

Expression of organic anion-transporting polypeptides IB3, IB1, and IA2 in human pancreatic cancer reveals a new class of potential therapeutic targets

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Background: Organic anion-transporting polypeptides (OATPs) are influx transporters that mediate intracellular uptake of selective endogenous and xenobiotic compounds. Identification of new molecular targets and discovery of novel targeted therapies is top priority for pancreatic cancer, which lacks any effective therapy.

Materials and methods: We studied expression of OATP 1A2, 1B1, and 1B3 in pancreatic cancer tissue and in cell lines. Formalin-fixed paraffin-embedded biopsy material of 12 human pancreatic cancers was immunohistochemically assessed for protein expression of the three studied influx transporters. Immunohistochemistry was evaluated by experienced pathologists and quantified by use of an automated image analysis system. BxPC-3 and MIA PaCa-2 pancreatic cancer cell lines were used to quantify transcripts of OATP 1B1 and 1B3.

Results: OATP 1A2, 1B1, and 1B3 proteins were found ubiquitously expressed in all studied cases. Quantification performed by HistoQuest system revealed that mean intensity was 53 for 1A2, 45 for 1B1, and 167 for OATP 1B1/1B3 on a range scale 0–250 units. At mRNA level, 1B1 and 1B3 were overexpressed in both studied cancer cell lines but not in normal pancreatic tissue.

Conclusion: OATPs 1A2, 1B1, and 1B3 are highly expressed in pancreatic adenocarcinoma. We suggest that expression of these transporters in pancreatic cancer justify research efforts towards discovery of novel therapeutics targeting OATPs.

Keywords: organic anion-transporting polypeptides, targeted therapy, transporter

Introduction

Pancreatic carcinoma is the most deadly among cancers, ranked fourth as a cause of cancer-related deaths in the economically developed world.^{1,2} Characteristically, in Europe, new diagnoses of pancreatic cancer almost equaled the deaths caused by this disease in 2008.³

Surgery remains the gold standard for the treatment of pancreatic cancer, if diagnosed early. However, even in cases of surgically resected tumors, the outcome remains poor, and adjuvant therapy can offer marginal benefits.^{2,4} In advanced pancreatic cancer, the outlook is even worse. Extensive research has failed to produce any therapy efficient enough to substantially extend the median survival of treated patients beyond 6 months. Currently available therapies remain palliative on their intent.^{5–7} Therefore, identification of new molecular targets and discovery of novel targeted therapies is top priority for pancreatic cancer research.

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The organic anion transporting polypeptides (OATPs) superfamily comprises 11 polypeptide molecules that share a largely common structure with 12 putative transmembrane regions and a large extracellular loop between the 9th and 10th transmembrane domains⁸ (Figure 1). They operate as influx transporters that mediate the transmembrane uptake of various endogenous and xenobiotic anion compounds. Besides their characteristic expression in normal tissues, OATPs have been found overexpressed in several cancers, and it was such data that prompted us to undertake investigation of these transporters as potential therapeutic targets.⁹ Regarding pancreatic cancer, Abe et al, showed expression of OATP 1B3 in human pancreatic cancer on mRNA and protein level in a single case.¹⁰ To our knowledge, this is the first study that systemically assessed the expression profile of three OATPs (1A2, 1B1, and 1B3) in pancreatic cancer.

