# ORIGINAL RESEARCH Statistical Sequential Experimentation: Preliminary Mixed Factorial Design, I-Optimal Mixture Design Then Finally Novel Design Space Expansion for Optimization of Tazarotene Cubosomes

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**Objective:** This study aims to illustrate the potential of sequential experimentation for statistically scientific based optimization of Tazarotene (TAZA) cubosomes.

**Methods:** Hot melt emulsification method was used for cubosomes preparation. A preliminary (3.2) mixed factorial design (MFD) was conducted to choose suitable types of stabilizer and surfactant that maximize entrapment efficiency (EE) and minimize particle size (PS). These chosen stabilizer and surfactant were to be used in the statistical design proposed for optimization of TAZA cubosomes (I-optimal mixture design) (IOMD). Glyceryl monooleate (GMO), stabilizer and surfactant amounts were the three mixture components (MixCs) studied in that design. Responses (EE, PS and drug percent released after 24 hours (Q24h)) were statistically analyzed. Numerical optimization using desirability function based on different responses' importance was used to find an IOMDoptimized formulation (IOMD-OF) with the predetermined characters. Then, a novel statistical methodology of design space expansion was adopted to enhance Q24h. Suitable models to express EE, PS and Q24h were elucidated over the expanded mixture design (EMD) space. Validity of derived models was verified via prediction intervals and percent deviations of actual values from predicted ones for all the EMD design points. EMD was then navigated to find EMD-OF.

**Results:** Analysis of MFD showed that Pluronic-F68 and polyvinyl alcohol were the best stabilizer and surfactant to be used. First stage optimization after IOMD analysis led to a formulation with unsatisfactory Q24h of 58.8%. After design space expansion adoption, re-analysis and re-optimization, a satisfactory EMD-OF having EE of 82.1%, PS of 273.0 nm and Q24h of 68.8% was found. **Conclusion:** Statistical sequential experimentation with the novel design space expansion approach proved to be a successful paradigm for enhancing TAZA cubosomes optimization. Thus, this paradigm is expected to have promising future applications in various pharmaceutical formulations optimization.

**Keywords:** statistically scientific based optimization, topical drug delivery, Box-Cox transformation, contour plots, Piepel's trace plots, hot melt emulsification

#### **Introduction**

<span id="page-0-0"></span>In order to treat a topical disease, drugs can be administered systemically or topically. Nowadays topical application of drugs is preferred over systemic administration. Systemic administration of the drug may expose it to first pass metabolism or being deactivated by stomach pH. Moreover, all body organs are subjected to the systemically administered drug which may result in undesired side effects. Although topical conventional dosage forms have been used for decades, they have drawbacks such as frequent dosing and skin irritation or damage.<sup>1</sup> To avoid several side effects of conventional topical dosage forms application, topical nano-carrier drug delivery systems have been developed. These nano-carriers can be classified into polymeric, lipid, and metallic nano-carriers according to their composition. Of these nano-carriers, lipid-based type is the most common in

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#### **Graphical Abstract**



#### <span id="page-1-1"></span><span id="page-1-0"></span>pharmaceutical research[.2](#page-15-1) These lipid-based nano-carriers can be subdivided into three different classes: vesicular carriers such as liposomes, $3$  niosomes, $4$  proniosomes, $5$  cerosomes, $6$  nano-particulate carriers<sup>7,[8](#page-15-7)</sup> and emulsion based carriers. $9-11$  $9-11$

<span id="page-1-5"></span>Cubosomes are cubic shaped liquid crystal dispersions composed of lipid layers, separating non-intersecting water channels. They are considered a promising type of nano-vesicles. Besides being a non-toxic, biocompatible and inexpensive drug delivery system, cubosomes' special structure enables the incorporation of both hydrophilic and lipophilic drugs. Also, cubosomes have a bioadhesive nature. They can form a thin layer on skin and mucosal surfaces. This allows convenient delivery and deposition of topical drugs with fair temporary protection from irritation.<sup>[12](#page-15-10)[,13](#page-15-11)</sup> So, cubosomes can be considered as one of the nano-drug delivery systems of choice for topical drug delivery. Cubosomes can be prepared from lipids such as phytantriol and glyceryl monooleate in the presence of steric stabilizers such as poloxamers, casein and lecithin.<sup>[14](#page-15-12)</sup> Surfactants such as PVA, Tweens<sup>15</sup> and Brijs<sup>16</sup> can also be added along with the aforementioned stabilizers. They have the advantage of being stable at room temperature.<sup>[12](#page-15-10)</sup>

<span id="page-1-8"></span><span id="page-1-7"></span><span id="page-1-6"></span><span id="page-1-4"></span><span id="page-1-3"></span><span id="page-1-2"></span>Tazarotene (TAZA) is a third-generation synthetic retinoid (vitamin A derivative) with a selective retinoic acid receptors (mainly RAR-β and RAR-γ) activity.<sup>[17](#page-15-15)</sup> It is used for topical treatment of several skin conditions such as acne, psoriasis and photo-aging. TAZA is present in market as  $0.1\%$  gel<sup>[6](#page-15-5)</sup> (Acnitaz<sup>®</sup>),  $0.1\%$  cream<sup>18</sup> (Tazarotene<sup>®</sup>) and  $0.1\%$ foam<sup>[19](#page-15-17)</sup> (Fabior<sup>®</sup>). It was reported that long-term treatment with TAZA in conventional dosage forms caused several side effects.<sup>10</sup> Market formulations were reported to have skin irritation side effects such as pruritus, redness and peeling.<sup>[20](#page-15-19)</sup>

