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ORIGINAL RESEARCH Prognostic value of long noncoding RNAs in gastric cancer: a meta-analysis

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Background: In the last few years, accumulating evidence has indicated that numerous long noncoding RNAs (lncRNAs) are abnormally expressed in gastric cancer (GC) and are associated with the survival of GC patients. This study aimed to conduct a meta-analysis on 19 lncRNAs (AFAP1 antisense RNA 1 [AFAP1-AS1], CDKN2B antisense RNA 1 [ANRIL], cancer susceptibility 15 [CASC15], colon cancer associated transcript 2 [CCAT2], gastric adenocarcinoma associated, positive CD44 regulator, long intergenic noncoding RNA [GAPLINC], H19, imprinted maternally expressed transcript [H19], HOX transcript antisense RNA [HOTAIR], HOXA distal transcript antisense RNA [HOTTIP], long intergenic non-protein coding RNA 673 [LINC00673], metastasis-associated lung adenocarcinoma transcript 1 [MALAT1], maternally expressed 3 [MEG3], promoter of CDKN1A antisense DNA damage activated RNA [PANDAR], Pvt1 oncogene [PVT1], SOX2 overlapping transcript [Sox2ot], SPRY4 intronic transcript 1 [SPRY4-IT1], urothelial cancer associated 1 [UCA1], X inactive specific transcript [XIST], ZEB1 antisense RNA 1 [ZEB1-AS1] and ZNFX1 antisense RNA 1 [ZFAS1]) to systematically estimate their prognostic value in GC.

Methods: The qualified literature was systematically searched in PubMed, Web of Science, Embase and Cochrane Database of Systematic Reviews (up to March 16, 2018), and one metaanalysis relating to the relationship between lncRNA expression and overall survival (OS) of GC patients was performed. The only evaluation criterion of survival results was OS.

Results: A total of 6,095 GC patients and 19 lncRNAs from 51 articles were included in the present study. Among the listed 19 lncRNAs, 18 lncRNAs (other than SPRY4-IT1) showed a significantly prognostic value (P < 0.05).

Conclusion: This meta-analysis suggested that the abnormally expressed lncRNAs (AFAP1-AS1, ANRIL, CASC15, CCAT2, GAPLINC, H19, HOTAIR, HOTTIP, LINC00673, MALAT1, MEG3, PANDAR, PVT1, Sox2ot, UCA1, XIST, ZEB1-AS1 and ZFAS1) were significantly associated with the survival of GC patients, among which AFAP1-AS1, CCAT2, LINC00673, PANDAR, PVT1, Sox2ot, ZEB1-AS1 and ZFAS1 were strong candidates in predicting the prognosis of GC patients.

Keywords: long noncoding RNA, gastric cancer, prognosis, meta-analysis

Introduction

In the last few years, accumulating evidence has indicated that numerous long noncoding RNAs (lncRNAs) are abnormally expressed in gastric cancer (GC) and are associated with the survival of GC patients.¹⁻¹¹³ GC is the fourth most diagnosed tumor type and the third most common origin of tumor-related death all over the world.^{114,115} Although the incidence and mortality of GC are declining, >24,590 individuals are diagnosed with GC per year, of which 10,720 die from GC in the USA.¹¹⁶ Although diagnosis and treatment strategies have been improved, the number of surviving cases remains low, since diagnosis

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often occurs in the late stages.^{116,117} Thus, the molecular characteristics about the carcinogenesis of GC and the recognition of new biomarkers for GC are urgently needed.

lncRNA is a new type of noncoding RNA that has a length of >200 nucleotides (nt) and lacks important open reading frameworks and can be divided into five main categories (sense, antisense, bidirectional, intronic and intergenic).¹¹⁸ Abundant evidence has demonstrated that lncRNAs play significant regulatory roles in tumor biology via various mechanisms affecting transcriptional and posttranscriptional levels.^{118–120} Currently, for both cell behavior and clinicopathological factors, significant advances with respect to lncRNA effects on GC have been discovered.¹²¹

On account of the obvious expression differences between normal and malignant tissues as well as causal roles of lncRNAs in cancer development, lncRNAs are now attracting increasing attention, which has led to numerous investigations of the correlation between lncRNA states and clinical results in GC. Nevertheless, most of these studies were performed with small samples, and there were inconsistently observed connections. Consequently, we conducted a meta-analysis to determine the accurate role of lncRNAs in the prognosis of GC patients, which possibly supplied us with new insights into the clinical value of combined detection in forecasting prognostic results and determining promising biomarkers in GC treatment strategies.

Methods

Literature search strategy

We basically performed a systematic selection of papers published in English from four databases (PubMed, Web of Science, Embase and Cochrane Database of Systematic Reviews). A comprehensive search was conducted using the subject term: lncRNA and gastric cancer. Two authors (Song Gao and Zhi-Ying Zhao) checked the titles and abstracts of the retrieved papers, and Yue Zhang reevaluated uncertain data. Figure 1 shows the flow diagram of the literature search and selection.

Inclusion criteria

We set up inclusion criteria for qualified papers, which were analyzed using our full-text assessment: 1) articles

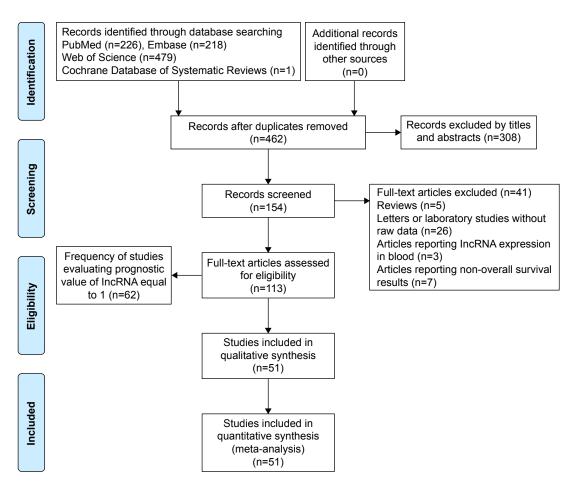


Figure I Flow diagram of the literature search and selection. Abbreviation: IncRNA, long noncoding RNA.

concerning the pertinence between lncRNA level in cancer tissues and prognosis of GC patients; 2) the survival results were estimated using overall survival (OS) and 3) full-text papers published in English.

Exclusion criteria

Articles that did not meet the abovementioned inclusion criteria, reviews, letters and laboratory studies without raw data were excluded. Articles of non-dichotomous lncRNA expression levels and frequency of studies evaluating prognostic value of lncRNAs equal to 1 were also excluded. If more than one paper had been published on the identical study cohort, only the most well-rounded investigation was selected for this research. In addition, if both of the univariate and multivariate outcomes were covered, only the latter were chosen, since they were adjusted for confounding factors.

Research frequency

Table 1 gives the frequency of investigations reporting prognosis of GC patients, which included the lncRNA name, frequency of researched lncRNA and reference.

Data extraction

The survival data were recovered from qualified articles independently by two authors (Song Gao and Zhi-Ying Zhao). Data extracted from them are as follows: researched lncRNA,

IncRNA	n	R	IncRNA	n	R	IncRNA	n	R
AC027119.1	1	1	HI9	4	29, 36–38	NEATI	1	83
AC138128.1	1	2	HAGLROS	1	39	NR_003573	1	7
ADAMTS9-AS2	1	1	HIFIA-AS2	1	40	OR3A4	1	84
AFAPI-ASI	2	3, 4	HOTAIR	9	41-59	OTUBI-isoform 2	1	85
AGAP2-ASI	1	5	HOTTIP	3	50-52	PANDAR	2	86, 87
AK023391	1	6	HOXA-AS2	1	53	PCATI	1	88
AK093735	1	7	HXAII-AS	1	54	PVTI	2	89, 90
AK123072	1	8	KRTI8P55	1	55	RP11-119F7.4	1	91
ANRIL	2	9, 10	LET	1	56	RP11-120K18.2	1	1
АТВ	1	11	LINC00052	1	57	RP11-389G6.3	1	1
BANCR	1	12	LINC00152	1	58	RP11-499F3.2	1	1
BC005927	1	13	LINC00628	1	59	RP11-789C1.1	1	92
BC032469	1	14	LINC00668	1	60	RPLPOP2	1	29
BC041951	1	15	LINC00673	2	61, 62	SLC26A4	1	29
BCAR4	1	16	LINC00675	1	63	SMIM10L2A	1	29
CASC2	1	17	LINC00982	1	64	SMIM10L2B	1	29
CASC15	2	18, 19	LINC01018	1	65	SNHGI	1	93
CCATI	1	20	LINC01234	1	66	SNHG6	1	94
CCAT2	2	21, 22	LINC01296	1	67	SNHG12	1	95
CHRDLI	1	23	LINC-ROR	1	68	SNORD116-4	1	29
CTD-2147F2.1	I	1	LINC-UBCI	I	69	Sox2ot	2	96, 97
DANCR	1	24	LOC100130476	1	70	SPRY4-IT I	2	98, 99
DLX6-ASI	1	1	LOC553137	1	65	TINCR	1	29
E2FI	1	25	MACCI	1	71	TTTY14	1	65
EGOT	1	26	MACCI-ASI	1	71	TUGI	1	100
FENDRR	1	27	MALATI	4	43, 72–74	UCAI	4	101-104
FEZFI-ASI	1	28	MANCR	1	75	VPS9D1-AS1	1	105
FOXD2-ASI	1	29	MEG3	2	76, 77	XIAP-ASI	1	106
FRLncl	I	30	MIR31HG	I	78	XIST	2	107, 108
GACAT3	1	31	MIR4435-2HG	I	65	XLOC_010235	1	92
GAPLINC	2	32, 33	MLK7-ASI	1	79	ZEBI-ASI	2	109, 110
GAS5	I	34	MLLT4-ASI	I	80	ZFASI	2	, 2
GBETI	I	35	MRUL	I	81	ZMATI	1	113
GCIncI	I	15	MTM	I	82			
		1						

