

Toll-Like Receptor 4 Polymorphisms (896A/G and I196C/T) as an Indicator of COVID-19 Severity in a Convenience Sample of Egyptian Patients

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Background: The clinical spectrum of COVID-19 is extremely variable. Thus, it is likely that the heterogeneity in the genetic make-up of the host may contribute to disease severity. Toll-like receptor (TLR)-4 plays a vital role in the innate immune response to SARS-CoV-2 infection. The susceptibility of humans to severe COVID-19 concerning TLR-4 single nucleotide polymorphisms (SNPs) has not been well examined.

Objective: The goal of this research was to investigate the association between TLR-4 (Asp299Gly and Thr399Ile) SNPs and COVID-19 severity and progression as well as the cytokine storm in Egyptian patients.

Methods: We genotyped 300 adult COVID-19 Egyptian patients for TLR-4 (Asp299Gly and Thr399Ile) SNPs using PCR-restriction fragment length polymorphism (PCR-RFLP). We also measured interleukin (IL)-6 levels by enzyme-linked immunosorbent assay (ELISA) as an indicator of the cytokine storm.

Results: The minor 299Gly (G) and 399Ile (T) alleles were associated with a significant ($P < 0.001$) positive risk of severe COVID-19 (OR = 3.14; 95% CI = 2.02–4.88 and OR = 2.75; 95% CI = 1.66–4.57), their frequency in the severe group were 71.8% (84/150) and 70.7% (58/150), respectively. We detected significant differences between TLR-4 (Asp299Gly, Thr399Ile) genotypes with regard to serum levels of IL-6. Levels of IL-6 increased significantly with the presence of the mutant 299Gly (G) and 399Ile (T) alleles to reach the highest levels in the Gly299Gly (GG) and the Ile399Ile (TT) genotypes (170 pg/mL (145–208.25) and 112 pg/mL (24–284.75), respectively).

Conclusion: The TLR-4 (Asp299Gly and Thr399Ile) minor alleles 299Gly (G) and 399Ile (T) are associated with COVID-19 severity, mortality, and the cytokine storm.

Keywords: COVID-19, Egypt, mortality, polymorphism, severity, Toll-like receptor

Introduction

Coronavirus disease 2019 (COVID-19) is a respiratory illness caused by severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2).¹ Even though most patients infected by SARS-CoV-2 are mild cases that can recover spontaneously with appropriate symptomatic treatment, some patients deteriorate rapidly, suffering from hyper-inflammation and cytokine storm, with an increased rate of hospitalization, intensive care admission, and high fatality.² Although there are known risk factors associated with severe diseases such as old age, high body mass index, diabetes mellitus, cardiovascular diseases, chronic chest diseases, and malignancies,^{3,4} disease progression can rapidly occur in some patients, even in

the absence of obvious risk factors. Thus, it is likely that other factors, such as the heterogeneity of the genetic makeup and the different immune responses of the host, rather than virus alone, may contribute to disease severity.

The innate immune system, the initial line of defense, is in charge of identifying structurally conserved patterns of organisms called pathogen-associated molecular patterns (PAMPs), as well as mounting an early pro-inflammatory response.⁵ Toll-like receptor 4 (TLR-4) is a critical pattern recognition receptor (PRR) that recognizes multiple PAMPs, from bacteria, mainly lipopolysaccharides (LPS), viruses, and other pathogens.⁶ TLR-4 also identifies certain damage-associated molecular patterns (DAMPs) generated by dead or lytic cells after host tissue injury or viral infection.⁵

Several studies conducted during the COVID-19 pandemic have found that dysregulated TLR-4 signaling may contribute to the initial failure of viral clearance and the development of COVID-19 severe clinical symptoms, namely adult respiratory distress syndrome (ARDS) and lethal respiratory failure.^{5–9}

Considering the importance of TLR-4 in initiating innate immunity against SARS-CoV-2, it is supposed that genetic heterogeneity in the TLR-4 gene might explain, at least in part, the broad spectrum of COVID-19 manifestations. Therefore, in this study, we assessed two common TLR-4 single nucleotide polymorphisms (SNPs), Asp299Gly and Thr399Ile, in Egyptian patients with COVID-19 and evaluated their frequency concerning disease severity, progression, and cytokine storm.

Methodology

Ethical Approval and Consent to Participate

The current study protocol was approved by the Research Ethics Committee (REC) of Ain Shams University Faculty of Medicine (FWA 00017585). We explained procedures to all participants and obtained their written informed consent before participation in the study. This study adheres to the Declaration of Helsinki.

Sample Size Calculation

We calculated sample size using STATA program version 17B setting the power at 80% and α error at 0.05. Suppose the expected odds ratio in the diseased group is 0.3 and the proportion of exposure in the control group is 0.09. In that case, the ratio of cases-to-controls is 0.21 according to

data revealed by Al-Qahtani et al, then 98 cases were needed in each group.

Study Settings and Subjects

In this cross-sectional study, we selected 150 severe and 150 non-severe COVID-19 adult patients. COVID-19 infection was confirmed by positive real-time reverse transcription-polymerase-chain reaction (RT-PCR) for respiratory swabs. Based on the World Health Organization (WHO) interim guidance, patients were diagnosed and categorized according to their disease severity.¹⁰ The severe group ($n = 150$) included patients with one or more of the following: respiratory distress, respiratory failure with mechanical ventilation, shock, organ failure syndrome, or ICU admission.

The non-severe group ($n = 150$) included patients with mild to moderate disease who did not match severe disease criteria, nonetheless, they had a positive nucleic acid test. We tracked each patient's outcome (hospital discharge or death). All participants were subjected to a thorough clinical and chest CT examination at the time of hospital admission.