Materials and methods

Tissue samples and anti-OATP antibodies

Formalin-fixed paraffin-embedded tissue samples of human pancreatic cancer were retrieved from the archives of the Department of Pathology, Chatzikosta General Hospital, Ioannina, Greece. The patients were diagnosed in the

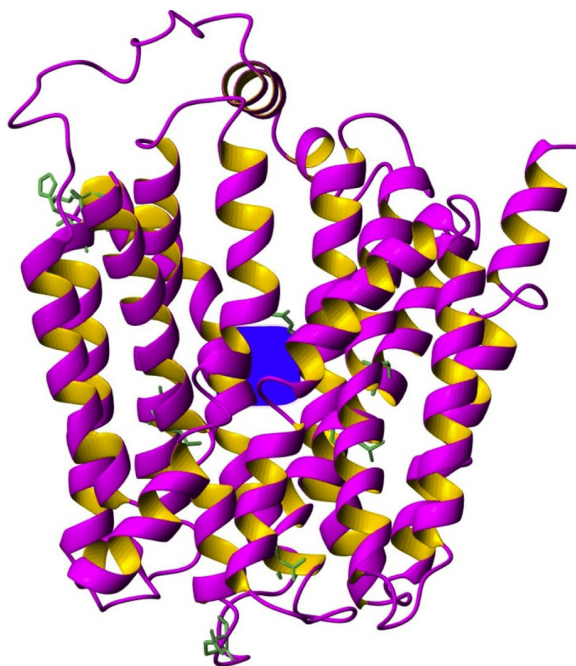


Figure 1 Ribbon representation of the three-dimensional model of organic anion-transporting polypeptide 1B3. TM domains, a probable substrate binding site and the conserved amino acid side chains. The model was built by Modeller program (San Francisco, CA, USA) by using the structure template of the *Escherichia coli* glycerol-3-phosphate transporter (PDB 1pw4).^{24,25} In blue color the potential location of the substrate binding site (the putative translocation pathway) according to former mutagenesis studies is indicated.²⁶ Conserved amino acid side-chains can also be seen on this figure.

period 2000–2008. Their median age was 69 years; six were female and six male. Histologically, eight cases were diagnosed as poorly differentiated pancreas adenocarcinomas, and four cases had intermediate differentiation.

The samples were assessed for expression of OATP 1B1 and 1B1/1B3 by using the mESL and mMDQ antibodies respectively (PROGEN Biotechnik, Heidelberg, Germany). Expression of OATP 1A2 was evaluated in 11 samples by polyclonal anti-OATP 1A2 antibody (Atlas Antibodies AB, Stockholm, Sweden). A polyclonal anti-OATP 1B3 antibody (Atlas Antibodies AB, Stockholm, Sweden) was also used that recognizes C-terminal region of OATP 1B3, on the aim to monitor the expression of the 1B3 transporter as a single entity. All antibodies were diluted with Dako REAL™ Antibody Diluent (DAKO, Code S2022) to the final working concentration (Table 1). The DAKO Autostainer/PT link system was used for the immunostaining process.

Cell lines

Two pancreatic cancer cell lines, BxPC-3 (CRL-1687TM) and MIA PaCa-2 (CRL-1420TM), were obtained from the American Tissue Culture Collection (ATCC, Manassas, VA) to be used in this study. MIA PaCa-2 originated from a male¹⁴ and BxPC-3 from a female donor.¹⁵ Cells were routinely cultured in RPMI-1640 medium (PAN Biotech, Aidenbach, Germany) supplemented with 10% fetal calf serum (Invitrogen Life Technologies, Paisley, Scotland) and 1% penicillin/streptomycin (Invitrogen) under standard culture conditions.

Immunohistochemistry

Tissue sections of 3–4 μm width were cut using a microtome and applied on microscope slides. Slides were incubated overnight at 65°C to enable optimal tissue–glass adhesion. Next, slides were immersed in DAKO's PT-Link containing preheated (65°C) target retrieval solution, at pH 9 (DAKO Code S2375), treated at 93°C for 20 minutes in order to achieve deparaffinization, rehydration, and heat-induced epitope-retrieval (HIER). After cooling back to 65°C, slides were inserted in DAKO Autostainer system for the rest of the immunohistochemistry procedure. The mESL and mMDQ antibodies were applied on slides for 70 minutes, while for the polyclonal anti-OATP1A2 and anti-OATP1B3 antibodies incubation time was set to 60 minutes. The endogenous peroxidase was blocked using Dako REAL peroxidase blocking solution (Code S2023) for 10 minutes. DAKO's special engineered Dextran backbone enriched with peroxidase molecules and goat secondary antibody molecules