Table I Research frequency of IncRNAs in GC

 $\textbf{Notes:} \ \text{Highlighted IncRNAs were included in the meta-analysis. n, number of research frequency; R, reference.}$

Abbreviations: AFAP1-AS1, AFAP1 antisense RNA I; ANRIL, CDKN2B antisense RNA I; CASC15, cancer susceptibility 15; CCAT2, colon cancer associated transcript 2; GAPLINC, gastric adenocarcinoma associated, positive CD44 regulator, long intergenic noncoding RNA; GC, gastric cancer; H19, H19, imprinted maternally expressed transcript; HOTAIR, HOX transcript antisense RNA; HOTTIP, HOXA distal transcript antisense RNA; LINC00673, long intergenic non-protein coding RNA 673; lncRNA, long noncoding RNA; MALAT1, metastasis-associated lung adenocarcinoma transcript 1; MEG3, maternally expressed 3; PANDAR, promoter of CDKN1A antisense DNA damage activated RNA; PVT1, Pvt1 oncogene; Sox2ot, SOX2 overlapping transcript; SPRY4-IT1, SPRY4 intronic transcript 1; UCA1, urothelial cancer associated 1; XIST, X inactive specific transcript; ZEB1-AS1, ZEB1 antisense RNA 1; ZFAS1, ZNFX1 antisense RNA 1.

first author's name, paper publication year, reference, patient's nationality, study design, histological type, patient number, neoplasm staging, cutoff value, detected method, follow-up period, survival analysis type, HRs and 95% CIs. The detailed data are shown in Table 2. If HR and 95% CI were not directly shown in the paper, data from survival curve were extracted. Disagreements were discussed with the third investigator (Yue Zhang).

Statistical analysis

Stata version 13.0 (StataCorp LP, College Station, TX, USA) was used for the whole meta-analysis. HR and 95% CI from GC patients were calculated on the basis of survival curve and patient number using Engauge Digitizer version 4.1 and Tierney's method.122 The random-effect model was used in the whole article because different histological type (frozen, formalin-fixed paraffin-embedded or undefined) from GC patients at different neoplasm staging, cutoff value and IncRNA detected method was used in the single study. The HR was considered significant if its P-value was <0.05 and 95% CI did not contain the value 1. Furthermore, the lncRNA was considered as a strong biomarker of prognosis, if its HR was >2. The Begg's funnel plot was used to estimate publication bias, and a two-tailed P-value <0.05 was considered as significant. The sensitivity analysis was performed to examine how sensitive the merged HR was if the single study was removed, and if the point of evaluation was outside the 95% CI after it was removed from the whole analysis, a single research was considered as excessive influence.

Results Meta-analysis

Table 3 gives the basic information of the merged metaanalysis for researched lncRNAs.

AFAPI antisense RNA I (AFAPI-ASI), CDKN2B antisense RNA I (ANRIL), cancer susceptibility I5 (CASCI5), colon cancer-associated transcript 2 (CCAT2), gastric adenocarcinoma associated, positive CD44 regulator, long intergenic noncoding RNA (GAPLINC) and H19, imprinted maternally expressed transcript (H19) demonstrated significantly prognostic value

Two articles^{3,4} reported the relationship between high AFAP1-AS1 expression and OS, indicating that GC patients with its high expression had significantly worse OS than those with its low expression (HR=2.47, 95% CI=1.41-4.30, P<0.01).

Table 2 Bas	Table 2 Basic information of included articles	icluded articles											
IncRNA	Study	Country/ source	Study design	Sample	Number	Stage	Cutoff	Method	Follow-up (months)	so	HR (L/H)	HR (H/L)	95% CI
AFAPI-ASI	Feng et al ³	China	Я	Frozen	16	>H	None	qRT-PCR	66	Multivariate		3.32	1.55-5.90
AFAPI-ASI	Qiao et al ⁴	China	R	Frozen	87	≥⊢	Median	qRT-PCR	60	Univariate		I.88	1.01-3.52
ANRIL	Zhang et al ⁹	China	R	Tissue	120	≥⊢	e	qRT-PCR	60	Multivariate		1.74	1.04-2.93
ANRIL	Deng et al ¹⁰	China	Ч	Tissue	100	>H	None	qRT-PCR	>60	Univariate		1.61	0.95–2.74
CASC15	Yao et al ⁱ⁸	China	Ч	Tissue	60	≥⊢	None	qRT-PCR	60	Univariate		2.33	I.I5-4.72
CASC15	Wu et al ¹⁹	China	Ч	Tissue	88	≥⊢	Mean	qRT-PCR	60	Univariate		1.70	0.84–3.47
CCAT2	Wang et al ²¹	China	Ч	Frozen	85	≥⊢	Mean	qRT-PCR	60	Multivariate		2.41	1.19–5.42
CCAT2	Wang et al ²²	China	Я	Frozen	108	>=	Median	qRT-PCR	66	Multivariate		2.11	1.44–3.20
GAPLINC	Hu et al ³²	China	Ч	Tissue	60	≥⊢	Median	RT-qPCR	>60	Multivariate		I.48	1.16–1.89
GAPLINC	Liu et al ³³	China	Ч	Frozen	33	None	2.03	qRT-PCR	60	Univariate		1.77	0.57-5.52
HI9	Li et al ³⁶	China	Ч	Frozen	74	>H	Mean	qRT-PCR	53	Univariate		2.26	0.58-8.86
HI9	Zhang et al ³⁷	China	Ч	Frozen	80	> -	Mean	qRT-PCR	60	Multivariate		I.14	1.01–1.29
HI9	Chen et al ³⁸	China	Ч	Tissue	128	> -	4.615	qRT-PCR	48	Multivariate		1.96	0.97–3.97
HI9	Li et al ²⁹	TCGA	R	Tissue	361	> -	None	Downloaded	>50	Univariate		1.79	1.26–2.53
HOTAIR	Endo et al ⁴¹	China I	Ч	Frozen	36	> -	0.1	qRT-PCR	>60	Univariate		0.95	0.21-4.31
		China II			32							5.12	0.96–27.18
HOTAIR	Xu et al ⁴²	China	Ж	Frozen	83	> -	None	RT-qPCR	>72	Multivariate		2.13	1.00-4.50
HOTAIR	Okugawa et al ⁴³	Japan	Ч	Frozen	150	>I–II	0.239	RT-qPCR	60	Multivariate		1.77	1.06–2.95

