

Upregulation of miR-137 Expression Suppresses Tumor Growth and Progression via Interacting with DNMT3a Through Inhibiting the PTEN/Akt Signaling in HCC

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Background: Downregulation of miR-137 regulates tumor growth in hepatocellular carcinoma (HCC). Yet, the underlying molecular mechanisms stay unclear.

Materials and Methods: miR-137 and DNA methyltransferase 3a (DNMT3a) expression levels were detected by Western blot, immunohistochemistry and qRT-PCR assays. Luciferase reporter and Western blot assays were also carried out to explore the correlation of miR-137 and DNMT3a. Flow cytometry assay, MTT analysis, transwell and wound healing assay were used to evaluate cell apoptosis, proliferation, as well as invasive and migratory abilities. Western blot was used to examine the caspase-3, cleaved caspase-3, PCNA, MMP-2, and MMP-7 protein levels, as well as PTEN/Akt signaling alternations. Methylation-specific PCR was applied to detect the PTEN promoter methylation status. Xenograft tumor assay, Western blot and immunohistochemistry analyses were taken to confirm the miR-137 regulation in vivo.

Results: Downregulation of miR-137, upregulation of DNMT3a, as well as an inverse correlation between them were observed in HCC clinical samples and cells. Moreover, miR-137 targeted directly and inhibited DNMT3a in HCC cells, which further retarded cell proliferative, migratory and invasive capabilities, while promoted apoptotic ones. Additionally, miR-137 overexpression inactivated the PTEN/Akt pathway in HCC cell by decreasing DNMT3a expression. Furthermore, miR-137 overexpression inhibited tumor growth in vivo in HCC via interacting with DNMT3a through inhibiting the PTEN/Akt cascades.

Conclusion: Our findings suggested that miR-137 inhibited HCC tumor growth and progression via interacting with DNMT3a and suppressing the PTEN/Akt signaling in vitro and in vivo.

Keywords: miR-137, DNMT3a, DNA methylation, hepatocellular carcinoma, the PTEN/Akt pathway

Introduction

Hepatocellular carcinoma (HCC), belonging to the commonest diagnosed carcinomas, makes up the second highest carcinoma-associated deaths with rapidly-increasing incidence and postoperative recurrence worldwide.¹ According to cancer statistical data in 2017, there were approximately 40,710 cases and 28,920 deaths due to HCC in the USA.¹ To date, hepatic transplantation, resection, and chemotherapy are common traditional methods for the treatment of early-stage HCC.² However, the advanced stage cases were counted most among the diagnosed ones due to the deficiency of distinct HCC biomarkers.³ In spite of the recent advances in

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fighting against HCC, the clinical outcomes remain unfavorable with only 15% 5-year overall survival rate.⁴ Hence, crystalizing the regulation and exploring novel effective treatments for HCC are in great need. microRNAs (miRNAs), small single-stranded non-coding RNAs, post-transcriptionally regulate the gene expression.⁵ Increasing experimental data have reported the association between miRNAs and pathological cellular processes (ie, proliferation, growth, cell cycle, apoptosis, and metastasis).⁶ It has been implicated that dysregulation of miRNAs is frequent in diverse carcinomas including HCC.⁷ Moreover, miRNAs are well documented to be closely implicated in either promoting or suppressing HCC carcinogenesis.⁸ Among these miRNAs, miR-137 has been confirmed to act as a tumor suppressor and be underexpressed in various carcinomas.^{9–11} Furthermore, studies also have implicated that the downregulation of miR-137 has a regulatory effect on HCC tumorigenesis.¹² However, the detailed mechanisms are not well elucidated.

Herein, our results confirmed the downregulation of miR-137 and observed upregulation of DNA methyltransferase 3a (DNMT3a), a miR-137 target, in HCC patients and cells. Mechanistically, overexpressing miR-137 inhibited HCC carcinogenesis via interacting with DNMT3a and suppressing the PTEN/Akt signaling. Our reports would provide great worth for understanding hepatocarcinogenesis mechanisms better.

Materials and Methods

Clinical Samples

HCC and paired peritumoral healthy tissues were obtained from twenty-nine HCC patients undergoing hepatectomy at the First Affiliated Hospital of Zhengzhou University from July 2017 to June 2018. After surgical resection, all tissues were immediately snap-frozen for RNA extraction. None of the involved patients accepted chemotherapy and/or radiotherapy prior to the surgery. The study protocols were approved by the Ethics Committee of First Affiliated Hospital of Zhengzhou University and conducted in accordance with the Declaration of Helsinki. The informed written consents were collected from every patient prior to tissue collection.

Cell Transfection and Treatments

HCC cell lines (Huh7, SMMC-7721, HepG2, Hep3B, and BEL-7404) and immortalized normal liver cell line LO2 were purchased from the American Type Culture

Collection (ATCC, Manassas, VA, USA). All cells were maintained with RPMI-1640 medium (Gibco, Grand Island, NY, USA) supplemented with 10% FBS (Thermo Fisher Scientific, Inc., Waltham, MA, USA), and fostered in a humidified incubator containing 5% CO₂. When growing to 70–80% confluence, SMMC7721 and HepG2 cells were transfected with miR-137 mimics (miR-137), miR-137 antagomir (anti-miR-137), respective negative controls (miR-NC, anti-miR-NC), or miR-137 + Lv-DNMT3a (GenePharma Shanghai, China) with Lipofectamine 3000 reagent (Invitrogen, Carlsbad, CA, USA). The transfected cells were harvested for further analyses.

Quantificational Real-Time Polymerase Chain Reaction (qRT-PCR)

TRIzol reagent (Invitrogen) was used to isolate total sample RNAs. The miScript reverse transcription kit (Bio-Rad Laboratories, Hercules, CA, USA) was used to synthesize the single strand cDNA. TaqMan MicroRNA Assay kit (Applied Biosystems, Foster City, CA, USA) was used to examine miR-137 expression. To determine DNMT3a mRNA expression, Scientific RevertAid First Strand cDNA Synthesis Kit (Thermo Fisher Scientific) was applied to synthesize cDNA. SYBR Premix Ex Taq II (Takara, Dalian, China) was used to carry out the subsequent qPCR analyses. StepOnePlus real-time PCR system (Applied Biosystems) was used to monitor all the reactions. The *U6* snRNA and *GAPDH* were employed as internal references for miR-137 and DNMT3a, respectively. The $2^{-\Delta\Delta C_t}$ method was used to compute the relative gene expression. Each experiment was repeated independently at least 3 times.