<span id="page-2-1"></span><span id="page-2-0"></span>Therefore, it was necessary to develop skin targeting formulations that can increase TAZA deposition into skin and reduce those annoying side effects. These formulations included liposomes,<sup>21</sup> cerosomes,<sup>[6](#page-15-5)</sup> proniosomal gel,<sup>[22](#page-16-1)</sup> microemulsions,<sup>10,[11](#page-15-9),[23](#page-16-2)</sup> spanlastic nano-vesicles,<sup>24</sup> nanoparticles,<sup>[25](#page-16-4)</sup> niosomal gel and nano-sponges.<sup>26</sup> TAZA was also formulated as in situ gels which were reported to be non-irritant to the skin.<sup>[27](#page-16-6)</sup>

<span id="page-2-2"></span>The use of statistical experimental designs, statistical modeling and optimization of responses based on derived models saves more time and effort than the traditional one-at-a-time experimentation.<sup>[28](#page-16-7)</sup> Also, the use of statistical designs enhances the experimental input-output quality. Statistical designs have been recently widely used in pharmaceutical research for formulation development and optimization. These designs vary from the parent factorial designs either full<sup>[29](#page-16-8)[,30](#page-16-9)</sup> or fractional,<sup>[31](#page-16-10)</sup> to response surface designs (RSD) such as Box-Behnken designs<sup>[32](#page-16-11)</sup> and central composite designs<sup>[16](#page-15-14)</sup> to mixture designs<sup>33</sup> and even combined mixture designs.<sup>[34](#page-16-13)</sup>

<span id="page-2-5"></span><span id="page-2-4"></span>Our study aims to move a step forward in the field of the use of statistics in pharmaceutical formulations development and optimization. In this study, statistically scientific based optimization is to be implemented. This is to be done through a statistical sequential experimentation strategy together with the novel approach of design space expansion for formulation optimization. Optimization of TAZA cubosomes for topical application will be performed as a practical example. To our knowledge, this is the first work to use design space expansion as a statistical tool for pharmaceutical formulation optimization.

#### **Theory**

In many recent pharmaceutical formulation studies, researchers adopted a certain statistical design. This design – whatever its type – is used for choosing which trials (design points) (DPs) will be done, then elucidation of a suitable polynomial model for representing each response. Simultaneous numerical optimization was then used to search for an optimized formulation based on the derived models. Though this traditional sequence is satisfactory to some extent, yet questions arise. How should the studied variables or components be chosen? What can we do if the optimization results were not satisfying enough? Shall we build a new design from scratch? Or what can we do? In this study, statistically scientific based optimization will be implemented through statistical sequential experimentation aided by design space expansion. This is to answer the previous questions and map how the best possible results can be reached while minimizing the amount of experimentation performed.

The proposed statistically scientific based optimization scheme in this study starts with a preliminary design for selecting suitable formulation parameters. This design should be a simple design of relatively few runs to screen suspected variables. Factorial designs whether full or fractional and their descendants such as Plackett-Burman or Taguchi designs fit very well for this job.<sup>[28](#page-16-7)</sup>

<span id="page-2-6"></span><span id="page-2-3"></span>After the selection of certain factors (independent variables) (IVs) and subsequent levels or ranges to be studied, the main statistical design for optimizing these IVs (whether they are process variables (PVs) or mixture components (MixCs) or both) should be implemented. It's preferable not to consume all the experimentation trials of the research in this design. Only about 70– 80% of experimentation trials are used to build this experimental design. In case of studying PVs where changing the level of one factor does not affect the others, a response surface design (RSD) may be used. This may be a Box-Behnken design, a central composite design, an optimal design, etc. While if the studied factors are MixCs, a mixture design (MixD) should be used. Although really important, MixDs are not as popular as RSDs. MixDs should be used when the studied IVs are proportions of a blend, where changes in one of the components can affect the relative amounts of others. Types of MixDs include simplex lattice design, simplex-centroid design, and various types of optimal designs.<sup>[34](#page-16-13)[,35](#page-16-14)</sup> In different types of simplex designs, there is a specified pattern for DPs (mixtures of specific composition to be formulated) selection. However, in optimal designs there is more flexibility in DPs selection. This DPs selection is dictated by a desire of achieving a certain property (optimality criterion). An algorithm is used to build the design in a manner that highly achieves this optimality criterion. There are different types of optimal designs according to the optimality criterion.<sup>28</sup> D-optimal designs are best for screening designs as they produce a design that best estimates factors' effects. A-optimal designs minimize the average variance of the polynomial coefficients. G-optimal designs minimize the maximum value of prediction variance in the design region. Integrated variance optimal designs (I-optimal designs) minimize the average prediction variance across the experimentation region. These optimal designs may be mandatory in some cases. These obligatory cases include unequal component ranges or multi-component constraints. In others, they are privileged for the ability of selecting a certain optimality criterion according to the study aims. Where it's required to study MixCs

and PVs simultaneously, Combined mixture designs<sup>34</sup> can be used, although their application in pharmaceutical research is still rare.

After implementation of the main optimization statistical design and elucidation of suitable models for navigation of design space, process or formulation optimization is performed either numerically or graphically. Now, some questions must be asked. Was the optimized formulation in the center of the design, or was it marginal? Has the optimized formulation achieved my predetermined constraints to a satisfactory degree or do I need to enhance certain response(s)? If the optimized formulation is marginal (that is, you can reach better results by choosing different levels or ranges or you need to improve a certain response in the optimized formulation), here comes the use of the saved 20–30% of the experimentation trials.

