a Open Access Full Text Article

ORIGINAL RESEARCH

Graphene-Based Nanoparticles as Potential Treatment Options for Parkinson's Disease: A Molecular Dynamics Study

This article was published in the following Dove Press journal: International Journal of Nanomedicine

amyloids could prevent their fibrillation.

Ehsan Alimohammadi Mohammad Khedri^{2,*} Ahmad Miri Jahromi³ Reza Maleki Milad Rezaian⁵

¹Neurosurgery Department, Kermanshah University of Medical Sciences, Kermanshah, Iran; ²Department of Chemical Engineering, Amirkabir University of Technology (Tehran Polytechnic), Tehran 1591634311, Iran; ³Department of Petroleum Engineering, Amirkabir University of Technology (Tehran Polytechnic), Tehran 1591634311, Iran; ⁴Department of Chemical Engineering, Sharif University of Technology, Tehran, Iran; ⁵Department of Pharmacology, School of Medicine, Shahid Beheshti University of Medical Sciences, Tehran 19839-63113, Iran

*These authors contributed equally to this work

Correspondence: Reza Maleki Department of Chemical Engineering, Sharif University of Technology, Tehran, Iran Email Rezamaleki96@gmail.com



Introduction: The study of abnormal aggregation of proteins in different tissues of the body has recently earned great attention from researchers in various fields of science. Concerning neurological diseases, for instance, the accumulation of amyloid fibrils can contribute to Parkinson's disease, a progressively severe neurodegenerative disorder. The most prominent features of this disease are the degeneration of neurons in the substantia nigra and accumulation of α -synuclein aggregates, especially in the brainstem, spinal cord, and cortical areas. Dopamine replacement therapies and other medications have reduced motor impairment and had positive consequences on patients' quality of life. However, if these medications are stopped, symptoms of the disease will recur even more severely. Therefore, the improvement of therapies targeting more basic mechanisms like prevention of amyloid formation seems to be critical. It has been shown that the interactions between monolayers like graphene and

Methods: For the first time, the impact of four types of last-generation graphene-based nanostructures on the prevention of α -synuclein amyloid fibrillation was investigated in this study by using molecular dynamics simulation tools.

Results: Although all monolayers were shown to prevent amyloid fibrillation, nitrogendoped graphene (N-Graphene) caused the most instability in the secondary structure of α synuclein amyloids. Moreover, among the four monolayers, N-Graphene was shown to present the highest absolute value of interaction energy, the lowest contact level of amyloid particles, the highest number of hydrogen bonds between water and amyloid molecules, the highest instability caused in α -synuclein particles, and the most significant decrease in the compactness of α -synuclein protein.

Discussion: Ultimately, it was concluded that N-Graphene could be the most effective monolayer to disrupt amyloid fibrillation, and consequently, prevent the progression of Parkinson's disease.

Keywords: α-synuclein, amyloid, graphene, Parkinson's disease, molecular dynamics

Introduction

According to the World Health Organization, recent evidence has revealed the accumulation of amyloid molecules to be the fundamental mechanism underlying many health care disorders. (KT= 2.479 kJ/mol) A significant number of diseases, including many neurodegenerative disorders, are known to be associated with the formation of stable, fibrillar protein aggregates called amyloids.^{1,3} These aggregations are made in specific steps, during which a specific protein or part of a protein

6887

International Journal of Nanomedicine downloaded from https://www.dovepress.com/ For personal use only

> CO 0 S © 2020 Alimohammadi et al. This work is published and licensed by Dove Medical Press Limited. The full terms of this license are available at https://www.dovepress.com/ terms.php and incorporate the Creative Commons. Attribution — Non Commercial (unported, v3.0) License (http://creativecommons.org/licenses/by-nc/3.0/). By accessing the work you hereby accept the Terms. Non-commercial uses of the work are permitted without any further permission from Dove Medical Press Limited, provided the work is properly attributed. For permission for commercial use of this work, please see paragraphs 4.2 and 5 of our Terms (https://www.dovepress.com/terms.php).

International Journal of Nanomedicine 2020:15 6887-6903

changes from its natural soluble state to insoluble form and eventually evolves into giant fibrillar proteins that accumulate in various tissues and organs of the body.^{4–6} Parkinson's disease, one of the common motor abnormalities, is the second most well-known neurodegenerative disorder after Alzheimer's.⁷ It is caused by the destruction of dopamine-producing cells in the substantia nigra of the brain's basal ganglia.⁸ As the brain's dopamine reserves decrease in proportionate to acetylcholine, symptoms of the disease appear. So far, there has been no definitive cure for Parkinson's, even though many drugs such as levodopa, amantadine, biperiden, and selegiline have been used to treat it^{9-13} As far as the patients take these drugs, their symptoms are controlled. If, however, these medications are stopped, symptoms of the disease will recur even more severely. This fact highlights the importance of developing medications that are targeted on more basic underlying mechanisms of the disease. It is believed that one pathophysiology beyond the dopamine-producing cell death involves the abnormal accumulation of amyloid fibrils in damaged cells. Amyloid fibrils consist of betasheeted α -synuclein monomers, in which the beta strands are perpendicular to the principal axis of the fibril. Alphasynuclein is a member of the synuclein proteins which

have critical intracellular functions. Even though the precise actions of natural α -synuclein protein in the nervous system are remained unclear, studies have revealed this protein to play critical roles in the synaptic transmission, including the synthesis, axonal transport, release, and recycling of neurotransmitters. Functional proteins usually do not form amyloids unless they lose their natural folding. The process can be self-generated, meaning that a misfolded protein can induce the same dysfunction in other similar proteins. The misfolded peptides can form soluble aggregates known as oligomers, which evolve into protofibrils, amyloid fibrils, and Lewy bodies, all of which have cytotoxic effects and lead to the death of neurons by a variety of intracellular mechanisms.^{14–17} Figure 1 illustrates the process of amyloid formation and Parkinson's disease development.

Proteins may bind to monolayers in natural or denatured states, depending on the surface properties of the protein (such as load, hydrophobicity, stability) and the monolayer (hydrophobicity, size, coating). The properties of monolayers can affect the self-assembly and biological functions of proteins.¹⁸ According to studies,^{19–23} monolayers can affect the fibrillation process with different mechanisms which could be listed as follows:



Figure I The mechanism underlying the development of Parkinson's disease.

- Monolayers can increase the number of hydrogen bonds between amyloid fibrils and water molecules, and therefore, block the active elongation site of the fibrils;¹⁸
- 2. Monolayers can prevent the fibrillation of proteins by disrupting their secondary structure;
- 3. The high local concentration of proteins on the surface of some monolayers can lead to the formation of critical nuclei, and in such cases, facilitate the formation of amyloid fibrils;¹⁸
- By their strong interactions with proteins, the monolayer can reduce the concentration of proteins in the solution, which will delay the appearance of critical nuclei, and hence, prevent fibril formation;¹⁸
- 5. Specific monolayers can act as chaperons.²⁴ Chaperons can bind to proteins through hydrophobic and electrostatic interactions, prevent their aggregation, and help them return to their original folding state.²⁵

Several studies have been performed to investigate the use of monolayers as potential agents to prevent amyloid fibrillation. In these studies, quantum dots (QDs),²⁰ gold nanoparticles,^{21–23} superparamagnetic iron oxide nanoparticles (SPION), graphene-based structures with positive and negative charges¹⁹ have been investigated.