Western Blot

Complete protease inhibitor Cocktail (Roche, Guangzhou, China) supplemented RIPA lysis buffer (Beyotime, Shanghai, China) was used to isolate sample proteins. Bicinchoninic acid protein assay kit (Peirce, Rockford, IL, USA) was employed to detect the concentration of total proteins. The extracted proteins were subjected to 10% SDS-PAGE, followed by electroblotting onto PVDF membranes (Millipore, Billerica, MA, USA). Blocked with 5% skimmed milk at 25°C for 2 h, the blots were incubated with the respective primary antibodies at 4°C overnight. Then, the blots were probed with corresponding HRP-conjugated secondary antibodies. Last, Enhanced Chemiluminescence Western

Blotting kit (Peirce) was used to visualize the immunoreactive protein bands. The following primary antibodies were used in this section: DNMT3a (Bioss antibodies, Woburn, MA, USA), PCNA, caspase-3, cleaved caspase-3, MMP-2, and MMP-7 (Abcam, Cambridge, UK); PTEN, Akt, p-Akt, and β -actin (Cell Signaling Technology, Beverly, MA, USA). Each experiment was repeated independently at least 3 times.

Luciferase Reporter Assay

PCR was used to synthesize 3'UTR of DMT3a where rested the putative miR-137 binding sites (DMT3a-WT) and its mutant (DMT3a-MUT). The pGL3-reporter luciferase vectors (Promega, Madison, WI, USA) were used to load the synthesized sequences. Next, miR-137, anti-miR-137, or respective controls and luciferase reporter constructs were co-transfected into SMMC7721 and Huh7 cells using Lipofectamine 3000 reagent (Invitrogen). Forty-eight hours post-delivery, Dual-Luciferase Reporter Assay System (Promega) was used to detect the luciferase activities. Each experiment was repeated independently at least 3 times.

MTT Assay

MTT assay was used to evaluate cell proliferative abilities. After transfection for 0, 1, 2 and 3 days, cells were added with 20 μ L of MTT reagent (5 mg/mL; Sigma, St Louis, MO, USA). After 4 h incubation, 150 μ L of DMSO was used to dissolve formazan in the cells. Last, a Microplate Reader (Bio-Tek, Winooski, VT, USA) was used to read the optical density at 490 nm wavelength. Each experiment was repeated independently at least 3 times.

Cell Apoptosis Assay

At 48 h post-treatment, the harvested cells were resuspended with binding buffer. And the cells were stained with Annexin V-fluorescein isothiocyanate and propidium iodide (BD Biosciences, San Diego, CA, USA) in the dark for 15 min. Then, a flow cytometer (FACSCalibur, BD Biosciences) was applied to assess the apoptotic cells. Each experiment was repeated independently at least 3 times.

Wound Healing Assay

Wound healing assay was used to assess cell migratory ability. The treated cells were cultured to reach 80% confluence. A 200 μ L sterile pipette tip was used to produce a uniform scratch on the cell monolayers. The generated scratch was photographed at 0 and 24 h with an inverse

microscope (Olympus, Tokyo, Japan). Each experiment was repeated independently at least 3 times.

Transwell Assay

The Transwell chambers (8- μ m pore size, Corning Incorporated, Corning, NY, USA) pre-coated with Matrigel (BD Biosciences) were used in this section. Upper compartments were added with cells suspended in 200 μ L serum-free medium, and the lower ones were added with 600 μ L medium supplemented with 10% FBS. Forty-eight hours post-incubation, the invaded cells in the lower compartment were fixed with ethanol and stained using crystal violet for 30 min. Last, an inverse microscope (Olympus) was used to count and record the invaded cells. Each experiment was repeated independently at least 3 times.

Methylation-Specific PCR (MSP)

Get pure DNA Kit (Dojindo Molecular Technologies, Gaithersburg, MD, USA) was used for extracting the cell genomic DNA. The bisulfite modification of genomic DNA for methylation analysis was conducted using an EpiTect Bisulfite kit (Qiagen Inc., Valencia, CA, USA). Then, the PTEN promoter methylation status was assessed using the EZ DNA Methylation Direct kit (Zymo Research, Orange, CA, USA) under the LightCycler 480 system (Roche Applied Science, Penzberg, Germany). Agarose gel electrophoresis was used to separate the PCR products and nucleic acid silver staining kit (Beijing Dingguo Changsheng Biotechnology Co., Ltd., Beijing, China) was used to stain them. Sequences for the methylated and unmethylated primers were listed as below: methylated PTEN, forward 5'-TTCGTTTCGTCGTCGTCGTATTT-3', 5'-GCCGCTTAAC TCTAAACCGCAA-3'; unmethylated PTEN, forward 5'-GTGTTGGTGGAGGTAGTTGTTT-3', reverse 5'-ACCA CTAAACTCTAAACCACAACCA-3'. Each experiment was repeated independently at least 3 times.

Xenograft Nude Mouse Models

Animal experiments strictly followed the Principles of Animal Care and Use of the First Affiliated Hospital of Zhengzhou University and were approved by the Ethics Committee of First Affiliated Hospital of Zhengzhou University. 9 nude mice (BALB/C, 6 weeks old, male, Slac Laboratory Animal Co. Ltd, China) were used to construct xenograft mouse models with right flank region subcutaneously injected with the transfected cells. The xenograft mouse models were sacrificed 4 weeks after inoculation. Xenografts were resected, pictured and

measured. Tumor volume was calculated according to the formula: Volume = (length × width²)/2. Each experiment was repeated independently at least 3 times.

Immunohistochemistry (IHC) Assay

Specimens were immersed in hydrogen peroxide at 37°C for 10 min, and blocked with goat serum at 25°C for 2 h. Then, specimens were probed with the primary (DNMT3a) and secondary antibodies strictly followed the manufacture's instruction. Last, 3,3'-diaminobenzidine (DAB) and hematoxylin were used to visualize the specimens. A light microscope (Olympus) was used for imaging the stained sections. DNMT3a expression was scored and the formula to calculate score was as follows: 4 (High Positive), 3+ (Positive), 2+ (Low Positive) and 1+ (Negative) based on the percentage of positive cells stained. Each experiment was repeated independently at least 3 times.