																														-						
1.16-7.03	1.04-2.06	1.06–3.77	0.45-5.14	0.82-4.05	0.97-4.38	0.52-2.05	1.19–2.23	1.01-4.54	1.12-5.06	I.38–2.83	I.35–4.68	1.03–1.85	0.92–2.58	0.99–3.75	0.88-4.54	1.13-12.06	2.70–3.54	1.07-4.10	I.05-4.93	I.28–3.30	1.24–6.43	0.31-1.57	I.08–5.75	1.22-4.52	0.72-1.73		1.07-7.96	0.95–3.38	1.32–2.26	I.40-4.42	1.41–3.96	I.27–5.84	I.52–2.49	I.25–6.84	0.96–6.17	ed transcript 2; ssed transcript; bidh excreasion
4.06 2.86	1.47	2.00	I.52	I.83	2.06	I.03	I.63	2.56	2.38	1.98	2.52	I.38	I.54			3.68	3.10	2.09	2.28	2.05	3.24			2.35	I.I.		2.92	I.80	1.72	2.49	2.36	2.72	1.95	2.57	2.43	ancer associat ternally expre
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Univariate	Multivariate	Multivariate	Univariate	Univariate	Univariate	Univariate	Univariate	Multivariate	Multivariate	Univariate	Univariate	Univariate	Univariate	Univariate	Univariate	Multivariate	Multivariate	Multivariate	Multivariate	Multivariate	Univariate	Multivariate	Univariate	Multivariate	Univariate		Multivariate	Univariate	Multivariate	Univariate	Multivariate	Univariate	Univariate	Multivariate	Univariate	usceptibility 15; CCA Dmnibus; H19, H19, in B // H) horord ratios
> 45 > 45	> 60	>60	>120	90	60	54	>I 50	66	>40	> 50	>60	>I 50	60	48	>60	36	84	36	48	> 84	>60	60	36	60	>116		36	60	06 <	54	72	90	>I50	60	36	CASCI5, cancer s ene Expression C
gRT-PCR	qRT-PCR	qRT-PCR	qRT-PCR	qRT-PCR	qRT-PCR	RT-qPCR	Downloaded	qRT-PCR	qRT-PCR	RT-qPCR	qRT-PCR	RT-qPCR	RT-qPCR	RT-qPCR	qRT-PCR	qRT-PCR	qRT-PCR	qRT-PCR	qRT-PCR	qRT-PCR	qRT-PCR	qRT-PCR	qRT-PCR	RT-qPCR	Downloaded		qRT-PCR	qRT-PCR	qRT-PCR	qRT-PCR	qRT-PCR	qRT-PCR	Downloaded	qRT-PCR	qRT-PCR	embedded tissues; C oding RNA; GEO, G
Median	Median	2.35	None	Mean	Median	Median	None	Median	2	None	None	None	0.985	Median	None	None	Mean	Median	Median	Median	Median	Median	Median	Median	Median		Median	None	Median	None	Median	Median	None	Median	Median	fixed paraffin- tergenic nonco
≥ ≥ 	≥I−III	≥⊢	None	≥⊢		Ē	≥⊢	×⊢	≥⊢	≥I−III	≥⊢	≥⊢	>I–II	>I–I<		≥⊢	≥⊢	≥⊢	≥⊢		None	×⊢	> -	≥⊢	≥⊢		>⊢I	>H	>H	>1-1		≥⊢	>I-I	≥⊢	≥⊢	n and formalin- gulator, long in
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Both	Tissue	Frozen	Tissue	FFPE	Frozen	Frozen	Tissue	Frozen	Tissue	Tissue	Tissue	FFPE	Frozen	Frozen	Frozen	Tissue	Tissue	Tissue	Tissue	Frozen	Tissue	Frozen	Frozen	Frozen	Tissue		Frozen	Frozen	Frozen	FFPE	Tissue	Frozen	Tissue	Frozen	Tissue	B antisense RN/ na associated, po
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China	China	China	China	China	China	China	GEO	China	China	TCGA	China	China	Japan	China	China	China	China	China	China	China	China	China	China	China	TCGA		China	China	China	China	China	China	KΜ	China	China	isense RNA 1; / SAPLINC, gastri
Llu et al Zhang et al ⁴⁵	Zhao et al ⁴⁶	Chen et al ⁴⁷	Feng and Huang ⁴⁸	Li et al ⁴⁹	Ye et al ⁵⁰	Yang et al ^{s I}	Zhao et al ⁵²	Ba et al ⁶¹	Huang et al ⁶²	Qi et al 72	Li et al ⁷³	Li et al ⁷⁴	Okugawa et al ⁴³	Sun et al ⁷⁶	Guo et al $^{\prime\prime}$	Ma et al ⁸⁶	Liu et al ⁸⁷	Kong et al ⁸⁹	Yuan et al ⁹⁰	Zhang et al%	Zou et al ⁹⁷	Peng et al ⁹⁸	Xie et al ⁹⁹	Zheng et al ¹⁰¹	Nasrollahzadeh-	Khakiani et al ¹⁰²	Zuo et al ¹⁰³	Gu et al ¹⁰⁴	Chen et al ¹⁰⁷	Ma et al ¹⁰⁸	Li et al ¹⁰⁹	Zhang et al ¹¹⁰		Zhang et al ^{III}	Nie et al ¹¹²	Abbreviations: AFAPI -ASI, AFAPI antisense RNA I; ANRIL, CDKN2B antisense RNA I; Both, frozen and formalin-fixed paraffin-embedded tissues; CASCI5, cancer susceptibility 15; CCAT2, colon cancer associated transcript 2: FFPE, formalin-fixed paraffin-embedded; GAPLINC, gastric adenocarcinoma associated, positive CD44 regulator, long intergenic noncoding RNA; GEO, Gene Expression Omnibus; H19, H19, imprinted maternally expressed transcript; HOTAIB HOX monocines and HOX discription and the HB (HI), heard region of the horoscine variance have been and for the fixed parafficient and the fixed parafficient parafficient and the fixed parafficient and the fixed parafficient and the fixed parafficient parafficient and the fixed parafficient parafficient and the fixed parafficient and the fixed parafficient and the fixed parafficient parafficient and the fixed parafficient and the fixed parafficient and the fixed parafficient and the fixed parafficient parafficient parafficient and the fixed parafficient parafficient and the fixed parafficien
HOTAIR	HOTAIR	HOTAIR	HOTAIR	HOTAIR	HOTTIP	HOTTIP	HOTTIP	LINC00673	LINC00673	MALATI	MALATI	MALATI	MALATI	MEG3	MEG3	PANDAR	PANDAR	PVTI	PVTI	Sox2ot	Sox2ot	SPRY4-IT I	SPRY4-IT I	UCAI	UCAI		UCAI	UCAI	XIST	XIST	ZEBI-ASI	ZEBI-ASI		ZFASI	ZFASI	Abbreviations: FFPE, formalin-fit

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Table 3 HR with 95% CI of IncRNA expression in GC

IncRNA	Number of articles	Included articles	HR	95% CI	Figure	P -value	Heterogeneity (Higgins I ² statistic)	Total patients
High AFAP1-AS1	2	3, 4	2.47	1.41-4.30	2	< 0.01	l ² =32.7%, P=0.22	178
High ANRIL	2	9, 10	1.68	1.16-2.43	2	<0.01	l ² =0.0%, P=0.84	220
High CASC15	2	18, 19	1.99	1.21-3.28	2	<0.01	l²=0.0%, P=0.54	148
High CCAT2	2	21, 22	2.17	1.53-3.09	2	< 0.0 I	l ² =0.0%, P=0.76	193
High GAPLINC	2	32, 33	1.49	1.18–1.89	2	<0.01	l ² =0.0%, P=0.76	123
High H19	4	29, 36–38	1.51	1.05-2.17	2	0.03	I ² =64.1%, P=0.04	643
High HOTAIR	9	41-49	1.93	1.53-2.43	3	< 0.0 I	1 ² =14.0%, P=0.31	794
High HOTTIP	3	50-52	1.57	1.20-2.05	6	<0.01	I ² =0.2%, P=0.37	540
High LINC00673	2	61,62	2.47	1.45-4.20	6	<0.01	l ² =0.0%, P=0.89	152
High MALAT I	4	43, 72–74	1.70	1.33-2.18	6	<0.01	I ² =29.7%, P=0.23	496
Low MEG3	2	76, 77	1.96	1.17-3.28	6	0.01	l ² =0.0%, P=0.95	206
High PANDAR	2	86, 87	3.11	2.72-3.55	6	<0.01	l ² =0.0%, P=0.79	246
High PVT I	2	89, 90	2.17	1.31-3.60	6	<0.01	l ² =0.0%, P=0.87	191
High Sox2ot	2	96, 97	2.30	1.52-3.46	7	<0.01	I ² =0.0%, P=0.35	287
Low SPRY4-IT I	2	98, 99	1.42	0.48-4.22	7	0.53	I ² =71.4%, P=0.06	236
High UCA1	4	101-104	1.73	1.12-2.68	7	0.01	<i>I</i> ² =45.5%, <i>P</i> =0.14	399
High XIST	2	107, 108	1.89	1.38–2.59	7	<0.01	l ² =23.4%, P=0.25	204
High ZEB1-AS1	2	109, 110	2.07	1.67–2.56	7	<0.01	l²=0.0%, P=0.62	831
High ZFASI	2	111, 112	2.51	1.34-4.69	7	< 0.0 I	I ² =0.0%, P=0.93	158

Abbreviations: AFAPI-ASI, AFAPI antisense RNA I; ANRIL, CDKN2B antisense RNA I; CASC15, cancer susceptibility 15; CCAT2, colon cancer associated transcript 2; GAPLINC, gastric adenocarcinoma associated, positive CD44 regulator, long intergenic noncoding RNA; GC, gastric cancer; H19, H19, imprinted maternally expressed transcript; HOTAIR, HOX transcript antisense RNA; HOTTIP, HOXA distal transcript antisense RNA; LINC00673, long intergenic non-protein coding RNA 673; lncRNA, long noncoding RNA; MALAT1, metastasis-associated lung adenocarcinoma transcript 1; MEG3, maternally expressed 3; PANDAR, promoter of CDKN1A antisense DNA damage activated RNA; PVT1, Pvt1 oncogene; Sox2ot, SOX2 overlapping transcript; SPRY4-IT1, SPRY4 intronic transcript 1; UCA1, urothelial cancer associated 1; XIST, X inactive specific transcript; ZEB1-AS1, ZEB1 antisense RNA 1; ZFAS1, ZNFX1 antisense RNA 1.

Two researches^{9,10} covered the connections between high ANRIL expression and OS, suggesting that GC patients with its high expression had significantly poorer OS than those with its low expression (HR=1.68, 95% CI=1.16–2.43, P<0.01).

Two investigations^{18,19} analyzed the associations between high CASC15 expression and OS, showing that GC patients with its high expression had significantly shorter OS than those with its low expression (HR=1.99, 95% CI=1.21–3.28, P<0.01).

Two studies^{21,22} focused on the correlation between high CCAT2 expression and OS, manifesting that GC patients with its high expression had significantly worse OS than those with its low expression (HR=2.17, 95% CI=1.53–3.09, P < 0.01).

Two papers^{32,33} paid attention to the pertinence between high GAPLINC expression and OS, demonstrating that GC patients with its high expression had significantly poorer OS than those with its low expression (HR=1.49, 95% CI=1.18–1.89, P<0.01).

Four literature^{29,36–38} described the relativity between high H19 expression and OS, proving that GC patients with its

high expression had significantly shorter OS than those with its low expression (HR=1.51, 95% CI=1.05–2.17, *P*=0.03; Figure 2).

HOX transcript antisense RNA (HOTAIR) demonstrated significantly prognostic value

Nine essays^{41–49} discussed the relation between high HOTAIR expression and OS, illuminating that GC patients with its high expression had significantly worse OS than those with its low expression (HR=1.93, 95% CI=1.53–2.43, P<0.01; Figure 3).

Publication bias

The Begg's funnel plot was used to estimate publication bias, and its *P*-value was 0.20, so there was no significant publication bias in the pooled analysis of OS about high HOTAIR expression (Figure 4).

Sensitivity analysis

The sensitivity analysis was performed to examine how sensitive the merged HR was if the single study was removed.