With the discovery of graphene, the use of twodimensional structures in nanomaterial-based systems came into consideration.²⁶⁻³⁰ Generally, each carbon atom can make covalent bonds with a maximum of four other carbon atoms, forming a three-dimensional network, known as the diamond. If, however, each carbon atom only bonds with three other carbon atoms, a twodimensional carbon allotrope is formed, called graphene. The surface area of graphene is much larger than that of carbon black and activated carbon. The hexagonal pattern of carbon atoms in graphene has turned this nanostructure into a giant aromatic molecule with unique electrical, optical, and thermal properties and excellent mechanical strength despite its small thickness and relatively low molecular weight. These features have facilitated the use of graphene in a variety of fields, including electronics, pharmaceutics, and the textile industry.³¹⁻³³ In order to improve its physicochemical properties as to fit nanomaterial practice better, the basic structure of graphene has undergone novel modifications through which new generations of this carbon allotrope have been developed over time. Second-generation graphene molecules were

established by chemical functionalization. This method has been vastly employed to improve the properties of this monolayer for many years. However, despite their advantageous features, chemically functionalized graphenes presented certain limitations as their potential utilities for innovative purposes began to emerge. Attempts to further adjust graphene molecules as to be suited for novel implementations gave rise to the emergence of lastgeneration carbon sheet allotropes or *-doped graphenes*. Such structures are designed as a proportion of carbon atoms in the mainstay composition are replaced with other elements such as bromine, nitrogen, phosphorus, etc.^{1,34,35}

Graphene-based nanomaterials like all of the engineered biomaterials, have been shown to have cytotoxic effects. However, in order to reach the toxic level, they have to be administered more than their toxic dose, so future experimental animal studies and clinical trials should be conducted in a way that the administered doses of graphene -based nanomaterials do not reach the toxic dose.^{36–40} Functionalized graphene, such as carboxylated graphene, has shown much lower toxicity and better hydrophilicity than pristine graphene. However, doped graphene represents a newer generation of graphene-based nanomaterials which has been shown to have more desirable chemical properties and better biocompatibility^{36,41–44}.

The implementation of graphene-based nanomaterials for the treatment of Parkinson's disease have been the topic of a number of previously published computational studies, some of which have also been backed up by experimental evidence. Such researches contributed to the development of the main concept of this study which was performed.^{19,36,41,44-47} This study has investigated four types of last-generation graphene-based nanostructures as potential agents to disrupt the process of α-synuclein amyloid formation in Parkinson's disease by using molecular dynamics (MD) simulation techniques. Different parameters involved in the interactions between α -synuclein proteins and the nanostructures have been analyzed. These monolayers include graphene, nitrogen-doped graphene (N-Graphene), phosphorus-doped graphene (P-Graphene), and graphene co-duped with bromine and nitrogen (BCN). Moreover, in order to validate the methods and algorithms used in this paper, an additional simulation of graphene polyglycerol interactions with α -synuclein protein was performed with conditions identical to a previously published similar study by Mohammad-Beigi et al, and the results were compared with each other.

Materials and Methods Molecular Dynamics Simulation

Molecular dynamics (MD) is a method for analyzing the physical motion of atoms and molecules. Molecular dynamics simulations produce information at the microscopic level (position and velocity of atoms). In simulating the molecular dynamics, atoms and molecules interact with each other over a period of time.⁴⁸ The trajectory of atoms and molecules is determined by the numerical solution of Newton's motion equation. A molecular system typically contains a large number of components, and the properties of such complex systems cannot be determined analytically. For this reason, numerical methods are used.

These data are converted to macroscopic values (such as pressure and energy) using statistical mechanics. MD and statistical mechanics link microscopic concepts and observable macroscopic quantities. In molecular dynamics simulations, Newton's second law is used to examine the temporal evolution of systems. By solving Newton's motion equations, the position of the atom (r_i) in a N atomic system is obtained:

$$m_i\left(\frac{d^2r_i}{dt^2}\right) = F_i, i = 1, \dots, N$$

 m_i is the mass of the atom, and F_i is the force that other atoms exert on atom *i*. In fact, in MD simulation methods with initial positions and velocities, the temporal evolution of the system can be examined. By integrating Newton's motion equations, a path is obtained that shows how the velocity, position, and acceleration of particles change over time. In molecular dynamics, to measure a physical quantity, the average of that quantity is calculated at the time of simulation in the system path.^{49–51}

Simulation Method

Alpha-protein-synuclein contains 140 amino acids. This protein is made up of alpha, beta, gamma-synuclein and synutrin structures. The amount of alpha-synuclein protein in the central nervous system (CNS) is very high. Alpha-synuclein is naturally present in the cell cytoplasm. In this study we used from molecular structure of the alpha-synuclein protein that has been studied by Ulmer et al.⁵²

The graphene structure is constructed by Nanotube_Modeler_1.7.9,^{53,54} with a size of 70 \AA \times 170 \AA in X-Y Cartesian coordinates. Then, with the help of Avogadro software, the molecular structures of BCN monolayers, a combination of nitrogen and bromine atoms with graphene carbon atoms, N-Graphene



Figure 2 This figure illustrates how α -synuclein proteins change their folding in the absence and presence of each monolayer: (A) Graphene; (B) N-Graphene; (C) P-Graphene; (D) BCN; (E) without monolayer.

monolayers, a combination of nitrogen atoms with graphene carbon atoms, P-Graphene monolayers, a combination of phosphorus atoms with graphene carbon atoms, and pure graphene were made. The molecular structures of nitrogen, bromine, and phosphorus-graphene graphene have been optimized by Gaussian 09 software.^{55,56}

To perform the simulation, the monolayer topology was designed using the x2top command and the amyloid topology by the pdb2gmx command for the optimized potentials for liquid simulations-All Atoms (OPLS-AA) force field.^{57,58} The simulation is performed in 4 steps, with energy level optimization and 50001fs time steps. The simulation system is coupled with the Berendsen thermostat, considering the equilibrium temperature of 310K in the nvt step. The execution time of the nvt step was 100 picoseconds.^{59–61} Moreover, the npt step is balanced by the Parrinello_Rahman algorithm in 10 nanoseconds at 1 bar.^{62–64} At the end of the simulation, with considering the cut-off radius is 1.4 nanometers and h_bond idone by the lincs algorithm in 40 nanoseconds. Last but not least, simulation images were taken by vmd software.



Figure 3 Changes in the α -synuclein structure during the simulation time in the presence of each monolayer.



Figure 4 Van der Waals, electrostatic, and total energy of interactions between α -synuclein protein and each monolayer versus time: (A) Graphene; (B) N- Graphene; (C) P- Graphene; (D) BCN.