COVID-19 Reporting and Data System (CO-RADS)

The CO-RADS score was assigned based on clinical data, laboratory test results, and chest CT records of patients. A CO-RADS of 0 indicates no infection; CO-RADS of 1 indicates a highly unlikely suspicion of infection with normal chest CT or non-infectious disease findings; a CO-RADS of 2 indicates low suspicion with chest CT findings of other infections; a CO-RADS of 3 indicates indeterminate COVID-19 suspicion with chest CT abnormalities indicating infection but not sure of COVID-19; a CO-RADS of 4 indicates high suspicion of infection with suspicious CT findings but not typical as multifocal consolidations, unilateral ground glass, or confluent; a CO-RADS of 5 indicates high suspicion with typical chest CT findings, whereas CO-RADS of 6 reflects positive RT-PCR SARS-CoV2 infection.¹¹

Chest Computed Tomography Severity Scoring (CT-SS)

The involvement of each lung lobe was visually assessed and rated from 0 to 5 (degree of involvement: 0–5%, 5–25%, 26–49%, 50–75%, and >75%). The individual lobar grades were summed to get the final score (range 0–25).^{12,13}

Routine Laboratory Investigations

From each participant on their admission, 4 mL of blood was collected via venipuncture under strict aseptic conditions and divided into two vacutainer tubes: 1) an EDTA-containing tube that was stored at -80°C until genotyping of TLR-4 by PCR restriction fragment length polymorphism (PCR-RFLP), and 2) a plain tube with no additives from which serum was collected after separation by centrifugation at $3500 \times g$ for 15 minutes. The separated sera were stored at -80°C until IL-6 analysis by enzyme-linked immunosorbent assay (ELISA).

Interleukin-6 Serum Level Measurement

Serum levels IL-6 were assessed using human ELISA kits supplied by CUSABIO Technology LLC, Houston, TX 77054, United States (Catalog Number: CSB-E04638h) following the manufacturer's instructions, with a measuring range of 7.8 pg/mL-500 pg/mL.

Molecular Analysis of TLR-4 SNPs

Two TLR-4 Exon SNPs rs4986790 (896A/G, Asp299Gly) and rs4986791 (1196C/T, Thr399Ile) were analyzed using the PCR-RFLP technique. Human DNA amplification from EDTA whole blood, without prior nucleic acid extraction, was performed using the Phusion™ Blood Direct PCR kit (Thermo Scientific™, USA, Cat. no.: MAN0012900) and primers for the human TLR-4 gene¹⁴ (Table 1).

According to manufacturer's instructions, the reaction mixture contained:

- 10 μL of $2\times$ Phusion Blood Direct PCR Master Mix.
- 1 μL of whole blood.
- 0.5 μM of each primer.
- 0.6 μL of 50 mM MgCl_2 in a 20 μL -final volume by RNase-Free Water.

The cycling conditions were as follows: initial denaturation at 98°C for 5 min, followed by 40 cycles of denaturation at 98°C for 1 s, annealing at 62°C (for Asp299Gly) or 60°C (for Thr399Ile) for 30 s, and extension at 72°C for 30 s, and final extension at 72°C for 5 min and cooling to 4°C . Subsequently, the 188bp (of Asp299Gly) and 124bp (of Thr399Ile) PCR products were treated with *Nco*I and *Hinf*I restriction enzymes (Thermo Scientific™, USA) overnight at 37°C . The digested products were separated by electrophoresis in a 2.5% agarose gel and detected by ethidium bromide using an Ultra-Violet Transilluminator (Syngene, Frederick, Maryland, USA). Lengths of Asp299Gly restriction fragments were 188bp in

Table 1 Sequences of Primers as Used for PCR and Restriction Enzymes and Length of the Restriction Fragments

TLR-4 SNPs	Primers	Annealing Temperature ($^{\circ}\text{C}$)	Restriction Enzyme	Product Length
rs4986790 (896A/G, Asp299Gly)	F: 5'AGCATACTTAGACTACTACCTCCATG-3' R: 5'-GAGAGATTTGAGTTTCAATGTGGG-3'	62°C	<i>Nco</i> I	-Asp299Asp (AA) 188bp -Gly299Gly (GG) 168 bp + 20 bp -Asp299Gly (AG) 188 bp + 168 bp + 20 bp
rs4986791 (1196C/T, Thr399Ile)	F: 5'-GGTTGCTGTTCTCAAAGTGATTTGGGAGAA-3' R: 5'-GGAAATCCAGATGTTCTAGTTGTTCTAAGCC-3'	60°C	<i>Hinf</i> I	- Thr399Thr (CC) 124bp - Ile399Ile (TT) 98 bp + 26 bp - Thr399Ile (CT) 124bp + 98 bp + 26 bp

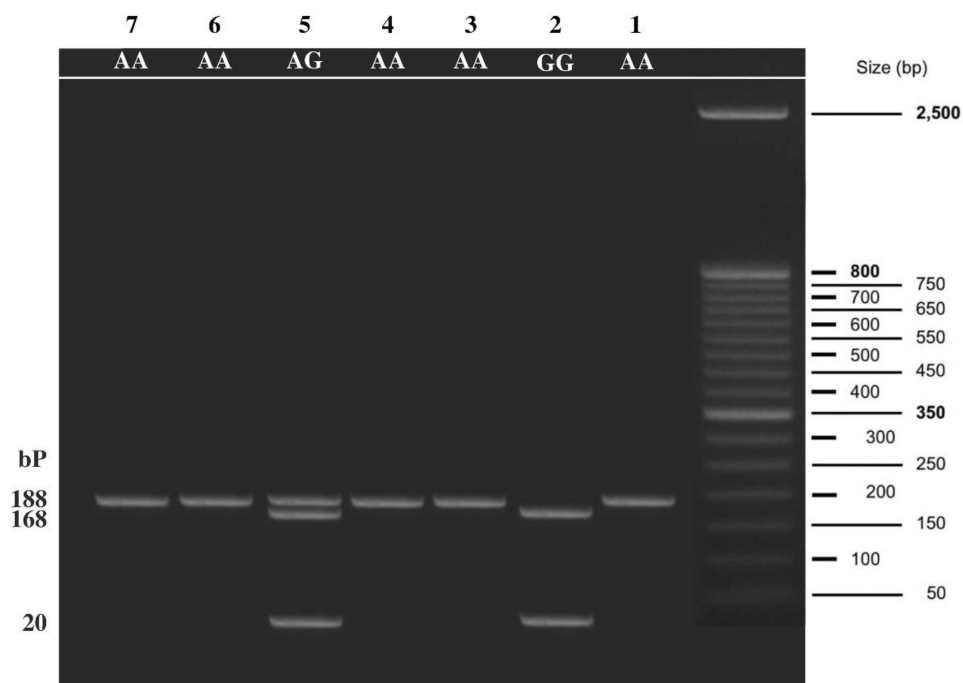


Figure 1 Agarose gel electrophoresis showing TLR-4 (Asp299Gly) genotyping by PCR-RFLP. Lane 1: wild Asp299Asp (AA) genotype at 188 bp (unrestricted); Lane 2: mutant Gly299Gly (GG) genotype at 168 and 20 bp; Lane 5: mutant Asp299Gly (AG) genotype at 188, 168 and 20bp.