Study ID	HR (95% CI)	% weigh
AFAP1-AS1	_	
Feng et al (2017) ³ (China)	3 .32 (1.55–5.90)	47.71
Qiao et al (2017) ⁴ (China)	1.88 (1.01–3.52)	52.29
Subtotal (<i>I</i> ² =32.7%, <i>P</i> =0.223)	2.47 (1.41-4.30)	100
ANRIL		
Zhang et al (2014) ⁹ (China)	1.74 (1.04–2.93)	51.12
Deng et al (2016) ¹⁰ (China)	1.61 (0.95–2.74)	48.88
Subtotal (/²=0.0%, P=0.837)	1.68 (1.16–2.43)	100
CASC15		
Yao et al (2017) ¹⁸ (China)	2.33 (1.15–4.72)	50.23
Wu et al (2018) ¹⁹ (China)	1.70 (0.84–3.47)	49.77
Subtotal (/²=0.0%, P=0.537)	1.99 (1.21–3.28)	100
CCAT2		o / = /
Wang et al (2015) ²¹ (China)	2.41 (1.19–5.42)	21.71
Wang et al (2016) ²² (China)	2.11 (1.44–3.20)	78.29
Subtotal (/2=0.0%, P=0.761)	2.17 (1.53–3.09)	100
GAPLINC		05 50
Hu et al (2014) ³² (China)	▲ 1.48 (1.16–1.89)	95.58
Liu et al (2016) ³³ (China)	1.77 (0.57–5.52)	4.42
Subtotal (/²=0.0%, P=0.763)	1.49 (1.18–1.89)	100
		0.40
Li et al (2014) ³⁶ (China)	2.26 (0.58–8.86)	6.12 44.41
Zhang et al (2014) ³⁷ (China)	1.14 (1.01–1.29)	
Chen et al (2016) ³⁸ (China)	1.96 (0.97–3.97)	16.86
Li et al (2016) ²⁹ (TCGA)	1.79 (1.26–2.53)	32.62
Subtotal (<i>I</i> ²=64.1%, <i>P</i> =0.039)	1.51 (1.05–2.17)	100
0.01 0.1 1	 10 100	

Figure 2 Forest plot of pooled analyses of OS in association with high AFAP1-AS1, ANRIL, CASC15, CCAT2, GAPLINC and H19 expression levels. Note: Weights are from random-effects analysis.

Abbreviations: AFAPI-AS1, AFAPI antisense RNA 1; ANRIL, CDKN2B antisense RNA 1; CASC15, cancer susceptibility 15; CCAT2, colon cancer associated transcript 2; GAPLINC, gastric adenocarcinoma associated, positive CD44 regulator, long intergenic noncoding RNA; H19, H19, imprinted maternally expressed transcript. OS, overall survival.

After this process, no individual study significantly affected the combined HR with 95% CI (Figure 5).

HOXA distal transcript antisense RNA (HOTTIP), long intergenic non-protein coding RNA 673 (LINC00673), metastasis-associated lung adenocarcinoma transcript I (MALATI), maternally expressed 3 (MEG3), promoter of CDKNIA antisense DNA damage activated RNA (PANDAR) and PvtI oncogene (PVTI) demonstrated significantly prognostic value The details are shown in Table 3 and Figure 6. SOX2 overlapping transcript (Sox2ot), urothelial cancer-associated I (UCAI), X inactive specific transcript (XIST), ZEBI antisense RNA I (ZEBI-ASI) and ZNFXI antisense RNA I (ZFASI) demonstrated significantly prognostic value The details are shown in Table 3 and Figure 7.

Discussion

Current situation

So far, the clinical treatment of GC remains limited. In the past score years, there has been little progress in both traditional

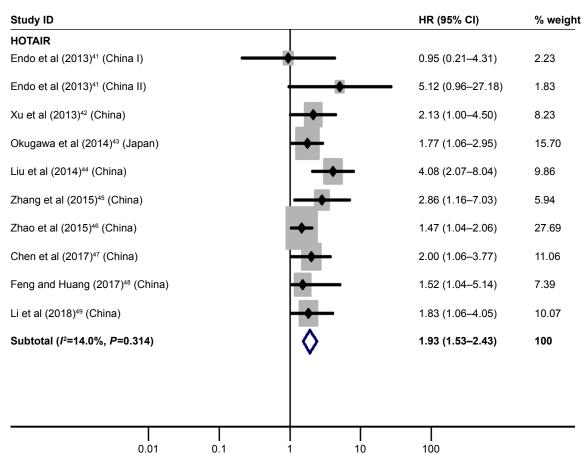


Figure 3 Forest plot of pooled analysis of OS in association with high HOTAIR expression levels.

Abbreviations: HOTAIR, HOX transcript antisense RNA; OS, overall survival.

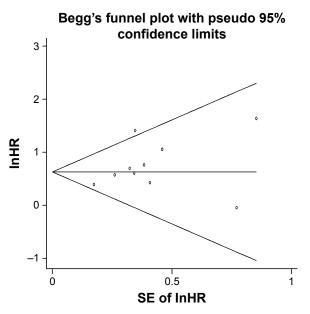
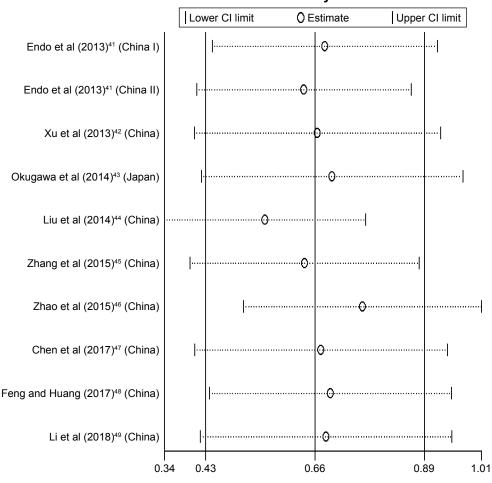


Figure 4 Beggs's funnel plot of publication bias for pooled analysis of OS in association with high HOTAIR expression levels. Abbreviations: HOTAIR, HOX transcript antisense RNA; OS, overall survival; and new treatment methods. Therefore, novel biomarkers that can improve the prognosis of GC patients are in need. Recently, there is an increasing evidence that lncRNAs can hinder the growth and metastasis of cancer. For example, Xu et al¹²³ reported that upregulating long stress-induced noncoding 5 (LSINCT5) significantly promoted the growth of the GC cell, while downregulating LSINCT5 suppressed its growth. Dan et al¹²⁴ conducted the cancer model experiments using mice, proving that MEG3 overexpression could suppress GC growth and metastasis in vivo by suppressing miR-21 expression. More importantly, several abnormally expressed lncRNAs have been discovered to touch upon the development of GC and perhaps possess prognostic potency in this illness. In view of the above consequences, we conducted this metaanalysis about the prognostic value of lncRNAs in GC.

Research finding

In the present research, a total of 51 articles reporting 19 lncRNAs, which were latent prognostic biomarkers and

SE. standard error.



Meta-analysis estimates, given named study is omitted

Figure 5 Sensitivity analysis of pooled analysis of OS in association with high HOTAIR expression levels. Abbreviations: HOTAIR, HOX transcript antisense RNA; OS, overall survival.

6,095 GC patients were included, among which 18 lncRNAs (except SPRY4 intronic transcript 1 [SPRY4-IT1]) manifested a significantly prognostic value. Meanwhile, strong heterogeneity was only shown in two (H19 and SPRY4-IT1) analyses about lncRNAs, during which there was no significant associations between SPRY4-IT1 expression and OS. Further analysis suggested that AFAP1-AS1, CCAT2, LINC00673, PANDAR, PVT1, Sox2ot, ZEB1-AS1 and ZFAS1 were strong candidates in predicting prognosis of GC patients.

Molecular mechanisms

Figure 8 shows the summary of lncRNAs with aberrant expression, potential targets and pathways included in this study. It is noteworthy that there existed inconsistent outcomes about expression of HOTTIP and SPRY4-IT1

compared with normal controls, so these two lncRNAs were not shown to be up or down expressed. Unexpected results were findings that CDKN1A was target of both CASC15 and PANDAR and KLF2 was target of both LINC00673 and ZFAS1. In addition, cell proliferation was the most related cell function of these lncRNAs.

Merits

The current study had several merits: 1) nearly all articles appraising the associations between OS of GC patients and lncRNA expression were searched and are clearly shown in Table 1; 2) most of our meta-analyses revealed no or low heterogeneity ($I^2 \le 50.0\%$), indicating relatively consistent results of the meta-analyses and 3) all the included studies had a relatively large sample size (≥ 30), decreasing the error of low sample size to some degree.

Study ID	HR (95% CI)	% weigh
НОТТІР		
Ye et al (2016) ⁵⁰ China	2.06 (0.97–4.38)	12.60
Yang et al (2017) ⁵¹ China	- 1.03 (0.52–2.05)	15.21
Zhao et al (2018) ⁵² GEO	► 1.63 (1.19–2.23)	72.19
Subtotal (<i>I</i> ² =0.2%, <i>P</i> =0.367)	> 1.57 (1.20-2.05)	100
LINC00673	_	
Ba et al (2017) ⁶¹ China	2.56 (1.01–4.54)	50.17
Huang et al (2017) ⁶² China	2.38 (1.12–5.06)	49.83
Subtotal (/2=0.0%, P=0.893)	2.47 (1.45–4.20)	100
MALAT1		
Qi et al (2016) ⁷² TCGA	← 1.98 (1.38–2.83)	30.23
Li et al (2017) ⁷³ China	2.52 (1.35–4.68)	13.29
Li et al (2017) ⁷⁴ China	- 1.38 (1.03–1.85)	38.45
Okugawa et al (2014) ⁴³ Japan	1.54 (0.92–2.58)	18.02
Subtotal (<i>I</i> ² =29.7%, <i>P</i> =0.234)	1.70 (1.33–2.18)	100
MEG3		
Sun et al (2014) ⁷⁶ China	↓ 1.93 (0.99–3.75)	60.28
Guo et al (2017) ⁷⁷ China	2.00 (0.88–4.54)	39.72
Subtotal (<i>I</i> ² =0.0%, <i>P</i> =0.947)	> 1.96 (1.17–3.28)	100
PANDAR		
Ma et al (2016) ⁸⁶ China	3.68 (1.13–12.06)	1.29
Liu et al (2018) ⁸⁷ China	◆ 3.10 (2.70-3.54)	98.71
Subtotal (I ² =0.0%, P=0.778)	3.11 (2.72–3.55)	100
PVT1		
Kong et al (2015) ⁸⁹ China	2.09 (1.07–4.10)	57.00
Yuan et al (2016) ⁹⁰ China	2.28 (1.05–4.93)	43.00
Subtotal (<i>I</i> ² =0.0%, <i>P</i> =0.868)	2.17 (1.31–3.60)	100
I I 0.01 0.1 1	I I 10 100	

Figure 6 Forest plot of pooled analyses of OS in association with high HOTTIP, LINC00673, MALATI, PANDAR, PVTI expression levels, or low MEG3 expression levels. Note: Weights are from random-effects analysis.