Root-Mean-Square Deviation Analysis

Root-Mean-Square Deviation (RMSD) values are calculated by Equation (2). In Equation (2), *n* represents the number of particles, $Po_i(t)$ and $Po_r(t)$ show the position of the particle *i* and the reference particle at time *t*.

$$RMSD = \sqrt{\frac{1}{n}\sum\limits_{i=1}^{n}\left(Po_{i}(t)-Po_{r}(t)\right)^{2}}$$

Energy Analysis

Van Der Walls (VDW) energy is calculated based on the Lenard-Jones equation (Equation 3). The energy of electrostatic interactions follows Columbus' law. Colum's law is shown in Equation 4.

$$\mathbf{V}_{\mathrm{vdw}} = 4 \in \left[\left(\frac{\sigma}{r} \right)^{12} - \left(\frac{\sigma}{r} \right)^{6} \right]$$

Table I Average Values for VDW, Electrostatic, and the Total Energy of Interactions Between α -Synuclein Protein and Monolayers During the Simulation Process Monolayer

Average Values	Graphene	N-Graphene	P-Graphene	BCN
Van der Waals Energy(KT) ¹ Electrostatic energy(KT)	-1494.07 28.785	-1067.486 -541.636	-323.186 -50.161	-503.986 -0.001
Total energy(KT)	-1465.28	-1609.122	-373.347	-503.987

$$U_{\{E\}}(r) = (4\pi\epsilon)^{-1} \frac{q_i q_j}{r_{ij}^2}$$

where *r* is the distance between these two particles, *q* is the amount of charge mentioned in the topology for each particle, *V* is the potential between the two atoms, and is the depth of the potential well. In this paper, energy analysis for different simulations is taken by mmpbsa software.^{65,66}

Gyration Radius

The gyration radius is calculated by Equation (5):

$$R_g = \sqrt{\frac{\sum_{i=1}^n r_i^2 m_i}{\sum_{i=1}^n m_i}}$$

where *n* is the number of particles, m_i , the mass of the particle *i*, and r_i indicate the distance of the particle *i* from the center of gravity. To measure the density of amyloid particles, an analysis of the radius of amyloid gyration was performed in all simulations. R_g , helps to understand protein density during the simulation, where for higher R_g , higher density is expected.

SASA

In this paper, the solvent accessible surface area (SASA) analysis for amyloid particles is taken in all simulations.

The contact level of amyloid particles in this paper is calculated by Equation 6.

$$ContactArea(t) = 0.5 \times (ca_0 - ca_t)$$

where ca_0 and ca_t show the amount of SASA analysis at zero and at time *t*, respectively. Mohammad-Beigi et al¹⁹ have simulated graphene polyglycerol and amyloid in water using the OPLS-AA force field by Gromacs. In this study, the simulation algorithms, force field, molecular structures and simulation time were selected according to Mohammad-Beigi et al¹⁹ and the LINCS algorithm with the cut-off radius of 1.4 nm was implemented.

Results and Discussion

Formation of α -Synuclein Amyloids

Figure 2–3 illustrate how amyloid formation is affected by monolayers during 40 nanoseconds simulation. In order to form amyloids, α -synuclein proteins first become misfolded and then gradually rise to amyloids. Figure 2E shows the misfolded form of α -synuclein protein at the end of the simulation in the absence of monolayers. As can be seen in Figure 2A-D, the presence of monolayers disrupts the misfolding of α -synuclein proteins, giving rise to other forms which are not suitable for amyloid formation.



Figure 5 Rg of α-synuclein proteins versus the simulation time in the presence of (A) Graphene; (B) N- Graphene; (C) P- Graphene; (D) BCN; (E) Without monolayers.

Table 2 Values of the Gyration Radius Variation for a	x-Synuclein
Proteins in the Presence of Each Monolayer	

Monolayer	Gyration Radius Variation (Rg0-Rg40)
Graphene	1.628
N-Graphene	0.683
P-Graphene	2.071
BCN	2.939
Without monolayer	3.074

Among these monolayers, N-Graphene provides better prevention of α -synuclein misfolding.

Evaluating the Interaction Energies

The amount and type of interaction energy are essential parameters as far as the interactions between monolayers are concerned. In general, the greater the absolute amount of interaction energy, the stronger are the interactions between the particles. This fact could play an important role in the prevention of amyloid formation. More absolute amounts of the interaction energy between the monolayers and α -synuclein proteins indicate better prevention of amyloid fibrillation. The type of interaction energy, whether electrostatic or VDW, is also essential.^{67–70}

Figure 4 shows the energies of interactions between α synuclein proteins and each of the monolayers versus the simulation time. These diagrams reveal the existence of considerable interactions between a-synuclein proteins and all monolayers. Negative energy values in most parts of the diagrams reveal the existence of attractive forces between the particles. It is also noted that VDW energy plays a vital role in the simulated interactions than electrostatic energy. Among these diagrams, Figure 4B and C present the highest and the lowest absolute energy values, respectively, which means that N-Graphene makes stronger interactions with α -synuclein proteins than other monolayers. The absolute total energy values in diagrams b and c increase monotonically, which reveals that N-Graphene and P-Graphene possess more stable interactions with increasing strength with α -synuclein proteins. On the other hand, the fluctuations in diagrams a and d reveal that the strength of interactions made between graphene and BCN and a-synuclein proteins suddenly changes during the simulation.

Table 1 shows the average values for electrostatic, VDW, and the total energies of the interactions between α -synuclein proteins and each of the monolayers. According to the values reported in the table, the highest absolute



Figure 6 The number of hydrogen bonds between the α -synuclein protein and water molecules versus time of simulation in the presence of (A) Graphene; (B) N-Graphene; (C) P-Graphene; (D) BCN; (E) Without monolayers.

Table 3 The Average Number of Hydrogen Bonds Between α -Synuclein Protein and Water Molecules During the Simulation Time in the Presence of Each Monolayer

Monolayer	Average Number of Hydrogen Bonds
Graphene	397.428
N-Graphene	400.994
P-Graphene	391.986
BCN	374.588
Without monolayer	366.923

interaction energy belongs to N-Graphene, followed by graphene, BCN, and P-Graphene. Based on Figure 4 and Table 1, it can be concluded that N-Graphene makes stronger bonds with α -synuclein proteins, and therefore, can prevent the formation of α -synuclein amyloids more effectively than the other monolayers.

Table 1

Evaluating the Compactness of α -Synuclein

In order to form critical nuclei and make aggregations, particles need to become more compact. A critical indicator of the compactness of protein structures is the Gyration Radius (R_g). The smaller the value of R_g , the more compact is the macromolecule being investigated. Therefore, concerning α -synuclein proteins, the variation

of R_g during the simulation indicates the possibility of prevention of amyloid formation.^{71–75}

Figure 5 shows R_g of α -synuclein proteins versus time with and without the presence of monolayers. According to the diagrams, R_g has been increased in the presence of monolayers, which means that they have decreased compactness of α -synuclein proteins, making the process of amyloid formation less likely. It is also noted that N-Graphene and graphene have caused higher values of R_g compared to P-Graphene and BCN.

Table 2 indicates the variation of R_g during 40 nanoseconds. This table shows that in the presence of monolayers, slighter decrease in R_g occurs, where among the monolayers, N-Graphene has the least variation, followed by graphene, P-Graphene, and BCN. Based on Figure 5 and Table 2, although all the monolayers can reduce the compactness of α -synuclein proteins and decrease the likelihood of amyloid formation, N-Graphene has the most desired effect followed by graphene, P-Graphene, and BCN Table 2.

Evaluating the Hydrogen Bonds

Hydrogen bonding is the strongest intermolecular bond.^{76–78} Concerning the formation of α -synuclein amyloids, the existence of abundant hydrogen bonds between α -synuclein



Figure 7 Different components of the secondary structure of α -synuclein protein during the simulation time (colours represent secondary structures) in the present of (A) Graphene; (B) N- Graphene; (C) P- Graphene; (D) BCN; (E) Without monolayers.

