

Cutaneous *Mycobacterium Abscessus* Infection Following Plastic Surgery: Three Case Reports

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Aim: *Mycobacterium abscessus* is ubiquitous in the environment and seldom causes infections in immunocompetent individuals. However, skin and soft tissue infections caused by *M. abscessus* have been reported in recent years. Additionally, the cutaneous infections or outbreaks post cosmetic surgery caused by *M. abscessus* have been increasing due to the popularity of plastic surgery. The main modes of transmission are through contaminated saline, disinfectants, or surgery equipment, as well as close contact between patients. This article describes three patients who were admitted to our hospital between November 2019 and October 2020. They presented with long-term non-healing wounds caused by *M. abscessus* infection after undergoing plastic surgery. Symptoms presented by the three patients included swelling, ulceration, secretion, and pain. After identification of *M. abscessus* with Ziehl-Neelsen staining and MALDI-TOF MS system, the patients were treated with surgical debridement and clarithromycin.

Conclusion: It is important to note that a long-term wound that does not heal, especially after plastic surgery, should raise suspicion for *M. abscessus* infection. The infection mechanism in these three patients may have been due to exposure to surgical equipment that was not properly sterilized or due to poor sterile technique by the plastic surgeon. To prevent such infections, it is important to ensure proper sterilization of surgical equipment and saline.

Keywords: *Mycobacterium abscessus*, cutaneous infection, plastic surgery, treatment, antibiotic resistance

Introduction

Skin and soft tissue infections caused by nontuberculous mycobacteria are on the rise. These ubiquitous acid-fast bacilli are found in the environment and primarily cause cutaneous infections after trauma, surgery, or cosmetic procedures. Abscesses, sporotrichoid nodules, or ulcers are common skin findings. Significant species implicated in these infections include *Mycobacterium marinum* and the rapidly growing mycobacterium: *Mycobacterium fortuitum*, *M. abscessus*, and *Mycobacterium chelonae*.¹ The three rapidly growing pathogenic species exhibit significant differences in their antimicrobial susceptibilities, making the identification of species crucial for clinical purposes.²

M. abscessus seldom leads to severe infections in immunocompetent individuals, but it has been reported to cause cutaneous infections in cases of trauma, tattoos, skin injuries, and surgery.³ It can also result in pulmonary or systemic infection in patients who are immunosuppressed.⁴ According to a 2021 review,⁵ the most common rapidly growing mycobacteria species causing skin and soft tissue infections was *M. abscessus* (184/475, 38.7%). Cases of skin and soft tissue infections caused by *M. abscessus* have been reported in various regions, including the USA, Taiwan, India, Switzerland, France, and Germany.⁵ In China, there have been sporadic reports of *M. abscessus* soft tissue infections following injections or plastic surgery.⁶⁻⁸ A total of 955 cases of *M. abscessus* soft tissue infections were reported in 60 studies by 2020.⁹ The infections were caused by trauma (76%), injections (54%), acupuncture (26%), and plastic surgery (15%). While 95% of patients fully recovered after treatment, two cases resulted in disseminated infections and death.⁹ It is worth noting that plastic surgery may increase the risk of *M. abscessus* infection.

Cutaneous infections caused by *M. abscessus* have been reported through various routes, including tattooing,¹⁰ liposuction,¹¹ skin transplantation,¹² rhytidectomy,¹³ hip augmentation,¹⁴ hair transplant,¹⁵ botulinum toxin injection,¹⁶ abdominal plastic surgery, and skin rejuvenation.¹⁷ Outbreaks of invasive infection have been reported in children's dental clinics and skin infections caused by acupuncture in traditional Chinese medical clinics.¹⁸ These infections were most common in departments specializing in cosmetic surgery, liposuction, and intravenous cell therapy. The transmission routes included contaminated saline, disinfectants, surgical instruments, and close contact with the patient.¹⁴

Due to *M. abscessus* resistance to various antibiotics, conventional antibiotic treatment may not prove effective and diagnosis and treatment are highly dependent on etiological findings. This article reports three cases of long-term *M. abscessus* cutaneous infection after plastic surgery. Results are presented to contribute to epidemiologic data and clinical treatments for *M. abscessus* soft tissue infection in future.