Abbreviations: GEO, Gene Expression Omnibus; HOTTIP, HOXA distal transcript antisense RNA; LINC00673, long intergenic non-protein coding RNA 673; MALATI, metastasis-associated lung adenocarcinoma transcript I; MEG3, maternally expressed 3; OS, overall survival; PANDAR, promoter of CDKNIA antisense DNA damage activated RNA; PVTI, PvtI oncogene; TCGA, The Cancer Genome Atlas.

Limitations

However, the limitations of this work could not be ignored: 1) only English papers were included in the present research, which may exclude potentially relevant articles; 2) most of the patients were from China, which cannot adequately represent the prognosis of global patients; 3) only the metaanalysis of HOTAIR was composed of nine articles,⁴¹⁻⁴⁹ and other merged analyses about lncRNAs were from relatively small article number (two to four) and 4) the papers omitted due to no mention of OS may provide a lot of information on which lncRNAs hold promise for a prognostic value.

Inspirations

This study left several inspirations for us: 1) lncRNAs were arranged in an alphabetical order as shown in Table 1, via which the recently research frequency could be distinctly seen by clinical workers and scientific researchers; 2) the detailed outcomes of OS from the pooled analyses are shown in Table 3, through which combined detection of lncRNAs might better predict the survival time of GC patients and 3) for the molecular mechanisms of the included lncRNAs, their connections are shown in Figure 8, which might play enlightening roles in future basic experiments on lncRNAs in GC.

Study ID	HR (95% CI)	% weigh
Sox2ot		
Zhang et al (2016) ⁹⁶ (China)	← 2.05 (1.28, 3.30)	75.13
Zou et al (2016) ⁹⁷ (China)	3.24 (1.24, 6.43)	24.87
Subtotal (<i>I</i> ² =0.0%, <i>P</i> =0.345)	2.30 (1.52, 3.46)	100
SPRY4-IT1		
Peng et al (2015) ⁹⁸ (China)	0.82 (0.31, 1.57)	50.43
Xie et al (2015) ⁹⁹ (China)	2 .49 (1.08, 5.75)	49.57
Subtotal (I ² =71.4%, P=0.062)	1.42 (0.48, 4.22)	100
UCA1		
Zheng et al (2015) ¹⁰¹ (China)	2 .35 (1.22, 4.52)	24.68
Nasrollahzadeh-Khakiani et al (2017) ¹⁰² (TCGA)	1.11 (0.72, 1.73)	35.70
Zuo et al (2017) ¹⁰³ (China)	2.92 (1.07, 7.96)	14.07
Gu et al (2018) ¹⁰⁴ (China)	1.80 (0.95, 3.38)	25.55
Subtotal (<i>I</i> ² =45.5%, <i>P</i> =0.138)	> 1.73 (1.12, 2.68)	100
XIST		
Chen et al (2016) ¹⁰⁷ (China)	► 1.72 (1.32, 2.26)	74.55
Ma et al (2017) ¹⁰⁸ (China)	2.49 (1.40, 4.42)	25.45
Subtotal (<i>I</i> ² =23.4%, <i>P</i> =0.253)	1.89 (1.38, 2.59)	100
ZEB1-AS1	_	
Li et al (2017) ¹⁰⁹ (China)	2.36 (1.41, 3.96)	17.14
Zhang et al (2018) ¹¹⁰ (China)	2.72 (1.27, 5.84)	7.85
Zhang et al (2018) ¹¹⁰ (KM)	• 1.95 (1.52, 2.49)	75.01
Subtotal (/²=0.0%, P=0.617)	2.07 (1.67, 2.56)	100
ZFAS1	_	
Zhang et al (2016) ¹¹¹ (China)	2 .57 (1.25, 6.84)	54.51
Nie et al (2017) ¹¹² (China)	2.43 (0.96, 6.17)	45.49
Subtotal (<i>I</i> ² =0.0%, <i>P</i> =0.931)	2.51 (1.34, 4.69)	100
0.01 0.1 1	 10 100	

Figure 7 Forest plot of pooled analyses of OS in association with high Sox2ot, UCA1, XIST, ZEB1-AS1, ZFAS1 expression levels, or low SPRY4-IT1 expression levels. Note: Weights are from random-effects analysis.

Abbreviations: OS, overall survival; Sox2ot, SOX2 overlapping transcript; SPRY4-IT1, SPRY4 intronic transcript 1; UCA1, urothelial cancer associated 1; XIST, X inactive specific transcript; ZEB1-AS1, ZEB1 antisense RNA 1; ZFAS1, ZNFX1 antisense RNA 1.

Conclusion

This meta-analysis suggested that the abnormally expressed lncRNAs (AFAP1-AS1, ANRIL, CASC15, CCAT2, GAPLINC, H19, HOTAIR, HOTTIP, LINC00673, MALAT1, MEG3, PANDAR, PVT1, Sox2ot, UCA1, XIST, ZEB1-AS1 and ZFAS1) were significantly associated with the survival of GC patients, among which AFAP1-AS1, CCAT2, LINC00673, PANDAR, PVT1, Sox2ot, ZEB1-AS1 and ZFAS1 were strong candidates in predicting prognosis of GC patients.

Author contributions

Yue Zhang contributed toward study concept and design. Song Gao and Zhi-Ying Zhao were involved in acquisition of data. Song Gao, Zhi-Ying Zhao and Rong Wu carried out analysis and interpretation of data. Yue Zhang performed drafting of the manuscript. Song Gao, Zhi-Ying Zhao, Rong Wu, Yue Zhang and Zhen-Yong Zhang assisted with revision of manuscript. Yue Zhang and Zhen-Yong Zhang helped in supervision of work. All authors read and approved the final manuscript. All authors contributed toward data analysis,

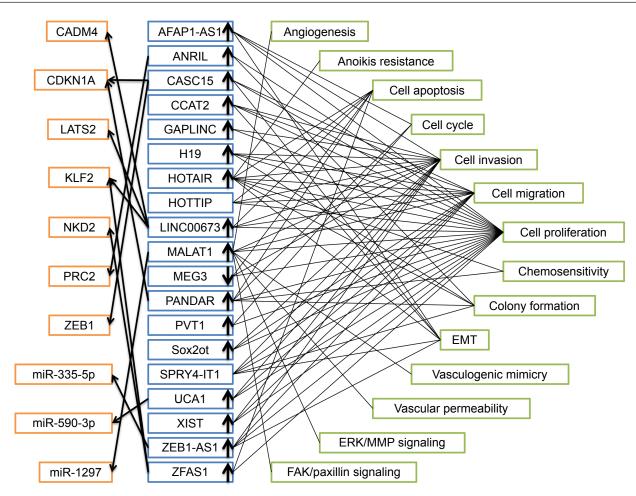


Figure 8 Summary of IncRNAs with altered expression, potential targets and pathways entered in this study. Abbreviations: AFAPI-ASI, AFAPI antisense RNA I; ANRIL, CDKN2B antisense RNA I; CASC15, cancer susceptibility 15; CCAT2, colon cancer associated transcript 2; EMT, epithelial–mesenchymal transition; GAPLINC, gastric adenocarcinoma associated, positive CD44 regulator, long intergenic noncoding RNA; H19, H19, imprinted maternally expressed transcript; HOTAIR, HOX transcript antisense RNA; HOTTIP, HOXA distal transcript antisense RNA; LINC00673, long intergenic non-protein coding RNA 673; IncRNA, long noncoding RNA; MALAT1, metastasis-associated lung adenocarcinoma transcript 1; MEG3, maternally expressed 3; PANDAR, promoter of CDKN1A antisense DNA damage activated RNA; PVT1, Pvt1 oncogene; Sox2ot, SOX2 overlapping transcript; SPRY4-IT1, SPRY4 intronic transcript 1; UCA1, urothelial cancer associated 1; XIST, X inactive specific transcript; ZEB1-AS1, ZEB1 antisense RNA 1; ZFAS1, ZNFX1 antisense RNA 1.

drafting and revising the paper and agree to be accountable for all aspects of the work.

Disclosure

The authors report no conflicts of interest in this work.