Table 4 The Secondary Structure of the α -Synuclein Protein in the Presence of Each Monolayer

Monolayer		Structure	Coil	B-Sheet	B-Bridge	Bend
Graphene	Total Average Percent	1,061,333 26.533 0.19	2,515,854 62.893 0.45	0 0 0	27,095 0.677 0	1,735,710 43.392 0.31
N-Graphene	Total Average Percent	437,537 10.938 0.08	3,760,125 94.001 0.67	112 0.003 0	14,666 0.366 0	1,093,673 27.341 0.2
P-Graphene	Total Average Percent	1,377,376 34.434 0.25	2,404,485 60.111 0.43	11,718 0.293 0	28,703 0.718 0.01	1,382,491 34.56 0.25
BCN	Total Average Percent	1,649,858 41.245 0.29	1,940,508 48.512 0.35	617 0.0154 0	62,656 1.566 0.01	1,627,225 40.679 0.29
Without monolayer	Total Average Percent	1,622,821 40.569 0.29	1,928,421 48.209 0.34	81,064 2.026 0.01	136,748 3.419 0.02	1,697,853 42.445 0.3
(B)						·
Monolayer			Turn	A-Helix	5-Helix	3-Helix
Graphene	Total Average Percent		847,265 21.181 0.15	186,973 4.674208 0.03	2316 0.057899 0	284,927 7.123 0.05
N-Graphene	Total Average Percent		351,453 8.786 0.06	71,306 1.783 0.01	1119 0.028 0	307,686 7.692 0.05
P-Graphene	Total Average Percent		979,013 24.475 0.17	357,942 8.948 0.06	36,683 0.917 0.01	399,105 9.977 0.07
BCN	Total Average Percent		1,187,575 29.688 0.21	399,010 9.975 0.07	50,215 1.255 0.01	332,334 8.308 0.06
Without monolayer	Total Average Percent		861,803 21.545 0.15	543,206 13.579 0.1	106,531 2.663 0.02	244,514 6.113 0.04
(C)						·
Monolayer			Turn+Bend+Coil	Helices+B-Sheet		
Graphene	Total Average Percent		5,098,829 127.468 0.91	474,216 11.855 0.08		
N-Graphene	Total Average Percent		5,205,251 130.128 0.93	380,223 9.505 0.06		
P-Graphene	Total Average Percent		4,765,989 119.147 0.85	805,448 20.136 0.14		

(Continued)

Table 4 (Continued).

Monolayer		Structure	Coil	B-Sheet	B-Bridge	Bend
BCN	Total Average Percent		4,755,308 118.879 0.85	782,176 19.554 0.14		
Without monolayer	Total Average Percent		4,488,077 112.199 0.79	975,315 24.3823 0.17		

proteins and water molecules can interrupt their movement in the aqueous media and disrupt the process of amyloid formation.

Figure 6 shows the number of hydrogen bonds versus time during the simulation. By comparing the diagrams, it is noticed that in the presence of N-Graphene, the number of hydrogen bonds between α -synuclein proteins and water molecules is substantially increased compared to other monolayers. In order to make a quantified comparison of the effectiveness of monolayers to increase the number of hydrogen bonds between α -synuclein proteins and water molecules, the average number of hydrogen bonds for each simulation is shown in Table 3 Although all monolayers increased the number of hydrogen bonds between α -synuclein proteins and water molecules, the most significant effect is for N-Graphene. Therefore N-Graphene is the best in preventing the amyloid formation and ceasing the progress of Parkinson's disease Table 3.

Evaluating the Secondary Structure of $\alpha\mathchar`-$ Synuclein

Intramolecular hydrogen bonds form the secondary structure of proteins. Depending on the position of the bonds and the types of involved amino acids, the secondary structure of proteins can be classified into several categories. The most important types of the secondary structure consist of the α -helix and β -sheet. Other less common structures include 3-helices, π -helices, β -bridges, turns, bends, and coils. One



Figure 8 RMSD values of α -synuclein protein versus the simulation time in the presence of (A) Graphene; (B) N- Graphene; (C) P- Graphene; (D) BCN; (E) Without monolayers.

Table 5	Values	s of the	Geometric	Means	of	α -Synuclein	RMSD
Derivati	ves in t	he Pres	ence of Eacl	h Mono	laye	er	

Monolayer	Geometric Mean of RMSD Derivative *10^3
Graphene	4.248
N-Graphene	5.758
P-Graphene	4.228
BCN	3.584
Without monolayer	3.452

single protein could have a combination of these secondary structures throughout its residues. Nevertheless, the secondary structure of a protein and its functionality can change in different cellular conditions.

The α -synuclein protein is naturally a membrane-bound protein consisting of 140 amino acids. The analysis of the structure of α -synuclein reveals that the entire length of the protein can be divided into three domains with different structures and functions. The N-terminal, consisting of amino acids 1–60, is an amphipathic domain that interacts with phospholipid membranes. It consists of two α -helices in the membrane-bound state, while parts of which may turn into random coils under certain circumstances. Amino acids 61–95 make the hydrophobic middle domain, also refers to non-amyloid component (NAC), which has a tendency to form β -sheets and is mostly involved in α -synuclein protein aggregation and amyloid formation. Finally, the C-terminal, consisting of amino acids 96–140, is the region that makes interactions with other proteins and causes the water solubility of the natural α -synuclein protein. This domain contains five prolines, which are responsible for the presence of coils, or the lack of secondary structure, in the C-terminal. During the process of aggregation and formation of oligomers, amyloid fibrils, and Lewy bodies, the secondary structures of α -synuclein proteins undergo significant transformations, resulting in an increased proportion of β -sheets and α helices, and decreased proportion of coils, bends, and turns.^{79–84}

In order to further investigate the effectiveness of monolayers in preventing amyloid formation, the secondary structure of α -synuclein proteins has been analyzed with and without monolayers. In Figure 7, different colours represent different structures. The numerical results are in Table 4 Decreased helices and β -sheets, and increased turns, bends, and coils favour the prevention of α -synuclein aggregation and amyloid formation. Based on Figure 7 and Table 4, it



Figure 9 RMSF values of α -synuclein protein versus the simulation time in the presence of (A) Graphene; (B) N- Graphene; (C) P- Graphene; (D) BCN; (E) Without monolayers.

Table 6 Average RMSF Values of α -Synuclein Protein Atoms in the Presence of Each Monolayer

Monolayer	Average of RMSF
Graphene	1.2686
N-Graphene	1.758
P-Graphene	1.056
BCN	1.021
Without monolayer	1.002

can be concluded that all monolayers have increased the proportion of turns, bends and coils, and reduced the proportion of helices and β -sheets, among which the most favourable result belongs to N-Graphene, followed by graphene, P-Graphene, and BCN.

Evaluating the Stability of α -Synuclein

The stability of proteins is an important indicator to understand the aggregation of particles and amyloid formation. α synuclein protein stability reduces the motility of the particles and creates the conditions for deformation in its chain. Any agent which causes an increase in the molecular motions of α -synuclein proteins interferes with the process of amyloid fibrillation. In order to assess the stability of α -synuclein proteins, the following two parameters have been analyzed.

Analysis of RMSD

RMSD is the most commonly used parameter to evaluate the stability of a system in MD simulations. The RMSD curve reflects fluctuations of the particle calculated at different points of the simulation time relative to a constant reference. The greater the oscillations of the particles, the more unstable the system and the greater the slope of the RMSD curve will be.^{85–90} Therefore, higher fluctuation rates reflected by the RMSD curve reveal better prevention of α -synuclein amyloid formation by the monolayer.