Instead of performing a new design from scratch to choose different levels of PVs or ranges of MixCs, a novel statistical approach can be adopted, which is expansion of the design space. The saved 20–30% of the experimentation trials is now used to change the levels of some factors or ranges of MixCs. This change is dictated by the direction which is believed to be towards a sweeter optimization spot or towards the increase or decrease of a certain response. This change leads to an expansion of design space without the need of building a new design. The new DPs performed in this extra space may be added in the same block as the initial DPs or may be added in a new block. This can be decided case by case by judging whether any of the constant factors have been changed for the new DPs.<sup>36</sup>

## <span id="page-3-0"></span>**Experimental details**

#### Materials

Tazarotene was kindly supplied by ENALTEC Labs, Navi Mumbai, India. Glyceryl monooleate (GMO), Pluronic® F127 (PL-F127), Pluronic F108 (PL-F108), Pluronic F68 (PL-F68), Brij® 53, and Polyvinyl alcohol (PVA; Mwt. 13,000–23,000, 87–89% hydrolyzed) were purchased from Sigma Aldrich®, Darmstadt, Germany. Methanol, Sodium chloride, Disodium hydrogen phosphate, Potassium dihydrogen phosphate and Sodium lauryl sulphate were purchased from Adwic, El-Nasr Pharmaceutical Chemicals Co., Cairo, Egypt. All other chemicals and solvents were of analytical grade. Water used was deionized, distilled water.

## Statistical Design

In this study mixed factorial design (MFD) (3.2) was chosen as the preliminary design. This design was chosen to select a stabilizer and a surfactant for formulation of TAZA loaded cubosomal dispersions. Three levels of stabilizer (PL-F68, PL-F108 and PL-F127) and two levels of surfactant (PVA and Brij 35) were incorporated into the study. Fixed amounts of GMO, TAZA and water were used. The prepared formulations were investigated for their entrapment efficiency percentage (EE) and particle size (PS).

On the basis of results obtained from the preliminary study, a statistical design was needed to search for the best proportion with which the components should be mixed to optimize a cubosomal formulation with desired characteristics. Hence an I-optimal mixture design (IOMD) was chosen as it can be the mixture design of choice when prediction across the experimental space is the main aim of the study. It was decided to use only 10 experimentation trials in this design to leave the possibility for design space expansion if needed. The components studied in the IOMD coded in different methods together with the responses studied and their constraints are shown in [Table 1](#page-4-0). [Figure 1](#page-4-1) shows the IOMD space with its 10 DPs (C1-C10). The detailed composition of the 10 cubosomal formulations (C1-C10) prepared accordingly is shown in [Table 2.](#page-5-0)

Expansion of design space in a certain required direction was finally adopted. Four extra trials were performed which represent the DPs at the two new vertices, the center of the new edge and the overall centroid of the expanded mixture design (EMD) space (see [Figure 1](#page-4-1)). As the experimental space was expanded in a certain direction, only some of the limits of the ranges of original space of the IOMD changed. The limits that changed to obtain the new EMD space are shown in italics in [Table 1](#page-4-0). The detailed compositions of the extra four DPs (C11-C14) are shown in [Table 2.](#page-5-0)

## Preparation of TAZA Cubosomes

TAZA cubosomes (TAZA-Cs) were prepared by melt dispersion emulsification method.<sup>[15](#page-15-13)</sup> Accurately weighed amounts of GMO, stabilizer and 10 mg TAZA were melted on a hot plate magnetic stirrer (Model MSH-20D, Wise Stir, Daihan Scientific Co. Ltd, Korea) at 70°C. Accurate amount of surfactant was dissolved in a certain amount of water and the



<span id="page-4-0"></span>

**Notes**: Altered limits for the expanded mixture design are shown in parentheses. Studied responses and their targeted constraints are also shown.

<span id="page-4-1"></span>

**Figure 1** I-optimal mixture design (IOMD) space - defined by the constraints 300≤A≤420, 120≤B≤240 and 200≤C≤320 - and its DPs (C1–C11) together with extra added DPs (C11–C14) to give the expanded mixture design (EMD) space defined by the constraints 300≤A≤510, 120≤B≤330 and 110≤C≤320. Where; A = Glyceryl monooleate amount (mg), B = Pluronic F68 amount (mg), and C = Polyvinyl alcohol amount (mg). Design points (DPs): O single trial, ● replicated trial, ◊ selected optimized formulation from IOMD (IOMD-OF), Δ selected optimized formulation from EMD (EMD-OF).

Std	A: GMO*	<b>B: PL-F68*</b>	C: PVA*	EE **	PS**	Q24h**	PDI**	$ZP**$
<b>CI</b>	420	120	200	$88.5 \pm 0.1$	207.9±11.5	$58.8 \pm 1.0$	$0.36 \pm 0.05$	$-40.1 \pm 0.3$
C <sub>2</sub>	360	180	200	$82.0 \pm 1.4$	$256.8 \pm 5.3$	$57.0 \pm 2.7$	$0.42 \pm 0.05$	$-33.5 \pm 1.5$
C <sub>3</sub>	360	180	200	$80.2$ ±1.1	$240.9 \pm 12.9$	$55.6 \pm 1.2$	$0.40 \pm 0.04$	$-38.1 \pm 0.4$
C <sub>4</sub>	300	240	200	$85.3 \pm 0.8$	$317.2 \pm 51.7$	$62.5 \pm 0.9$	$0.47 \pm 0.01$	$-33.5 \pm 1.7$
C <sub>5</sub>	340	160	240	$79.5 \pm 1.4$	$245.1 \pm 6.1$	$60.9 \pm 7.2$	$0.50 \pm 0.04$	$-36.9 \pm 1.7$
C <sub>6</sub>	340	160	240	$81.1 \pm 0.4$	$251.5 \pm 2.7$	$60.8 \pm 4.1$	$0.38 \pm 0.03$	$-31.9 \pm 0.6$
C7	360	120	260	$87.2 \pm 0.6$	$203.6 \pm 15.7$	$56.3 \pm 2.9$	$0.29 \pm 0.02$	$-35.5 \pm 1.3$
C8	360	120	260	$89.6 \pm 0.4$	$202.9 \pm 12.5$	$54.1 \pm 6.9$	$0.30 \pm 0.03$	$-50.7 \pm 0.6$
C9	300	180	260	$86.3 \pm 2.0$	$258.8 \pm 41.0$	$56.7 \pm 2.8$	$0.43 \pm 0.05$	$-41.4 \pm 0.2$
C10	300	120	320	$87.3 \pm 1.0$	$204.8 \pm 1.9$	$51.9 \pm 2.8$	$0.41 \pm 0.01$	$-35.1 \pm 4.5$
<b>CII</b>	510	120	110	$83.7 \pm 2.6$	$209.7 \pm 12.5$	$42.5 \pm 2.6$	$0.43 \pm 0.05$	$-36.5 \pm 0.9$
C12	405	225	110	$81.7 \pm 0.9$	$236.8 \pm 1.7$	$58.1 \pm 6.2$	$0.47 \pm 0.02$	$-46.5 \pm 1.8$
C13	300	330	110	$82.1 \pm 1.1$	$273.0 \pm 11.2$	$68.8 \pm 6.6$	$0.47 \pm 0.04$	$-36.8 \pm 0.6$
C14	370	190	180	$76.6 \pm 0.4$	$229.9 \pm 4.4$	$61.0 \pm 6.9$	$0.50 \pm 0.01$	$-37.3 \pm 0.5$