References

- Li F, Huang C, Li Q, Wu X. Construction and comprehensive analysis for dysregulated long non-coding RNA (IncRNA)-associated competing endogenous RNA (ceRNA) network in gastric cancer. *Med Sci Monit*. 2018;24:37–49.
- Chen X, Sun J, Song Y, et al. The novel long noncoding RNA AC138128.1 may be a predictive biomarker in gastric cancer. *Med Oncol.* 2014;31(11):262.
- Feng Y, Zhang Q, Wang J, Liu P. Increased IncRNA AFAP1-AS1 expression predicts poor prognosis and promotes malignant phenotypes in gastric cancer. *Eur Rev Med Pharmacol Sci.* 2017;21(17):3842–3849.
- 4. Qiao CF, Zhang Y, Jin L, Du XG, Qiao ZJ. High expression of lncRNA AFAP1-AS1 promotes cell proliferation and invasion by inducing epithelial-to-mesenchymal transition in gastric cancer. *Int J Clin Exp Pathol.* 2017;10(1):393–400.

- Qi F, Liu X, Wu H, et al. Long noncoding AGAP2-AS1 is activated by SP1 and promotes cell proliferation and invasion in gastric cancer. *J Hematol Oncol.* 2017;10(1):48.
- Huang Y, Zhang J, Hou L, et al. LncRNA AK023391 promotes tumorigenesis and invasion of gastric cancer through activation of the PI3K/ Akt signaling pathway. *J Exp Clin Cancer Res.* 2017;36(1):194.
- Fan ZY, Liu W, Yan C, et al. Identification of a five-lncRNA signature for the diagnosis and prognosis of gastric cancer. *Tumour Biol.* 2016; 37(10):13265–13277.
- Yang Z, Wang R, Zhang T, Dong X. Hypoxia/lncRNA-AK123072/ EGFR pathway induced metastasis and invasion in gastric cancer. *Int J Clin Exp Med.* 2015;8(11):19954–19968.
- Zhang EB, Kong R, Yin DD, et al. Long noncoding RNA ANRIL indicates a poor prognosis of gastric cancer and promotes tumor growth by epigenetically silencing of miR-99a/miR-449a. *Oncotarget*. 2014;5(8):2276–2292.
- Deng W, Wang J, Zhang J, Cai J, Bai Z, Zhang Z. TET2 regulates LncRNA-ANRIL expression and inhibits the growth of human gastric cancer cells. *IUBMB Life*. 2016;68(5):355–364.
- Saito T, Kurashige J, Nambara S, et al. A long non-coding RNA activated by transforming growth factor-β is an independent prognostic marker of gastric cancer. *Ann Surg Oncol.* 2015;22(suppl 3): S915–S922.

- Li L, Zhang L, Zhang Y, Zhou F. Increased expression of LncRNA BANCR is associated with clinical progression and poor prognosis in gastric cancer. *Biomed Pharmacother*. 2015;72:109–112.
- Liu X, Wang Y, Sun L, et al. Long noncoding RNA BC005927 upregulates EPHB4 and promotes gastric cancer metastasis under hypoxia. *Cancer Sci.* 2018;109(4):988–1000.
- Lü MH, Tang B, Zeng S, et al. Long noncoding RNA BC032469, a novel competing endogenous RNA, upregulates hTERT expression by sponging miR-1207-5p and promotes proliferation in gastric cancer. *Oncogene*. 2016;35(27):3524–3534.
- Sun TT, He J, Liang Q, et al. LncRNA GClnc1 promotes gastric carcinogenesis and may act as a modular scaffold of WDR5 and KAT2A complexes to specify the histone modification pattern. *Cancer Discov*. 2016;6(7):784–801.
- Wang L, Chunyan Q, Zhou Y, et al. BCAR4 increase cisplatin resistance and predicted poor survival in gastric cancer patients. *Eur Rev Med Pharmacol Sci.* 2017;21(18):4064–4070.
- Zhou J, Huang H, Tong S, Huo R. Overexpression of long non-coding RNA cancer susceptibility 2 inhibits cell invasion and angiogenesis in gastric cancer. *Mol Med Rep.* 2017;16(4):5235–5240.
- Yao XM, Tang JH, Zhu H, Jing Y. High expression of LncRNA CASC15 is a risk factor for gastric cancer prognosis and promote the proliferation of gastric cancer. *Eur Rev Med Pharmacol Sci.* 2017;21(24):5661–5667.
- Wu Q, Xiang S, Ma J, et al. Long non-coding RNA CASC15 regulates gastric cancer cell proliferation, migration and epithelial mesenchymal transition by targeting CDKN1A and ZEB1. *Mol Oncol.* 2018;12(6): 799–813.
- Liu JN, Shangguan YM. Long non-coding RNA CARLo-5 upregulation associates with poor prognosis in patients suffering gastric cancer. *Eur Rev Med Pharmacol Sci.* 2017;21(3):530–534.
- Wang CY, Hua L, Yao KH, Chen JT, Zhang JJ, Hu JH. Long non-coding RNA CCAT2 is up-regulated in gastric cancer and associated with poor prognosis. *Int J Clin Exp Pathol.* 2015;8(1):779–785.
- 22. Wang YJ, Liu JZ, Lv P, Dang Y, Gao JY, Wang Y. Long non-coding RNA CCAT2 promotes gastric cancer proliferation and invasion by regulating the E-cadherin and LATS2. *Am J Cancer Res.* 2016;6(11): 2651–2660.
- Pei YF, Zhang YJ, Lei Y, Wu DW, Ma TH, Liu XQ. Hypermethylation of the CHRDL1 promoter induces proliferation and metastasis by activating Akt and Erk in gastric cancer. *Oncotarget*. 2017;8(14): 23155–23166.
- Hao YP, Qiu JH, Zhang DB, Yu CG. Long non-coding RNA DANCR, a prognostic indicator, promotes cell growth and tumorigenicity in gastric cancer. *Tumour Biol*. 2017;39(6):1010428317699798.
- Xu TP, Wang YF, Xiong WL, et al. E2F1 induces TINCR transcriptional activity and accelerates gastric cancer progression via activation of TINCR/ STAU1/CDKN2B signaling axis. *Cell Death Dis.* 2017;8(6):e2837.
- Peng W, Wu J, Fan H, Lu J, Feng J. LncRNA EGOT promotes tumorigenesis Via Hedgehog Pathway in gastric cancer. *Pathol Oncol Res.* 2017.
- 27. Xu TP, Huang MD, Xia R, et al. Decreased expression of the long non-coding RNA FENDRR is associated with poor prognosis in gastric cancer and FENDRR regulates gastric cancer cell metastasis by affecting fibronectin1 expression. *J Hematol Oncol.* 2014;7:63.
- Wu X, Zhang P, Zhu H, Li S, Chen X, Shi L. Long noncoding RNA FEZF1-AS1 indicates a poor prognosis of gastric cancer and promotes tumorigenesis via activation of Wnt signaling pathway. *Biomed Pharmacother*. 2017;96:1103–1108.
- 29. Li CY, Liang GY, Yao WZ, et al. Integrated analysis of long non-coding RNA competing interactions reveals the potential role in progression of human gastric cancer. *Int J Oncol.* 2016;48(5):1965–1976.
- Chong DQ, Shan JL, Yang CS, Wang R, Du ZM. Clinical prognostic value of A FOXM1 related long non-coding RNA expression in gastric cancer. *Eur Rev Med Pharmacol Sci.* 2018;22(2):417–421.
- Feng L, Zhu Y, Zhang Y, Rao M. LncRNA GACAT3 promotes gastric cancer progression by negatively regulating miR-497 expression. *Biomed Pharmacother*. 2018;97:136–142.

- Hu Y, Wang J, Qian J, et al. Long noncoding RNA GAPLINC regulates CD44-dependent cell invasiveness and associates with poor prognosis of gastric cancer. *Cancer Res.* 2014;74(23):6890–6902.
- Liu L, Zhao X, Zou H, Bai R, Yang K, Tian Z. Hypoxia promotes gastric cancer malignancy partly through the HIF-1α dependent transcriptional activation of the long non-coding RNA GAPLINC. *Front Physiol*. 2016;7:420.
- 34. Sun M, Jin FY, Xia R, et al. Decreased expression of long noncoding RNA GAS5 indicates a poor prognosis and promotes cell proliferation in gastric cancer. *BMC Cancer*. 2014;14:319.
- Yang F, Xue X, Zheng L, et al. Long non-coding RNA GHET1 promotes gastric carcinoma cell proliferation by increasing c-Myc mRNA stability. *FEBS J.* 2014;281(3):802–813.
- Li H, Yu B, Li J, et al. Overexpression of lncRNA H19 enhances carcinogenesis and metastasis of gastric cancer. *Oncotarget*. 2014;5(8): 2318–2329.
- Zhang EB, Han L, Yin DD, Kong R, De W, Chen J. c-Myc-induced, long, noncoding H19 affects cell proliferation and predicts a poor prognosis in patients with gastric cancer. *Med Oncol.* 2014;31(5):914.
- Chen JS, Wang YF, Zhang XQ, et al. H19 serves as a diagnostic biomarker and up-regulation of H19 expression contributes to poor prognosis in patients with gastric cancer. *Neoplasma*. 2016;63(2): 223–230.
- Chen JF, Wu P, Xia R, et al. STAT3-induced lncRNA HAGLROS overexpression contributes to the malignant progression of gastric cancer cells via mTOR signal-mediated inhibition of autophagy. *Mol Cancer*. 2018;17(1):6.
- 40. Chen WM, Huang MD, Kong R, et al. Antisense long noncoding RNA HIF1A-AS2 is upregulated in gastric cancer and associated with poor prognosis. *Dig Dis Sci.* 2015;60(6):1655–1662.
- Endo H, Shiroki T, Nakagawa T, et al. Enhanced expression of long non-coding RNA HOTAIR is associated with the development of gastric cancer. *PLoS One*. 2013;8(10):e77070.
- 42. Xu ZY, Yu QM, Du YA, et al. Knockdown of long non-coding RNA HOTAIR suppresses tumor invasion and reverses epithelial-mesenchymal transition in gastric cancer. *Int J Biol Sci.* 2013;9(6):587–597.
- Okugawa Y, Toiyama Y, Hur K, et al. Metastasis-associated long noncoding RNA drives gastric cancer development and promotes peritoneal metastasis. *Carcinogenesis*. 2014;35(12):2731–2739.
- 44. Liu XH, Sun M, Nie FQ, et al. Lnc RNA HOTAIR functions as a competing endogenous RNA to regulate HER2 expression by sponging miR-331-3p in gastric cancer. *Mol Cancer*. 2014;13:92.
- 45. Zhang ZZ, Shen ZY, Shen YY, et al. HOTAIR long noncoding RNA promotes gastric cancer metastasis through suppression of poly r(C)-binding protein (PCBP) 1. *Mol Cancer Ther.* 2015;14(5): 1162–1170.
- 46. Zhao W, Dong S, Duan B, et al. HOTAIR is a predictive and prognostic biomarker for patients with advanced gastric adenocarcinoma receiving fluorouracil and platinum combination chemotherapy. *Am J Transl Res.* 2015;7(7):1295–1302.
- 47. Chen WM, Chen WD, Jiang XM, et al. HOX transcript antisense intergenic RNA represses E-cadherin expression by binding to EZH2 in gastric cancer. *World J Gastroenterol.* 2017;23(33):6100–6110.
- Feng X, Huang S. Effect and mechanism of lncRNA HOTAIR on occurrence and development of gastric cancer. J Cell Biochem. 2017.
- Li H, Li J, Zhang B, Zeng H. Long-chain non-coding RNA HOTAIR expression in tissue samples correlates with gastric cancer survival. *Int J Clin Exp Med.* 2018;11(2):856–862.
- Ye H, Liu K, Qian K. Overexpression of long noncoding RNA HOTTIP promotes tumor invasion and predicts poor prognosis in gastric cancer. *Onco Targets Ther.* 2016;9:2081–2088.
- Yang Y, Ma B, Yan Y, et al. Long non-coding RNA HOXA transcript at the distal tip as a biomarker for gastric cancer. *Oncol Lett.* 2017;14(1): 1068–1072.
- 52. Zhao R, Zhang Y, Zhang X, et al. Exosomal long noncoding RNA HOTTIP as potential novel diagnostic and prognostic biomarker test for gastric cancer. *Mol Cancer*. 2018;17(1):68.