Figure 8 shows the RMSD curves of α -synuclein proteins in the presence and absence of monolayers. Apparent instability of the system is noted in the first 15 nanoseconds of all simulations, as reflected by a sudden surge at the first five nanoseconds, followed by moderate oscillations in the next ten nanoseconds of simulation. Figure 8C (P-Graphene), D (BCN), and -E (without monolayers) maintain the instability of α -synuclein proteins after nanosecond 15; however, in Figure 8A (Graphene) and B (N-Graphene), fluctuations exist until the end of the simulation process. Therefore, it can be concluded that graphene and N-Graphene cause more sustained instability in the system.Besides, the geometric means of the derivatives of RMSD are shown in Table 5, which shows that N-Graphene has the highest value with the most significant instability of α -synuclein particles. Hence,



Figure 10 The contact area of α-synuclein proteins versus time in the presence of (A) Graphene; (B) N- Graphene; (C) P- Graphene; (D) BCN; (E) Without monolayers.

N-Graphene is the best to prevent the formation of amyloids, followed by graphene, P-Graphene, and BCN.

Table 5

Analysis of RMSF

In the RMSF diagram, the average fluctuation during the whole simulation process is shown for each atom. Higher RMSF values indicate more instability of the system and hence, better prevention of a-synuclein amyloid formation by monolayers.^{81,91–94} The RMSF curves of α synuclein proteins in the presence and absence of each monolayer are shown in Figure 9. A comparison of the curves reveals that all monolayers increase RMSF values, while N-Graphene and graphene cause the most instability.Furthermore, to compare the effectiveness of each monolayer, the average RMSF values are shown in Table 6, which shows the highest value for N-Graphene, followed by graphene, P-Graphene, and BCN. It can be concluded that N-Graphene has the highest potential can prevent amyloid fibrillation more effectively by disrupting the stability of the system.

Evaluating the Contact Area of α -Synuclein Molecules

Amyloid formation increases if α -synuclein proteins are in close contact. So, the contact surface of α -synuclein proteins indicates the probability of amyloid formation. SASA is an essential parameter for evaluating the contact surface of biomolecules.^{95–97} In order to analyze the effects of monolayers on preventing amyloid fibrillation, SASA values have been calculated, and the contact area of α -synuclein proteins plotted as a function of time (Figure 10 and Table 7), which shows that N-Graphene has the least values of the contact area.

Table 7 shows the average contact area of α -synuclein proteins during the simulations. The values of the table reveal that although all monolayers reduce the contact surface of α -synuclein proteins and disrupt amyloid formation, the most desirable effect is seen in the simulation of N-Graphene, followed by graphene, P-Graphene, and BCN.

Validation Analysis

Mohammad-Beigi et al¹⁹ simulated the molecular dynamics of graphene polyglycerol and amyloid in water. Ultimately, as a validation of the algorithms used in this paper, an additional simulation of the van der Waals energy of interactions between graphene polyglycerol and α -synuclein proteins have been performed with conditions identical to a previously published study by Mohammad-Beigi et al.¹⁹

Table 7 Average Values of Contact Area for α -Synuclein Proteins During the Simulation in the Presence of Each Monolayer

Monolayer	Average Contact Area (nm\S ² \N)
Graphene	6.262
N-Graphene	-5.271
P-Graphene	11.229
BCN	26.277
Without monolayer	27.237

The results are shown in Figure 11 and indicate that the values of the curve produced by this paper and Mohammad-Beigi et al¹⁹ posses similar results.

Conclusion

Alpha-synuclein proteins can aggregate and rise to oligomers, amyloids, and Lewy bodies, which is a vital sign of Parkinson's disease pathophysiology. Preventing this pathological cascade has been the target of many conducted studies so far. In this study, the effects of four graphene-based nanostructures on preventing α -synuclein amyloid formation were investigated. For this purpose, important parameters involved in the interactions of α synuclein proteins with themselves and with the monolayers were analyzed. Ultimately, it was concluded that, although all monolayers changed the evaluated parameters in the way which helps in the prevention of α -synuclein amyloid formation, the most significant results were reported for N-Graphene and graphene. N-Graphene and graphene are the most effective particle for the prevention



Figure 11 Comparison of the values of VDW between the α -synuclein protein and Graphene polyglycerol versus time in two different studies; black curve: the current study; red curve: the previously published study of Mohammad-Beigi et al.¹⁹

of amyloid fibrillation and hence, ceasing the progression of Parkinson's disease.

Such results could have important implications in the future of Parkinson's disease treatment. For such a purpose, small-scale computational studies must be followed by gross-scale simulations in which more variables and factors are being considered as part of the simulation process. The clinical implementation of such a potential treatment option for Parkinson's disease undoubtedly requires experimental in-vitro analyses followed by animal studies which would be strongly suggested by the authors of this paper. Ultimately, the results of such experimental studies must be put into a series of single or double-blind clinical trials in which already-proven effective nanomaterials could be administered as therapeutic agents or used as carries for conventional medications of Parkinson's disease.

Disclosure

The authors report no conflicts of interest for this work.

References

- Yavarpour-Bali H, Pirzadeh M, Ghasemi-Kasman M. Curcumin-loaded nanoparticles: a novel therapeutic strategy in treatment of central nervous system disorders. *Int J Nanomed.* 2019;14:4449–4460. doi:10.2147/IJN.S208332
- Xiong B, Zhou Y, Zhao Y, et al. The use of nitrogen-doped graphene supporting Pt nanoparticles as a catalyst for methanol electrocatalytic oxidation. *Carbon N Y.* 2013;52:181–192. doi:10.1016/j. carbon.2012.09.019
- Ross C, Taylor M, Fullwood N, Allsop D. Liposome delivery systems for the treatment of Alzheimer's disease. *Int J Nanomed*. 2018;13:8507–8522. doi:10.2147/IJN.S183117
- Lee CC, Nayak A, Sethuraman A, Belfort G, McRae GJ. A three-stage kinetic model of amyloid fibrillation. *Biophys J*. 2007;92:3448–3458. doi:10.1529/biophysj.106.098608
- Javdani N, Rahpeyma SS, Ghasemi Y, Raheb J. Effect of superparamagnetic nanoparticles coated with various electric charges on αsynuclein and β-amyloid proteins fibrillation process. *Int J Nanomed.* 2019;14:799–808. doi:10.2147/IJN.S190354
- Zand Z, Afarinesh Khaki P, Salihi A, et al. Cerium oxide NPs mitigate the amyloid formation of α-synuclein and associated cytotoxicity. *Int J Nanomed*. 2019;14:6989–7000. doi:10.2147/IJN.S220380
- Bi CC, Wang A, Chu Y, et al. Intranasal delivery of rotigotine to the brain with lactoferrin-modified PEG-PLGA nanoparticles for Parkinson's disease treatment. *Int J Nanomed.* 2016;11:6547–6559. doi:10.2147/IJN.S120939
- Yan X, et al. Lactoferrin-modified rotigotine nanoparticles for enhanced nose-to-brain delivery: LESA-MS/MS-based drug biodistribution, pharmacodynamics, and neuroprotective effects. *Int J Nanomed.* 2018;13:273–281. doi:10.2147/IJN.S151475
- Pedrosa Carrasco AJ, Timmermann L, Pedrosa DJ. Management of constipation in patients with Parkinson's disease. *Npj Park Dis.* 2018;4:1–10.
- Qu M, Lin Q, He S, et al. A brain targeting functionalized liposomes of the dopamine derivative N-3,4-bis(pivaloyloxy)-dopamine for treatment of Parkinson's disease. J Control Release. 2018;277:173–182. doi:10.1016/j.jconrel.2018.03.019