<span id="page-5-0"></span>**Table 2** Composition and Characterization of Cubosomes Prepared According to Different Design Points of I-Optimal Mixture Design (C1-C10) and Extra Points Added in Expanded Design Space (C11–C14)

**Notes**: \*Studied mixture components amounts: A: Glyceryl monooleate (GMO) (mg); B: Pluronic acid F68 (PL-F68) (mg); C: Polyvinyl alcohol (PVA) (mg). \*\*Measured Responses. Data are duplicate measurements of each trial (mean ±SD).

**Abbreviations**: EE, Entrapment efficiency; PS, Particle size; PDI, Polydispersity index; Q24h, drug percent released after 24 hours; ZP, zeta potential.

<span id="page-5-1"></span>aqueous solution was heated to the same temperature and added dropwise to the molten lipid mixture under continuous stirring at 950 rpm. TAZA-Cs were formed as milk-like dispersions. All formulations were left to cool gradually at room temperature. Then they were homogenized by intermittent probe sonication for 10 minutes (5 s on and 5 s off).<sup>[37](#page-16-16)</sup> TAZA-Cs were kept at room temperature for further investigations.

## Characterization of Prepared Cubosomes

#### EE and Drug Loading (DL)

<span id="page-5-2"></span>Entrapment efficiency of TAZA-Cs was determined by separating the un-entrapped free drug. One mL of each formulation was centrifuged for 1 hour at 4°C and 22,000 rpm in an ultra-cooling centrifuge (Model 3–30KS, Sigma Laborzentrifugen GmbH, Osterode am Harz, Germany). The supernatant containing free TAZA was diluted with methanol and spectrophotometrically measured (UV-1601 PC shimadzu, Kyoto, Japan) at  $\lambda_{\text{max}}$  = 351.5 nm. EE was calculated using the following formula: $38$ 

$$
EE = \left(\frac{TAZA_{Total} - TAZA_{Supernatant}}{TAZA_{Total}}\right) \times 100\tag{1}
$$

<span id="page-5-3"></span>Drug loading for TAZA-Cs was calculated using the following equation:<sup>[39](#page-16-18)[,40](#page-16-19)</sup>

$$
DL = \left(\frac{\text{Amount of TAZA entrapped}}{\text{Cubosomes weight}}\right) \times 100\tag{2}
$$

Where cubosomes weight is the weight of the lipid mixture of the formulation.

#### PS and Polydispersity Index (PDI)

In order to determine PS and PDI of the formulations, TAZA-Cs were appropriately diluted with water and measured by Malvern zetasizer (Model ZEN3600, Malvern Instrument Ltd, Worcestershire, England). The measurements were carried out at  $25^{\circ}C^{41}$ 

#### ZP

<span id="page-6-1"></span><span id="page-6-0"></span>Determination of ZP depends on measuring the mobility of charged moieties in an electric field.<sup>[41](#page-16-20)</sup> Malvern zetasizer was used to measure ZP at a potential of  $\pm$  150 mV via a laser Doppler anemometer.<sup>42</sup> TAZA-Cs were diluted and measured for ZP at 25°C.

#### Q24h

<span id="page-6-3"></span><span id="page-6-2"></span>The in-vitro TAZA release from TAZA-Cs was conducted in a horizontal shaking water bath (GFL, Gesellschaft Laboratories, Berlin, Germany). One mL of each formulation was placed in a dialysis bag (12,000–14,000 molecular weight cut off). The release medium was 50 mL hydro-alcoholic solution (PBS of  $pH = 7.4$ : methanol in ratio 1:1) containing 0.05% sodium lauryl sulphate at 37°C  $\pm$  0.5 under agitation speed of 50 rpm.<sup>[21](#page-16-0)</sup> The solubility of TAZA in water is very low (practically insoluble in water)<sup>[43](#page-16-22)</sup> as it is a highly lipophilic drug (Log P= 5.96).<sup>[11](#page-15-9)</sup> Hydroethanolic solution was used as a release medium to achieve the sink conditions.<sup>21,[44](#page-16-23)</sup> Samples of 3 mL were withdrawn at time intervals 0.5, 1, 1.5, 2, 3, 4, 6, 8 and 24 hours. The withdrawn aliquots were replaced by 3 mL of the dissolution medium each time. The amounts of TAZA released ( $\mu$ g/mL) were spectrophotometrically measured at  $\lambda_{\text{max}} = 351.5$  nm. For all formulations,  $Q_n$  was calculated according to the following formula:<sup>[38](#page-16-17)</sup>

$$
Q_n = \frac{\left(C_n \times V_r + \sum_{i=1}^{n-1} C_i \times V_s\right)}{\text{Initial drug amount}} \times 100\tag{3}
$$

Where,  $Q_n$  represents the cumulative percent of TAZA released,  $C_n$  is the receptor medium current concentration at n<sup>th</sup> sample, V<sub>r</sub> is the receptor medium volume, V<sub>s</sub> is the volume of each sample removed for analysis, and  $\sum_{n=1}^{n-1}$  $\sum_{i=1} C_i$ denotes the summed total of the previously measured concentrations. Q24h was determined for each formulation.