the wild Asp299Asp (AA) genotype, 168 bp + 20 bp in the mutant Gly299Gly (GG) genotype, and 188 bp + 168 bp + 20 bp in the heterozygous mutant Asp299Gly (AG) genotype (Figure 1) while in Thr399Ile restriction fragment lengths were 124bp in the wild Thr399Thr (CC) genotype, 98 bp + 26 bp in the mutant Ile399Ile (TT) genotype, and 124bp + 98 bp + 26 bp in the heterozygous mutant Thr399Ile (CT) genotype (Table 1 and Figure 2).¹⁵

Statistical Analysis

We used IBM SPSS Statistics (V. 26.0, IBM Corp., 2019) to analyze the data. The median, percentiles, numbers, and percentages were used to represent the data. With respect to the comparisons, the Chi-square, Wilcoxon Rank Sum, and Kruskal-Wallis tests were utilized. Fisher's exact test was used to analyze the associations of TLR4 SNPs with COVID-19 clinical characteristic severity and in-hospital mortality, as well as odds ratios (OR) and 95% confidence intervals (CI) were used to interpret the results. At <0.05, the probability of error is significant.

Results

Characteristics of the Study Subjects

In this study we selected 150 severe and 150 non-severe COVID-19 adult patients. Patients in the severe group

were significantly older ($P < 0.001$). The male-to-female ratio did not differ significantly between the two groups ($P = 0.288$). The most common related co-morbidities were diabetes and hypertension. Eighty percent (120/150) of the severe group had a severe CT score, compared to only 12% (18/150) of the non-severe group. There was no significant difference in ICU admission rates between the study patients ($P = 0.533$). The severe patients' median (IQR) hospital stay was significantly longer than non-severe patients (19 days (14–22) vs 8 days (5–14); $P < 0.001$). Severe patients had a substantially higher in-hospital death rate than non-severe patients (25.3% (38/150) vs 6.7% (10/150); $P < 0.001$) and showed the highest serum levels of IL-6 (54 pg/mL (30–88) vs 17 pg/mL (11–23.5); $P < 0.001$). The baseline features of the included patients are shown in Table 2.

TLR-4 SNPs and Risk of Severe COVID-19

Table 3 shows the correlations between TLR-4 Asp299Gly and Thr399Ile SNPs (genotypes, alleles and haplotypes) and the risk of severe COVID-19. The wild Asp299Asp (AA) and Thr399Thr (CC) genotypes were the most common genotypes among the study groups and were associated with a significant negative risk of severe COVID-19

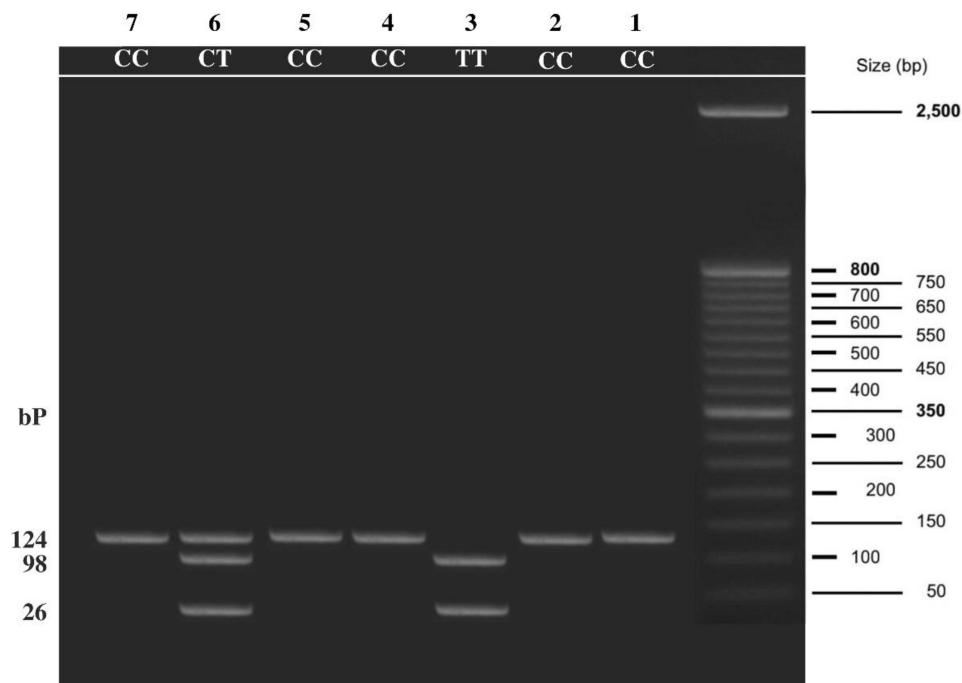


Figure 2 Agarose gel electrophoresis showing TLR-4 (Thr399Ile) genotyping by PCR-RFLP. Lane 1: wild Thr399Thr (CC) genotype at 124 bp (unrestricted); Lane 3: mutant Ile399Ile (TT) genotype at 98 and 26 bp; Lane 6: mutant Thr399Ile (CT) genotype at 124, 98 and 26 bp.

(Asp299Asp (AA): OR 0.28; 95% CI 0.17–0.47; $P < 0.001$ and Thr399Thr (CC): OR 0.27; 95% CI 0.15–0.49; $P < 0.001$). The homozygous mutant Gly299Gly (GG) genotype was detected in 6.7% (10/150) of the severe patients but not in any non-severe patients. On the contrary, the Ile399Ile (TT) genotype was found in 4.0% (6/150) of the non-severe patients and 5.3% (8/150) of the severe patients, but it did not represent a significant risk for severe COVID-19 (OR 0.73; 95% CI 0.25–2.18; $P = 0.584$). Frequency of the heterozygous mutant Asp299Gly (AG) and Thr399Ile (CT) genotypes were significantly higher in the severe group compared to the non-severe (42.7% and 28.0% vs 22.0% and 8.0%, respectively), both of which were associated with a significant positive risk of severe COVID-19 (Asp299Gly (AG) genotype: OR 2.63; 95% CI 1.59–4.36; $P < 0.001$ and Thr399Ile (CT) genotype: OR 4.47; 95% CI 2.24–8.90; $P < 0.001$).