Statistical Analysis

SPSS17.0 software (IBM, Chicago, IL, USA) was employed for the statistics. The measurement data were shown as mean ± standard deviation (SD) and analyzed by Student's *t*-test or analysis of variance (ANOVA). The counting data were analyzed with the Chi-square test. *P* < 0.05 was rendered statistically significant.

Results

A Decrease of miR-137 and an Increase of DNMT3a Were Observed in HCC Tissues and Cells

The miR-137 and DNMT3a expression profiles in HCC tissues and cells were determined with Western blot, immunohistochemistry and qRT-PCR analyses. The miR-137 expression was significantly lowly expressed (Figure 1A), while DNMT3a mRNA expression (Figure 1B) and DNMT3a protein expression were dramatically upregulated (Figure 1C) in HCC tissues with respective to that in their healthy counterparts. DNMT3a expression status (IHC score 2+) was found in 75% of HCC tissues and DNMT3a expression status (IHC score 1) was detected in 80% of healthy counterparts (data not shown). Further, a negative correlation between the expression of miR-137 and DNMT3a in HCC tissues was described by Spearman correlation analysis (Figure 1D). Meanwhile, miR-137 expression was also abnormally suppressed (Figure 1E), while both DNMT3a mRNA and protein levels were exceptionally higher (Figure 1F and G) in HCC cell lines (Hep3B, SMMC-7721, HepG2, Huh7, and BEL-7404) relative with that in immortalized normal liver cell line LO2. Meanwhile, the analyses of miR-137 expression and the clinicopathological characteristics of HCC patients (Table 1) implicated a great significance of the correlation between the decreased miR-137 and tumor size

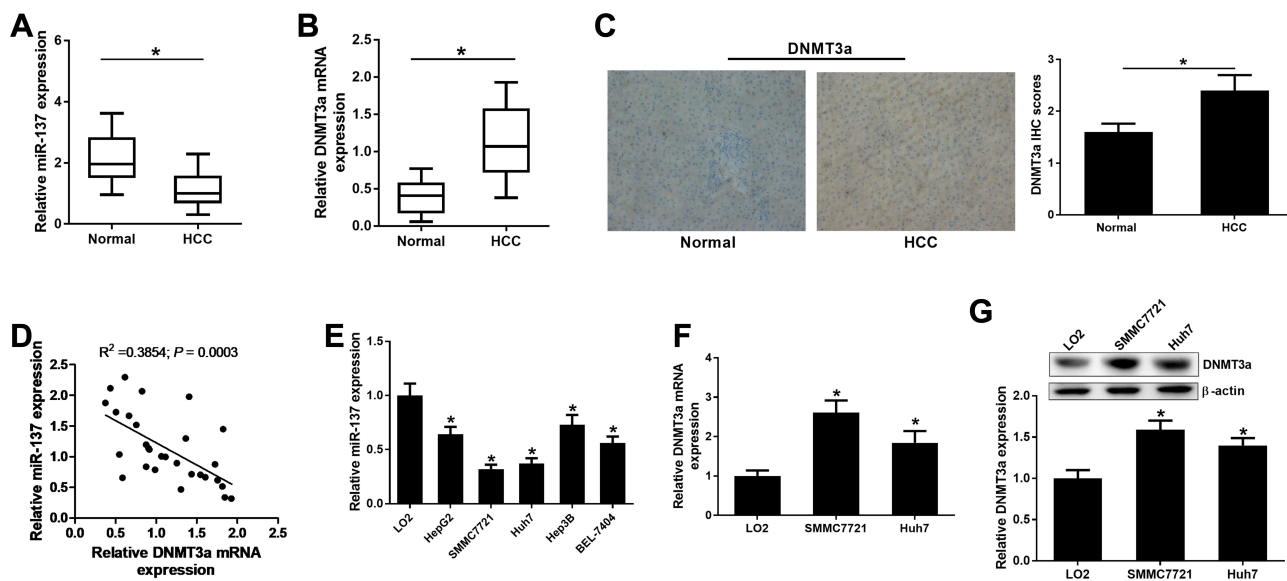


Figure 1 miR-137 and DNMT3a expression in HCC clinical specimens and cells. qRT-PCR analyses for miR-137 (A) and DNMT3a were (B) expressions in 29 paired HCC and their healthy counterparts. (C) Representative pictures (left) of IHC staining for DNMT3a expression and DNMT3a IHC scores (right) in HCC tissues and their healthy counterparts. (D) miR-137 was inversely correlated with DNMT3a mRNA level in HCC tissues. (E) qRT-PCR analyses for miR-137 level in different cell lines (Hep3B, HepG2, SMMC-7721, Huh7, and BEL-7404, and LO2). (F and G) DNMT3a mRNA and protein levels in HCC cell lines (SMMC-7721, Huh7, and LO2) were valued using qRT-PCR and Western blot, respectively. **P* < 0.05.

Table 1 Relationship Between miR-137 Expression and Clinicopathological Features of HCC Patients

Features		n	miR-137 Expression		P value	χ^2
			High	Low		
Age	<50	16	7	9	0.774	0.083
	≥ 50	13	5	8		
Gender	Male	12	6	6	0.428	0.627
	Female	17	6	11		
Tumor size (cm)	<5	13	8	5	0.047	3.948
	≥ 5	16	4	12		
Liver cirrhosis	Yes	22	8	14	0.331	0.945
	No	7	4	3		
Metastasis	Yes	16	4	12	0.047	3.948
	No	13	8	5		
TNM stage	I/II	13	9	4	0.006	7.535
	III/IV	16	3	13		
HBsAg status	Positive	19	8	11	0.913	0.012
	Negative	10	4	6		

(medium value, $P = 0.047$), metastasis ($P = 0.047$) and advanced TNM stage ($P = 0.006$). These speculated that there may be a close relationship between abnormal expressed miR-137 and DNMT3a in HCC progression and metastasis.

miR-137 Directly Targeted and Repressed DNMT3a in HCC Cells

Aiming at exploring the miR-137 functions in HCC, miR-137, anti-miR-137, or respective controls was transfected into SMMC7721 and Huh7 cells. As demonstrated by qRT-PCR assay, a notable increase of miR-137 expression was found after delivery of miR-137 (Figure 2A and B) in SMMC7721 and Huh7 cells. miR-137 was drastically decreased following introduction with anti-miR-137 in cells (Figure 2C and D). Interestingly, a dramatic reduction in DNMT3a after miR-137 delivery, while a noticeable elevation following anti-miR-137 delivery in HCC cells was revealed by Western blot analysis (Figure 2E and F). Furthermore, bioinformatics analyses by online software TargetScan showed that DNMT3A was a putative miR-137 target (Figure 2G). To verify the hypothesis, luciferase reporter assay was employed. As demonstrated by the luciferase reporter assay, introducing miR-137 greatly inhibited DNMT3a-WT luciferase activity in HCC cells, while inhibiting miR-137 conspicuously promoted (Figure 2H and I). However, both treatments failed to alter

the luciferase activity in DNMT3a-MUT groups (Figure 2H and I). Thus, miR-137 interacting with DNMT3a directly in HCC cells was proved.