Materials and Methods

Culture and Identification of *M. Abscessus*

Samples of drainage fluid and wound secretion were cultured on blood agar, chocolate agar, and MacConkey agar (Autobio, China) and then incubated in a CO₂ incubator (Thermo Fisher Scientific, USA) at 35°C for several days. Ziehl-Neelsen staining was performed for preliminary examination. The *M. abscessus* strain was identified using the Autof ms 1000 MALDI-TOF MS system (Autobio, China) according to manufacturer's instructions.

Antimicrobial Susceptibility Testing

The susceptibility of the *M. abscessus* isolate to 12 antimicrobials was tested using the Sensititre RAPMYCO2 plate (Thermo Fisher Scientific, USA) according to the manufacturer's instructions. After incubation, the minimal inhibitory concentration (MIC) value was measured and the results were interpreted as susceptible, intermediate, or resistant based on the Clinical and Laboratory Standards Institute guideline M24-A2.¹⁹ Quality control was performed using *M. peregrinum* ATCC 700686.

Case I

Clinical Information

A 32-year-old female underwent bilateral augmentation mammoplasty with silicone implants in November 2019. Subsequently, she developed a wound infection on the left side with yellowish-white purulent discharge, although no pathogenic organisms were isolated from the discharge. Despite removing the bilateral breast implants in December 2019, the bilateral axillary wound continued to exude and did not heal. Ten days following the removal procedure, red and blue light anti-inflammatory was administered. In addition to increasing blood supply to the wound surface and reducing excessive inflammatory response, red and blue lights have been shown to inhibit multiple bacteria, accelerate wound healing, and reduce levels of pathogenic bacteria on the wound surface.²⁰ However, there was a persistent bilateral axillary wound dehiscence with a purulent yellow-green discharge. The wound exhibited erosive changes, and a 4 cm subcutaneous cavity was detected in the direction from the right armpit towards the breast (Figure 1).

Chest MRI revealed an encapsulated effusion in the posterior space of both pectoralis major muscles. Following admission, the patient underwent debridement, deformity correction, and pedicled composite tissue flap plasty on January 22, 2020. Necrotic tissue at the wound's edge was excised, though no pathogens could be isolated from the tissue sample.

The patient's right wound exhibited inadequate healing post-operation, and mild light yellow exudates were observed upon wound compression. Levofloxacin (0.5g QD, inv) was administered as an empirical treatment. On February 5, a chest MRI revealed multiple lesions near the upper marginal muscular space and beneath the armpit. A second debridement of the right armpit and negative pressure drainage for the ventricular septal defect (VSD) were performed on February 6 due to unsatisfactory results from dressing changes. Cefoperazone/sulbactam (1.5g BID, inv) and metronidazole (100mL, 3 days) were used for antibiotic therapy.

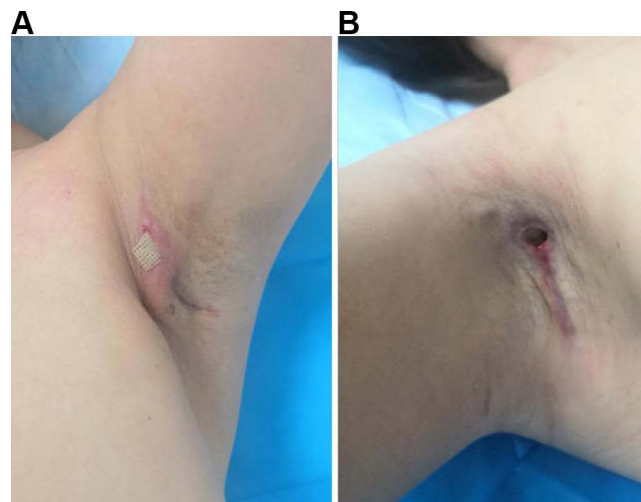


Figure 1 Bilateral axillary wound showed dehiscence at admission. **(A)** Despite the removal of bilateral breast implants and the use of red and blue light anti-inflammatory treatment, there was still evidence of bilateral axillary wound dehiscence with purulent yellow-green discharge. **(B)** A subcutaneous cavity was visible in the right armpit.