## Testing the Validity of the Elucidated Polynomial Models

In order to test the validity of the proposed statistical models for the whole EMD, 95% two-sided prediction intervals (95% PIs) for the predicted values of the investigated responses were calculated. It is an objective and statistically suitable method for model validity testing. The prediction interval (PI) is the range within which a single future response for a specified combination of PVs/MixCs is supposed to lie. All prepared 14 combinations (C1-C14) were inspected to find whether the actual values for their measured responses fall within the 95% PIs or not. Also percent deviation of actual responses measured from predicted ones were calculated according to the following equation.<sup>38</sup>

$$
Y_m = 100 \times \left| \frac{Y_m \text{predicted} - Y_m \text{actual}}{Y_m \text{predicted}} \right| \tag{4}
$$

#### Development of Optimized Formulation

<span id="page-6-4"></span>Optimization of TAZA-Cs was performed using the multi-response numerical optimization technique.<sup>45</sup> Desirability function based on the individual desirability  $(d_m)$  of each response  $(Y_m)$  was used as the basis of optimization in many studies[.29,](#page-16-8)[30](#page-16-9) In this research an extra feature of this function was used which is different levels of importance for responses to be optimized. A certain response (Q24h) was chosen to have higher importance than other responses as the release of the optimized formulation was the most important criterion. Q24h was to be given an importance value of 3 and the other responses a value of one. Design expert software (V.12, Stat-Ease Inc., MN, USA) (DX) software offers this by changing the importance given to each response from one to five based on the following formula:

$$
D = (d_1^{r_1} \ldots d_n^{r_n})^{\frac{1}{\sum r_i}} \tag{5}
$$

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where D is the desirability of the formulation ranging from zero to one,  $d_m$  is the individual desirability of each response ranging from zero to one, m is the number of responses to be optimized,  $r_i$  is the importance given to each response ranging from one to five.

At first IOMD was navigated and an optimized formulation within this space (IOMD-OF) was found. After design space expansion for enhancing the properties of IOMD-OF, another optimization process was performed to find the optimized formulation of the whole expanded mixture design (EMD-OF).

## Investigation of Morphological Characters of EMD-OF

The morphological characters of the EMD-OF were investigated by transmission electron microscope (TEM) (Jeol, Jem-2100, Tokyo, Japan). EMD-OF was diluted with water then a drop of the diluted dispersion was placed on a copper grid coated with carbon, negatively stained with 1% phosphotungstic acid  $(w/v)$  and left to dry in air for 5 minutes. The stained sample was then placed into TEM for imaging. TEM was operated at 80 kv at room temperature and the morphological characters EMD-OF were inspected at proper magnifications.<sup>[46](#page-16-25)</sup>

## <span id="page-7-0"></span>Statistical Analyses

Design expert software® (V.12, Stat-Ease Inc., Minneapolis, MN, USA) was used for analysis of the MFD and construction of the IOMD and its subsequent analysis and finding IOMD-OF. It was used for navigation of the expanded design space as well as finding EMD-OF.

## **Results and Discussion**

#### Preliminary Parameters Selection

Upon exploring the different factors and interactions affecting different responses PL-F68 and PVA were respectively selected from the stabilizers and surfactants studied in the MFD as this would result in the maximized EE and minimized PS.

## IOMD Results and Analysis

<span id="page-7-1"></span>Composition and characterization of formulations prepared according to the IOMD (C1-C10) are listed in [Table 2.](#page-5-0) Release profiles of TAZA from these trials and from TAZA suspension are shown in [Figure 2.](#page-8-0) The chosen ranges for MixCs studied (stabilizer and surfactant) led to a wide range of characterization results. EE ranged from 79.5%  $\pm$  1.4 to 89.6%  $\pm$  0.4, PS ranged from 202.9 nm  $\pm$  12.5 to 317.2 nm  $\pm$  51.7, Q24h ranged from 51.9%  $\pm$  2.8 to 62.5%  $\pm$  0.9, PDI ranged from 0.29 nm  $\pm$  0.02 to 0.50  $\pm$  0.04 and ZP ranged from −31.9 mV  $\pm$  0.6 to −50.7mV  $\pm$  0.6. All the formulations had a PDI of 0.5 or less indicating narrow size distribution.<sup>47</sup> As for ZP which gives an indication about the physical stability of the system, all the formulations carried negative charges. This is mainly due to the presence of ionized carboxylic groups of GMO on their surfaces. It can also be due to the ionized water molecules surrounding the cubosomes.[48](#page-16-27) ZP of all prepared formulations was more negative than −30 mv. It was reported that an absolute ZP value more than 30 mV is sufficient for good repulsion and physical stability.<sup>[49](#page-16-28)[,50](#page-16-29)</sup> So, all the prepared Cs are considered to have an acceptable stability against aggregations. Thus, optimization was based on selecting a formulation that maximizes EE and Q24h and minimizes PS.