The minor 299Gly (G) and 399Ile (T) alleles were associated with a significant positive risk of severe COVID-19 (OR 3.14; 95% CI 2.02–4.88; $P < 0.001$ and OR 2.75; 95% CI 1.66–4.57; $P < 0.001$, respectively), their frequency in the severe group were 71.8% (84/150) and 70.7% (58/150), respectively.

Furthermore, haplotype analysis was used to examine the additive effect of both SNPs. The frequency of the Asp299-

Thr399 (A-C) haplotype was 57.1% in the non-severe group and 42.9% in the severe group; they were associated with a significant negative risk of severe COVID-19 (OR 0.32; 95% CI 0.21–0.47; $P < 0.001$). We found significant positive associations between- Asp299-Ile399 (A-T), Gly299-Thr399 (G-C) and Gly299-Ile399 (G-T) haplotypes and the risk of severe COVID-19 with a higher frequency in the severe group compared to the non-severe cohort (Asp299-Ile399 (A-T): 65.0% vs 35.0%; OR: 1.93; 95% CI: 0.99–3.79; $P = 0.049$), (Gly299-Thr399 (G-C): 69.3% vs 30.7%; OR: 2.52; 95% CI: 1.50–4.24; $P < 0.001$) and (Gly299-Ile399 (G-T): 76.2% vs 23.8%; OR: 3.46; 95% CI: 1.67–7.17; $P < 0.001$), respectively.

TLR-4 (Asp299Gly and Thr399Ile) Genotypes and Levels of IL-6

Regarding IL-6 levels, we found significant differences between TLR-4 Asp299Gly and Thr399Ile genotypes. The lowest IL-6 levels were associated with the wild Asp299Asp (AA) and Thr399Thr (CC) genotypes (31 pg/mL (17–46) and 47.5 pg/mL (25–78), respectively) and significantly increased with the presence of the mutant 299Gly (G) and 399Ile (T) alleles to reach the highest levels in the Gly299Gly (GG) and the Ile399Ile (TT) genotypes (170 pg/mL (145–208.25) and 112 pg/mL (24–284.75), respectively) (Figure 3).

Table 2 Baseline Characteristics of COVID-19 Patients Included (Non-Severe vs Severe)

Variable		Non- Severe	Severe	p- value
		(n=150)	(n=150)	
Age (years)	Median (IQR)	42 (30–58)	55 (40–67)	< 0.001
Sex n, (%)	Male	55 (36.7%)	86 (57.3%)	0.288
	Female	95 (63.3%)	64 (42.7%)	
Comorbidities n, (%)	DM	60 (40.0%)	79 (52.7%)	0.028
	HTN	42 (28.0%)	78 (52.0%)	< 0.001
	IHD	11 (7.3%)	34 (22.7%)	0.064
	CLD	38 (25.3%)	22 (14.7%)	0.005
	CKD	4 (2.7%)	11 (7.3%)	< 0.001
Symptoms n, (%)	Cough	112 (74.7%)	106 (70.7%)	0.437
	Diarrhea	12 (8.0%)	22 (14.7%)	0.069
	Dyspnea	104 (69.3%)	112 (74.7%)	0.304
	Fever	100 (66.7%)	100 (66.7%)	1.000
	Fatigue	128 (85.3%)	138 (92.0%)	0.069
	Arthralgia	103 (68.7%)	120 (80.0%)	0.025
	Myalgia	108 (72.0%)	116 (77.3%)	0.288
	Rhinitis	18 (12.0%)	6 (4.0%)	0.011
	Loss of taste	14 (9.3%)	10 (6.7%)	0.395
	Sore throat	18 (12.0%)	26 (17.3%)	0.192
CT-SS	Median (IQR)	5 (0–15)	20 (18–21)	< 0.001
CORADS	Median (IQR)	3 (1–5)	5 (5–6)	< 0.001
IL-6 (pg/mL)	Median (IQR)	17 (11–23.5)	54 (30–88)	< 0.001
ICU admission n, (%)	Positive	44 (29.3%)	49 (32.7%)	0.533
	Negative	106 (70.7%)	101 (67.3%)	
Hospital stay (days)	Median (IQR)	8 (5–14)	19 (14–22)	< 0.001
Fate	Discharged	140 (93.3%)	112 (74.7%)	< 0.001
	Died	10 (6.7%)	38 (25.3%)	

Note: Bold values denote statistical significance at the $P < 0.05$ level.

Abbreviations: CO-RADS, COVID-19 Reporting and Data System; CT-SS, Chest Computed Tomography Severity Scoring; ICU, Intensive care unit; IL-6, Interleukin-6.

Discussion

Data on immunological modulation, signaling pathways, and proinflammatory cytokines in SARS-CoV-2 infection can be used as a relevant platform for future pharmacological and vaccine studies.¹⁶ In the present study, which included 300 Egyptian COVID-19 patients, we found that severe COVID-19 was associated with TLR-4 (Asp299Gly and Thr399Ile) SNPs. The minor 299Gly (G) and 399Ile (T) alleles have been highly linked to severe COVID-19 and elevated IL-6 levels. Moreover, haplotype analysis showed that the co-segregation of both mutations was associated with a significantly a high risk for severe disease.