miR-137 Overexpression Resulted in a Decrease in HCC Cell Proliferation and an Increase in Apoptosis by Decreasing DNMT3a Expression

To clarify whether miR-137 executed its biological function via DNMT3a, miR-137, miR-NC, or miR-137 + Lv-DNMT3a were delivered into SMMC7721 and Huh7 cells. The inhibitory effect of increased expression of miR-137 on DNMT3a expression was partly restored following reintroduction with Lv-DNMT3a in SMMC7721 and Huh7 cells (Figure 3A). MTT assays exhibited that upregulation of miR-137 distinctly impeded proliferation of SMMC7721 and Huh7 cells relative to control group, whereas this effect was effectively abolished by DNMT3a overexpression (Figure 3B). Also, flow cytometry analyses revealed a substantial enhancement in apoptosis in SMMC7721 (Figure 3C) and Huh7 (Figure 3D) following miR-137 overexpression, while augmented expression of DNMT3a partially attenuated this effect. Moreover, the protein expression of cell proliferation marker PCNA was apparently curbed in miR-137-overexpressing SMMC7721 (Figure 3E) and Huh7 (Figure 3F) cells. That effect of miR-137 overexpression was obviously reversed following DNMT3a overexpression. Furthermore, overexpression of miR-137 markedly increased cleaved caspase-3 level in SMMC7721 (Figure 3G) and Huh7 (Figure 3H) cells. However, those effects were remarkably reversed by forced expression of DNMT3a. However, miR-137 or DNMT3a overexpression produced little alternation in caspase-3 level. These data suggested that HCC cell proliferation was retarded and apoptosis was promoted because of the miR-137 overexpression induced DNMT3a inhibition.

Enforced miR-137 Expression Retarded HCC Cell Migratory and Invasive Traits by Decreasing DNMT3a Expression

Next, the suppressive effect of miR-137 on HCC cell migratory abilities was revealed by wound healing assays (Figure 4A and B). The inhibitory roles of miR-137 in HCC cell invasion were also manifested by Transwell assays (Figure 4C and D). However, the above inhibitory effect was effectively reversed by ectopic DNMT3a expression (Figure 4A–D). MMPs are