## <span id="page-7-3"></span><span id="page-7-2"></span>Optimization Based on IOMD

The measured values of the three responses; EE, PS and Q24h were individually analyzed. Reduced polynomial models based on maximizing prediction  $R^2$  were derived. EE, PS and Q24h were modeled with suitable prediction  $R^2$  of 0.8501, 0.9432 and 0.9028 respectively. Contour plots for the EE, PS and Q24h over the IOMD space are shown in [Figure 3A-C.](#page-9-0) Numerical optimization - giving higher importance of 3 to Q24h - based on these models was performed. This optimization resulted in a formulation having the components of the previously performed trial C1. This formulation  $(C1 = IOMD-OF)$  resulted in EE of 88.7%. PS of 206.5 nm and Q24h of 58.8% and desirability of 0.753. IOMD-OF is shown in [Figure 3D](#page-9-0) having the highest possible desirability.

<span id="page-8-0"></span>

Figure 2 Release profiles of TAZA from trials (CI - C14) compared with TAZA release from suspension.

## Design Space Expansion

This IOMD-OF had unsatisfactory Q24h, and here comes the importance of not using all experimentation trials to be done in the main design. Expansion of design space was done in an attempt to increase Q24h. The direction of design expansion was dictated by the two vertices, one at which maximum Q24h was found and the other showing maximum desirability in the IOMD (see arrows in [Figure 3C](#page-9-0) and [D](#page-9-0)). These directions dictate an increment in the upper limits of A: GMO and B: PL-F68 and a reduction in lower limit of C: PVA. These limits changes are shown in parentheses in [Table 1](#page-4-0). Expanded design space is shown in [Figure 1](#page-4-1) in which the DPs of the IOMD (C1–C10) and the extra added points (C11–C14) are clarified. The whole EMD with its 14 DPs  $(C1-C14)$  was navigated to model each response individually. Composition and characterization of the extra added points  $(C1-$ C14) are listed in [Table 2](#page-5-0). Release profiles of TAZA from these trials and from TAZA suspension are shown in [Figure 2](#page-8-0).

<span id="page-9-0"></span>

**Figure 3** Contour plots showing the changes in (**A**) Entrapment efficiency, (**B**) Particle size, (**C**) Drug percent released after 24h (Q24h) and (**D**) desirability over the I-optimal mixture design space. Arrows represent regions of maximum: (**C**) Q24h and (**D**) desirability, ✖ represents the selected optimized formulation based on this design (IOMD-OF).

#### EMD Results and Analysis

EE was adequately modeled using reduced three-factor interaction model with prediction  $R^2$  of 0.7342. PS and Q24h responses could not be directly modeled. Box-Cox transformations were recommended by DX for these responses.

<span id="page-10-1"></span>Box-Cox transformation was introduced by George Box and David Cox in 1964 to transform the studied data into normal shape.<sup>28</sup> Design expert software has the ability to suggest and conduct those transformations for some responses when ordinary modeling is not adequate enough.<sup>[51,](#page-16-30)[52](#page-16-31)</sup> This transformation minimizes the natural log (ln) of the residual sum of squares by selecting a certain value for power transformation (λ) to which all results of a response are raised. The required transformations for the responses of PS and Q24h are shown in [Figure 4](#page-10-0). PS and Q24h were recommended to be raised to the power of −1.62 and 2.24 respectively. These recommended transformations were applied leading to adequate modeling of PS and Q24h with prediction  $\mathbb{R}^2$  of 0.9225 and 0.8145 respectively. Final model terms for each response and their coefficient estimates appearing in the final equations derived from EMD analysis in addition to their p-values are illustrated in [Table 3.](#page-11-0) This table shows final models' evaluations too. From this table it is clear that all measured responses were significantly affected by the studied mixture components. Alpha was set at 0.05, and coefficients with p value <0.05 (shown in bold) are considered to significantly affect the respective response. As for model evaluation, all the derived models showed relatively small relative mean square error (RMSE) and coefficient of variation percent (CV%). Model adequacy was also verified by the high values of different calculated regression coefficients  $(R^2)$ ;  $R^2$ , Adjusted  $R^2$  and Prediction  $R^2$ . Prediction  $R^2$  is the most relevant regression coefficient in optimization studies. Prediction  $R^2$  determines the ability of the derived model to predict the responses' values for a new formula within the design space even if its mixture combination has not been investigated in the design. Prediction  $\mathbb{R}^2$  values for the three studied responses were 0.7342, 0.9225 and 0.8145 for EE, PS and Q24h respectively. These values were high enough and were in good accordance with the Adjusted  $\mathbb{R}^2$  values of 0.7873, 0.9255 and 0.8410 respectively.

#### Testing the Validity of the Developed Polynomial Models

In order to check the validity of results obtained from the EMD, the actual results of the three investigated responses for each of the 14 TAZA-Cs were compared with the predicted values developed by the software. Comparison took place via percentage deviation calculation. Maximum % deviation for EE, PS and Q24h were 3.8%, 7.3% and 6.3% respectively.

<span id="page-10-0"></span>

Figure 4 Box-Cox plot for data transformation showing In residual sum of squares (In residual SS) without transformation and after recommended transformation for the responses: (**A**) Particle size, (**B**) Drug percent released after 24h.