- Xie M, Sun M, Zhu YN, et al. Long noncoding RNA HOXA-AS2 promotes gastric cancer proliferation by epigenetically silencing P21/PLK3/ DDIT3 expression. *Oncotarget*. 2015;6(32):33587–33601.
- Sun M, Nie F, Wang Y, et al. LncRNA HOXA11-AS promotes proliferation and invasion of gastric cancer by scaffolding the chromatin modification factors PRC2, LSD1, and DNMT1. *Cancer Res.* 2016;76(21): 6299–6310.
- Ma B, Wang J, Song Y, et al. Upregulated long intergenic noncoding RNA KRT18P55 acts as a novel biomarker for the progression of intestinal-type gastric cancer. *Onco Targets Ther.* 2016;9:445–453.
- Zhou B, Jing XY, Wu JQ, Xi HF, Lu GJ. Down-regulation of long noncoding RNA LET is associated with poor prognosis in gastric cancer. *Int J Clin Exp Pathol*. 2014;7(12):8893–8898.
- 57. Shan Y, Ying R, Jia Z, et al. LINC00052 promotes gastric cancer cell proliferation and metastasis via activating the Wnt/β-catenin signaling pathway. *Oncol Res.* 2017;25(9):1589–1599.
- Chen WM, Huang MD, Sun DP, et al. Long intergenic non-coding RNA 00152 promotes tumor cell cycle progression by binding to EZH2 and repressing p15 and p21 in gastric cancer. *Oncotarget*. 2016;7(9): 9773–9787.
- Zhang ZZ, Zhao G, Zhuang C, et al. Long non-coding RNA LINC00628 functions as a gastric cancer suppressor via long-range modulating the expression of cell cycle related genes. *Sci Rep.* 2016;6:27435.
- 60. Zhang E, Yin D, Han L, et al. E2F1-induced upregulation of long noncoding RNA LINC00668 predicts a poor prognosis of gastric cancer and promotes cell proliferation through epigenetically silencing of CKIs. *Oncotarget*. 2016;7(17):23212–23226.
- Ba MC, Long H, Cui SZ, et al. Long noncoding RNA LINC00673 epigenetically suppresses KLF4 by interacting with EZH2 and DNMT1 in gastric cancer. *Oncotarget*. 2017;8(56):95542–95553.
- Huang M, Hou J, Wang Y, et al. Long noncoding RNA LINC00673 is activated by SP1 and exerts oncogenic properties by interacting with LSD1 and EZH2 in gastric cancer. *Mol Ther*. 2017;25(4):1014–1026.
- Zeng S, Xie X, Xiao YF, et al. Long noncoding RNA LINC00675 enhances phosphorylation of vimentin on Ser83 to suppress gastric cancer progression. *Cancer Lett.* 2018;412:179–187.
- Fei ZH, Yu XJ, Zhou M, Su HF, Zheng Z, Xie CY. Upregulated expression of long non-coding RNA LINC00982 regulates cell proliferation and its clinical relevance in patients with gastric cancer. *Tumour Biol.* 2016;37(2):1983–1993.
- 65. Miao Y, Sui J, Xu SY, Liang GY, Pu YP, Yin LH. Comprehensive analysis of a novel four-lncRNA signature as a prognostic biomarker for human gastric cancer. *Oncotarget*. 2017;8(43):75007–75024.
- Chen X, Chen Z, Yu S, et al. Long noncoding RNA LINC01234 functions as a competing endogenous RNA to regulate CBFB expression by sponging miR-204-5p in gastric cancer. *Clin Cancer Res.* 2018;24(8): 2002–2014.
- Qin QH, Yin ZQ, Li Y, Wang BG, Zhang MF. Long intergenic noncoding RNA 01296 aggravates gastric cancer cells progress through miR-122/MMP-9. *Biomed Pharmacother*. 2018;97:450–457.
- Zou Z, Ding Q, Li P, et al. Overexpression of lincRNA-ROR predicts poor prognosis in patients with gastric cancer. *Int J Clin Exp Pathol*. 2016;9(9):9467–9472.
- Hu Y, Pan J, Wang Y, Li L, Huang Y. Long noncoding RNA linc-UBC1 is negative prognostic factor and exhibits tumor pro-oncogenic activity in gastric cancer. *Int J Clin Exp Pathol.* 2015;8(1):594–600.
- Guo W, Dong Z, Shi Y, et al. Methylation-mediated downregulation of long noncoding RNA LOC100130476 in gastric cardia adenocarcinoma. *Clin Exp Metastasis.* 2016;33(5):497–508.
- Zhao Y, Liu Y, Lin L, et al. The lncRNA MACC1-AS1 promotes gastric cancer cell metabolic plasticity via AMPK/Lin28 mediated mRNA stability of MACC1. *Mol Cancer*. 2018;17(1):69.
- Qi Y, Ooi HS, Wu J, et al. MALAT1 long ncRNA promotes gastric cancer metastasis by suppressing PCDH10. *Oncotarget*. 2016;7(11): 12693–12703.
- Li J, Gao J, Tian W, Li Y, Zhang J. Long non-coding RNA MALAT1 drives gastric cancer progression by regulating HMGB2 modulating the miR-1297. *Cancer Cell Int.* 2017;17:44.

