

Downregulation of microRNA-122 promotes proliferation, migration, and invasion of human hepatocellular carcinoma cells by activating epithelial–mesenchymal transition

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Objective: To investigate the effects of microRNA-122 (miR-122) on proliferation, migration, and invasion in human hepatocellular carcinoma (HCC) cells by activating epithelial–mesenchymal transition (EMT) pathways.

Methods: miR-122 mimics, miR-122 inhibitors, relevant control oligonucleotides, and *Wnt1* were transfected into HepG2 and huh7 cell lines which were then divided into six groups: miR-122 group, anti-miR-122 group, miR-negative control (NC) group, anti-miR-NC group, miR-122 + *Wnt1* group, and miR-122 + vector group. The miR-122 expressions and mRNA expressions of *Wnt1* and EMT-related genes (*E-cadherin*, *vimentin*, β -*cadherin*, and *N-cadherin*) were quantified by quantitative real-time polymerase chain reaction (qRT-PCR). Protein expression levels of *Wnt1*, *E-cadherin*, *vimentin*, β -*cadherin*, and *N-cadherin* were measured by Western blot. Cell proliferation, migration, and invasion were evaluated using 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay, wound-healing assay, and Transwell assay, respectively.

Results: Dual luciferase reporter gene results showed that *Wnt1* is a direct target gene of miR-122 in both HepG2 and huh7 cell lines. Compared to miR-NC, anti-miR-NC, and miR-122 + *Wnt1* groups, miR-122 expression was markedly higher in the miR-122 group and miR-122 + vector group, but was sharply decreased in anti-miR-122 group (both $P < 0.05$), and the mRNA and protein levels of *Wnt1*, *vimentin*, β -*cadherin*, and *N-cadherin* decreased significantly; also *E-cadherin* increased, and cell proliferation, migration, and invasion decreased in the miR-122 group and miR-122 + vector group (all $P < 0.05$), but the situation was totally reversed in the anti-miR-122 group (all $P < 0.05$).

Conclusion: Downregulation of miR-122 promoted proliferation, migration, and invasion of human HCC cells by targeting *Wnt1* and regulating *Wnt*/ β -catenin pathway which activated the EMT pathways.

Keywords: microRNA-122, HepG2 cells, epithelial mesenchymal transition, *Wnt1*, β -*cadherin*, proliferation, migration, invasion

Introduction

Hepatocellular carcinoma (HCC) is the third leading cause of cancer-related deaths worldwide, and the mortality rates are mainly related to late diagnosis and high tumor recurrence after surgery.¹ The 5-year survival rates vary widely between 27% and 78%, depending on the underlying disease course, and frequent metastasis are observed in clinics. Despite promising therapeutic strategies for HCC, surgical resection or liver transplant remains the best options currently available.² Several studies explored the

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underlying mechanisms of HCC progression with a view of understanding key pathways driving HCC and to uncover novel therapeutic targets.^{2,3} Epithelial–mesenchymal transition (EMT) is a cellular program that enables epithelial cells to lose epithelial properties and acquire mesenchymal phenotype, gaining the ability to invade neighboring tissue or undergo distant metastasis, and the process involves loss of epithelial cell markers (eg, E-cadherin, occludin, and mucin) and the overexpression of mesenchymal markers (eg, N-cadherin, fibronectin, and vimentin).⁴ EMT is a critical step in HCC metastasis, and is closely related with intrahepatic migration, extensive invasion, and poor survival of HCC patients.⁵ Alteration in microRNA (miRNA) expression influences tumor growth and metastasis at multiple stages of tumor progression, and deregulated miRNA expression is also found in hepatocarcinogenesis, modulating tumor cell growth, apoptosis, metastasis and invasion, and aggravating HCC.⁶

miRNAs are 21- to 25-nucleotide-long noncoding RNA molecules that regulate gene expression by binding to complementary sequences in the 3' untranslated region to promote translational repression or mRNA degradation.⁷ Up to 30% of genes are regulated by miRNAs in this manner, and important biological pathways, including EMT, are targets of miRNA regulation.⁸ Some miRNAs function as tumor suppressors by inhibiting EMT, while others serve as oncogenes disrupting the proper assembly of epithelial architecture, leading to tumorigenesis.⁹ Growing evidence suggests that miRNAs are essential for maintaining the balance between EMT and mesenchymal–epithelial transition (MET) (the reverse process of EMT), and MET can be induced to effectively control tumor growth and progression.^{10,11} miR-122 is the most abundant miRNA in the liver, accounting for an estimated 70% of the total miRNA population, and the maintenance of liver homeostasis is regulated by miR-122.¹² Importantly, miR-122 expression is either undetectable or very low in hepatic carcinoma during hepatic differentiation, intrahepatic metastasis, and invasion.¹³ It was reported that miR-122 could act as a tumor suppressor by targeting several pathways, including Wnt/ β -catenin signaling pathway.¹⁴ However, the detailed mechanisms remain unknown. In this study, we examined the associations between miR-122 and human HCC cell proliferation, migration, and invasion, and further studied the role of EMT in human HCC cell migration and invasion.

Materials and methods

Cell collection and culture

Human HCC cell lines HepG2 and huh7 were obtained from the Department of Oncology, The Affiliated Jiangyin Hospital, School of Medicine, Southeast University,

Jiangyin, People's Republic of China. Cell suspension (1 mL) was cultured in Roswell Park Memorial Institute-1640 medium (HyClone, Logan, UT, USA) supplemented with heat-inactivated 10% fetal bovine serum (FBS), 1,000 U/mL penicillin, and 100 mg/mL streptomycin and maintained in a 5% CO₂ humidified incubator at 37°C. Cells were digested with pancreatin after they reached 80%–90% confluence and passaged using standard methods.¹⁵ All the cell lines were provided from participants in our hospital after they signed the informed consent. The study was approved by the ethics committee of Affiliated Jiangyin Hospital, School of Medicine, Southeast University.

Dual luciferase reporter gene activity analysis

LipofectAMINE 2000 (Invitrogen, Carlsbad, CA, USA) was used to combine luciferase reporter vector pWnt1-Wt and pWnt1-Mut with miR-122 mimics or miR-negative control (NC) respectively, and cotransfection of the combination was conducted in HepG2 and huh7 cells; simultaneously, pRL-TK vector (containing HSV cytosine kinase promoter region, weakly expressing *Renilla reniformis* luciferase in a variety of cells, and purchased from Cyagen Biosciences Inc., Santa Clara, CA, USA) was cotransfected as a standard internal control. Specific transfection steps followed the LipofectAMINE2000 operating instructions; and the transfected plasmid was 0.8 μ g, the miRNA final concentration was 40 μ mol/L, and the number of transfected cells was 1×10^5 (subsequent transfection experiments used the same plasmid mass and cell numbers). After transfection for 36 hours, the cells were harvested and treated according to Promega Dual-Luciferase® Reporter activity assay kit instructions (Fitchburg, WI, USA), and cell luciferase activity was detected by a single photon detector. The relative luciferase activity was calculated as firefly luciferase activity value/*R. reniformis* luciferase activity. The experiment was repeated three times.