Table 1 Antibodies and technical data used for immunohistochemistry

Ab symbol	OATP target	Clonality	Host species	Immunoglobulin subclass	Working dilution	Incubation time (minutes)	Epitope target	Company	Cat #
mMDQ	IB1/IB3	Monoclonal	Mouse	IgG1	1:10	70	MDQHQLNKTAESASSEKKTTRRC for IB3 (N-terminus) MDQNQHNLKTAEQPSENKKTTRYC for IB1 (N-terminus)	Progen Biotechnik	651140
mESL	IB1	Monoclonal	Mouse	IgM	1:5	70	ESLNKHKHFVPSAGADSETHC (C-terminus)	Progen Biotechnik	651139
pIA2	IA2	Polyclonal	Rabbit	IgG	1:250	60	SSWGINTSYEGIPQDLYVENDIFADCNVDCNCPKSIWDP VCGNNGLSYLSACLACGCEISIGTGINMVFQNC CIQTSG NSSAVLGLCLDKGPDC (C-terminus) QGKDTKASDNERYMDEANLEFLNNGEHFV PSAGTDSKTCNLD MQDNAAA (C-terminus)	Atlas Antibodies	HPA027537
pIB3	IB3	Polyclonal	Rabbit	IgG	1:250	60		Atlas Antibodies	HPA004943

Abbreviation: OATP, organic anion-transporting polypeptide.

against rabbit and mouse immunoglobulins (Dako REAL™ EnVision™/HRP, Rabbit/Mouse ENV, Code Code K5007) was applied on slides for 20 minutes followed by a 5 minute Dako REAL™ DAB+ Chromogen (DAKO, Code K5007) detection system and a 2 minute hematoxyline (QS H-3404 Vector Laboratories) treatment. Finally, sections were dehydrated with alcohol/xylene baths and stabilized with mounting medium.

Immunohistochemistry evaluation

Immunostaining intensity was assessed by two experienced pathologists, who graded it, using endothelial cells and tissue macrophages as internal controls (Table 2). Quantitation was done by HistoQUEST (TissueGnostics, Vienna, Austria) system in values of gray from 0 to 250^{11,12} (Figures 2 and 3).

SLCO1A2, SLCO1B1, and SLCO1B3 mRNA expression quantification

Total RNA was isolated from both cell lines grown to subconfluency using the Trizol reagent (Invitrogen). We also used human liver RNA (purchased from Stratagene La Jolla, USA) and human pancreas RNA (purchased from Clontech, Saint-Germain-en-Laye, France) for control purposes.

The concentration, purity, and integrity of RNA samples were determined on a Nanodrop ND-1000 (Kisker-Biotech, Steinfurt, Germany) and by agarose gel electrophoresis.

Reverse transcription of total RNA to cDNA (2 µg) was done by using the high capacity cDNA reverse transcription kit (Applied Biosystems, Foster City, USA) as recommended by the manufacturer. The quantification of SLCO/OATP mRNA transcripts was done by using TaqMan® real-time

Table 2 Immunohistochemical expression of OATPs. Score scale (0, 1+, 2+, 3+) reflecting antibody product expression intensity

Patient number	Sex	Age	Differentiation	pIA2	mESL	mMDQ
1	F	71	Intermediate	+	+	+++
2	F	67	Poor	++	+	+++
3	F	74	Poor	+++	+	++
4	M	56	Poor	+++	+	++
5	M	80	Intermediate	++	+	++
6	F	55	Poor	++	+	++
7	F	78	Poor	+++	+	++
8	F	70	Poor	+++	+	+++
9	M	72	Intermediate	+++	+	++
10	M	67	Poor	++	+	++
11	M	65	Poor	++	+	++
12	M	70	Intermediate	NA	+	++

Abbreviations: F, female; M, male; NA, not applicable; OATP, organic anion-transporting polypeptide.