- Kim K, Bohnen NI, Müller MLTM, Lustig C. Compensatory dopaminergic-cholinergic interactions in conflict processing: evidence from patients with Parkinson's disease. *Neuroimage*. 2019;190:94–106. doi:10.1016/j.neuroimage.2018.01.021
- McKinley JW, Shi Z, Kawikova I, et al. Dopamine deficiency reduces striatal cholinergic interneuron function in models of Parkinson's disease. *Neuron.* 2019;103(1056–1072.e6):1056–1072.e6. doi:10.1016/j. neuron.2019.06.013
- Moayeri A, Darvishi M, Amraei M. Homing of super paramagnetic iron oxide nanoparticles (SPIONs) labeled adipose-derived stem cells by magnetic attraction in a rat model of parkinson's disease. *Int J Nanomed.* 2020;15:1297–1308. doi:10.2147/IJN.S238266
- 14. Li J, Uversky VN, Fink AL. Effect of familial Parkinson's disease point mutations A30P and A53T on the structural properties, aggregation, and fibrillation of human α-synuclein. *Biochemistry*. 2001;40:11604–11613. doi:10.1021/bi010616g
- Wu WH, Sun X, Yu Y-P, et al. TiO2 nanoparticles promote β-amyloid fibrillation in vitro. *Biochem Biophys Res Commun.* 2008;373:315–318. doi:10.1016/j.bbrc.2008.06.035
- Härd T, Lendel C. Inhibition of amyloid formation. J Mol Biol. 2012;421:441–465. doi:10.1016/j.jmb.2011.12.062
- Gazit E. A possible role for π-stacking in the self-assembly of amyloid fibrils. FASEB J. 2002;16:77–83. doi:10.1096/fj.01-0442hyp
- Colvin VL, Kulinowski KM. Nanoparticles as catalysts for protein fibrillation. *Proc Natl Acad Sci U S A.* 2007;104:8679–8680. doi:10.1073/pnas.0703194104
- Mohammad-Beigi H, et al. Mechanistic understanding of the interactions between nano-objects with different surface properties and α-synuclein. *ACS Nano*. 2019;13:3243–3256. doi:10.1021/acsnano.8b08983
- Waxman EA, Giasson BI. Molecular mechanisms of α-synuclein neurodegeneration. *Biochim Biophys Acta - Mol Basis Dis.* 1792;616–624:2009.
- Gaumet M, Vargas A, Gurny R, Delie F. Nanoparticles for drug delivery: the need for precision in reporting particle size parameters. *Eur J Pharm Biopharm*. 2008;69:1–9. doi:10.1016/j. ejpb.2007.08.001
- 22. Yang JA, Lin W, Woods WS, George JM, Murphy CJ. α-Synuclein's adsorption, conformation, and orientation on cationic gold nanoparticle surfaces seeds global conformation change. *J Phys Chem B*. 2014;118:3559–3571. doi:10.1021/jp501114h
- Álvarez YD, Fauerbach JA, Pellegrotti JV, et al. Influence of gold nanoparticles on the kinetics of α-synuclein aggregation. *Nano Lett.* 2013;13:6156–6163. doi:10.1021/nl403490e
- Sanfins E, Dairou J, Rodrigues-Lima F, Dupret JM. Nanoparticle-protein interactions: from crucial plasma proteins to key enzymes. J. Phys. Conf. Ser. 2011;304:012039. doi:10.1088/1742-6596/304/1/012039
- Vertegel AA, Siegel RW, Dordick JS. Silica nanoparticle size influences the structure and enzymatic activity of adsorbed lysozyme. *Langmuir*. 2004;20:6800–6807. doi:10.1021/la0497200
- 26. Bond M, Pitt M, Akoh J, et al. The effectiveness and cost-effectiveness of methods of storing donated kidneys from deceased donors: a systematic review and economic model. *Health Technol Assess (Rocky)*. 2009;13.
- 27. Wearable T, Kidney A. for End-Stage Renal Disease. 2005;149:325–333.
- Kuitunen A, Vento A, Suojaranta-Ylinen R, Pettilä V. Acute renal failure after cardiac surgery: evaluation of the RIFLE classification. *Ann Thorac Surg.* 2006;81:542–546. doi:10.1016/j.athoracsur.2005.07.047
- 29. Lassen M, Gallus A, Raskob G. New England Journal NFL.pdf. N Engl J Med. 2010;363:2487–2498. doi:10.1056/NEJMoa1006885
- Davenport A. Dialysis: a wearable dialysis device: the first step to continuous therapy. *Nat Rev Nephrol*. 2016;12:512–514. doi:10.1038/ nrneph.2016.100
- Huang L, Cao Y, Diao D. N-doped graphene sheets induced high electrochemical activity in carbon film. *Appl Surf Sci.* 2019;470:205–211. doi:10.1016/j.apsusc.2018.11.075

- 32. Science S Ac ce us pt. 2019.
- Khan FA, Almohazey D, Alomari M, Almofty SA. Impact of nanoparticles on neuron biology: current research trends. *Int J Nanomed*. 2018;13:2767–2776. doi:10.2147/IJN.S165675
- 34. Yang L, Jiang S, Zhao Y, et al. Boron-doped carbon nanotubes as metal-free electrocatalysts for the oxygen reduction reaction. *Angew. Chemie - Int. Ed.* 2011;50:7132–7135. doi:10.1002/anie.201101287
- 35. Yang DS, Bhattacharjya D, Inamdar S, Park J, Yu JS. Phosphorus-doped ordered mesoporous carbons with different lengths as efficient metal-free electrocatalysts for oxygen reduction reaction in alkaline media. J Am Chem Soc. 2012;134:16127–16130. doi:10.1021/ja306376s
- 36. Carrero-Sánchez JC, Elías AL, Mancilla R, et al. Biocompatibility and toxicological studies of carbon nanotubes doped with nitrogen. *Nano Lett.* 2006;6:1609–1616. doi:10.1021/nl060548p
- Baldea I, Olteanu D, Filip GA, et al. Cytotoxicity mechanisms of nitrogen-doped graphene obtained by electrochemical exfoliation of graphite rods, on human endothelial and colon cancer cells. *Carbon*. 2020;158:267–281. doi:10.1016/j.carbon.2019.12.011
- Liao C, Li Y, Tjong SC. Graphene nanomaterials: synthesis, biocompatibility, and cytotoxicity. *Int. J. Mol. Sci.* 2018;19(11):3564. doi:10.3390/ijms19113564
- Pelin M, Fusco L, León V, et al. Differential cytotoxic effects of graphene and graphene oxide on skin keratinocytes. *Sci Rep.* 2017;7:1–12.
- Liao KH, Lin YS, MacOsko CW, Haynes CL. Cytotoxicity of graphene oxide and graphene in human erythrocytes and skin fibroblasts. ACS Appl Mater Interfaces. 2011;3:2607–2615. doi:10.1021/am200428v
- Kim D, Yoo JM, Hwang H, et al. Graphene quantum dots prevent αsynucleinopathy in Parkinson's disease. *Nat. Nanotechnol.* 2018;13:812–818. doi:10.1038/s41565-018-0179-y
- Banerjee AN. Graphene and its derivatives as biomedical materials: future prospects and challenges. *Interface Focus*. 2018;8. doi:10.1098/rsfs.2017.0056
- Lee H, Paeng K, Kim IS. A review of doping modulation in graphene. Synth Met. 2018;244:36–47. doi:10.1016/j. synthmet.2018.07.001
- 44. Wang H, Maiyalagan T, Wang X. Review on recent progress in nitrogen-doped graphene: synthesis, characterization, and its potential applications. *ACS Catal.* 2012;2:781–794. doi:10.1021/cs200652y
- 45. Ysselstein D, Krainc D. Untangling alpha synuclein fibrils by graphene quantum dots. *Mov Disord*. 2018;33:1673. doi:10.1002/ mds.27511
- 46. Schroer ZS, Wu Y, Xing Y, et al. Nitrogen-sulfur-doped graphene quantum dots with metal ion-resistance for bioimaging. ACS Appl Nano Mater. 2019;2:6858–6865. doi:10.1021/acsanm.9b01309
- 47. Yang Y, Asiri AM, Tang Z, Du D, Lin Y. Graphene based materials for biomedical applications. *Mater Today.* 2013;16:365–373. doi:10.1016/j.mattod.2013.09.004
- 48. Frenkel D, Smit B, Tobochnik J, McKay SR, Christian W. Understanding molecular simulation. *Comput Phys.* 1997;11:351. doi:10.1063/1.4822570
- 49. Huang J, Lemkul JA, Eastman PK, Mackerell AD. Molecular dynamics simulations using the drude polarizable force field on GPUs with OpenMM: implementation, validation, and benchmarks. *J Comput Chem.* 2018;1–8. doi:10.1002/jcc.25339
- 50. Jones JE, et al. On the determination of molecular fields. —II. From the equation of state of a gas. *Proc R Soc London Ser A, Contain Pap a Math Phys Character.* 1924;106:463–477.
- Rezvantalab S, Keshavarz Moraveji M, Khedri M, et al. An insight into the role of riboflavin ligand in the self-assembly of poly(lactic-co -glycolic acid)-based nanoparticles – a molecular simulation and experimental approach. *Soft Matter*. 2020;16(22):5250–5260. doi:10.1002/9783527682300
- 52. Ulmer TS, Bax A, Cole NB, Nussbaum RL. Structure and dynamics of micelle-bound human α-synuclein. J Biol Chem. 2005;280:9595–9603. doi:10.1074/jbc.M411805200