Construction of eukaryotic expression vector p-EGEP-C3-Wnt-1

Gene Bank was used to retrieve the full-length *Wnt-1* gene sequence (NM-005430), and the upstream primer and downstream primer was designed and synthesized by Shanghai Invitrogen Biotechnology Company (Shanghai, People's Republic of China), and Hind III and Kpn I restriction sites were introduced. The upstream primer was 5'-GCAAGCTTATGAGGTGGCTCCTGCC-3' (Hind III) and the downstream primer was 3'-GCGGTACCTAATTGGCAATCTCTTCGAAGTC-5' (Kpn I). Polymerase chain reaction (PCR) conditions were as follows: predenaturation

at 94°C for 5 minutes; a total of 30 cycles of denaturation at 94°C for 40 seconds, annealing at 58°C for 40 seconds, extension at 72°C for 60 seconds; and final extension at 72°C for 10 minutes. The PCR-amplified products were run on a 1.0% agarose gel electrophoresis, and the results were analyzed using the image processing system of Tanon GIS 2010, Shanghai, People's Republic of China. The PCR-amplified target fragment *Wnt-1* and p-EGEP-C3 plasmid were digested by Hind III and Kpn I, and the digested *Wnt-1* and p-EGEP-C3 plasmid fragments were run on a 1.0% agarose gel electrophoresis and collected and linked by T4 DNA ligase (Takara Bio, Shiga, Japan) at 16°C overnight, and the product was transformed into *Escherichia coli* DH5h competent cells which were cultured in Luria–Bertani agar culture plates containing kanamycin (50 mg/L). Ten monoclonal clones were randomly selected, and the colonies were used as templates to conduct PCR initial screening. The eukaryotic expression vector was named p-EGEP-C3-Wnt-1. Correct positive clones were identified and incubated overnight in Luria–Bertani culture medium containing kanamycin (50 mg/L). The recombinant plasmids were extracted and used as templates for PCR and double restriction enzyme digestion identification, and positive clones were sent to Shanghai Invitrogen Biotechnology Company to carry out sequencing.

Cell groups and transfections

In brief, 2×10^5 cells were cultured in 2 mL Dulbecco's Modified Eagle's Medium (DMEM) containing 10% FBS in a six-well plate. Cells were divided into six groups: 1) miR-122 group, transfected with miR-122 mimics; 2) anti-miR-122 group, transfected with miR-122 inhibitor; 3) miR-NC group, transfected with miR NC; 4) anti-miR-NC group, transfected with anti-miR NC; 5) miR-122 + Wnt1 group (cotransfected with miR-122 mimics and p-EGEP-C3-Wnt1 plasmid); and 6) miR-122 + vector group (cotransfected with miR-122 mimics and blank plasmid). The cells were incubated overnight to achieve 50%–60% confluence, and Lipofectamine™ 2000 reagent was used for transfections (the manufacturer's instructions were followed). The miR-122

Table 1 The transfection sequences in miR-122, anti-miR-122, miR-NC, and anti-miR-NC groups

Groups	Sequences
miR-122	Sense: 5'-UGGAGUGUGACAAUGGUGUUUG-3' Antisense: 5'-AACACCAUUGUCACACUCCAUU-3'
Anti-miR-122	5'-CAAACACCAUUGUCACACUCCA-3'
miR-NC	Sense: 5'-UUCUCCGAACGUGUCACGUTT-3' Antisense: 5'-ACGUGACACGUUCGGGAATT-3'
Anti-miR-NC	5'-CAGUACUUUUGUGUAGUACAA-3'

Abbreviations: miR-122, microRNA-122; NC, negative control.

sequences are shown in Table 1 (Shanghai GenePharma Co., Ltd, Shanghai, People's Republic of China). The transfected cells were placed in a thermostatic incubator at 37°C in 5% CO₂ for 6 hours. After culture medium was replaced, the excitation light wavelength of the laser confocal microscopy was adjusted to cy3 fluorescence, and laser confocal microscopy was employed to identify transfected cells.

RNA extraction and quantitative real-time PCR

Total RNA was extracted using RNA Iso-plus reagent (Takara Bio). Ultraviolet spectrophotometer was used for measuring RNA concentration and purity. The reverse transcription PCR process was performed using PrimeScript RT Reagent Kit according to the manufacturer's protocol.¹⁶ Primers were designed by Primer 5.0 (Primer-E, Ltd., Plymouth, United Kingdom), (Table 2), and the reverse primer for miR-122 and U6 PCR used as internal control was the Uni-miR qPCR Primer offered by the kit.¹⁷ The reverse transcription reaction contained 25 µL of total RNA and miRNA PrimeScript RT Enzyme Mix, and the 20 µL real-time PCR (RT-PCR) was performed in the ABI PRISM 7300 Fast Real-Time PCR System (Ambion, Foster City, CA, USA), with the conditions: 95°C for 5 minutes, 95°C for 10 seconds, and 40 cycles at 60°C for 20 seconds. The reaction for Wnt1- and EMT-related genes was performed under conditions of 95°C for 10 seconds, 95°C for 30 seconds, and 40 cycles at 60°C for 30 seconds. The products were separated by electrophoresis on 0.2% agarose gels. The results of RT-PCR were expressed as $2^{-\Delta\Delta Ct}$.

Table 2 The sequences of forward primers and reverse primers of miR-122, U6, Wnt1, E-cadherin, vimentin, β-cadherin, N-cadherin, and GAPDH

Genes	Primer sequences
miR-122	F: 5'-TTGAATTCTAACACCTTCGTGGCTACAGAG-3' R: 5'-TTAGATCTCATTTATCGAGGGAAGGATTG-3'
U6	F: 5'-CTCGCTTCGGCAGCACA-3' R: 5'-AACGCTTCACGAATTTGCGT-3'
Wnt1	F: 5'-TGGCTGGGTTTCTGCTACG-3' R: 5'-TCGAGGACGTCGGACT-3'
E-cadherin	F: 5'-CGGTGGTCAAAGAGCCCTTACT-3' R: 5'-TGAGGGTTGGTGCAACAACGTCGTTA-3'
Vimentin	F: 5'-TGAGTACCGGAGACAGGTCGAG-3' R: 5'-TAGCAGCTTCAACGCAAAGTTC-3'
β-Cadherin	F: 5'-ATTGTCCACGCTGGATTTC-3' R: 5'-GACCCAAATGTCGCAGTCAG-3'
N-cadherin	F: 5'-GTGCCATTAGCCAAGGGAATTCAGC-3' R: 5'-GCGTTCCTGTTCCACTCATAGGAGG-3'
GAPDH	F: 5'-ACAGTCAGCCGCATCTTCTT-3' R: 5'-GACAAGCTTCCCGTTCTCAG-3'

Abbreviations: miR-122, microRNA-122; GAPDH, glyceraldehyde phosphate dehydrogenase; F, forward; R, reverse.