The patient's bilateral wounds in the axillary region exhibited poor healing progress after the second operation, particularly on the right side. Observations revealed the presence of pale red, bloody drainage fluid and some erosion on the left armpit. Additionally, the patient experienced swelling, pain, and ulceration with purulent secretions on the left breast fold (**Figure 2**). To address the non-healing wound that persisted unexpectedly, the third axillary debridement and VSD negative pressure drainage were performed on March 4. The drainage fluid on the right exhibited dark red coloration with flocculent necrosis, while the left was clear. After obtaining culture results, levofloxacin (0.5g QD, inv) and vancomycin (0.5g BID, inv) were prescribed as antibiotics.

Twenty days after the third surgery, the drainage tubes were removed, revealing a small amount of clear yellowish exudate at the drainage site. Prolonged hospitalization led to damage of the pectoralis major muscle and limitations in both upper limb mobility. Shortly after, a dark red area with a diameter of approximately 3cm surfaced in the right armpit (**Figure 3**). On March 13, the wound secretion was subjected to Ziehl-Neelsen staining, which resulted in a grade of ++++. Considering the possibility of skin tuberculosis, treatment with moxifloxacin and anti-tuberculosis quadruple drugs was initiated. However, the patient's wound still healed poorly and the symptoms did not improve.

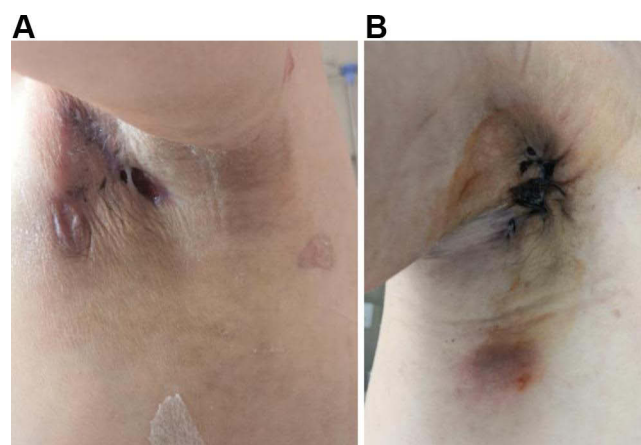


Figure 2 Skin ulceration and yellow pus after the second operation. **(A)** On the left breast fold, there was swelling, ulceration, and purulent secretions. **(A–B)** The bilateral axillary region showed poor healing after a second debridement and negative pressure drainage for VSD.



Figure 3 Dark redness, swelling, and pain are present on the right side following the third surgical drainage procedure.

Etiological Examination

The drainage fluid and wound secretion samples were taken and examined for pathogenic organisms during or after the debridement surgery. Sequentially, *Staphylococcus lugdunensis*, *Enterococcus faecalis*, *Staphylococcus aureus*, and *Corynebacterium jejuni* were isolated, and antibiotics were modified based on culture results. On March 15 and 31, the drainage fluid on the right side tested positive for Ziehl-Neelsen staining (Table 1). Small colonies were identified in drainage fluid sample after 3 days of culture. After 4 days of culture, rough and colorless colonies of *M. abscessus* were isolated (Figure 4). Antimicrobial susceptibility testing showed resistance to multiple antibiotics including imipenem, levofloxacin, and cefoxitin. A combination therapy of clarithromycin and tigecycline was recommended.

Case Follow-Up

The patient was readmitted on April 10 due to non-healing wounds located in both axillary and inframammary folds. An area measuring 4 cm in diameter and dark red in color was observed under the right axilla. Additionally, the breast MRI disclosed that there was encapsulated effusion in the posterior region of both pectoralis major muscles. The treatment regimen was altered to include rifampicin + ethambutol + clarithromycin + linezolid on April 14 subsequent to the culture outcomes. Incision and drainage were performed on the right precordial abscess and bilateral axillary chronic ulcer due to bilateral axillary erosion ulcer. A chest CT scan revealed the presence of inflammatory granulomas in the lungs, ruling out tuberculosis. The treatment plan was modified on April 19 based on antimicrobial susceptibility outcomes to include clarithromycin (0.5g BID, oral) and tigecycline (50mg Q12H, inv). However, tigecycline was