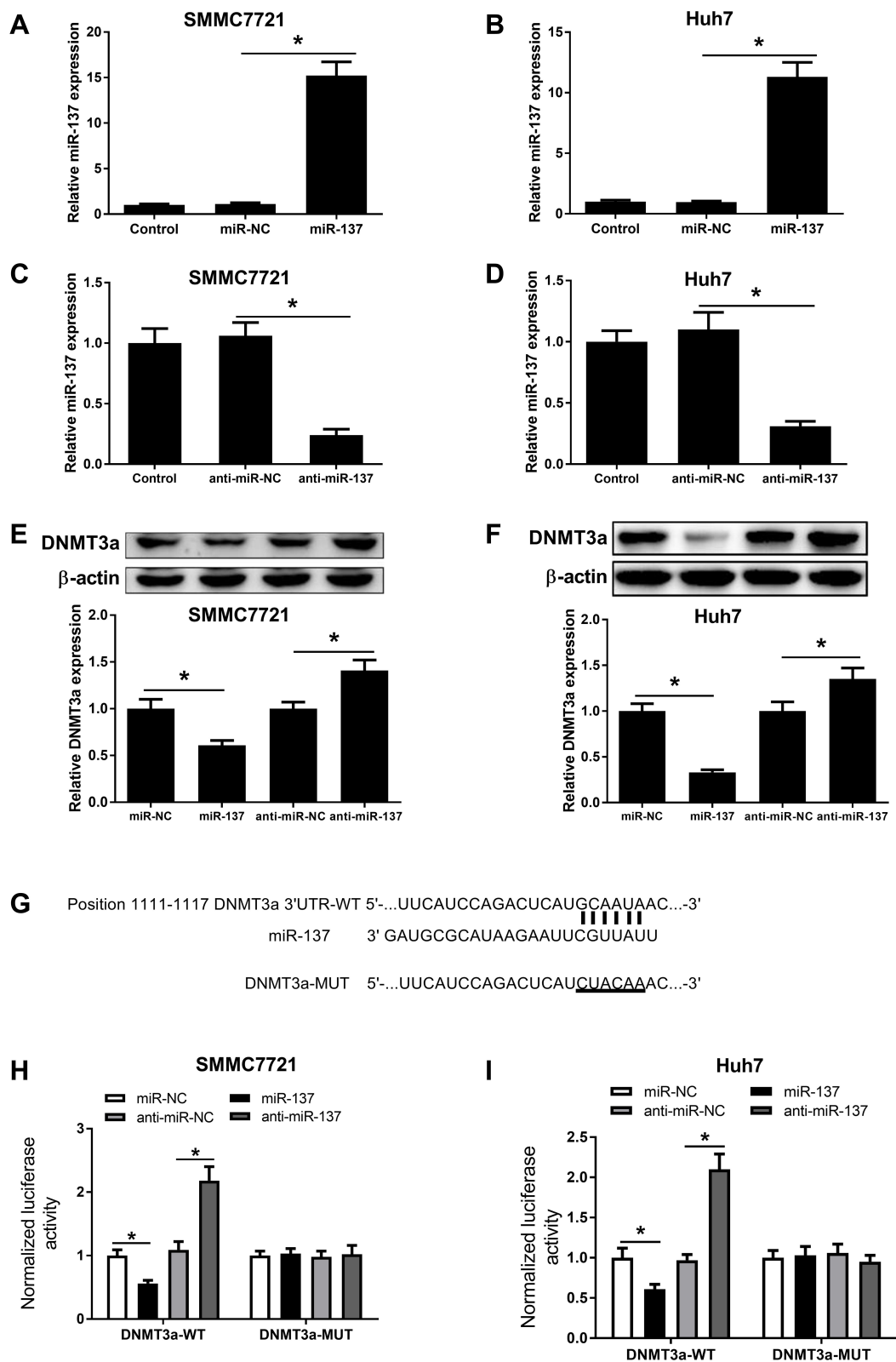


Figure 2 miR-137 targeted DNMT3a directly in HCC cells. miR-137 expression in SMMC7721 (A) and Huh7 (B) cells received miR-NC or miR-137 transfection, as well as SMMC7721 (C) and Huh7 (D) cells received anti-miR-NC or anti-miR-137 infection. Western blot analyses for DNMT3a protein level in SMMC7721 (E) and Huh7 (F) cells after delivery of miR-137, anti-miR-137, or corresponding controls. (G) The potential miR-137 targeting site in DNMT3a 3'UTR. Luciferase activity in SMMC7721 (H) and Huh7 (I) cells after miR-137, anti-miR-137, or matched controls and DNMT3a-WT or DNMT3a-MUT co-transfection. * $P < 0.05$.

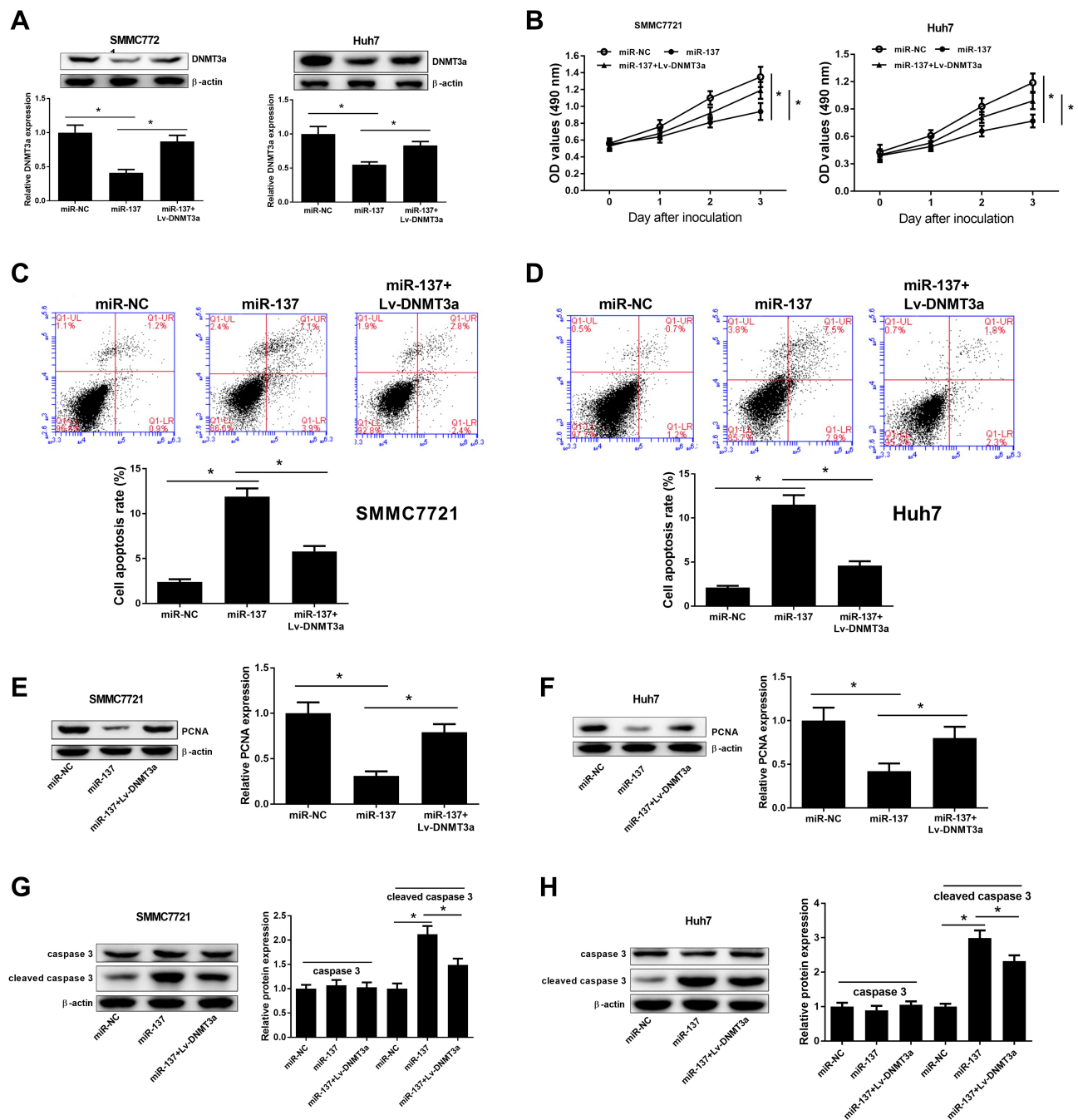


Figure 3 Effects of miR-137 or combined with DNMT3a on cell proliferation and apoptosis in HCC cells. SMMC7721 and Huh7 cells received miR-137, miR-NC, or miR-137 + Lv-DNMT3a delivery. (A) Western blot analyses for DNMT3a protein levels in the treated HCC cells. (B) MTT assays for cell proliferation of the treated HCC cells, at days 0, 1, 2, and 3. (C and D) Flow cytometry analyses for apoptosis in the treated HCC cells. (E and F) Western blot analyses for PCNA protein levels in the treated HCC cells. (G and H) Western blot analyses for caspase-3 and cleaved caspase-3 protein levels in the treated HCC cells. * $P < 0.05$.

well known for participating in tumorigenesis regulation.¹³ The miR-137 overexpression resulted in a marked decrease of MMP-2 and MMP-7 levels, which was partially overturned following reintroduction with Lv-DNMT3a (Figure 4E and F). Together, the enforced miR-137 expression retarded HCC cell migratory and invasive abilities via interacting with DNMT3a.

miR-137 Overexpression Inactivated the PTEN/Akt Pathway in HCC Cell by Decreasing DNMT3a Expression

The participation of PTEN/Akt signaling in HCC progression has been acknowledged in various studies.^{14,15} To confirm whether there was any biological regulation of miR-137