Western blot analysis

After 48-hour transfection, cell debris was removed by centrifugation, and total protein was extracted using RIPA cell lysis buffer. Protein concentration was determined by bicinchoninic acid (BCA) Protein Assay Kit (Pierce, Rockford, IL, USA) and proteins were separated by sodium dodecyl sulfate–polyacrylamide gel electrophoresis and blotted onto polyvinylidene difluoride membranes. The membranes were blocked with bovine serum albumin for 1 hour, and subsequently incubated with primary antibodies overnight at 4°C (Wnt1, 1:1,000; E-cadherin, 1:1,000; vimentin, 1:1,000; β-cadherin, 1:2,000; and N-cadherin, 1:400). After three washes with tris-buffered saline and Tween 20, the blots were incubated with secondary antibody conjugated to horseradish peroxidase (1:10,000) (Chemicon International, Inc., Billerica, MA, USA) at 37°C for 1 hour, and visualized by electrogenerated chemiluminescence (Amersham, Freiburg, Germany). The images were analyzed by Sigma Pro 5.0 software (SPSS Inc., Chicago, IL, USA). Glyceraldehyde-3-phosphate dehydrogenase (GAPDH) was used as an internal reference. Relative protein content = targeted protein gray value/GAPDH gray value. Samples in each group were analyzed three times.

MTT assay

After cell transfection for 24 hours, 1 mL culture medium suspended cells were diluted fivefold to be seeded in 96-well plates (each group with six duplicates) at 24, 48, 72, and 96 hours, respectively. 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT; 10 μL, 5 mg/mL, Sigma, St Louis, MO, USA) was added into each well, and cells were incubated at 37°C in a 5% CO₂ incubator and cultured for 4 hours. The cells were removed out and observed using an inverted microscope, and purple crystals were observed. Medium was discarded and added with 100 μL/well dimethylsulfoxide (Sigma) and the violet crystal was dissolved by oscillation. The absorbances of each well at 24, 48, 72, and 96 hours were measured at a wavelength of 490 nm by a microplate reader, and the cell growth curves were analyzed and drawn.

Wound-healing assay

A total of 1×10⁶ cells were seeded into six-well plates (each group with six duplicates) and incubated with mitomycin for 1 hour to inhibit cell division. The six-well plates were scratched with three parallel vertical lines using 200 μL pipettes.¹⁸ The cells were washed three times with phosphate-buffered saline and incubated in serum-free DMEM. Cell motility was evaluated by observing, at intervals of 0, 24,

and 48 hours, the movement of cells into the scratch area using a microscope. The images were recorded, and eight fields of imaging view were measured.

$$\text{Migration rate} = \left(1 - \frac{\text{distance at other time points}}{\text{distance at 0 hour}} \right) \times 100\% \quad (1)$$

Transwell invasion and migration assays

Matrigel was diluted 1:8 with the precooled serum-free DMEM and stored overnight at 4°C. The Transwell chamber coated with Matrigel and the uncoated Transwell chamber (8 μm pore, Corning Inc., Lowell, MA, USA) were used for invasion and migration assays, respectively.¹⁹ Cells were transfected with the desired plasmids, and trypsinized after 48-hour transfection. Each upper chamber was seeded with 1×10⁵ cells suspended in 100 μL of serum-free DMEM. The lower chamber was filled with DMEM supplemented with 10% FBS. After incubation at 37°C for 24 hours, the non-invading cells were gently removed with a soft cotton swab. The cells that invaded to the bottom chamber were fixed with 4% paraformaldehyde for 15 minutes, stained with 0.1% crystal violet, and counted by a light microscope at ×200 magnification. Five random fields were selected in the middle portion and the peripheral portion, and cells passing through the 8 μm pore were counted and calculated, which represented the invasion ability of cells. Cell migration assay procedure was similar to invasion assay, but the incubation lasted for 24 or 48 hours; nonmigratory cells on upper filters were also removed using cotton swab, and cells that had migrated on the underside of filters were fixed, stained, and counted.

Statistical analysis

Statistical analyses were performed using SPSS software (Version 19.0; SPSS). The measurement data are presented as mean ± standard deviation. Least Significant Difference-*t*-test was applied to compare differences. The comparison between groups was performed by applying one-way analysis of variance. All experiments were repeated three times. Differences were considered significant at $P < 0.05$.

Results

Detection results of dual luciferase reporter gene

miRNA target gene software predicted that *Wnt1* may be the target gene of miR-122 (Figure 1A). To confirm that *Wnt1* is a direct target gene of miR-122, luciferase reporter vector reconstructed plasmids, pWnt1-Wt and pWnt1-Mut, that were inserted with *Wnt1* mRNA 3'-untranslated region.

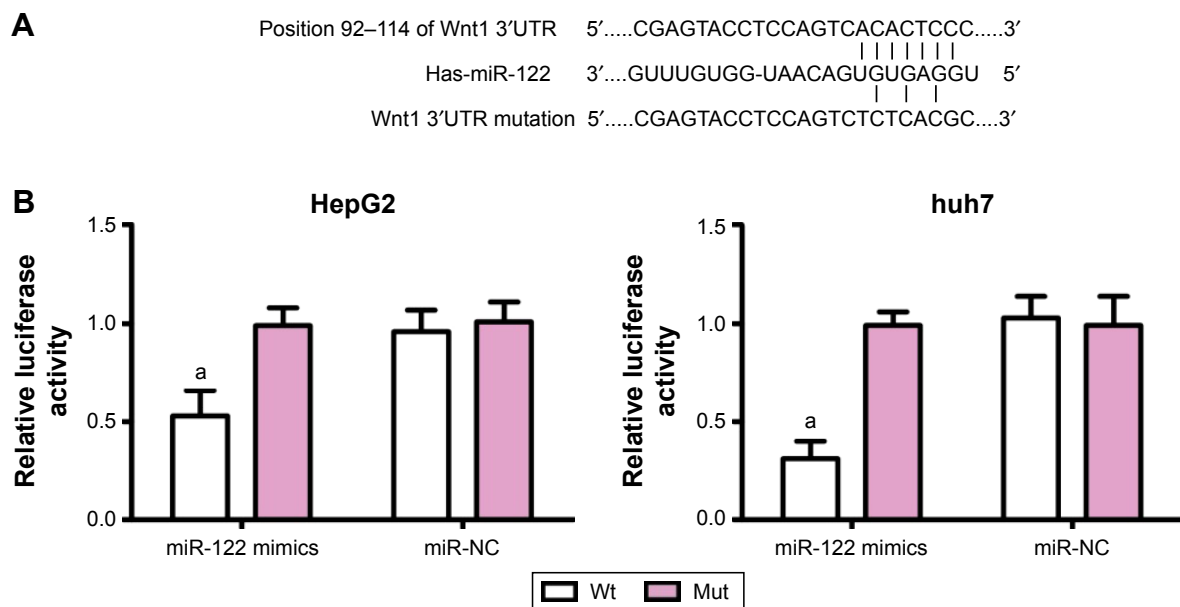


Figure 1 miR-122 targeting *Wnt1*.