The TLR-4 gene is made up of four exons located on chromosome 9q32–33. TLR-4 protein is polymorphic, containing 12 single amino acid variations that occur infrequently. Arbor and colleagues described two polymorphic SNPs: +896A/G substitution, which replaces a conserved aspartic acid residue with glycine at amino acid (aa) 299 +1196C/T substitution, which replaces a threonine with isoleucine at (aa) 399.¹⁷ Both SNPs have been linked to reduced sensitivity to inhaled endotoxin. Furthermore, transfected cells with the minor allele of these SNPs had a decreased sensitivity to LPS and significantly lower production of the TLR-4 protein on the surface of epithelial cells in a cell culture

Table 3 Frequency of TLR-4 Asp299Gly and Thr399Ile SNPs (Genotypes, Alleles, and Haplotypes) and Their Association with the Risk of Severe COVID-19

TLR-4 Polymorphism		Non-Severe	Severe	P-value	Severe COVID-19 Risk		P-value	
		(n=150)	(n=150)		Odds Ratio	95% CI		
rs4986790 genotypes	Asp299Asp (AA)	117	76	< 0.001	0.28	(0.17–0.47)	< 0.001	
		(78.0%)	(50.7%)					
	Asp299Gly (AG)	33	64		2.63	(1.59–4.36)		
		(22.0%)	(42.7%)					
	Gly299Gly (GG)	0.0	10		NC			
		(0.0%)	(6.7%)					
rs4986791 genotypes	Thr399Thr (CC)	132	100	< 0.001	0.27	(0.15–0.49)	< 0.001	
		(88.0%)	(66.7%)					
	Thr399Ile (CT)	12	42		4.47	(2.24–8.90)		
		(8.0%)	(28.0%)					
	Ile399Ile (TT)	6	8		0.73	(0.25–2.18)		0.584
		(4.0%)	(5.3%)					
rs4986790 alleles	Asp Allele (A)	267	216	< 0.001	0.31	(0.20–0.49)	< 0.001	
		(55.3%)	(44.7%)					
	Gly Allele (G)	33	84		3.14	(2.02–4.88)		
		(28.2%)	(71.8%)					
rs4986791 alleles	Thr Allele (C)	276	242	< 0.001	0.36	(0.21–0.60)	< 0.001	
		(53.3%)	(46.7%)					
	Ile Allele (T)	24	58		2.75	(1.66–4.57)		
		(29.3%)	(70.7%)					
Haplotype	A-C (Asp299-Thr399)	253	190	< 0.001	0.32	(0.21–0.47)	< 0.001	
		(57.1%)	(42.9%)					
	A-T (Asp299-Ile399)	14	26		1.93	(0.99–3.79)		0.049
		(35.0%)	(65.0%)					
	G-C (Gly299-Thr399)	23	52		2.52	(1.50–4.24)		< 0.001
		(30.7%)	(69.3%)					
	G-T (Gly299-Ile399)	10	32		3.46	(1.67–7.17)		< 0.001
		(23.8%)	(76.2%)					

Note: Bold values denote statistical significance at the P < 0.05 level.

model.¹⁸ The generation of cytokines and inflammatory mediators like fibrinogen is also affected by these SNPs.¹⁹

A multitude of studies have investigated the association between these SNPs and different diseases. Although it seems that the minor alleles of TLR-41196C>T and

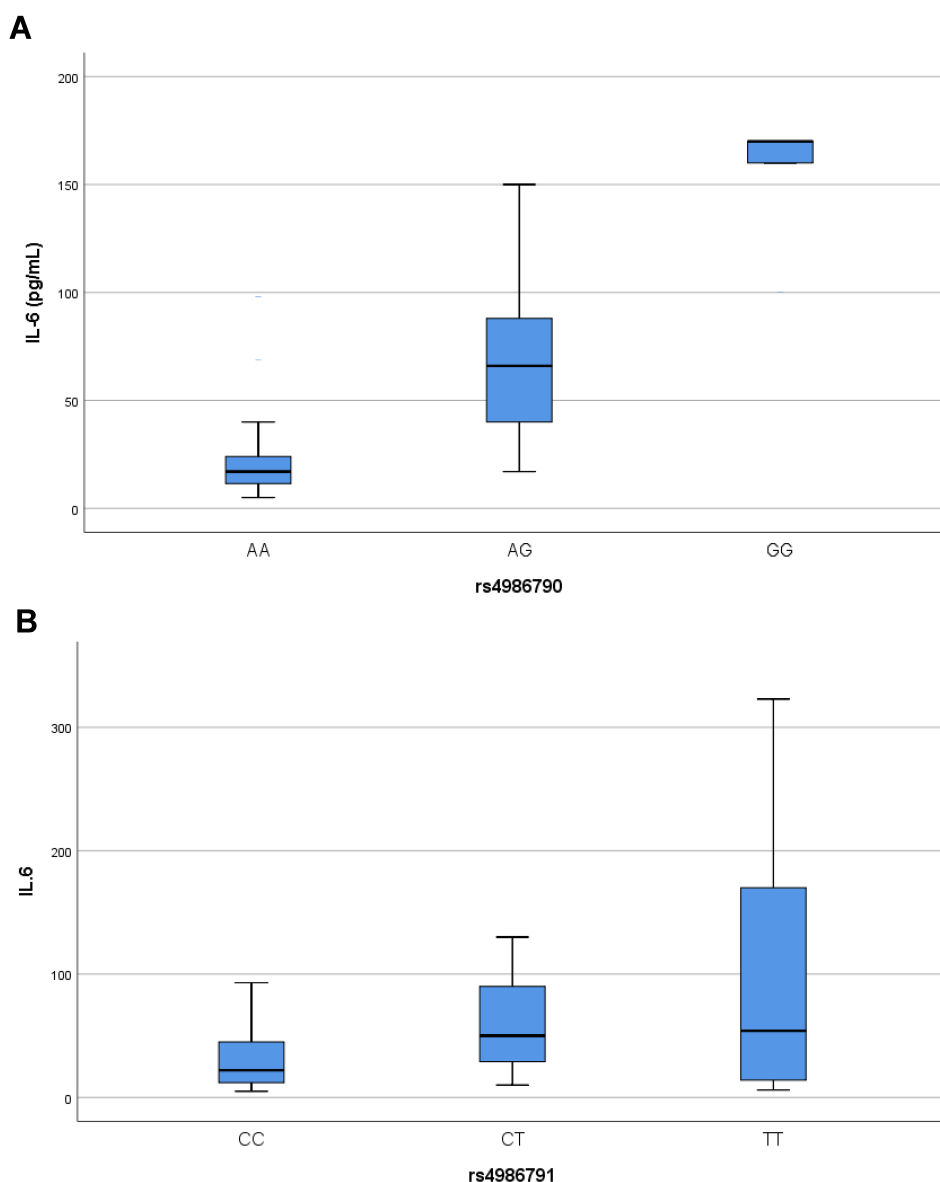


Figure 3 Box plot representing the range, median, and quartiles of IL-6 levels in pg/mL according to the different TLR-4 genotypes (tested by Kruskal–Wallis test) (A) rs4986790: Asp299Asp (AA):31 (17–46); Asp299Gly (AG):80 (60.25–96.75); Gly299Gly (GG): 170 (145–208.25); $P < 0.001$ (B) rs4986791: Thr399Thr (CC): 47.5 (25–78); Thr399Ile (CT): 76 (40–100); Ile399Ile (TT): 112 (24–284.75); $P=0.013$.