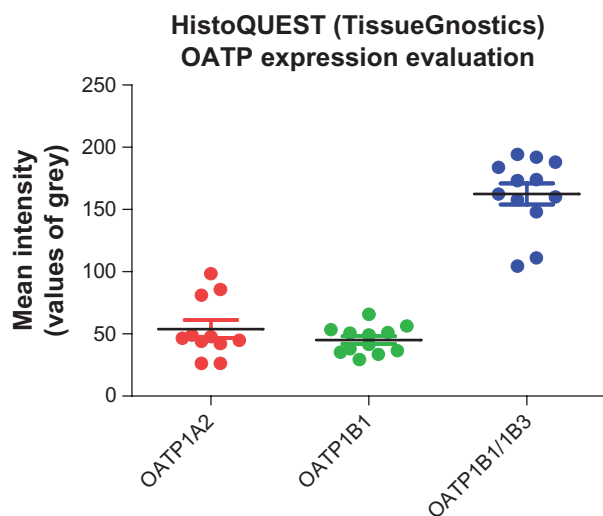


Figure 2 OATP immunohistochemical expression intensity as was assessed by HistoQUEST (TissueGnostics) automated immunohistochemistry analysis system. The Y-axis represents mean intensity measured in values of grey scale from 0 to 250. **Abbreviation:** OATP, organic anion-transporting polypeptide.

PCR method (Applied Biosystems). The following TaqMan assays were used: hs00272374_m1 for OATP 1A2 and hs00251986_m1 for OATP 1B3. For accurate normalization of real time RT-PCR data we included four reference genes in the analysis, namely 18S rRNA (PN 4310893E; Applied

Biosystems), ACTB (PN 4326315E; Applied Biosystems), CYC1 (PrimerDesign Ltd, Southampton, UK), and UBC (PrimerDesign Ltd). The target gene amplification mixture contained 5 μ L 2 \times TaqMan[®] Gene Expression PCR Master Mix (Applied Biosystems), 0.5 μ L of the appropriate Gene Expression Assay, 10 ng template cDNA diluted in 2.5 μ L nuclease free water, and 2 μ L nuclease free water. 5'–3' exonuclease activity of the DNA polymerase was measured with the ABI 7900HT Fast real time PCR system was equipped with SDS 2.3 software (Applied Biosystems). All samples were amplified in duplicates. Results were imported into Prism GraphPad for further analysis. Specimens exhibiting cycle threshold (Ct) values higher than 38 were considered to be negative, and comparable cDNA amounts in the experimental samples were calculated according to Hellemans et al.¹³ Values are expressed as n-fold difference according to a calibrator set to value 1. As a calibrator, RNA pooled from a panel of human tissue samples and cell lines analyzed was taken, and expression values in the calibrator were set to 1.

Results

Immunohistochemistry

The three studied polypeptides were found ubiquitously expressed in all studied biopsy cases. Both methods used confirmed extensive immunostaining of the entire cancer cell tissue with the antibodies used (Table 2; Figures 2 and 4).

Specifically, OATP 1A2 expression signal was weak in one sample and moderate to strong in all others. The HistoQUEST quantification analysis returned a mean intensity of 53.88 units with a standard error (SE) of 7.19. OATP 1B1