Notes: (A) TargetScan predicted that *Wnt1* was a target gene of miR-122; (B) dual-luciferase reporter gene detection confirmed that *Wnt1* was a target gene of miR-122 in HepG2 and huh7 cells. Compared with the miR-122-mimics + pWnt1-Mut, $^*P < 0.001$.

Abbreviations: miR-122, microRNA-122; NC, negative control; Wt, wild-type; Mut, mutated.

Luciferase activity test results showed that in HepG2 and huh7 cells with the cotransfected miR-122 mimics and recombinant plasmids pWnt1-Wt or pWnt1-Mut, miR-122 mimics had no significant effect on the intensity of luciferase activity in the mutant pWnt1-Mut group, but decreased its intensity by approximately 47% in the wild-type pWnt1-Wt group of HepG2 cells and by approximately 69% in huh7 cells, with statistically significant differences (both $P < 0.001$) (Figure 1B); however, miR-NC had no obvious effects on the luciferase activity intensity of pWnt1-Mut and pWnt1-Wt groups in the HepG2 and huh7 cells.

Expressions of miR-122 and mRNA expressions of *Wnt1*, *E-cadherin*, *vimentin*, β -*cadherin*, and *N-cadherin*

The expression of miR-122 in miR-NC, miR-122, anti-miR-NC, anti-miR-122, miR-122 + vector, and miR-122 + Wnt1 groups was 1.03 ± 0.15 , 4.22 ± 0.20 , 1.02 ± 0.16 , 0.41 ± 0.12 , 4.40 ± 0.12 , and 0.97 ± 0.10 , respectively, in HepG2 cells ($F = 459.0$, $P < 0.01$), while it was 1.05 ± 0.13 , 7.09 ± 1.13 , 1.01 ± 0.11 , 0.25 ± 0.03 , 6.85 ± 1.29 , and 0.97 ± 0.10 , respectively, in huh7 cells. There was no obvious difference in miR-122 expression among miR-NC group, anti-miR-NC group, and miR-122 + Wnt1 group (all $P < 0.05$), but miR-122 expressions in miR-122 group and miR-122 + vector group were markedly higher, and the level in anti-miR-122 group was lower than the miR-NC group, anti-miR-NC

group, and miR-122 + Wnt1 group (all $P < 0.05$) (Figure 2). The mRNA levels of *Wnt1*, *E-cadherin*, *vimentin*, β -*cadherin*, and *N-cadherin* were not statistically different among miR-NC group, anti-miR-NC group, and miR-122 + Wnt1 group (all $P < 0.05$); when compared with miR-NC group, anti-miR-NC group, and miR-122 + Wnt1 group, *Wnt1*, *vimentin*, β -*cadherin*, and *N-cadherin*, the mRNA levels were significantly increased and *E-cadherin* mRNA level was decreased in miR-122 group and miR-122 + vector group; however, in anti-miR-122 group, *Wnt1*, *vimentin*, β -*cadherin*, and *N-cadherin*, mRNA levels were upregulated and *E-cadherin* mRNA level was downregulated (all $P < 0.05$), as shown in Figure 3.

Protein levels of *Wnt1*, *E-cadherin*, *vimentin*, β -*cadherin*, and *N-cadherin*

As seen in Figure 4, protein levels of *Wnt1*, *E-cadherin*, *vimentin*, β -*cadherin*, and *N-cadherin* were not different among miR-NC group, anti-miR-NC group, and miR-122 + Wnt1 group (all $P < 0.05$). However, protein levels of *Wnt1*, *vimentin*, β -*cadherin*, and *N-cadherin* in miR-122 group and miR-122 + vector group were significantly decreased, and *E-cadherin* protein level was increased, while the protein levels of *Wnt1*, *vimentin*, β -*cadherin*, and *N-cadherin* were higher and of *E-cadherin* was lower in anti-miR-122 group in comparison to miR-NC group, anti-miR-NC group, and miR-122 + Wnt1 group (all $P < 0.05$).

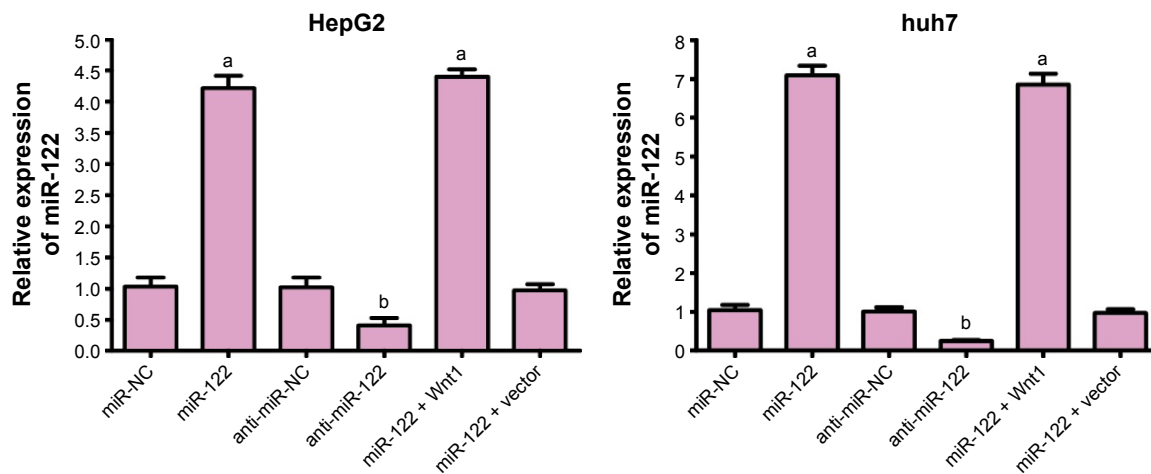


Figure 2 qRT-PCR detecting miR-122 detection levels after transfection in HepG2 and huh7 cells.

Notes: Compared with the miR-NC group, $^*P<0.05$; compared with the anti-miR-NC group, $^bP<0.05$.