896A>G SNPs predispose to several pathologies such as; gram-negative septic shock,²⁰ premature birth,²¹ Crohn's disease,²² chronic osteomyelitis,²³ urinary tract infection in children,²⁴ respiratory syncytial virus (RSV) bronchiolitis in infants,²⁵ Candida sepsis,²⁶ chronic sarcoidosis,²⁷ severe H-pylori infection and duodenal ulceration,²⁸ some reports described a protective role for these SNPs against the development of diseases, such as; gastric lymphoma,²⁹ neuropathy in diabetic patients,³⁰ atherosclerosis with increased carotid artery compliance in young adults,¹⁹ acute allograft rejection,³¹ and Alzheimer's disease.³² In addition, minor alleles have not been directly linked to type 2 diabetes, but

they may raise the risk of diabetic nephropathy through affecting levels of triglycerides and blood sugar.³³

The SARS-CoV-2 genome encodes four proteins; spike (S), membrane (M), envelope (E), and nucleocapsid (N). The virus invades host cells by interacting its S protein with the angiotensin-converting enzyme-2 (ACE-2) receptor on the host's cell membrane. It then enters the cell by endocytosis and uses its RNA and host machinery for replication.³⁴ Because the ACE-2 is found in most vital organs and serves as a receptor for COVID-19, SARS-CoV-2 affects most organs, including the lungs, causing inflammation and fibrosis, which may affect vital functions, either temporarily or

permanently, and sometimes result in death.³⁵ Binding of viral S glycoprotein to ACE-2 on type II alveolar cells, which is responsible for the synthesis of pulmonary surfactant, results in cell lysis, exposing the extracellular binding sites of TLR-4 on lung epithelial cells. The exposed TLR-4 interacts with the viral S protein, facilitating viral entry to the neighboring cells, either directly or indirectly, through an increase in cell surface expression of ACE-2 by interferons.⁵ TLR-4-S protein interaction cause downstream signaling events towards the MYD88 dependent pro-inflammatory pathway, rather than the TRIF/TRAM dependent interferon pathway, with the production of innate pro-inflammatory cytokines (IL-1, IL-6, tumor necrosis factor- α , and type 1 interferon α , β) by macrophages, stimulating a robust innate immune response, and deflecting the adaptive immunity into a T-helper (Th)1/Th2 response in an attempt to clear the virus.⁵ In addition, an exaggerated Th2 cytokine response, especially IL-6, mediated by eosinophils, basophils, and mast cells, has been linked to immune response dysregulation in COVID-19, which further exacerbates the inflammatory reaction.³⁶

Thus, by binding TLR-4, SARS-CoV-2 can trigger a rampant antiviral inflammatory state; if uncontrolled, especially in genetically predisposed individuals, it can ultimately end into a profound inflammatory reaction characterized by an increase in cytokines chemokines, and interferons, known as cytokine storm.³⁷ The cytokine storm plays a crucial role in severe COVID-19 pathogenesis; it can lead to apoptosis of epithelial and endothelial cells, vascular leakage, and finally can end up into acute lung injury and ARDS, with fatal outcomes.³⁸ Interestingly, in murine models of ARDS, including SARS-CoV-2, genetic inactivation of the TLR-4 gene was associated with the reduction in ARDS development.⁸ Furthermore, myocarditis, thrombosis, and multi-organ failure in severe COVID-19 cases have been linked to TLR-4-SARS-CoV-2 interaction, with aberrant signaling in genetically predisposed patients, resulting in a hyper-inflammatory state.⁸

Finally, considering the well-established role of TLR-4 in the immunological responses to SARS-CoV-2, mutations in the TLR-4 gene may explain, at least in part, the broad spectrum of COVID-19 manifestations, as well as the unexplained progression of the disease in some patients. In addition, since TLR-4 antagonists have previously been trialed in sepsis and other antiviral contexts,⁵ TLR-4 appears to be a promising therapeutic target in genetically predisposed COVID-19 patients. Also, as several vaccines targeting SARS-CoV-2, S glycoprotein have been developed, the SNP within the TLR-4 gene can affect the TLR4-S protein interaction, and thus constituting a significant determinant of immune

responses COVID-19 vaccines. Testing for SNP may help identify suitable immunization strategies for those individuals with incomplete responses to a particular vaccine strategy due to their genetic background.³⁹

This study has limitations. First, the disproportionate sample size between the different genotypes and alleles hindered us from comparing their various clinical, laboratory, and radiological characteristics. Second, many coexisting non-genetic risk factors for COVID-19 severity were present in our patients and could have affected our results. Third, our study was conducted in a single hospital setting, offering a small representative population. Fourth, we measured only IL-6 as an indicator of the cytokine storm. More comprehensive studies on the correlation of TLR-4 SNPs with other inflammatory cytokines and the intracellular pathways are still required to understand better the role of TLR-4 gene mutations in COVID-19 pathogenesis.

Conclusions

Our study found that the TLR-4 SNPs (Asp299Gly) and (Thr399Ile) were associated with COVID-19 severity, cytokine storm, and mortality. Testing for TLR-4 SNPs may be helpful for early prediction of the course of the disease and early identification of patients who may benefit from the introduction of TLR-4 antagonists in the treatment regimen.

Abbreviations

ACE-2, angiotensin-converting enzyme-2; ARDS, adult respiratory distress syndrome; CO-RADS, COVID-19 Reporting and Data System; COVID-19, Corona virus disease 2019; CT, computed tomography; CT-SS, Chest Computed Tomography Severity Scoring; DAMPs, Damage-associated molecular patterns; EDTA, ethylenediaminetetraacetic acid; ELISA, enzyme-linked immunosorbent assay; ICU, intensive care unit; LPS, lipopolysaccharide; PAMPs, pathogen-associated molecular patterns; PCR-RFLP, polymerase-chain reaction-restriction fragment length polymorphism; PRR, pattern recognition receptor; RT-PCR, real-time reverse transcription-polymerase-chain reaction; SARS-CoV-2, severe acute respiratory syndrome coronavirus-2; SNPs, single nucleotide polymorphisms; Th, T-helper; TLR-4, toll-like receptor 4; WHO, World Health Organization (WHO).