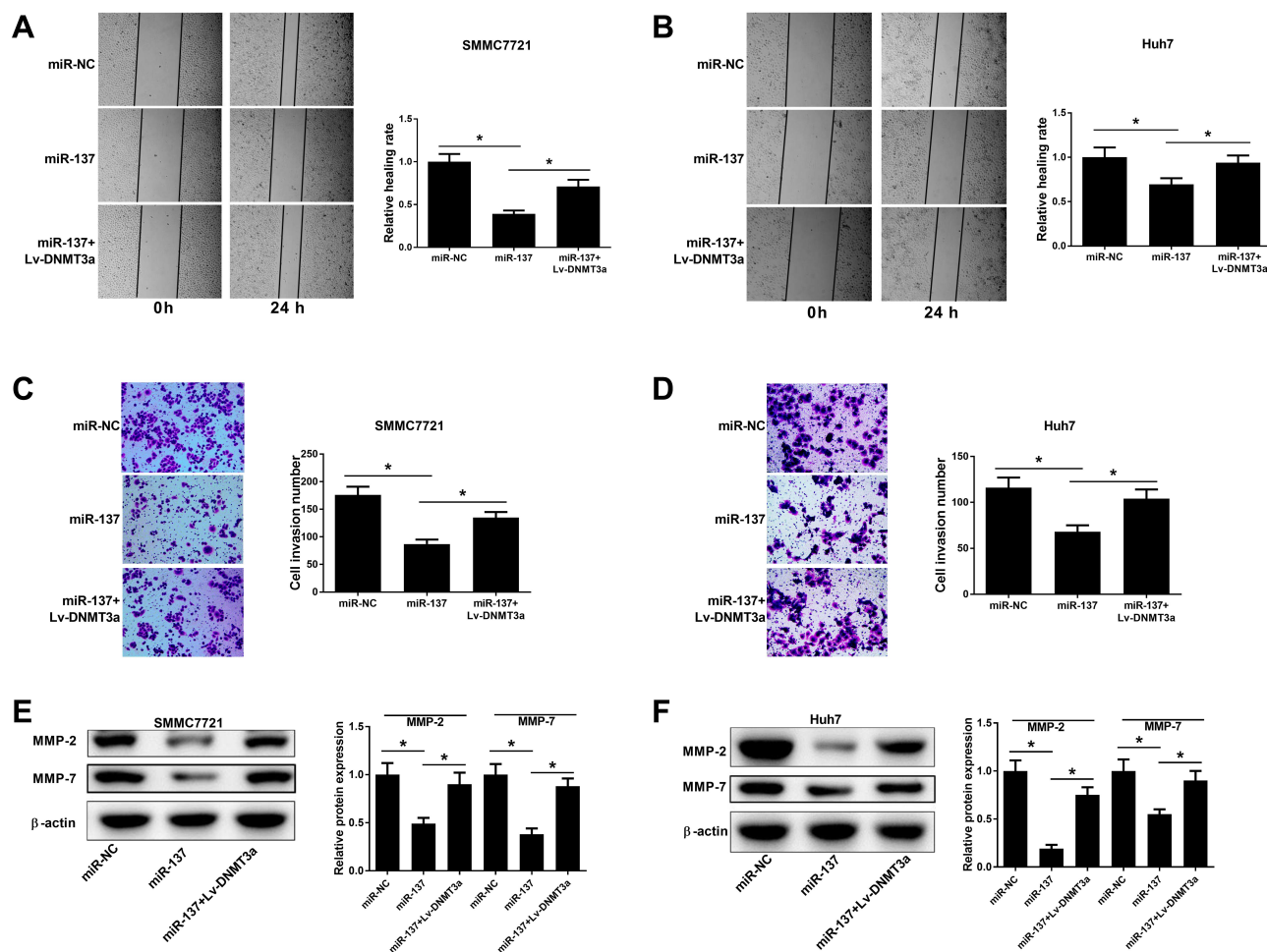


Figure 4 Effects of miR-137 or combined with DNMT3a on cell migration and invasion in HCC cells. Cells were delivered with miR-137, miR-NC, or miR-137+Lv-DNMT3a. (A–D) Wound healing analyses and Transwell invasion analyses for HCC cell migratory and invasive abilities. (E and F) Western blot analyses for MMP-2 and MMP-7 protein levels in the transfected cells. * $P < 0.05$.

on PTEN signaling in HCC cells, Western blot analysis was employed. The PTEN protein level was obviously increased upon miR-137 delivery, while DNMT3a overexpression considerably attenuated this effect in HCC cells (Figure 5A). Next, we further analyzed whether the increased PTEN expression was attributed to the demethylation of PTEN promoter region by miR-137. Moreover, the CpG sites of PTEN promoter were successfully demethylated following miR-137 overexpression in SMMC7721 and Huh7 cells, which was evidently recuperated by DNMT3a overexpression (Figure 5B and C). These results suggested that the increased PTEN expression was due to the demethylation of its CpG sites. Further, the involvement of the Akt pathway in miR-137 regulation on HCC cells was analyzed using Western blot. We observed an inhibition of Akt phosphorylation by miR-137 in HCC cells (Figure 5D and E), while this effect was partially restored by DNMT3a

overexpression. These results manifested that miR-137 overexpression inactivated the PTEN/Akt pathway in HCC cells by decreasing DNMT3a expression.

miR-137 Overexpression Inhibited Tumor Growth in vivo in HCC via Interacting with DNMT3a and Inhibiting the PTEN/Akt Cascades

To verify the influence of miR-137 or along with DNMT3a in HCC in vivo, xenograft models were generated. Four weeks, mice were killed and xenografts were excised and analyzed. The tumors from the SMMC-7721-miR-137 group presented significant decreased weight and volume compared with their counterparts, which was dramatically reversed following reintroduction with Lv-DNMT3a (Figure 6A–C). Moreover, IHC staining and