Abbreviations: qRT-PCR, quantitative real-time polymerase chain reaction; miR-122, microRNA-122; NC, negative control.

miR-122 in proliferation of HepG2 and huh7 cells

MTT results showed that compared with the miR-NC, anti-miR-NC, and miR-122 + Wnt1 groups, the cell proliferation rate was obviously slowed down in miR-122 group and miR-122 + vector group of HepG2 and huh7 cells, while it was significantly speeded up in anti-miR-122 group after cells were seeded for 48 hours (all $P<0.05$). The cell proliferation rate showed no statistical significance in the miR-NC group, anti-miR-NC group, and miR-122 + Wnt1 group (all $P>0.05$) (Figure 5).

miR-122 in migration capacity of HepG2 and huh7 cells

Wound-healing assay showed that HepG2 cell migration rate was not different significantly among miR-NC group, anti-miR-NC group, and miR-122 + Wnt1 group (all $P>0.05$). Nevertheless, we found that the migration rate was suppressed in miR-122 group and miR-122 + vector group, and the rate was dramatically augmented in anti-miR-122 group, compared to the miR-NC group, anti-miR-NC group, and miR-122 + Wnt1 group (all $P<0.05$) (Table 3). Transwell migration assay showed that the number of cells migrating through the membranes in miR-NC, miR-122, anti-miR-NC, anti-miR-122, miR-122 + vector, and miR-122 + Wnt1 groups was 197.17 ± 13.69 , 133.44 ± 10.63 , 207.67 ± 12.58 , 270.00 ± 7.66 , 147.31 ± 17.6 , and 199.96 ± 10.87 , respectively, in HepG2 cells ($F=35.58$, $P<0.01$) (Figure 6) and was 237.33 ± 17.65 , 143.69 ± 13.76 , 229.38 ± 14.30 , 299.31 ± 10.56 , 151.00 ± 13.35 , and 242.63 ± 12.11 , respectively, in huh7 cells ($F=27.73$, $P<0.001$) (Figure 7). There was no difference in

migrated HepG2 and huh7 cell numbers among the miR-NC group, the anti-miR-NC group, and the miR-122 + Wnt1 group (all $P>0.05$). However, the number of migrated HepG2 and huh7 cells was significantly decreased in miR-122 group and miR-122 + vector group, but increased in anti-miR-122 group (all $P<0.05$).

miR-122 in invasion capacity of HepG2 and huh7 cells

The number of HepG2 cells invading through Matrigel and across the membranes in miR-NC, miR-122, anti-miR-NC, anti-miR-122, miR-122 + vector, and miR-122 + Wnt1 groups was 163.70 ± 16.33 , 108.33 ± 15.08 , 160.33 ± 17.35 , 229.17 ± 15.81 , 105.31 ± 11.21 , and 167.41 ± 13.42 , respectively ($F=55.90$, $P<0.001$) (Figure 8), while the number of huh7 cells invading through Matrigel and across the membranes in miR-NC, miR-122, anti-miR-NC, anti-miR-122, miR-122 + vector, and miR-122 + Wnt1 groups was 209.63 ± 12.87 , 109.76 ± 12.39 , 210.01 ± 11.17 , 355.50 ± 13.11 , 97.01 ± 11.60 , and 203.09 ± 11.55 , respectively ($F=175.6$, $P<0.001$) (Figure 9). There was no obvious difference in invasion capacity among miR-NC group, anti-miR-NC group, and miR-122 + Wnt1 group (all $P>0.05$). However, the invasion capacity was significantly inhibited in miR-122 group and miR-122 + vector group, and the capacity was dramatically promoted in anti-miR-122 group compared to the miR-NC group, anti-miR-NC group, and miR-122 + Wnt1 group (all $P<0.05$).

Discussion

In the present study, we found that the proliferation rate and the migration rate were dramatically augmented in

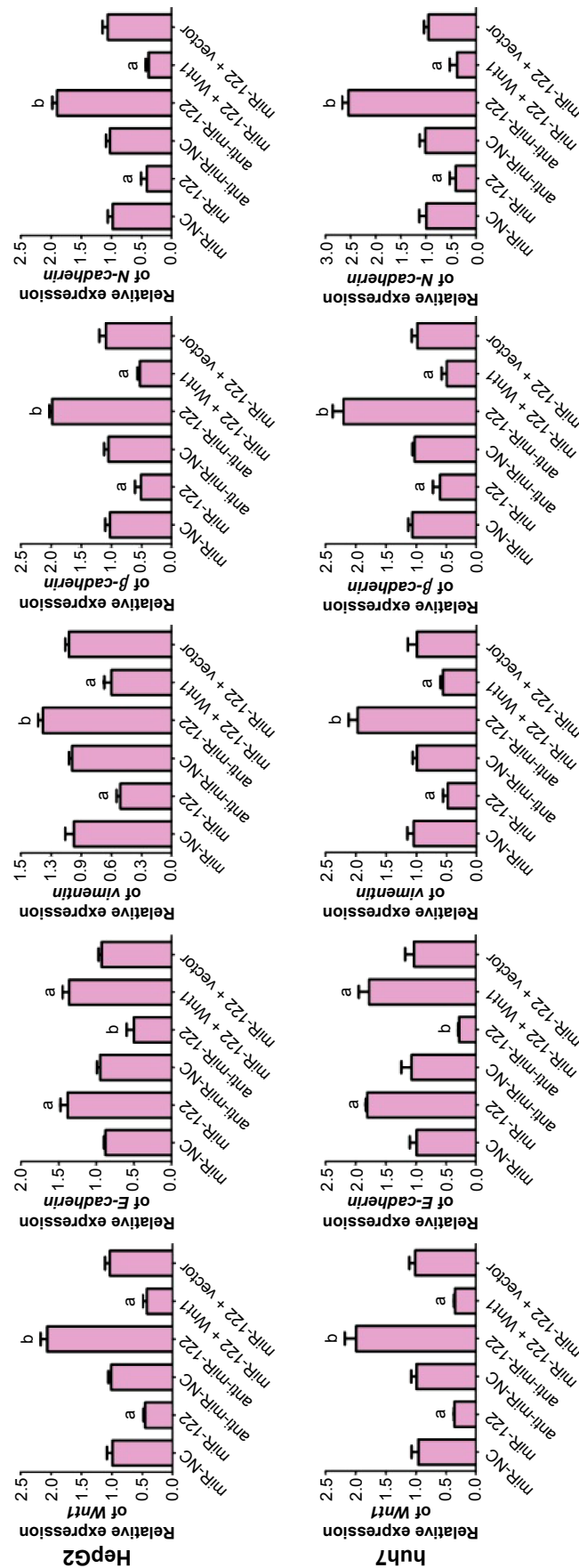


Figure 3 qRT-PCR detecting mRNA expressions of Wnt1- and EMT-related genes.
Notes: Compared with the miR-NC group, * $p < 0.05$; compared with the anti-miR-NC group, $^{#}p < 0.05$.
Abbreviations: qRT-PCR, quantitative real-time polymerase chain reaction; EMT, epithelial-mesenchymal transition; mRNA, messenger RNA; miR-122, microRNA-122; NC, negative control.