Table 1 Infection-Related Laboratory Test Results

Date ^a	White Blood Cell Count (10 ⁹ /L)	Neutrophil Count (10 ⁹ /L)	CRP (mg/L)	Culture of Drainage Fluid or Wound Secretion Samples	Ziehl-Neelsen Staining	T-Spot
January 21	7.13	5.74	- ^b	-	-	-
January 29	8.30	6.06	17.00	-	-	-
February 13	-	-	-	<i>S. lugdunensis</i> + <i>E. faecalis</i>	-	-
March 04	-	-	-	<i>S. aureus</i> + <i>Acinetobacter baumannii</i> ^c	-	-
March 05	14.49	13.34	-	-	-	-
March 11				<i>C. jejuni</i>		
March 15			-	-	+++	-
March 31	6.98	5.31	-	<i>M. abscessus</i> + <i>S. lugdunensis</i> + <i>Corynebacterium</i> spp.	+	Negative
April 21	4.46	2.70	-	-	-	-
May 14	5.39	3.79	-	-	-	-

Notes: ^aThe 1st, 2nd, and 3rd operation was performed on January 22, February 6, and March 4, respectively. ^bNot detected. ^cThe result was obtained after the drainage fluid was injected and incubated into blood culture bottles. The laboratory test results during hospitalization indicated a persistent infection, as evidenced by the continuously elevated total count of white blood cells and neutrophils, as well as a CRP level of 700mg/L. Only after Ziehl-Neelsen staining tested positive was the pathogenic bacteria *M. abscessus* recognized and isolated. *Mycobacterium tuberculosis* infection was ruled out as the Tuberculosis SPOT (T-Spot) test was negative. Adjusted therapy was administered based on antimicrobial susceptibility testing after March 31st, and the infection was controlled as evidenced by decreasing cell count and negative Ziehl-Neelsen staining results.

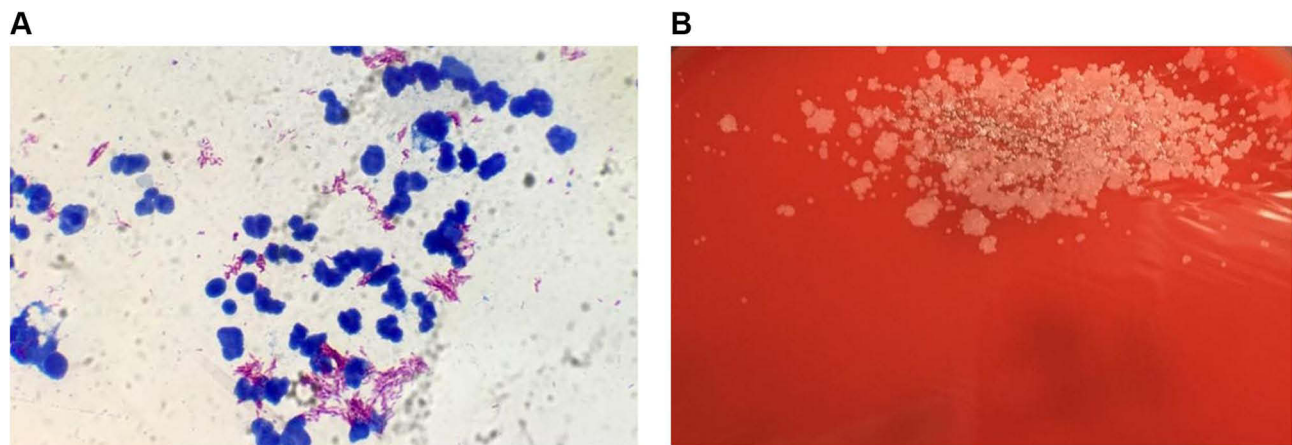


Figure 4 Ziehl-Neelsen staining and isolate of *Mycobacterium abscessus* on blood agar. **(A)** Ziehl-Neelsen staining was used to test drainage fluid and wound secretion samples, which showed a positive result with red bacillus against a blue background. **(B)** After culturing for several days, rough and colorless colonies with varying sizes of *M. abscessus* were isolated.

discontinued due to nausea and vomiting side-effects. The patient was discharged on April 29 after the wound on the right submammary fold had healed without bleeding, exudation, or secretions.