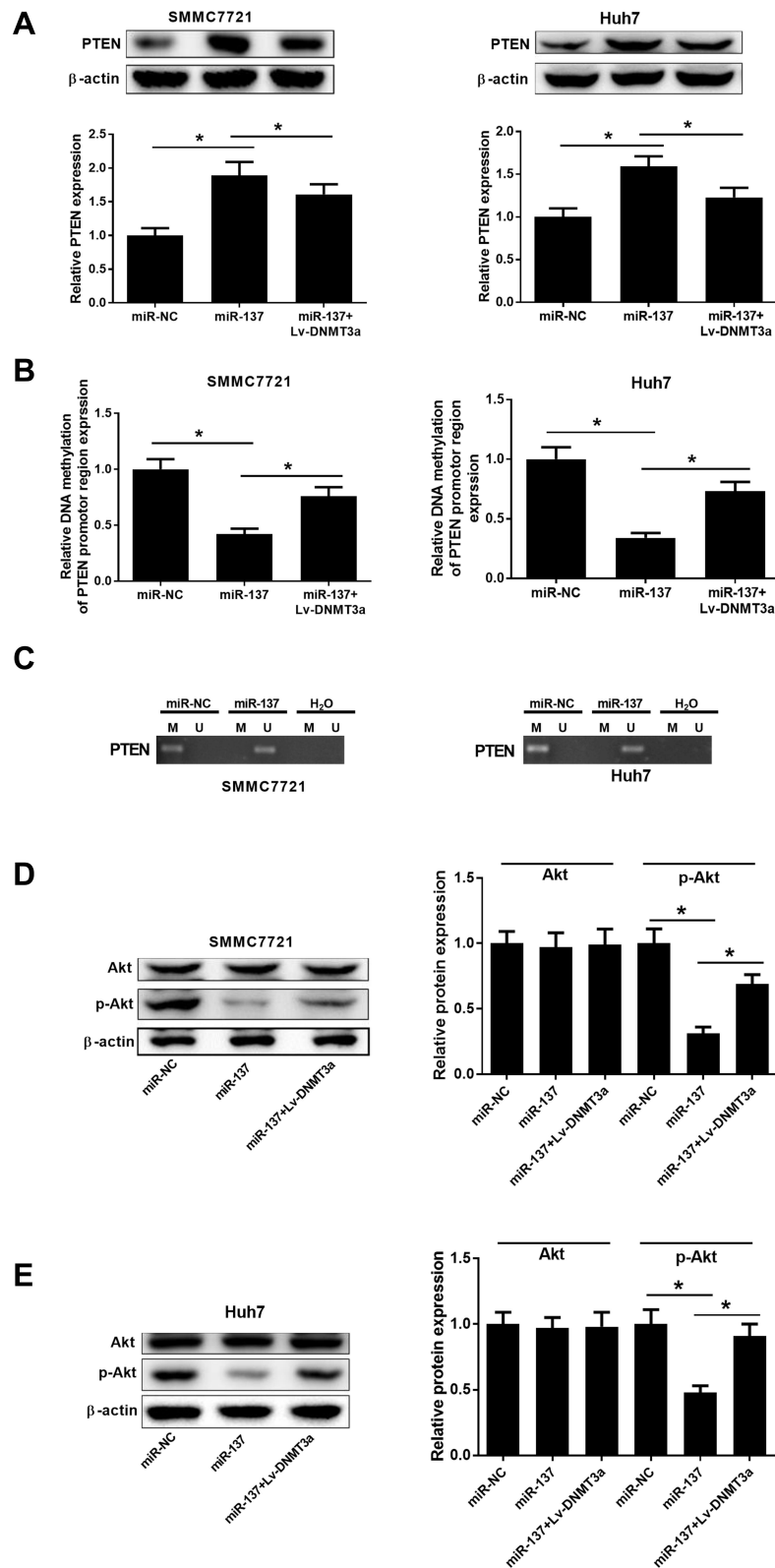


Figure 5 Effects of miR-137 or combined with DNMT3a on the PTEN/Akt pathway in HCC cells. SMMC7721 and Huh7 cells received miR-137, miR-NC, or miR-137 + Lv-DNMT3a transfection. **(A)** Western blot analyses for PTEN protein levels in the treated cells. **(B and C)** MSP was conducted to examine the extent of methylation status of PTEN promoter region in the treated cells. 'M' and 'U' lanes represented PCR products obtained with methylation-specific and unmethylation-specific primers. H₂O was used as a negative control. **(D and E)** Western blot analyses for Akt and p-Akt protein levels in the treated HCC cells. **P* < 0.05.