<b>Terms</b>	EE**		$PS^{-1.62}**$		$Q24h^{2.24**}$						
	<b>Coefficient</b>	p-value	<b>Coefficient</b>	p-value	<b>Coefficient</b>	p-value					
A*	83.4	0.0146	0.00017	< 0.0001	4591.5	< 0.0001					
B*	82.2	0.0146	0.00011	< 0.0001	13,080.2	< 0.0001					
$C^*$	87.1	0.0146	0.00018	0.0001	6880.6	< 0.0001					
AC	11.7	0.1160	.	$\cdots$	11,383.8	0.0026					
BC	.	$\cdots$	$-0.00021$	0.0001	$\cdots$	$\cdots$					
<b>ABC</b>	$-202.0$	0.0002	$\cdots$	$\cdots$	.	$\cdots$					
Model type	Reduced special cubic		Reduced quadratic		Reduced quadratic						
<b>Model evaluation</b>											
<b>RMSE</b>	1.76		8.04E-06		781.29						
<b>C.V. %</b>	2.1		5.52		8.81						
$R^2$	0.8528		0.9427		0.8777						
Adjusted R <sup>2</sup>	0.7873		0.9255		0.841						
<b>Prediction <math>R^2</math></b>	0.7342		0.9225		0.8145						

<span id="page-11-0"></span>**Table 3** Coefficient Estimates for Different Model Terms – Appearing in the Final Equations from the Expanded Mixture Design – for Each Response and Their Significance, Together with Models' Types and Evaluation

**Notes**: \*Studied mixture components amounts: A: Glyceryl monooleate (GMO) (mg); B: Pluronic acid F68 (PL-F68) (mg); C: Polyvinyl alcohol (PVA) (mg). \*\*Measured Responses. p-values < 0.05 indicating significant coefficients are shown in bold.

**Abbreviations**: EE, entrapment efficiency; PS, particle size; Q24h, drug percent released after 24 hours; ZP, zeta potential; RMSE, relative mean square error; CV%, coefficient of variation percent.

All the results of the three responses for all formulations were found to fall within 95% PI. This ensures the validity and reliability of the derived models over the EMD space.

## Effect of MixCs Used on the Studied Responses

For illustrating the effects of the studied MixCs on each of the studied responses, contour plots and Piepel's trace plots were used. In contour plots, the response under study is shown as lines of equal value. These lines show the variation of this response all over the design space. In Piepel's trace plots, the reference blend of values (1/3, 1/3, 1/3) for the three MixCs is the main point in the plot. A line is drawn for each MixC. The X axis for each line represents the deviation of this MixC from the reference blend in L-pseudo values. As each MixC has L-pseudo values ranging from 0 to 1, its deviation from reference blend ranges from  $-1/3$  to 2/3. Tracing each MixC is moving with this MixC from its base (L-pseudo value = 0 and deviation =  $-1/3$ ) passing through reference blend (L-pseudo value =  $1/3$  and deviation = 0) till we reach its peak (L-pseudo value = 1 and deviation  $= 2/3$ ). Upon tracing the increment in each MixC, the values of the two other MixCs decrease simultaneously but keeping their ratio the same as in their reference blend (1:1). The response under study is shown on the Y axis.

#### EE and DL

<span id="page-11-3"></span><span id="page-11-2"></span><span id="page-11-1"></span>EE is a valuable parameter in the assessment of nano-drug delivery systems. It shows the capability of the prepared nano-system to carry the drug. EE represents the portion of drug incorporated into the nano-carrier from the initial amount used. For accurate EE determination, the unentrapped drug needs to be separated from the nano-carrier containing the entrapped drug.<sup>[53](#page-17-0)</sup> The higher the EE the more drug delivered into the skin. This can be beneficial in the treatment of some topical diseases such as psoriasis which need high levels of drug into the skin.<sup>[54](#page-17-1)</sup> Also, high EE means less irritation to the skin surface that may occur when there is a direct contact with the drug.<sup>[55](#page-17-2)</sup>

<span id="page-12-0"></span>

**Figure 5** Contour plots and Piepel's trace plots showing the changes in Entrapment efficiency (**A** and **B**), Particle size (**C** and **D**), Drug percent released after 24h (**E** and **F**) over the expanded mixture design space.

From [Figure 5A](#page-12-0) and [B](#page-12-0) illustrating  $Y_1$  (EE), it can be seen that for all the studied components, the reference blend showed nearly the least entrapment efficiency, with increasing the EE upon decreasing or increasing any of the studied MixCs. Before the reference blend, increasing GMO leads to an increase in the hydrophobicity of cubosomes and accordingly production of smaller PS. This may result in decreasing EE with increasing GMO before reference blend. As for reduction in EE with increasing of PVA and PL-F68 before the reference blend, this may be due to increasing solubility of TAZA in the aqueous phase. After the reference blend point, the increase of any component led to increasing EE. Increasing EE with increasing GMO amount can be attributed to the lipophilic nature of TAZA so it tends to be more entrapped when GMO increases as it became strongly attracted to the hydrophobic domain of the dispersion. Increasing EE with increasing either PL-F68 or PVA may be attributed to forming a coat around the formed cubosomes to stabilize them. This coat might contribute to the increased EE by retaining more drug with higher amounts of PL-F68 or PVA.<sup>56[,57](#page-17-4)</sup> DL for all the studied formulations ranged from 1 to 1.2%.

#### <span id="page-13-1"></span>PS

<span id="page-13-2"></span>PS is a crucial parameter to be investigated for nano-carriers, as it has an influence on EE, drug release profiles, stability, mucoadhesion and cellular uptake of nano-systems. $58,59$  $58,59$  $58,59$ 