- 53. Mortazavifar A, Raissi H, Akbari A. DFT and MD investigations on the functionalized boron nitride nanotube as an effective drug delivery carrier for Carmustine anticancer drug. *J Mol Liq.* 2019;276:577–587. doi:10.1016/j.molliq.2018.12.028
- 54. Shafiei F, Hashemianzadeh SM, Bagheri Y. Insight into the encapsulation of gemcitabine into boron- nitride nanotubes and gold cluster triggered release: a molecular dynamics simulation. J Mol Liq. 2019;278:201–212. doi:10.1016/j.molliq.2019.01.020
- 55. Wang L, Xu J, Wang X, Cheng Z, Xu J. Facile preparation of N-rich functional polymer with porous framework as QCM sensing material for rapid humidity detection. *Sensors Actuators B Chem.* 2019;288:289–297. doi:10.1016/j.snb.2019.02.058
- Chen W, He J, Jiang Y, Zhang H, Zhou X. Experimental and theoretical studies on the atmospheric degradation of 1, 1, 2, 2, 3, 3, 4-heptafluorocyclopentane. *Atmos Environ*. 2019;196:38–43. doi:10.1016/j.atmosenv.2018.09.057
- 57. Robertson MJ, Qian Y, Robinson MC, Tirado-Rives J, Jorgensen WL. Development and Testing of the OPLS-AA/M force field for RNA. *J Chem Theory Comput.* 2019;15:2734–2742. doi:10.1021/acs. jctc.9b00054
- Dalgicdir C, Van Der Vegt NFA. Improved temperature behavior of PNIPAM in water with a modified OPLS model. *J Phys Chem B*. 2019;123:3875–3883. doi:10.1021/acs.jpcb.9b01644
- Itliong JN, Villagracia ARC, Moreno JLV, et al. Investigation of reverse ionic diffusion in forward-osmosis-aided dewatering of microalgae: a molecular dynamics study. *Bioresour. Technol.* 2019;279:181–188. doi:10.1016/j.biortech.2019.01.109
- Kyrychenko A, Blazhynska MM, Slavgorodska MV, Kalugin ON. Stimuli-responsive adsorption of poly(acrylic acid) onto silver nanoparticles: role of polymer chain length and degree of ionization. *J Mol Liq.* 2019;276:243–254. doi:10.1016/j.molliq.2018.11.130
- Herrera-Rodríguez AM, Miletić V, Aponte-Santamaría C, Gräter F. Molecular dynamics simulations of molecules in uniform flow. *Biophys J.* 2019;116:1579–1585. doi:10.1016/j.bpj.2018.12.025
- Zhong W, Cai Y, Tománek D. Computer simulation of hydrogen embrittlement in metals. *Nature*. 1993;362:435–437. doi:10.1038/362435a0
- Martoňák R, Donadio D, Oganov AR, Parrinello M. Crystal structure transformations in SiO2 from classical and ab initio metadynamics. *Nat Mater.* 2006;5:623–626. doi:10.1038/nmat1696
- 64. Carretero-González R, Kevrekidis PG, Kevrekidis IG, Maroudas D, Frantzeskakis DJ. A Parrinello-Rahman approach to vortex lattices. *Phys Lett Sect a Gen at Solid State Phys.* 2005;341:128–134.
- Wang E, Sun H, Wang J, et al. End-point binding free energy calculation with MM/PBSA and MM/GBSA: strategies and applications in drug design. *Chem. Rev.* 2019;119:9478–9508. doi:10.1021/ acs.chemrev.9b00055
- 66. Greene D, Qi R, Nguyen R, Qiu T, Luo R. Heterogeneous dielectric implicit membrane model for the calculation of MMPBSA binding free energies. J Chem Inf Model. 2019;59:3041–3056. doi:10.1021/ acs.jcim.9b00363
- Astrand P, Wallqvist A, Karlström G. Intermolecular interactions of urea and water. J Chim Phys. 1991;88:2457–2464. doi:10.1051/jcp/ 1991882457
- Damghani T, Sedghamiz T, Sharifi S, Pirhadi S. Critical c-Metinhibitor interactions resolved from molecular dynamics simulations of different c-Met complexes. J Mol Struct. 2020;1203:127456. doi:10.1016/j.molstruc.2019.127456
- 69. Dehury B, Behera SK, Mahapatra N. Structural dynamics of Casein Kinase I (CKI) from malarial parasite Plasmodium falciparum (Isolate 3D7): insights from theoretical modelling and molecular simulations. J Mol Graph Model. 2017;71:154–166. doi:10.1016/j. jmgm.2016.11.012
- Maleki R, Khoshoei A, Ghasemy E, Rashidi A. Molecular insight into the smart functionalized TMC-Fullerene nanocarrier in the pH-responsive adsorption and release of anti-cancer drugs. J Mol Graph Model. 2020;100:107660. doi:10.1016/j.jmgm.2020.107660