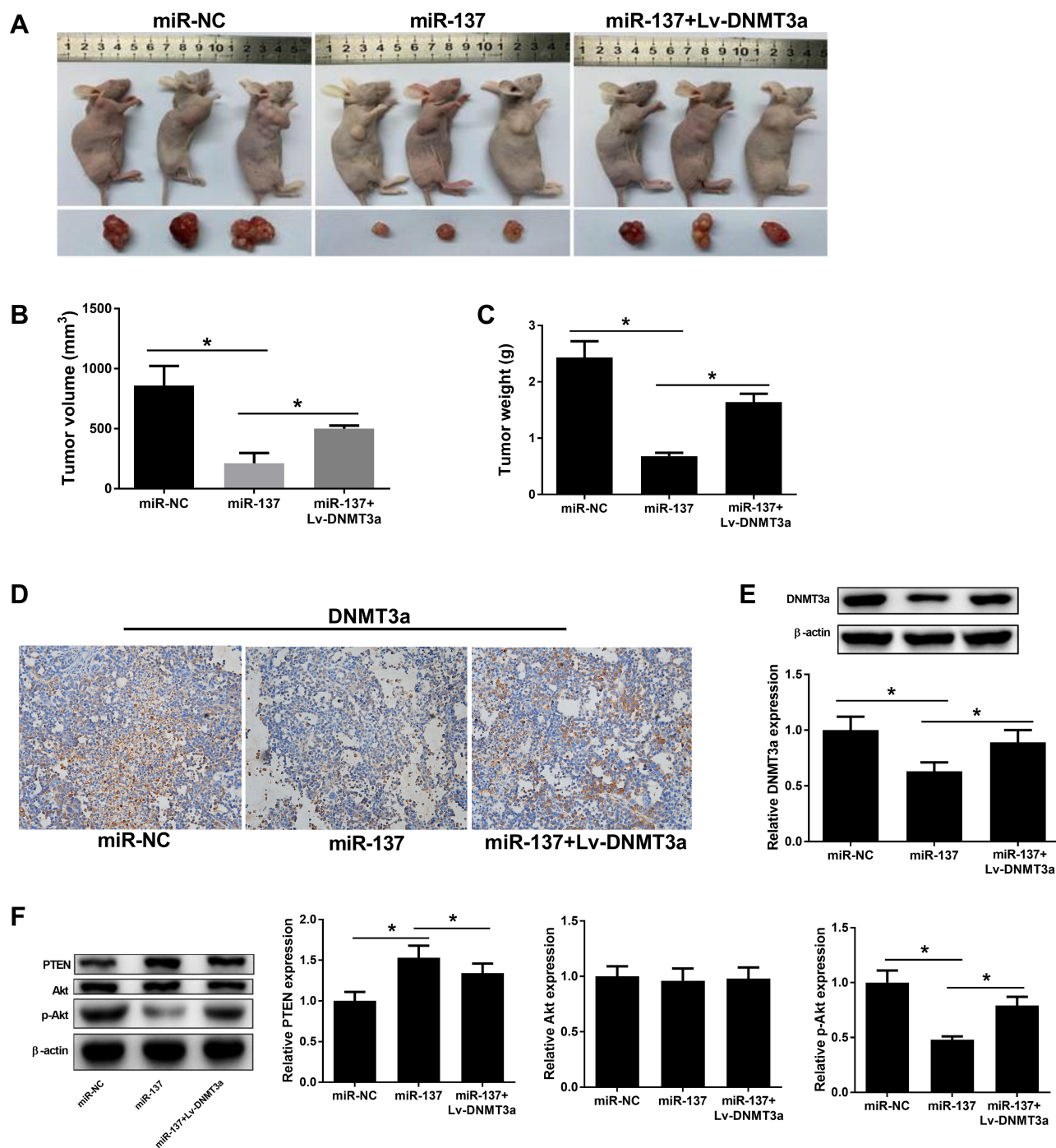


Figure 6 Effects of miR-137 or combined with DNMT3a on HCC xenograft growth. The mice right flank regions were subcutaneously injected with miR-NC, miR-137, or miR-137 + Lv-DNMT3a introduced SMMC7721 cells. (A) 4 weeks post-inoculation, tumor tissues were resected and pictured. Tumor volume (B) and weight (C) in different transfection groups. (D) DNMT3a expression in the xenografts by IHC staining. (E and F) Western blot analyses for DNMT3a, PTEN, p-Akt and Akt protein levels in the resected tumor tissues. * $P < 0.05$.

Western blot assays showed that DNMT3a expression was markedly reduced in tumors overexpressing miR-137 (Figure 6D and E). And, those roles of miR-137 were abolished by overexpressing DNMT3a (Figure 6D and E). Furthermore, overexpressing miR-137 dramatically

increased PTEN protein level and suppressed the phosphorylation of Akt in xenografted tumors (Figure 6F). However, the increased expression of DNMT3a drastically overturned those effects (Figure 6F). Collectively, these results suggested that miR-137 inhibited HCC tumor

growth in vivo by targeting DNMT3a through suppressing the PTEN/Akt pathway.

Discussion

HCC belongs to the lethal human malignancies and leads to a huge burden on families and society due to its high metastasis and recurrence rate.¹ It is widely believed that the abnormally expressed miRNAs are considered as important regulators in carcinogenesis.¹⁶ Hence, understanding miRNAs characteristics better would be helpful for understanding and controlling HCC better. Our study aimed at clarifying how miR-137 inhibited the HCC malignancy. Herein, we observed downregulation of miR-137 in HCC cells and tissues. DNMT3a was highly expressed in HCC cells and tissues and confirmed to be directly regulated by miR-137. miR-137 overexpression induced HCC cell apoptosis, as well as suppressed proliferative, migratory and invasive abilities via interacting with DNMT3a. Mechanistically, miR-137 inactivated the PTEN/Akt cascades in HCC cells via inhibiting DNMT3a.

Recently, miR-137 has attracted increasing attention due to its tumor-suppressing effects by suppressing cell proliferation, migration, and promoting apoptosis in diverse malignancies.^{17–19} In contrast, miR-137 upregulation and functioning as an oncogene in bladder cancer and squamous cell carcinoma has been reported in recent studies.^{20,21} Here, we observed a decrease in miR-137 expression in HCC and demonstrated its suppressive effect on cell proliferative, migratory, and invasive abilities, as well as promotive effect on apoptosis, which was in accordance with the previous reports.^{12,22–24} More interestingly, our study found that miR-137 interacted with DNMT3a directly in HCC cells. The DNMT3a belongs to the DNA methyltransferases (DNMTs) family, which is crucial for carcinogenesis because of their epigenetic regulating abilities.^{25,26} Reportedly, DNMT3a is related to de novo methylation.²⁷ As an oncogene, DNMT3a is ubiquitously overexpressed in many tumors such as pancreatic cancer,²⁸ gastric cancer,²⁹ and HCC,³⁰ and implicated in tumorigenesis, differentiation and metastasis.³¹ Consistently, the overexpression of DNMT3a in HCC cells and tissues was observed in the present study. Moreover, the regulation of miR-137 overexpression on HCC cell proliferative, apoptotic, migratory and invasive capabilities was effectively abrogated by ectopic DNMT3a expression, suggesting that miR-137 overexpression inhibited HCC carcinogenesis via suppressing DNMT3a.

The PTEN, a well-studied tumor suppressor, regulates cell proliferation, survival, metastasis and energy metabolism.³² It is believed that the PTEN level is dependent on the methylation status of its promoter, which is priorly subjected to DNMT3a methylation.³³ Promoter hypermethylation or silencing of PTEN is considered as the major mechanism for PTEN epigenetic inactivation.³⁴ The aberrant DNA methylation induced PTEN silence has been well documented in many cancers including HCC.³⁵ DNMT3a depletion inhibits HCC growth and progression via demethylating PTEN promoter.³⁰ PTEN acts as a key inhibitor of the Akt pathway,³⁶ and downregulation of PTEN induces hyperactivation of the Akt pathway, which contributes to carcinogenesis in various cancers.³⁷ Our study proved that the CpG sites of PTEN promoter were successfully demethylated following restoration of miR-137 expression, thus resulting in increased PTEN expression. Furthermore, miR-137 overexpression ultimately inactivated the Akt pathway in HCC cells, which was mediated by downregulating DNMT3a. That miR-137 overexpression inhibited HCC carcinogenesis in vivo via interacting with DNMT3a and inhibiting the PTEN/Akt pathway has also been validated in our study.

Conclusion

In conclusion, our study demonstrated that the suppressive effect of miR-137 on HCC tumor growth was through targeting DNMT3a and suppressing the PTEN/Akt signaling. Also, targeting miR-137/DNMT3a/PTEN/Akt cascades may be a potential therapeutic strategy for HCC, which would be helpful for HCC control and treatment.

Disclosure

The authors declare no competing or financial interests.

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