- Li Y, Wu Z, Yuan J, et al. Long non-coding RNA MALAT1 promotes gastric cancer tumorigenicity and metastasis by regulating vasculogenic mimicry and angiogenesis. *Cancer Lett.* 2017;395:31–44.
- 75. Chen F, Tian Y, Pang EJ, Wang Y, Li L. MALAT2-activated long noncoding RNA indicates a biomarker of poor prognosis in gastric cancer. *Cancer Gene Ther*. 2015.
- 76. Sun M, Xia R, Jin F, et al. Downregulated long noncoding RNA MEG3 is associated with poor prognosis and promotes cell proliferation in gastric cancer. *Tumour Biol*. 2014;35(2):1065–1073.
- 77. Guo W, Dong Z, Liu S, et al. Promoter hypermethylation-mediated downregulation of miR-770 and its host gene MEG3, a long noncoding RNA, in the development of gastric cardia adenocarcinoma. *Mol Carcinog.* 2017;56(8):1924–1934.
- Nie FQ, Ma S, Xie M, Liu YW, De W, Liu XH. Decreased long noncoding RNA MIR31HG is correlated with poor prognosis and contributes to cell proliferation in gastric cancer. *Tumour Biol.* 2016;37(6): 7693–7701.
- Quan Y, Zhang Y, Lin W, et al. Knockdown of long non-coding RNA MAP3K20 antisense RNA 1 inhibits gastric cancer growth through epigenetically regulating miR-375. *Biochem Biophys Res Commun.* 2018;497(2):527–534.
- Lai Y, Xu P, Liu J, et al. Decreased expression of the long non-coding RNA MLLT4 antisense RNA 1 is a potential biomarker and an indicator of a poor prognosis for gastric cancer. *Oncol Lett.* 2017;14(3):2629–2634.
- Wang Y, Zhang D, Wu K, Zhao Q, Nie Y, Fan D. Long noncoding RNA MRUL promotes ABCB1 expression in multidrug-resistant gastric cancer cell sublines. *Mol Cell Biol*. 2014;34(17):3182–3193.
- Lin Z, Lai S, He X, et al. Decreased long non-coding RNA MTM contributes to gastric cancer cell migration and invasion via modulating MT1F. *Oncotarget*. 2017;8(57):97371–97383.
- Fu JW, Kong Y, Sun X. Long noncoding RNA NEAT1 is an unfavorable prognostic factor and regulates migration and invasion in gastric cancer. J Cancer Res Clin Oncol. 2016;142(7):1571–1579.
- Guo X, Yang Z, Zhi Q, et al. Long noncoding RNA OR3A4 promotes metastasis and tumorigenicity in gastric cancer. *Oncotarget*. 2016;7(21): 30276–30294.
- Wang YQ, Zhang QY, Weng WW, et al. Upregulation of the non-coding RNA OTUB1-isoform 2 contributes to gastric cancer cell proliferation and invasion and predicts poor gastric cancer prognosis. *Int J Biol Sci.* 2016;12(5):545–557.
- Ma P, Xu T, Huang M, Shu Y. Increased expression of LncRNA PANDAR predicts a poor prognosis in gastric cancer. *Biomed Pharmacother*. 2016;78:172–176.
- Liu J, Ben Q, Lu E, et al. Long noncoding RNA PANDAR blocks CDKN1A gene transcription by competitive interaction with p53 protein in gastric cancer. *Cell Death Dis.* 2018;9(2):168.
- Bi M, Yu H, Huang B, Tang C. Long non-coding RNA PCAT-1 overexpression promotes proliferation and metastasis in gastric cancer cells through regulating CDKN1A. *Gene.* 2017;626:337–343.
- Kong R, Zhang EB, Yin DD, et al. Long noncoding RNA PVT1 indicates a poor prognosis of gastric cancer and promotes cell proliferation through epigenetically regulating p15 and p16. *Mol Cancer*. 2015;14:82.
- Yuan CL, Li H, Zhu L, Liu Z, Zhou J, Shu Y. Aberrant expression of long noncoding RNA PVT1 and its diagnostic and prognostic significance in patients with gastric cancer. *Neoplasma*. 2016;63(3):442–449.
- Sun J, Song Y, Chen X, et al. Novel long non-coding RNA RP11-119F7.4 as a potential biomarker for the development and progression of gastric cancer. *Oncol Lett.* 2015;10(1):115–120.
- Song W, Liu YY, Peng JJ, et al. Identification of differentially expressed signatures of long non-coding RNAs associated with different metastatic potentials in gastric cancer. *J Gastroenterol*. 2016;51(2):119–129.
- Hu Y, Ma Z, He Y, Liu W, Su Y, Tang Z. LncRNA-SNHG1 contributes to gastric cancer cell proliferation by regulating DNMT1. *Biochem Biophys Res Commun*. 2017;491(4):926–931.
- 94. Yan K, Tian J, Shi W, Xia H, Zhu Y. LncRNA SNHG6 is associated with poor prognosis of gastric cancer and promotes cell proliferation and EMT through epigenetically silencing p27 and sponging miR-101-3p. *Cell Physiol Biochem.* 2017;42(3):999–1012.

Gao et al

- Zhang H, Lu W. LncRNA SNHG12 regulates gastric cancer progression by acting as a molecular sponge of miR-320. *Mol Med Rep.* 2018; 17(2):2743–2749.
- Zhang Y, Yang R, Lian J, Xu H. LncRNA Sox2ot overexpression serves as a poor prognostic biomarker in gastric cancer. *Am J Transl Res.* 2016;8(11):5035–5043.
- Zou JH, Li CY, Bao J, Zheng GQ. High expression of long noncoding RNA Sox2ot is associated with the aggressive progression and poor outcome of gastric cancer. *Eur Rev Med Pharmacol Sci.* 2016;20(21): 4482–4486.
- Peng W, Wu G, Fan H, Wu J, Feng J. Long noncoding RNA SPRY4-IT1 predicts poor patient prognosis and promotes tumorigenesis in gastric cancer. *Tumour Biol.* 2015;36(9):6751–6758.
- Xie M, Nie FQ, Sun M, et al. Decreased long noncoding RNA SPRY4-IT1 contributing to gastric cancer cell metastasis partly via affecting epithelial-mesenchymal transition. *J Transl Med.* 2015;13:250.
- 100. Zhang E, He X, Yin D, et al. Increased expression of long noncoding RNA TUG1 predicts a poor prognosis of gastric cancer and regulates cell proliferation by epigenetically silencing of p57. *Cell Death Dis.* 2016;7:e2109.
- 101. Zheng Q, Wu F, Dai WY, et al. Aberrant expression of UCA1 in gastric cancer and its clinical significance. *Clin Transl Oncol.* 2015; 17(8):640–646.
- 102. Nasrollahzadeh-Khakiani M, Emadi-Baygi M, Nikpour P. Augmented expression levels of lncRNAs ecCEBPA and UCA1 in gastric cancer tissues and their clinical significance. *Iran J Basic Med Sci.* 2017;20(10):1149–1158.
- 103. Zuo ZK, Gong Y, Chen XH, et al. TGFβ1-induced LncRNA UCA1 upregulation promotes gastric cancer invasion and migration. DNA Cell Biol. 2017;36(2):159–167.
- 104. Gu L, Lu LS, Zhou DL, Liu ZC. UCA1 promotes cell proliferation and invasion of gastric cancer by targeting CREB1 sponging to miR-590-3p. *Cancer Med.* 2018;7(4):1253–1263.
- 105. Chen M, Wu X, Ma W, et al. Decreased expression of lncRNA VPS9D1-AS1 in gastric cancer and its clinical significance. *Cancer Biomark*. 2017;21(1):23–28.
- 106. Cai J, Wang D, Bai ZG, Yin J, Zhang J, Zhang ZT. The long noncoding RNA XIAP-AS1 promotes XIAP transcription by XIAP-AS1 interacting with Sp1 in gastric cancer cells. *PLoS One*. 2017;12(8): e0182433.
- 107. Chen DL, Ju HQ, Lu YX, et al. Long non-coding RNA XIST regulates gastric cancer progression by acting as a molecular sponge of miR-101 to modulate EZH2 expression. *J Exp Clin Cancer Res.* 2016;35(1):142.
- Ma L, Zhou Y, Luo X, Gao H, Deng X, Jiang Y. Long non-coding RNA XIST promotes cell growth and invasion through regulating miR-497/ MACC1 axis in gastric cancer. *Oncotarget*. 2017;8(3):4125–4135.

- Li Y, Wen X, Wang L, et al. LncRNA ZEB1-AS1 predicts unfavorable prognosis in gastric cancer. Surg Oncol. 2017;26(4):527–534.
- 110. Zhang LL, Zhang LF, Guo XH, Zhang DZ, Yang F, Fan YY. Downregulation of miR-335-5p by long noncoding RNA ZEB1-AS1 in gastric cancer promotes tumor proliferation and invasion. *DNA Cell Biol.* 2018;37(1):46–52.
- 111. Zhang JJ, Chen JT, Yao KH, Hua L, Wang CY, Hu JH. Up-regulated expression of long non-coding RNA ZFAS1 associates with aggressive tumor progression and poor prognosis in gastric cancer patients. *Int J Clin Exp Pathol.* 2016;9(2):2059–2063.
- Nie F, Yu X, Huang M, et al. Long noncoding RNA ZFAS1 promotes gastric cancer cells proliferation by epigenetically repressing KLF2 and NKD2 expression. *Oncotarget*. 2017;8(24):38227–38238.
- 113. Lai Y, Xu P, Li Q, et al. Downregulation of long noncoding RNA ZMAT1 transcript variant 2 predicts a poor prognosis in patients with gastric cancer. *Int J Clin Exp Pathol*. 2015;8(5):5556–5562.
- McLean MH, El-Omar EM. Genetics of gastric cancer. Nat Rev Gastroenterol Hepatol. 2014;11(11):664–674.
- Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A. Global cancer statistics, 2012. *CA Cancer J Clin.* 2015;65(2):87–108.
- Siegel RL, Miller KD, Jemal A. Cancer statistics, 2015. CA Cancer J Clin. 2015;65(1):5–29.
- 117. GBD 2013 Mortality and Causes of Death Collaborators. Global, regional, and national age-sex specific all-cause and cause-specific mortality for 240 causes of death, 1990–2013: a systematic analysis for the Global Burden of Disease Study 2013. *Lancet*. 2015;385(9963): 117–171.
- 118. Ponting CP, Oliver PL, Reik W. Evolution and functions of long noncoding RNAs. *Cell*. 2009;136(4):629–641.
- Cheetham SW, Gruhl F, Mattick JS, Dinger ME. Long noncoding RNAs and the genetics of cancer. *Br J Cancer*. 2013;108(12):2419–2425.
- Spizzo R, Almeida MI, Colombatti A, Calin GA. Long non-coding RNAs and cancer: a new frontier of translational research? *Oncogene*. 2012;31(43):4577–4587.
- 121. Li T, Mo X, Fu L, Xiao B, Guo J. Molecular mechanisms of long noncoding RNAs on gastric cancer. *Oncotarget*. 2016;7(8):8601–8612.
- Parmar MK, Torri V, Stewart L. Extracting summary statistics to perform meta-analyses of the published literature for survival endpoints. *Stat Med.* 1998;17(24):2815–2834.
- 123. Xu MD, Qi P, Weng WW, et al. Long non-coding RNA LSINCT5 predicts negative prognosis and exhibits oncogenic activity in gastric cancer. *Medicine (Baltimore)*. 2014;93(28):e303.
- 124. Dan J, Wang J, Wang Y, et al. LncRNA-MEG3 inhibits proliferation and metastasis by regulating miRNA-21 in gastric cancer. *Biomed Pharmacother*. 2018;99:931–938.

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