- Kharazian B, Lohse SE, Ghasemi F, et al. Bare surface of gold nanoparticle induces inflammation through unfolding of plasma fibrinogen. *Sci. Rep.* 2018;8:1–9. doi:10.1038/s41598-018-30915-7
- 72. Ajori S, Ansari R, Haghighi S. A molecular dynamics study on the buckling behavior of cross-linked functionalized carbon nanotubes under physical adsorption of polymer chains. *Appl Surf Sci.* 2018;427:704–714. doi:10.1016/j.apsusc.2017.08.049
- Shariatinia Z, Mazloom-Jalali A. Chitosan nanocomposite drug delivery systems designed for the ifosfamide anticancer drug using molecular dynamics simulations. J Mol Liq. 2019;273:346–367. doi:10.1016/j.molliq.2018.10.047
- 74. Xu WS, Carrillo JMY, Lam CN, Sumpter BG, Wang Y. Molecular dynamics investigation of the relaxation mechanism of entangled polymers after a large step deformation. ACS Macro Lett. 2018;7:190–195. doi:10.1021/acsmacrolett.7b00900
- 75. Khoshoei A, Ghasemy E, Poustchi F, Shahbazi MA, Maleki R. Engineering the pH-sensitivity of the graphene and carbon nanotube based nanomedicines in smart cancer therapy by grafting trimetyl Chitosan. *Pharm Res.* 2020;37:1–13.
- 76. Fang H, Tang Z, Liu X, Huang Y, Sun CQ. Discriminative ionic capabilities on hydrogen-bond transition from the mode of ordinary water to (Mg, Ca, Sr)(Cl, Br) 2 hydration. *J Mol Liq.* 2019;279:485–491. doi:10.1016/j.molliq.2019.01.147
- 77. Zhong Y, Chen Y, Feng X, et al. Hydrogen-bond facilitated intramolecular proton transfer in excited state and fluorescence quenching mechanism of flavonoid compounds in aqueous solution. *J. Mol. Liq.* 2020;302:112562. doi:10.1016/j.molliq.2020.112562
- Bu R, Xiong Y, Wei X, Li H, Zhang C. Hydrogen bonding in CHON-containing energetic crystals: a review. *Cryst Growth Des.* 2019;19:5981–5997. doi:10.1021/acs.cgd.9b00853
- 79. Miceli M, Muscat S, Morbiducci U, Cavaglià M, Deriu MA. Ultrasonic waves effect on S-shaped β-amyloids conformational dynamics by non-equilibrium molecular dynamics. J Mol Graph Model. 2020;96:107518. doi:10.1016/j.jmgm.2019.107518
- Turner M, Mutter ST, Kennedy-Britten OD, Platts JA. Molecular dynamics simulation of aluminium binding to amyloid-β and its effect on peptide structure. *PLoS One*. 2019;14:1–14. doi:10.1371/ journal.pone.0217992
- Nemaysh V, Luthra PM, Advances RSC. Computational analysis revealing that K634 and T681 mutations modulate the 3D-structure of PDGFR- b and lead to sunitinib resistance †. 2017:37612–37626. doi:10.1039/c7ra01305a
- 82. Turner M, Mutter ST, Kennedy-Britten OD, Platts JA. Replica exchange molecular dynamics simulation of the coordination of Pt(ii)-Phenanthroline to amyloid-β. *RSC Adv.* 2019;9:35089–35097. doi:10.1039/C9RA04637B
- Uversky VN, Li J, Fink AL. Metal-triggered structural transformations, aggregation, and fibrillation of human α-synuclein: a possible molecular link between parkinson's disease and heavy metal exposure. J Biol Chem. 2001;276:44284–44296. doi:10.1074/jbc. M105343200
- Murray IVJ, Giasson BI, Quinn SM, et al. Role of α-synuclein carboxy-terminus on fibril formation in vitro. *Biochemistry*. 2003;42:8530–8540. doi:10.1021/bi027363r

International Journal of Nanomedicine

Publish your work in this journal

The International Journal of Nanomedicine is an international, peerreviewed journal focusing on the application of nanotechnology in diagnostics, therapeutics, and drug delivery systems throughout the biomedical field. This journal is indexed on PubMed Central, MedLine, CAS, SciSearch[®], Current Contents[®]/Clinical Medicine, Journal Citation Reports/Science Edition, EMBase, Scopus and the Elsevier Bibliographic databases. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit http://www.dovepress.com/ testimonials.php to read real quotes from published authors.

Submit your manuscript here: https://www.dovepress.com/international-journal-of-nanomedicine-journal

- Khezri A, Karimi A, Yazdian F, et al. Molecular dynamic of curcumin/chitosan interaction using a computational molecular approach: emphasis on biofilm reduction. *Int. J. Biol. Macromol.* 2018;114:972–978. doi:10.1016/j.ijbiomac.2018.03.100
- Saini RK, Thakur H, Goyal B. Effect of Piedmont mutation (L34V) on the structure, dynamics, and aggregation of Alzheimer's Aβ40 peptide. J Mol Graph Model. 2020;97:107571. doi:10.1016/j. jmgm.2020.107571
- Bertaccini EJ, Tradelia JR, Lindane E. Normal-mode analysis of the glycine alpha1 receptor by three separate methods. *J Chem Inf Model*. 2007;47:1572–1579. doi:10.1021/ci600566j
- 88. Hasanzade Z, Raissi H. Investigation of graphene-based nanomaterial as nanocarrier for adsorption of paclitaxel anticancer drug: a molecular dynamics simulation study. *J Mol Model*. 2017;23:1–8. doi:10.1007/s00894-017-3207-1
- Rahmatabadi SS, Sadeghian I, Ghasemi Y, Sakhteman A, Hemmati S. Identification and characterization of a sterically robust phenylalanine ammonia-lyase among 481 natural isoforms through association of in silico and in vitro studies. *Enzyme Microb Technol.* 2019;122:36–54. doi:10.1016/j.enzmictec.2018.12.006
- 90. Farhadian S, Shareghi B, Asgharzadeh S, Rajabi M, Asadi H. Structural characterization of α-chymotrypsin after binding to curcumin: spectroscopic and computational analysis of their binding mechanism. J Mol Liq. 2019;289:111111. doi:10.1016/j. molliq.2019.111111
- Chairatana P, Niramitranon J, Pongprayoon P. Dynamics of human defensin 5 (HD5) self-assembly in solution: molecular simulations/ insights. *Comput Biol Chem.* 2019;83:107091. doi:10.1016/j. compbiolchem.2019.107091
- Liu J, Wei B, Che C, et al. Enhanced stability of manganese superoxide dismutase by amino acid replacement designed via molecular dynamics simulation. *Int. J. Biol. Macromol.* 2019;128:297–303. doi:10.1016/j.ijbiomac.2019.01.126
- 93. Zaboli M, Raissi H, Zaboli M, Farzad F, Torkzadeh-Mahani M. Stabilization of D-lactate dehydrogenase diagnostic enzyme via immobilization on pristine and carboxyl-functionalized carbon nanotubes, a combined experimental and molecular dynamics simulation study. *Arch Biochem Biophys.* 2019;661:178–186. doi:10.1016/j. abb.2018.11.019
- Rezapour N, Rasekh B, Mofradnia SR, et al. Molecular dynamics studies of polysaccharide carrier based on starch in dental cavities. *Int. J. Biol. Macromol.* 2019;121:616–624. doi:10.1016/j.ijbiomac.2018.10.027
- Pereira GRC, Da Silva ANR, Do Nascimento SS, De Mesquita JF. In silico analysis and molecular dynamics simulation of human superoxide dismutase 3 (SOD3) genetic variants. J Cell Biochem. 2019;120:3583–3598. doi:10.1002/jcb.27636
- Arifuzzaman M, Mitra S, Das R, et al. In silico analysis of nonsynonymous single-nucleotide polymorphisms (nsSNPs) of the SMPX gene. Ann. Hum. Genet. 2020;84:54–71. doi:10.1111/ahg.12350
- 97. Da Silva ANR, Pereira GRC, Moreira LGA, Rocha CF, De Mesquita JF. SOD1 in amyotrophic lateral sclerosis development – in silico analysis and molecular dynamics of A4F and A4V variants. *J Cell Biochem.* 2019;120:17822–17830. doi:10.1002/jcb.29048

Dovepress