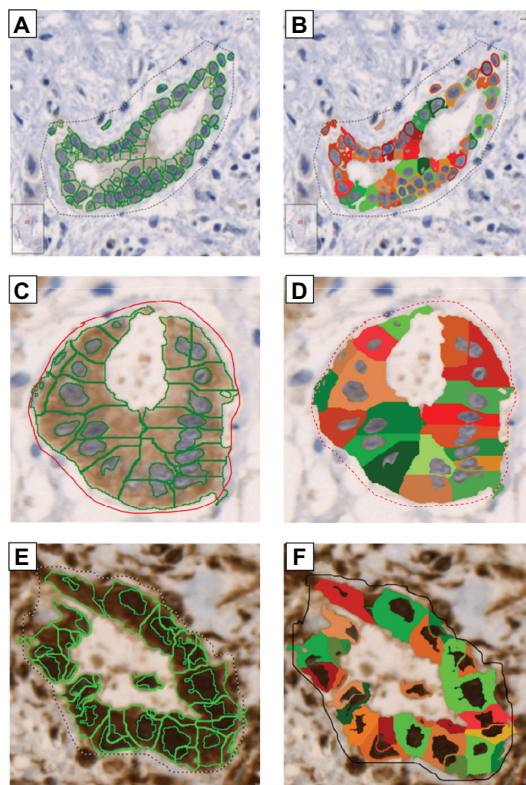


Figure 3 HistoQUEST (TissueGnostics) expression analysis. Pictures **A**, **C**, and **E** represent the identification and segmentation of cells and nuclei. Pictures **B**, **D**, and **F** demonstrate the diaminobenzidine signal intensity for each cell in the studied area.

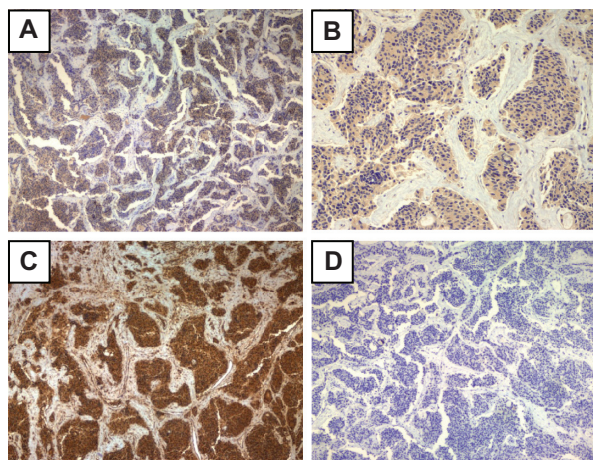


Figure 4 Immunohistochemical staining of pancreatic adenocarcinoma tissue sample from a 77-year-old female patient diagnosed with poorly differentiated adenocarcinoma of the head of the pancreas. **A**) p1A2 antibody ($\times 40$), **B**) mESL antibody ($\times 100$), **C**) mMDQ antibody ($\times 200$), **D**) p1B3 antibody ($\times 40$).

was found to be weakly expressed in all 12 cases with mean intensity 45.10 units (SE 3.15). Immunostaining for mMDQ (OATP 1B1/1B3) was proved the most intense. Nine cases demonstrated moderate expression, three cases stained strong, and HistoQUEST assessment showed a mean intensity of 167.90 units (SE 8.27). Interestingly, the anti-OATP 1B3 p1B3 antibody failed to detect expression of its nominal target in the same cancer tissue material (Figure 4), although it stained normal human liver tissue, which was used in this case as a positive control for p1B3 antibody (Figure 5).

Cell lines

OATP 1B1 and 1B3 mRNA expression in the two cell lines was comparable to that in normal liver, which was taken as a control, because both these transporters are considered “liver-specific”. Their mRNA expression, however, in normal pancreas was either undetectable (OATP 1B1) or 30–60 times lower than in normal liver (OATP 1B3) (Figure 6).