On May 13, the patient was admitted to the hospital for the third time due to redness, swelling, and pain in the left armpit. A 2 cm diameter red and swollen area was observed in the left armpit, and a small amount of pale yellowish exudate was noted in the right armpit wound. Treatment for the left armpit chronic ulcer wound repair and incision and drainage of the abscess were carried out. The individual was discharged on May 16 after showing improvement. She received 6 months of continued oral treatment with clarithromycin (0.5g BID, oral) before the wound fully healed.

Case 2

A 51-year-old female underwent a facial rhytidectomy in August 2020. Four days after the surgery, she experienced local redness, swelling, and pus. Antibiotic treatment with Levofloxacin and Omeprazole was administered after incision and drainage. Due to inadequate improvement, the patient was admitted to our hospital on October 18, 2020.

During the admission examinations, significant swelling of the bilateral cheeks, ears, and neck was observed (Figure 5A). The patient had multiple masses that were tender upon palpation, with a sense of fluctuation. Multiple visible cracks were present on the face and neck, with white pus discharge. A dark red, uneven scar was visible on the

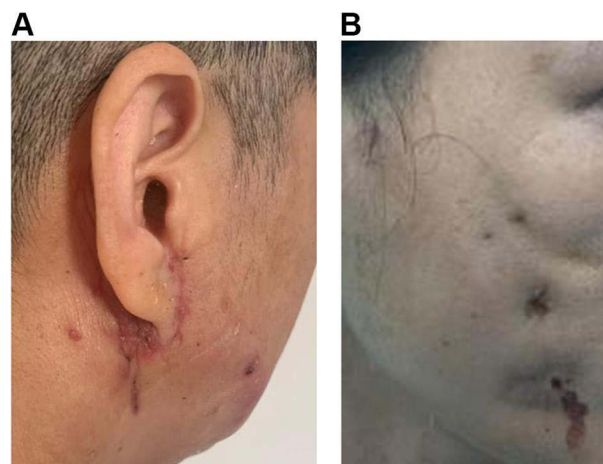


Figure 5 Facial redness and swelling of Case 2 **(A)** and Case 3 **(B)**. Swelling and cracks were visible on the cheeks, ears, and neck. Both patients had a visible scar on their face and ear, which appeared dark red and uneven.

face and ear. The mouth appeared slightly crooked on the left side while smiling. The craniocerebral CT revealed swelling with air in the bilateral maxillofacial area and subcutaneous tissue of the cheek, as well as thickening and local damage to the maxillofacial skin, suggesting an infectious lesion and local abscess.

Laboratory tests revealed a high-sensitivity C-reactive protein (hCRP) level of 9.08 mg/L and a procalcitonin (PCT) level of 0.187 ng/mL. The patient's clinical symptoms and epidemiological history indicated a strong likelihood of non-tuberculous mycobacterial infection. Therefore, empiric treatment was initiated with imipenem, amikacin, and doxycycline, while wound secretions were collected for bacterial culture.

The patient underwent surgical procedures for facial ulcer debridement, foreign body removal, and facial defect flap transfer repair on October 21. A week later, the wound secretion specimen showed the isolation of *M. abscessus*. The isolate showed sensitivity to clarithromycin, amikacin, and linezolid, as well as intermediate susceptibility to cefoxitin while being resistant to ciprofloxacin, doxycycline, tobramycin, and trimethoprim/sulfamethoxazole. After analyzing the antibiotic susceptibility, the medication was altered to include amikacin, clarithromycin, and linezolid.