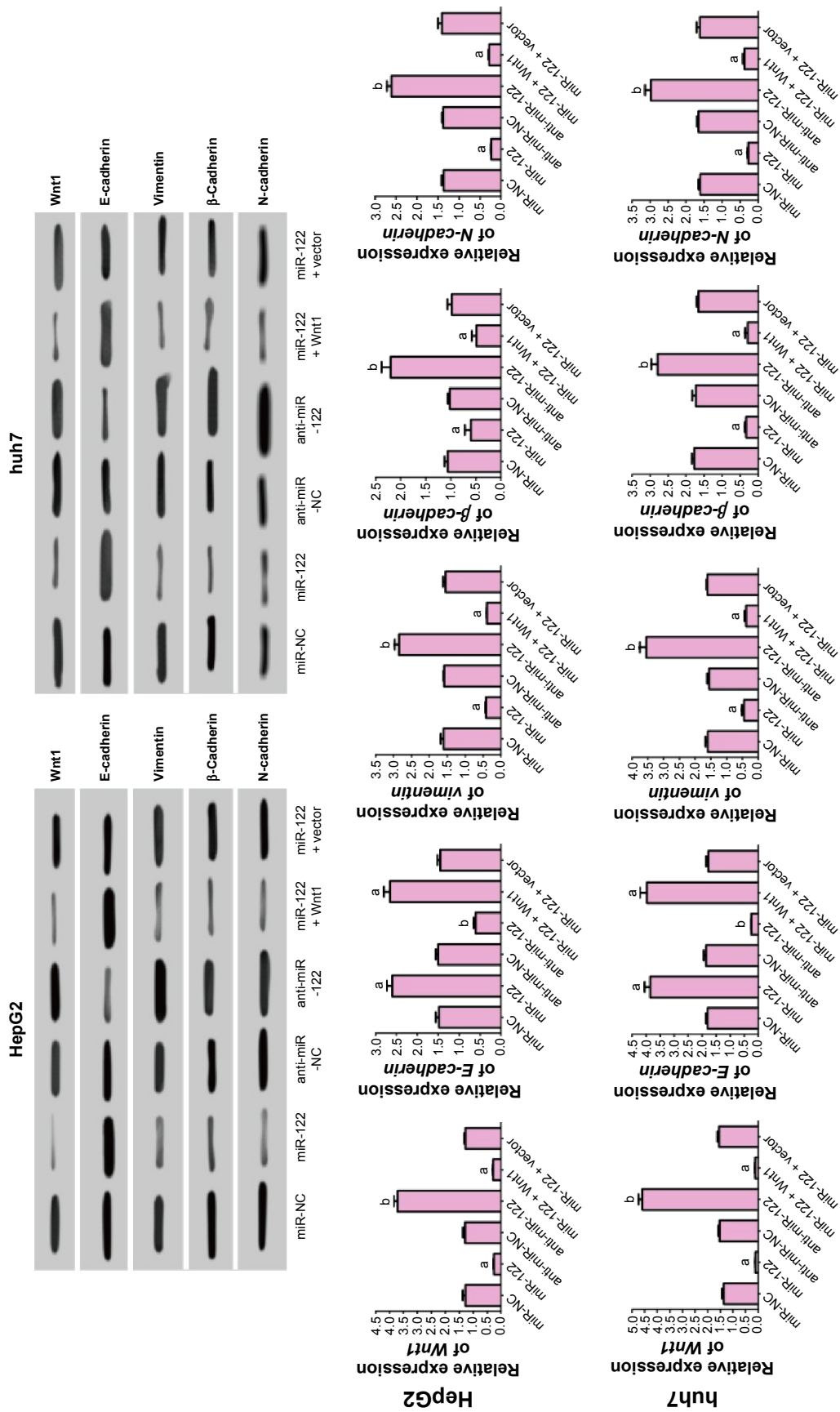


Figure 4 Western blot detecting Wnt1 protein and EMT-associated proteins. **Notes:** Compared with the miR-NC group, * $P < 0.05$; compared with the anti-miR-NC group, [#] $P < 0.05$. **Abbreviations:** EMT, epithelial-mesenchymal transition; miR-122, microRNA-122; NC, negative control.

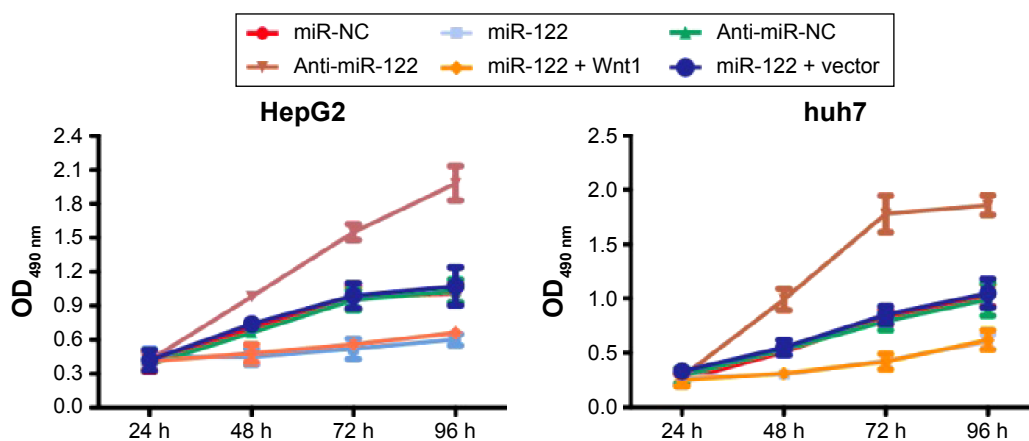


Figure 5 MTT assay detecting the effects of miR-122 on cell proliferation of HepG2 and huh7 cells.

