, in this study, the current CD4⁺ T-cell count was increased as the duration of HAART increased. Similar results were reported from studies in Cameroon and China, in which the CD4⁺ T-cell count continually increased along with HAART duration, even a long time after HAART initiation.^{4,5}

Finally, the current study indicates that patients with an initial regimen of first-line drugs, specifically d4T-3TC-NVP, showed a significantly increased current CD4⁺ T-cell count compared to those patients who started with an initial regimen of TDF-3TC-EFV. d4T-3TC-NVP used to be given to children before this combination was banned. In the current study, the age of study participants who were started with the initial regimen d4T-3TC-NVP was <18 years. It is known that children have a high baseline CD4⁺ T-cell count physiologically. Therefore, an increased current CD4⁺ T-cell count among patients with initial regimen of d4T-3TC-NVP may be due to those participants having been children at the baseline. However, a further study is needed to ascertain this claim.

This study has some limitations. First, baseline values for viral load were not tested because of the unavailability of viral load testing facilities in Ethiopia before 2017. Second, we could not control the confounding effect of drug resistance on HIV strains because of the unavailability of drug resistance testing facilities. In addition, the impact of survival bias was not assessed. As this study was conducted in only one referral hospital, it may not reflect the trends in immunological response and determinants of current CD4⁺ T-cell count in the wider community. Despite these limitations, the findings of this study may provide insights into the trends in the immunological response after initiation of HAART and determinants of the current CD4⁺ T-cell count.

Conclusions

The trend in immunological response did not increase linearly throughout the HAART duration. Sex, type of initial regimen, baseline adherence, baseline CD4⁺ T-cell count, viral load, and duration on HAART were independent determinants of current CD4⁺ T-cell count. These determinants could be addressed by regular monitoring of HIV patients on HAART using both virological and immunological tests. During monitoring, special attention needs to be paid to males. Further studies need to be

conducted in a wider community and at multiple ART centers to determine whether there are differences in immunological response and associated factors of current CD4⁺ T-cell count in different set-ups.

Abbreviations

ABC, abacavir; AIDS, acquired immune deficiency syndrome; ART, antiretroviral therapy; ARV, antiretroviral; AZT, zidovudine; CD4, cluster of differentiation; d4T, stavudine; ddl, didanosine; EFV, efavirenz; HAART, highly active antiretroviral therapy; HIV, human immunodeficiency virus; LPV/R, lopinavir/ritonavir; NVP, nevirapine; RNA, ribonucleic acid; 3TC, lamivudine; TDF, tenofovir disoproxil fumarate; UOGRH, University of Gondar Referral Hospital.

Data Sharing Statement

All the data (SPSS) sets generated and/or analyzed in this study are available from the corresponding author on reasonable request (ayeale.gizachew@yahoo.com).

Ethics Approval and Consent to Participate

This study was conducted in accordance with the Declaration of Helsinki after ethical clearance was obtained from the University of Gondar, School of Biomedical and Laboratory Sciences Ethical Review Committee, and an official letter was submitted to University of Gondar Referral Hospital administration prior to data collection with Ref. No. SBMLS/678/09. Written informed consent was obtained from each study participant after explaining the purpose and objective of the study. Patients who were not willing to participate in the study were not forced to participate. All the data and samples obtained from them were kept confidential by using codes instead of any personal identifiers and meant only for the purpose of the study. The laboratory results from the study participants were communicated to their physicians for appropriate management.

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Author Contributions

All authors contributed to data analysis, drafting or revising the article, gave final approval of the version to be published, and agree to be accountable for all aspects of the work.

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Disclosure

The authors declare that they do not have any conflicts of interest.

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