However, the patient's wound healed poorly, and there was white purulent exudation with bleeding at the incision on the left side of the neck. On November 10, wound resection and debridement of subcutaneous tissue on the head, face, and neck skin were performed. The drainage fluid on the left was pale yellow with floccules, and the fluid on the right was clear. The dressing was changed regularly after the operation, and the sutures were removed 9 days later. Due to side effects of tinnitus, amikacin was discontinued. Subcutaneous tissue from the face and neck underwent its third debridement and drainage procedure on December 2. After 20 days, the incision sites on the face and neck had healed well, resulting in the patient's discharge. Further recovery took place over a duration of 6 months, aided by oral administration of clarithromycin.

Case 3

A 37-year-old woman received facial line carving and autologous fat grafting at a cosmetic clinic 6 months prior to admission. Four months prior to admission, she underwent multiple injections of lysozyme and hormones into the lower eyelid. Two months prior to admission, she developed localized facial redness and swelling, and subsequently underwent a foreign body removal procedure. Despite treatment with cephalosporin, imipenem, levofloxacin, and penicillin in multiple institutions, there was no significant improvement in the local redness, swelling, or pus (Figure 5B). On October 15, 2020, the patient was admitted to the Department of Burn and Plastic Surgery at our hospital.

Admission examination revealed multiple localized masses on both cheeks with evident redness, swelling, and fluctuation. Additionally, four incisions on the right side of the face were unhealed with minimal discharge, and there was a slight deviation in the left angle of the mouth during smiling with visible tooth exposure. *M. abscessus* was isolated from the wound secretion specimen at another hospital, but its antimicrobial susceptibility was unknown. Empiric treatment was initiated with imipenem, amikacin, levofloxacin, and doxycycline.

On October 21, surgery was conducted to repair a chronic facial ulcer and remove a foreign body. Following the surgery, wound secretion was collected for bacterial culture and antibiotic susceptibility testing. The *M. abscessus* isolate exhibited sensitivity to clarithromycin, amikacin, cefoxitin, doxycycline, and linezolid. As such, treatment consisted of a combination of linezolid, amikacin, and doxycycline. Most of the symptoms had improved 1 month later; however, the patient's left cheek exhibited local redness and swelling with a visible break observed. White purulent secretions and a small amount of exudative bleeding were found from the break, thus requiring a second debridement on December 2. Following 12 days, the antibiotics regimen was altered to include clarithromycin, doxycycline, and levofloxacin. The patient was discharged with antibiotics and her wound healed after 5 months of antibiotic therapy.

Discussion

The initial *M. abscessus* isolate was obtained from a patient with a knee abscess in 1952.²¹ *M. abscessus* and *M. chelonae* were initially classified as the same species and were later identified as an independent species in 1992. *Mycobacterium massiliense* and *Mycobacterium bolletii*, two new subspecies, were subsequently discovered and combined with *M. abscessus* to form the *Mycobacterium abscessus* complex (MABC).²² MABC is present widely in the environment, including in water tanks, hot water pipes, swimming pools, medical equipments such as endoscopes, and indoor dust.²³

MABC is resistant to high concentrations of chlorine and other disinfectants, such as alkaline glutaraldehyde. Furthermore, its high-temperature resistance and biofilm formation enable it to survive in aquatic environments and infect wounds that are exposed to improperly disinfected surgical equipment or contaminated water from showers and faucets.²⁴ Cases of *M. abscessus* infection have been reported after plastic surgery procedures such as silicone injection, liposuction, abdominoplasty, bilateral mastopexy, botulinum toxin injections, and breast augmentation.²⁵ The infection is most likely caused by a lack of proper surgical equipment sterilization or poor sterile technique by the surgeon.²⁶ Although, in our three cases, *M. abscessus* was not isolated from the environment of the cosmetic institution, it is suspected that the infection was a result of inadequate disinfection of medical equipment or contaminated sterile saline, based on their epidemiological history.