Abbreviations: MTT, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide; miR-122, microRNA-122; h, hours; NC, negative control; OD, optical density.

anti-miR-122 group. Transwell migration assay showed that the number of HepG2 and huh7 cells migrating through the membranes was significantly increased in anti-miR-122 group. Transwell invasion assay showed that the invasion capacity was dramatically promoted in anti-miR-122 group. miR-383 might inhibit HCC cell proliferation partially via downregulating APRIL expression.²⁰ Overexpression of miR-224 has been reported to be related with the malignant phenotype of HepG2 cells and played an important role in regulating the migration and invasion of HepG2 cells.²¹ Besides, it has been shown that miR-1246, by downregulating *CADMI*, a well-defined tumor suppressor gene, enhances migration and invasion in HCC cells.²²

In this study, our main finding demonstrates that downregulation of miR-122 might regulate Wnt/ β -catenin pathway, which is a key step in metastasis and invasion of hepatic carcinoma cells. The role of Wnt/ β -catenin in HCC was widely studied, and it was suggested that a combination treatment of conventional chemotherapy agents and

Wnt/ β -catenin pathway inhibitors could derive effective antitumor response.²³ Some miRNAs including miR-122, miR-221, miR-1, and miR-21 have been investigated for their functions in suppressing cell cycle, cell invasion and metastasis, and cell proliferation and apoptosis.²⁴ In this context, miR-122 is identified as tumor suppressor, inducing apoptosis in HCC via directly targeting Wnt/ β -catenin signaling.²⁵ Our dual luciferase reporter gene activity analysis validated that *Wnt1* is a direct target gene of miR-122. Overexpression of miR-122 is associated with negative expression of *Wnt1* gene in HepG2 and huh7 cells, thereby decreasing the activities of Wnt/ β -catenin pathway by affecting downstream molecules such as E-cadherin, N-cadherin, and vimentin which are related with the EMT process.^{26,27} The trends of EMT-related gene and protein expressions of miR-122 + Wnt1 group were as the same of those in miR-NC group and anti-miR-NC group, which further verified that miR-122 targeted on *Wnt1* to regulate the expressions of EMT-related genes and proteins. During EMT, epithelial

Table 3 Wound-healing assay detecting cell migration rate in HepG2 and huh7 cells at different time points

Groups	Migration rate (%)			
	HepG2 cells		huh7 cells	
	24 hours	48 hours	24 hours	48 hours
miR-NC	17.10±2.01	24.33±3.20	10.11±1.02	29.36±3.00
miR-122	6.21±0.23 ^a	14.10±1.65 ^a	4.09±0.21 ^a	16.70±1.96 ^a
Anti-miR-NC	17.22±3.17	25.01±2.99	9.99±1.13	28.97±5.31
Anti-miR-122	30.96±7.03 ^b	40.44±6.15 ^b	26.87±4.33 ^b	47.65±6.11 ^b
miR-122 + vector	4.96±0.11	13.15±1.33	3.98±0.33	15.43±1.11
miR-122 + Wnt1	17.31±2.96 ^a	24.56±2.27 ^a	11.00±0.98 ^a	30.01±4.11 ^a
F	22.15	26.28	56.92	25.32
P	<0.001	<0.001	<0.001	<0.001

Notes: Compared with the miR-NC group, ^aP<0.05; compared with the anti-miR-NC group, ^bP<0.05.

Abbreviations: miR-122, microRNA-122; NC, negative control.

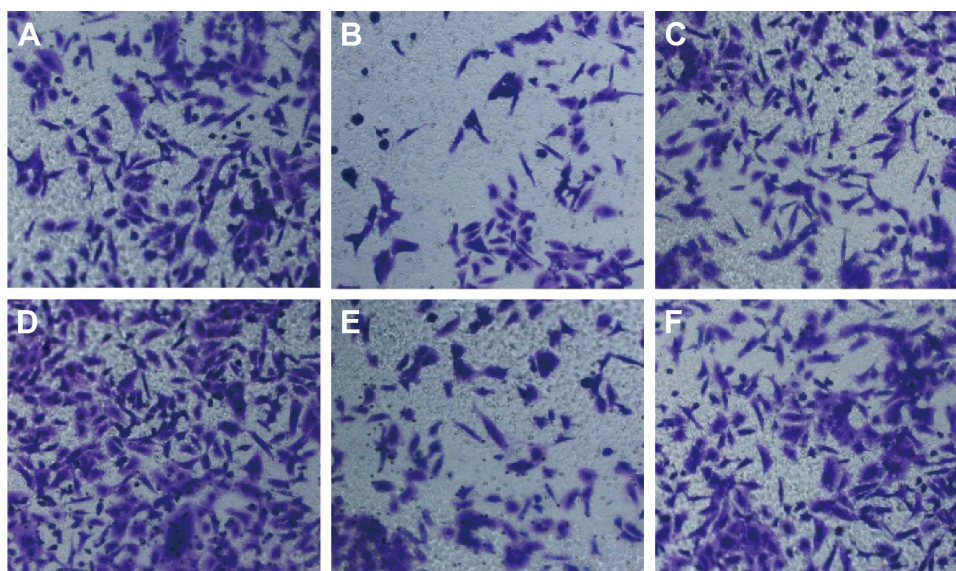


Figure 6 Transwell assay detecting migration capacity of HepG2 cells (A: miR-NC group; B: miR-122 group; C: anti-miR-NC group; D: anti-miR-122 group; E: miR-122 + vector group; F: miR-122 + Wnt1 group).

Note: Magnification was $\times 200$.

Abbreviations: miR-122, microRNA-122; NC, negative control.

markers are lost and mesenchyme-like phenotypes are gained, interfering with intercellular adhesions and promoting fibroblastic properties, which play important roles in the metastatic and invasive behavior of tumors.²⁸ The complex process of EMT is considered a pivotal step permitting metastasis and invasion in various cancers.^{29,30} EMT is accompanied by loss of cell adhesion proteins such as E-cadherin and gain expression of mesenchymal markers such as N-cadherin, β -cadherin, and vimentin.³¹ E-cadherin

is expressed in most epithelial cells, and downregulation of E-cadherin is a hallmark of EMT process.³² Several pathways exist in various tumors to disrupt E-cadherin adhesion junctions. For example, E-cadherin expression is directly suppressed by transcriptional repressors such as Snail, ZEB1, and ZEB2 or through indirect repressors such as Twist, Goosecoid, FoxC2, and E2.2, which indirectly reduce E-cadherin transcription.^{33,34} The cadherin/cadherin-based adhesion system affects EMT process, and Wnt/ β -catenin