<span id="page-13-3"></span>The response of PS is illustrated in [Figure 5C](#page-12-0) and [D.](#page-12-0) As for GMO, the higher the amount of GMO used the lower the PS. This may be due to increasing the hydrophobic domain of the cubosomal dispersion and therefore the surface free energy decreased leading to smaller PS.<sup>[60](#page-17-7)</sup> This is in contrary with Younes et al. and El-Deeb et al. Results of both studies stated that increasing GMO led to increase in PS of Sertaconazole and Brimonidine tartrate cubosomes respectively. The findings of these studies could be because of the increased dispersion viscosity which might have hindered formation of smaller cubosomes.<sup>[15,](#page-15-13)[16](#page-15-14)</sup> PS showed a parabolic increase with increasing amount of PL-F68. This may be attributed to the intercalation of the increased amounts of PL-F68 in between the cubosomal bilayers. These findings are in agreement with the results reported by El-Deeb et al. It was found that high PL-F68 amounts in respect to GMO amounts caused a significant increase in particle size of Brimonidine tartrate cubosomes. This was justified that the triblock copolymer molecules of the poloxamer were accumulated into the bilayer causing its expansion and PS enlargement.<sup>[15](#page-15-13)</sup> PS showed a parabolic increase with increasing amount of PVA with maximum PS at reference blend then PS decreases with further increment in the amount of PVA. The increase in PS before reaching the reference blend may be attributed to PVA adsorption on cubosomal surface. Also, it can be due to the elevation of aqueous phase viscosity that may hinder the dispersion and subdivision of droplets.<sup>[56](#page-17-3)</sup> The reduction of PS with increasing PVA after reference blend may be explained on the basis that the PVA as a surfactant can minimize particles coalescence leading to smaller cubosomes. This explanation was also adopted by Esposito et al. They reported that increasing the PVA concentration led to PS reduction of topical cubosomes due to minimizing particles coalescence. $61$ 

#### <span id="page-13-4"></span><span id="page-13-0"></span>Q24h

<span id="page-13-6"></span><span id="page-13-5"></span>The in vitro drug release studies reflect the quality of nano-carriers. It can also predict the nano-carrier in vivo behaviour and performance.<sup>62,[63](#page-17-10)</sup> It was reported that nano-carriers intended for topical delivery may be embedded into the stratum corneum and drugs are supposed to be released and diffuse through the remaining deep skin layers.<sup>[64](#page-17-11)</sup> This shows how much the evaluation and determination of drug percent released is crucial. Accordingly, Q24h was selected to be the most important response incorporated into the design.

<span id="page-13-8"></span><span id="page-13-7"></span>The response of Q24h is illustrated in [Figure 5E](#page-12-0) and [F.](#page-12-0) Generally, the increase of GMO or PVA amounts resulted in a decrease in Q24h. On the contrary the increase in PL-F68 led to an increase in Q24h. Because TAZA is a highly lipophilic drug (log  $P=5.96$ ), it prefers to be retained into the lipophilic cubosomes rather than diffusing into aqueous release media. GMO was reported to cause release retardation for lipophilic drugs.<sup>[65](#page-17-12)</sup> So, increasing GMO led to a logic decrease in Q24h due to the elevation of hydrophobic portion of cubosomal dispersion. These findings were similar to those reported by Elgindy et al. as amounts of progesterone released after 24 hours from progesterone-loaded cubosomes decreased with increasing GMO amount.<sup>[48](#page-16-27)</sup> Similar results were mentioned by Sherif et al. where the percentage released of alpha lipoic acid from cubosomes decreased by increasing GMO amounts.<sup>[66](#page-17-13)</sup> The increase in O24h with increasing PL-F68 may be due to increasing TAZA solubility in the release media with increasing PL-F68 proportion. This may help the drug diffusion through the <span id="page-14-1"></span>medium and increase Q24h. This finding is contrary to Morsi et al., who found that elevated poloxamer-407 concentration led to decrease in percentage silver sulfadiazine released from the cubosomes. The explanation for this is that at low poloxamer concentrations, the polymer molecules adhere to the cubosomal surface and fewer are incorporated into its internal structure. This led to formation of cubosomes with double diamond (cD) patterns. But, at higher poloxamer concentrations, it participates in the internal cubic matrix leading to a primitive (cP) structure which is characterized by more restricted drug release than cD patterns.<sup>[67](#page-17-14)</sup> PVA, as previously mentioned, can make a coat surrounding the cubosomes. This may result in increasing the distance through which the drug diffuses. Therefore, at higher PVA amounts, fewer drug amounts are released. This is in agreement with the results found by Aboud et al. where the increase in PVA concentration decreased Sildenafil release rates from the prepared cubosomes.[56](#page-17-3)

## Optimization Based on EMD

Numerical optimization - giving higher importance of 3 to Q24h - based on the models derived for EMD was performed. This optimization resulted in a formulation having the components of the previously performed DP C13. This formulation (C13 = EMD-OF) resulted in EE of 82.1 $\pm$ 1.1%, PS of 273.0 $\pm$ 11.2 nm and Q24h of 68.8 $\pm$ 6.6% and desirability of 0.650. EMD-OF is shown in [Figure 1.](#page-4-1)

## Investigation of Morphological Characters of EMD-OF

Morphological examination of stained EMD-OF using TEM - [Figure 6](#page-14-0) - showed the irregular angular cubic shapes of the prepared optimized formulation. PS was also in harmony with those measured by zetasizer.

<span id="page-14-0"></span>

Figure 6 Transmission electron microscope morphological examination for stained expanded mixture design optimized formulation showing its cubic shape.

## **Conclusion**

Statistical sequential experimentation was adopted successfully in this research for optimizing Tazarotene cubosomes. This was done through three successive steps. First step: A preliminary mixed factorial design for stabilizer and surfactant choice. Second step: Construction of the main experimental design (I-optimal mixture design) (IOMD) in which amounts of chosen stabilizer and chosen surfactant were simultaneously changed with amount of lipid used. Optimization of a formulation with predetermined characters of different levels of importance was then performed over the IOMD. As this optimization did not result in a formulation with satisfying in-vitro release, we moved to the third step. Third step: Novel design space expansion was implemented. This design space expansion allows changing ranges of components to be used. Navigating the new expanded mixture design space led to finding a new optimized formulation with satisfying properties. Thus this sequence of work enabled re-optimization without re-building a new design from scratch. So, this epitome is expected to have promising future applications in various pharmaceutical formulations optimization.

## **Disclosure**

The authors report no conflicts of interest in this work.

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