Skin infections caused by *M. abscessus* are commonly present with erythema, purplish-red nodules, and painless abscesses. Clinical symptoms encompass cellulitis, papular lesions, purple discolored nodules, abscesses, drainage sinus, subcutaneous nodules (nodular pseudoerythema), and ulcers.²⁷ Soft tissue infections caused by *M. abscessus* generally result in multiple skin lesions, while sporadic *Mycobacterium* infections tend to result in a single skin lesion.²⁸

In our cases, patients presented with non-healing wounds accompanied by facial redness, purulent discharge, exudate, swelling, pain, and ulceration, which are common symptoms of *M. abscessus* infection. The increasing prevalence of plastic surgery has led to a rise in related infections, making it an issue that requires attention. A thorough physical examination of patients with a history of plastic surgery and persistent non-healing wounds should include a careful evaluation of any abnormal clinical manifestations. Recurrent or unhealed subcutaneous abscesses, even after adequate debridement and antibiotic treatment, should be monitored for timely diagnosis and treatment of *M. abscessus* infection. To prevent infection, it is essential to properly sterilize all surgical equipment and saline, adequately manage post-operative wounds, and provide patients with education on wound care.¹⁴

Due to the limited number of cases and clinical trials, recommended treatment options for *M. abscessus* soft tissue infection primarily follow guidelines for lung infections, with multiple antibiotic options available for clinical use. The inherent multidrug resistance induced resistance to macrolides and disparities between in vitro drug sensitivity testing and clinical responses all contribute to the complexity of treatment. The in vitro susceptibility testing results of *M. abscessus* to macrolides and tigecycline are relatively reliable, followed by clofazimine, bedaquiline, amikacin, imipenem, and cefoxitin.²⁹ There was only one prospective non-random observational study on the treatment of soft tissue infections caused by *M. abscessus*.³⁰ This study compared the therapeutic effects of oral clarithromycin+moxifloxacin with that of clarithromycin+amikacin from 2007 to 2008. The former treatment had a shorter time of clinical improvement (17 weeks) compared to the latter (20 weeks). However, the study only recruited 52 patients, and patients who received oral clarithromycin+amikacin had more severe symptoms, which might have introduced bias into the results.

The standard approach to treating *M. abscessus* infection is through debridement coupled with antimicrobials. Given the high resistance of *M. abscessus* to antibiotics, a combined therapy is required. However, the most effective treatment regimen and course is not entirely clear.³¹ For a duration of at least 4.5 months, oral clarithromycin or moxifloxacin should be maintained, with an average course of treatment lasting 6–8 months.³²

In our study, the second patient underwent three surgeries with debridement and was treated with amikacin, clarithromycin, and linezolid during hospitalization. Additionally, oral administration of clarithromycin aided in her total recovery. For a third patient, the antibiotics was changed from a combination of linezolid, amikacin, and doxycycline to clarithromycin, doxycycline, and levofloxacin. Treatment for *M. abscessus* infection is complex. Physicians should exercise caution and adjust or alter therapy in a timely manner based on patient reactions.

The treatment for *M. abscessus* infection is constrained by multiple factors such as drug tolerance, toxicity, and drug interferences. Cefoxitin and imipenem exhibit potent bacteriostasis activity against *M. abscessus* in vitro. However, when used alone, they lack bactericidal effect. Hence, a combined therapy is proposed to accomplish synergistic bactericidal activity.³³ Although tigecycline exhibited effective bactericidal activity in vitro and in vivo, 90% of patients experienced adverse reactions including nausea and vomiting.³⁴ Studies indicate that treatment with azithromycin, amikacin, and imipenem significantly improved bacterial clearance rates among patients with pulmonary infections caused by *M. abscessus*.³⁵ Furthermore, drugs that have lower toxicity, such as clofazimine,³⁶ exhibit a synergistic effect with

amikacin, omacycline,³⁷ and tedizolomide.³⁸ These medications show promise in treating skin and soft tissue infections and can be considered for use in treating *M. abscessus* infections.

Due to the slower growth of *M. abscessus* compared to general pathogenic bacteria, it can be easily overlooked by traditional bacterial culture. Case 1 was hospitalized for over 100 days before a diagnosis was made. Throughout her hospitalization, samples of wound secretions were collected for etiological examination, and antibiotics were changed several times in response to the results. Despite undergoing five debridement operations and receiving multiple antibiotics, the wound still healed poorly. Long-term bed confinement causes difficulty in lifting the upper limbs and imposes significant physiological, psychological, and financial burdens. Valuable evidence for targeted treatment and prevention of adverse effects resulting from long-term anti-tuberculosis treatment was only provided after the Ziehl-Neelsen staining results showed positive and *M. abscessus* were isolated. Similar cases have also been reported in Switzerland³⁹ and Denmark.⁴⁰ *M. abscessus* was isolated from the wound specimens 10 years and 2 years after breast augmentation, respectively, indicating that diagnosing and treating such cases is extremely challenging.