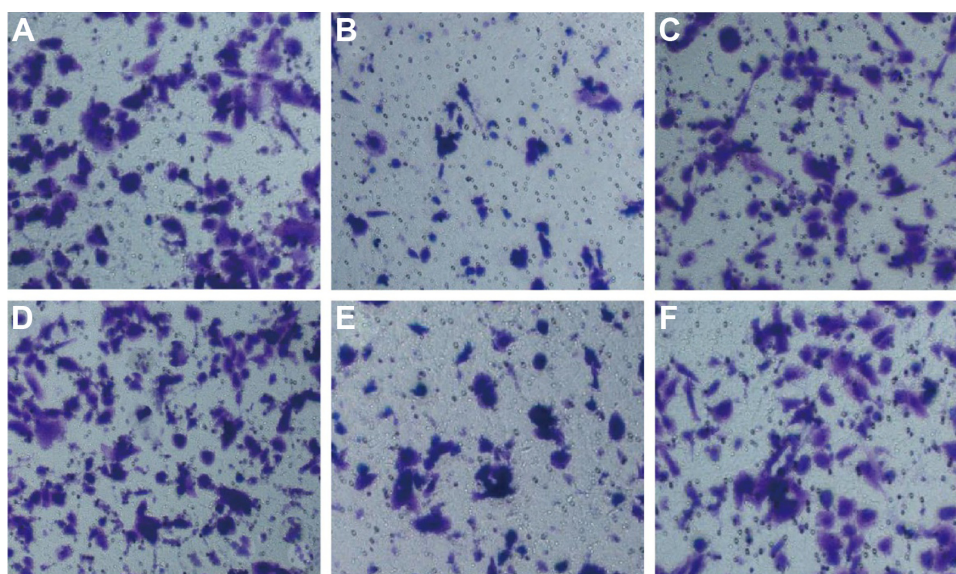


Figure 7 Transwell assay detecting migration capacity of huh7 cells (A: miR-NC group; B: miR-122 group; C: anti-miR-NC group; D: anti-miR-122 group; E: miR-122 + vector group; F: miR-122 + Wnt1 group).

Note: Magnification was $\times 200$.

Abbreviations: miR-122, microRNA-122; NC, negative control.

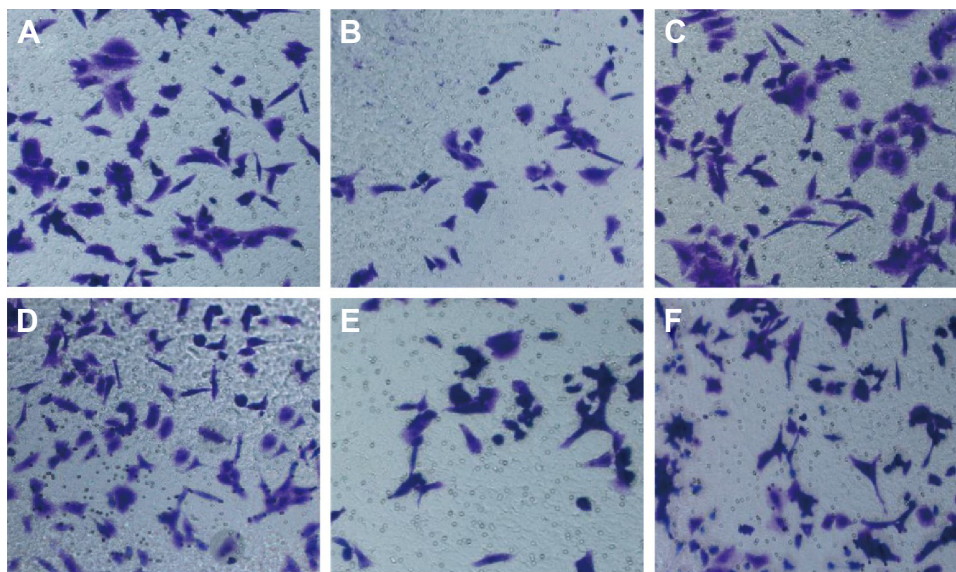


Figure 8 Transwell assay detecting invasion capacity of HepG2 cells (A: miR-NC group; B: miR-122 group; C: anti-miR-NC group; D: anti-miR-122 group; E: miR-122 + vector group; F: miR-122 + Wnt1 group).

Note: Magnification was $\times 200$.

Abbreviations: miR-122, microRNA-122; NC, negative control.

activity is capable of releasing β -cadherin to disassemble adherens junctions.³⁵ The loss of Wnt1 and β -cadherin is related to the loss of E-cadherin and subsequent induction of EMT, but suppression of Wnt/ β -catenin can stimulate epithelial differentiation and delay EMT.^{36,37} Translocation of Wnt/ β -catenin into the nucleus acts as critical regulator in tumor cell survival and proliferation.^{38,39} Initiation of aberrant β -catenin and E-cadherin signaling contributes to the

early steps of the EMT process to promote tumorigenesis and metastasis.⁴⁰ In line with our finding, Wang et al⁴¹ found that miR-122 triggers MET activation and suppresses HCC motility and invasion by targeting RhoA.

Taken together, we provide evidence that downregulation of miR-122 plays a key role in proliferation, migration, and invasion of HepG2 and huh7 cells by regulating Wnt/ β -catenin pathway, which activated the EMT pathway.

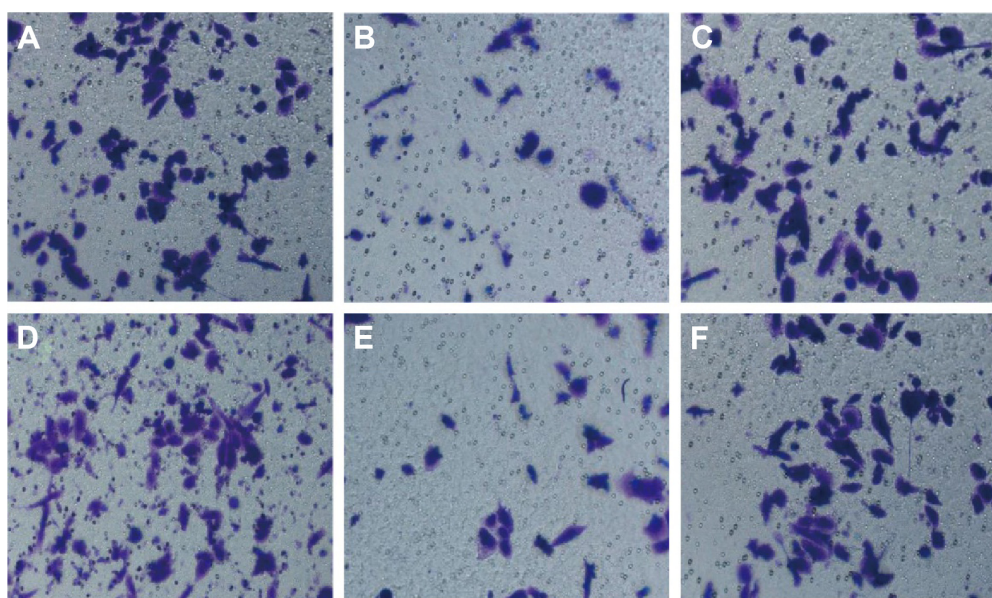


Figure 9 Transwell assay detecting invasion capacity of huh7 cells (A: miR-NC group; B: miR-122 group; C: anti-miR-NC group; D: anti-miR-122 group; E: miR-122 + vector group; F: miR-122 + Wnt1 group).

Note: Magnification was $\times 200$.

Abbreviations: miR-122, microRNA-122; NC, negative control.

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Disclosure

The authors report no conflicts of interest in this work.

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