Data Sharing Statement

All the data needed to support the findings will be provided upon request.

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Disclosure

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References

- Cascella M, Rajnik M, Aleem A, Dulebohn SC, Di Napoli R. Features, evaluation, and treatment of coronavirus (COVID-19). In: *StatPearls [Internet]*. Treasure Island (FL): StatPearls Publishing; January, 2021. PMID: 32150360.
- Cron RQ, Caricchio R, Chatham WW. Calming the cytokine storm in COVID-19. *Nat Med*. 2021;27(10):1674–1675. PMID: 34480126. doi:10.1038/s41591-021-01500-9
- Tan L, Wang Q, Zhang D, et al. Lymphopenia predicts disease severity of COVID-19: a descriptive and predictive study. *Signal Transduct Target Ther*. 2020;5(1):33. PMID: 32296069; PMCID: PMC7100419. doi:10.1038/s41392-020-0148-4
- Fouad SH, Allam MF, Ibrahim S, et al. ICU admission of COVID-19 patients: identification of risk factors. *Egypt J Anaesth*. 2021;37(1):202–207. doi:10.1080/11101849.2021.1919433
- Aboudounya MM, Heads RJ. COVID-19 and Toll-Like Receptor 4 (TLR4): SARS-CoV-2 may bind and activate TLR4 to increase ACE2 expression, facilitating entry and causing hyperinflammation. *Mediators Inflamm*. 2021;2021:8874339. PMID: 33505220; PMCID: PMC7811571. doi:10.1155/2021/8874339
- Satoh T, Akira S. Toll-like receptor signaling and its inducible proteins. *Microbiol Spectr*. 2016;4(6). PMID: 28084212. doi:10.1128/microbiolspec.MCHD-0040-2016
- Kate Gadaneq L, Qaradakhi T, Renee McSweeney K, Ashiana Ali B, Zulli A, Apostolopoulos V. Dual targeting of Toll-like receptor 4 and angiotensin-converting enzyme 2: a proposed approach to SARS-CoV-2 treatment. *Future Microbiol*. 2021;16:205–209. PMID: 33569984; PMCID: PMC7885526. doi:10.2217/fmb-2021-0018
- Onofrio L, Caraglia M, Facchini G, Margherita V, Placido S, Buonerba C. Toll-like receptors and COVID-19: a two-faced story with an exciting ending. *Future Sci OA*. 2020;6(8):FSO605. PMID: 32974046; PMCID: PMC7434222. doi:10.2144/fsoa-2020-0091
- Conte C. Possible link between SARS-CoV-2 infection and Parkinson's disease: the role of toll-like receptor 4. *Int J Mol Sci*. 2021;22(13):7135. PMID: 34281186; PMCID: PMC8269350. doi:10.3390/ijms22137135
- World Health Organization. Coronavirus disease (COVID-19) Weekly Epidemiological Update and Weekly Operational Update. Available from: <https://www.who.int/emergencies/diseases/novel-coronavirus-2019/situation-reports>. Accessed November 22, 2021.
- Prokop M, Van Everdingen W, Van Rees Vellinga T, et al. CO-RADS : a categorical CT assessment scheme for patients suspected of having COVID-19 definition and evaluation. *Radiology*. 2020;296(2):E97–E104. PMID: 32339082; PMCID: PMC7233402. doi:10.1148/radiol.2020201473
- Francone M, Iafrafe F, Masci GM, et al. Chest CT score in COVID-19 patients: correlation with disease severity and short-term prognosis. *Eur Radiol*. 2020;30(12):6808–6817. PMID: 32623505; PMCID: PMC7334627. doi:10.1007/s00330-020-07033-y
- Pan F, Ye T, Sun P, et al. Time course of lung changes at chest CT during recovery from coronavirus disease 2019 (COVID-19). *Radiology*. 2020;295(3):715–721. PMID: 32053470; PMCID: PMC7233367. doi:10.1148/radiol.2020200370
- Senhaji N, Diakitè B, Serbati N, Zaid Y, Badre W, Nadifi S. Toll-like receptor 4 Asp299Gly and Thr399Ile polymorphisms: new data and a meta-analysis. *BMC Gastroenterol*. 2014;14:206. PMID: 25492126; PMCID: PMC4279599. doi:10.1186/s12876-014-0206-x
- Folwaczny M, Glas J, Török HP, Limbersky O, Folwaczny C. Toll-like receptor (TLR) 2 and 4 mutations in periodontal disease. *Clin Exp Immunol*. 2004;135(2):330–335. PMID: 14738464; PMCID: PMC1808953. doi:10.1111/j.1365-2249.2004.02383.x
- Zolfaghari Emameh R, Nosrati H, Eftekhari M, Falak R, Khoshmirsafa M. Expansion of single cell transcriptomics data of SARS-CoV infection in human bronchial epithelial cells to COVID-19. *Biol Proced Online*. 2020;22:16. PMID: 32754004; PMCID: PMC7377208. doi:10.1186/s12575-020-00127-3
- Trejo-de la O A, Hernández-Sancén P, Maldonado-Bernal C. Relevance of single-nucleotide polymorphisms in human TLR genes to infectious and inflammatory diseases and cancer. *Genes Immun*. 2014;15(4):199–209. PMID: 24622688. doi:10.1038/gene.2014.10
- Arbour NC, Lorenz E, Schutte BC, et al. TLR4 mutations are associated with endotoxin hyporesponsiveness in humans. *Nat Genet*. 2000;25(2):187–191. PMID: 10835634. doi:10.1038/76048
- Kiechl S, Lorenz E, Reindl M, et al. Toll-like receptor 4 polymorphisms and atherosclerosis. *N Engl J Med*. 2002;347(3):185–192. PMID: 12124407. doi:10.1056/NEJMoa012673
- Barber RC, Aragaki CC, Rivera-Chavez FA, Purdue GF, Hunt JL, Horton JW. TLR4 and TNF-alpha polymorphisms are associated with an increased risk for severe sepsis following burn injury. *J Med Genet*. 2004;41(11):808–813. PMID: 15520404; PMCID: PMC1383768. doi:10.1136/jmg.2004.021600
- Lorenz E, Hallman M, Marttila R, Haataja R, Schwartz DA. Association between the Asp299Gly polymorphisms in the Toll-like receptor 4 and premature births in the Finnish population. *Pediatr Res*. 2002;52(3):373–376. PMID: 12193670. doi:10.1203/00006450-200209000-00011
- Browning BL, Huebner C, Petermann I, et al. Has toll-like receptor 4 been prematurely dismissed as an inflammatory bowel disease gene? Association study combined with meta-analysis shows strong evidence for association. *Am J Gastroenterol*. 2007;102(11):2504–2512. PMID: 17850411. doi:10.1111/j.1572-0241.2007.01463.x
- Montes AH, Asensi V, Alvarez V, et al. The Toll-like receptor 4 (Asp299Gly) polymorphism is a risk factor for Gram-negative and haematogenous osteomyelitis. *Clin Exp Immunol*. 2006;143(3):404–413. PMID: 16487238; PMCID: PMC1809620. doi:10.1111/j.1365-2249.2005.03002.x
- Karoly E, Fekete A, Banki NF, et al. Heat shock protein 72 (HSPA1B) gene polymorphism and Toll-like receptor (TLR) 4 mutation are associated with increased risk of urinary tract infection in children. *Pediatr Res*. 2007;61(3):371–374. Erratum in: *Pediatr Res*. 2007 Oct;62(4):482. PMID: 17314700. doi:10.1203/pdr.0b013.e318030d1f4
- Tulic MK, Hurrelbrink RJ, Prêle CM, et al. TLR4 polymorphisms mediate impaired responses to respiratory syncytial virus and lipopolysaccharide. *J Immunol*. 2007;179(1):132–140. PMID: 17579031. doi:10.4049/jimmunol.179.1.132
- Van der Graaf CA, Netea MG, Morrè SA, et al. Toll-like receptor 4 Asp299Gly/Thr399Ile polymorphisms are a risk factor for Candida bloodstream infection. *Eur Cytokine Netw*. 2006;17(1):29–34. PMID: 16613760.
- Pabst S, Baumgarten G, Stremmel A, et al. Toll-like receptor (TLR) 4 polymorphisms are associated with a chronic course of sarcoidosis. *Clin Exp Immunol*. 2006;143(3):420–426. PMID: 16487240; PMCID: PMC1809614. doi:10.1111/j.1365-2249.2006.03008.x