Discussion

The need for discovery of new molecular targets and the development of innovative targeted therapeutics in pancreatic cancer is indisputably illustrated by the fact that of the 36 orphan-designated products for pancreatic cancer by the United States Food and Drug Administration, none received marketing approval on this indication during the last 25 years.¹⁶ We considered that expression of organic anion-transporting polypeptides in a wide range of cancers along with their unique capacity for energy-independent intracellular transport of xenobiotics makes them reasonable targets for potential development of novel cancer therapeutics against hard-to-treat cancers.^{17–20}

We opted to investigate the expression of OATP 1A2, 1B1, and 1B3 in pancreatic cancer, because of their common and distinctive ability to facilitate intracellular uptake of microcystin cyclopeptides, which are natural toxins produced from cyanobacteria.²¹ Microcystins exert

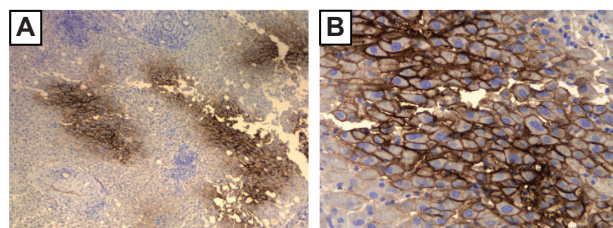


Figure 5 Immunostaining with p1B3 of normal liver tissue with moderate steatosis of a 75-year-old individual who underwent subtotal hepatectomy because of a car accident trauma, shows cell membrane signal with increasing intensity towards central vein regions. **A)** $\times 40$, **B)** $\times 200$.

OATP 1B3 mRNA expression

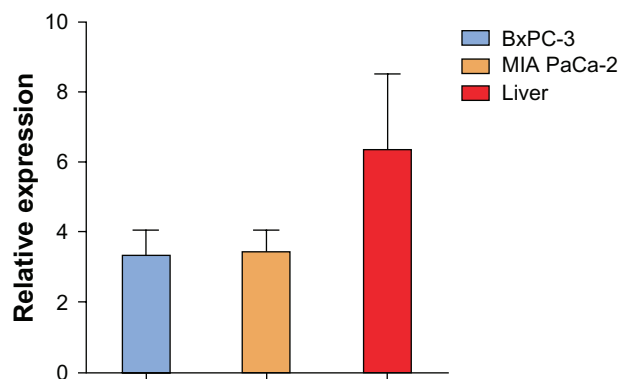


Figure 6 OATP 1B3 mRNA expression in BxPC-3, MIA PaCa-2 cancer cell lines and in human liver. Values are expressed as n-fold difference to the calibrator (pooled from the human tissue samples and cell lines) taken as value 1.

Abbreviation: OATP, organic anion-transporting polypeptide.

cytotoxic effects through depletion of glutathione, generation of reactive oxygen species, and strong inhibition of protein phosphatases 1 and 2A.²² We surmise that these natural toxins are of potential pharmacological interest because they offer opportunities to engineer analogs optimized to induce selective cancer toxicity in OATP expressing tumors.⁹

The OATPs investigated in this study were all found to be ubiquitously expressed in pancreatic cancer: OATP 1A2, 1B1, and 1B3 protein expression was documented in biopsy samples and OATP 1B1 and 1B3 mRNA in cell lines. Quantification analysis showed an increased expression of 1B1/1B3 compared with 1A2 and 1B1, which indicates enhanced expression potential for OATP 1B3 in these tumors. It should be noted that the inability to detect OATP 1B3 C-terminal region with p1B3 antibody opposes the strong positive mMDQ signal in the same samples, which confirmed the presence of the 24 amino acid N-terminal epitope of OATP 1B3. Interestingly, we obtained similar results in a colon cancer study in which we identified an abundant presence of 1B3 protein when studied with mMDQ monoclonal antibody but not with the p1B3 antibody.²³ We speculate that these findings might possibly be related to unknown mutation near the C-terminal end of 1B3 polypeptide in cancers. This is currently under investigation in our laboratory.

Finally, we consider that demonstration of OATP 1B3 and 1B1 mRNA expression in BxPC-3 and MIA PaCa-2 human pancreatic cell lines flags them as appropriate candidates for in-vitro studying of OATP targeted anticancer compounds.

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Disclosure

The authors report no conflicts of interest in this work.

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