For skin and soft tissue infections that occur after trauma, surgery, and plastic surgery, particularly those that fail to respond to traditional antibiotic treatment, and exhibit symptoms such as abscesses, ulcers, and nodules resembling sporotrichosis,¹ it is crucial to widen the pathogen screening scope beyond routine pathogenic examinations. The identification of subspecies in *M. boletii* and *M. abscessus* is vital for guiding clinical treatment due to the presence of the inducible macrolide resistance gene, *erm*,⁴¹ while *M. massiliense* lacks *erm*.⁴¹ Commonly used methods for subspecies identification include MALDI-TOF mass spectrometry⁴² and molecular techniques such as sequencing 16S rRNA, *hsp65*, and *rpoB* regions.⁴³ In our study, the identification of *M. abscessus* in three cases depended on MALDI-TOF MS system. However, the growth of *M. abscessus* takes several days. To increase the efficiency of treating *M. abscessus* infection, it is more desirable to combine MALDI-TOF mass spectrometry with molecular techniques. The secretion from the wound can be sequenced for testing the 16S rRNA, *hsp65*, and *rpoB* regions.

Accurate identification of MABC subspecies is crucial for appropriate treatment due to their varying antimicrobial resistance properties. While MALDI-TOF MS is the main identification method in clinical laboratories for most NTMs, it remains a challenge to differentiate closely related species such as MABC subspecies. However, a combination of MALDI-TOF MS and machine learning has shown a high accuracy in identifying MABC subspecies, although the geographic origin of the strains can impact the protein spectra.^{44,45}

In addition to identifying subspecies, it is necessary to genotype macrolide susceptibility to guide precise treatment for clinical *M. abscessus* infections. The real-time multiplex assay allows for both the distinguishing of MABC subspecies and the determination of its susceptibility to macrolides. Compared to the 3 to 14 days needed for clarithromycin phenotypic susceptibility testing, this assay can provide clinically significant treatment information in less than 3 hr.⁴⁶ Multiple PCR methods have been established and have shown excellent capacity for identifying MABC subspecies and detecting clarithromycin resistance due to *erm* and 23S *rrl* genes, particularly when mixed genetic profiles are present.^{46–49}

Despite the advantages of PCR methods, genetic tests require additional reagents and reaction time for subspecies identification and detection of antibiotic resistance. In contrast, using machine learning algorithm to detect MABC subspecies based on MALDI-TOF spectra does not require any additional cost or reaction time. Therefore, the combination of MALDI-TOF MS and real-time PCR offers high cost-effectiveness, making it a more favourable option for real-world deployment. Expanding diagnostic capabilities in health-care facilities would greatly improve patient care by enabling the administration of appropriate antibiotic treatment and antimicrobial stewardship programs for *M. abscessus* infections.

Conclusion

This article details three cases of *M. abscessus* cutaneous infection after plastic surgery, and presents etiological evidence for treatment. Medical professionals should consider this rare organism as a possible cause for long-term non-healing wounds following postoperative incision and immediately perform etiological examination to determine diagnosis. For treatment, surgery combined with long-term antibiotics is recommended. Additional attention should be given to surgical instruments and saline disinfection in the plastic surgery operating room. Due to the relatively low incidence of

M. abscessus cutaneous infection and the prolonged course of treatment, further study of its pathogenicity and treatment guidelines are necessary for long-term follow-up.

Ethical Approval

All aspects of the study were performed in accordance with national ethics regulations and approved by the Ethics Committee of Shenzhen Hospital of Southern Medical University. Written informed consent was obtained from each patient for publication of this case reports and any accompanying images.

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Disclosure

The authors declare no conflicts of interest in this work.

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