28. Trejo-de la O A, Torres J, Pérez-Rodríguez M, et al. TLR4 single-nucleotide polymorphisms alter mucosal cytokine and chemokine patterns in Mexican patients with Helicobacter pylori-associated gastroduodenal diseases. *Clin Immunol.* 2008;129(2):333–340. PMID: 18755634. doi:10.1016/j.clim.2008.07.009
29. Hellmig S, Fischbach W, Goebeler-Kolve ME, Fölsch UR, Hampe J, Schreiber S. Association study of a functional Toll-like receptor 4 polymorphism with susceptibility to gastric mucosa-associated lymphoid tissue lymphoma. *Leuk Lymphoma.* 2005;46(6):869–872. PMID: 16019531. doi:10.1080/1042819050086451
30. Rudofsky G Jr, Reismann P, Witte S, et al. Asp299Gly and Thr399Ile genotypes of the TLR4 gene are associated with a reduced prevalence of diabetic neuropathy in patients with type 2 diabetes. *Diabetes Care.* 2004;27(1):179–183. PMID: 14693986. doi:10.2337/diacare.27.1.179
31. Palmer SM, Burch LH, Mir S, et al. Donor polymorphisms in Toll-like receptor-4 influence the development of rejection after renal transplantation. *Clin Transplant.* 2006;20(1):30–36. PMID: 16556150. doi:10.1111/j.1399-0012.2005.00436.x
32. Minorette P, Gazzaruso C, Vito CD, et al. Effect of the functional toll-like receptor 4 Asp299Gly polymorphism on susceptibility to late-onset Alzheimer's disease. *Neurosci Lett.* 2006;391(3):147–149. PMID: 16157451. doi:10.1016/j.neulet.2005.08.047
33. Khaghanzadeh N, Naderi N, Pournasrollah N, Farahbakhsh E, Kheirandish M, Samiei A. TLR4 polymorphisms (896A>G and 1196C>T) affect the predisposition to diabetic nephropathy in type 2 diabetes mellitus. *Diabetes Metab Syndr Obes.* 2020;13:1015–1021. PMID: 32308451; PMCID: PMC7138628. doi:10.2147/DMSO.S238942
34. Malik YA. Properties of Coronavirus and SARS-CoV-2. *Malays J Pathol.* 2020;42(1):3–11. PMID: 32342926.
35. Zolfaghari Emameh R, Falak R, Bahreini E. Application of system biology to explore the association of neprilysin, angiotensin-converting enzyme 2 (ACE2), and carbonic anhydrase (CA) in pathogenesis of SARS-CoV-2. *Biol Proced Online.* 2020;22:11. PMID: 32572334; PMCID: PMC7302923. doi:10.1186/s12575-020-00124-6
36. Roncati L, Nasillo V, Lusenti B, Riva G. Signals of Th2 immune response from COVID-19 patients requiring intensive care. *Ann Hematol.* 2020;99(6):1419–1420. PMID: 32382776; PMCID: PMC7205481. doi:10.1007/s00277-020-04066-7
37. Brandão SCS, Ramos JOX, Dompieri LT, et al. Is Toll-like receptor 4 involved in the severity of COVID-19 pathology in patients with cardiometabolic comorbidities? *Cytokine Growth Factor Rev.* 2021;58:102–110. PMID: 32988728; PMCID: PMC7505161. doi:10.1016/j.cytogfr.2020.09.002
38. Tang Y, Liu J, Zhang D, Xu Z, Ji J, Wen C. Cytokine storm in COVID-19: the current evidence and treatment strategies. *Front Immunol.* 2020;11:1708. PMID: 32754163; PMCID: PMC7365923. doi:10.3389/fimmu.2020.01708
39. Pellegrino P, Falvella FS, Cheli S, Perrotta C, Clementi E, Radice S. The role of Toll-like receptor 4 polymorphisms in vaccine immune response. *Pharmacogenomics J.* 2016;16(1):96–101. PMID: 25823688. doi:10.1038/tpj.2015.21

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