

Mesenchymal Stem Cell–Derived Exosomes in Various Chronic Liver Diseases: Hype or Hope?

Lujian Zhu^{1,*}, Qin Wang^{1,*}, Maodong Guo², Hao Fang³, Ting Li⁴, Yin Zhu⁵, Huimian Jiang⁶, Peiguang Xiao², Minli Hu²

¹Department of Infectious Diseases, Affiliated Jinhua Hospital, Zhejiang University School of Medicine, Jinhua, People's Republic of China; ²Department of Gastroenterology, Affiliated Jinhua Hospital, Zhejiang University School of Medicine, Jinhua, People's Republic of China; ³Department of Traumatology, Affiliated Jinhua Hospital, Zhejiang University School of Medicine, Jinhua, People's Republic of China; ⁴Department of Emergency Medicine, the First Affiliated Hospital, Zhejiang University School of Medicine, Hangzhou, People's Republic of China; ⁵Department of Infectious Diseases, Taizhou Enze Medical Center (Group), Enze Hospital, Taizhou, People's Republic of China; ⁶Department of Infectious Diseases, the First Affiliated Hospital of Ningbo University, Ningbo, People's Republic of China

*These authors contributed equally to this work

Correspondence: Minli Hu, Department of Gastroenterology, Affiliated Jinhua Hospital, Zhejiang University School of Medicine, Jinhua, People's Republic of China, Email huminlignmc@126.com

Abstract: Chronic liver conditions are associated with high mortality rates and have a large adverse effect on human well-being as well as a significant financial burden. Currently, the only effective treatment available for the effects of liver failure and cirrhosis resulting from the progression of several chronic liver diseases is liver transplantation carried out at the original location. This implies that developing novel and effective treatments is imperative. Regenerative medicine has long been associated with stem cell therapy. Mesenchymal stem cells (MSCs), a type of cell with great differentiation potential, have become the preferred source for stem cell therapy. According to recent studies, MSCs' paracrine products—rather than their capacity for differentiation—play a significant therapeutic effect. MSC exosomes, a type of extracellular vesicle (MSC-EV), came into view as the paracrine substances of MSCs. According to research, MSC exosomes can maintain tissue homeostasis, which is necessary for healthy tissue function. All tissues contain them, and they take part in a variety of biological activities that support cellular activity and tissue regeneration in order to preserve tissue homeostasis. The outcomes support the use of MSCs and the exosomes they produce as a therapeutic option for a range of diseases. This review provides a brief overview of the source of MSC-EVs and outlines their physiological roles and biochemical capabilities. The elucidation of the role of MSC-EVs in the recovery and repair of hepatic tissues, as well as their contribution to maintaining tissue homeostasis, is discussed in relation to different chronic liver diseases. This review aims to provide new insights into the unique roles that MSC-EVs play in the treatment of chronic liver diseases.

Keywords: mesenchymal stem cells, exosomes, liver disease, immunomodulation, tissue homeostasis

Introduction

Numerous pathogenic factors, such as viruses, chemicals, and autoimmune reactions, and the excessive consumption of alcohol can result in both acute and chronic liver damage, which leads to inflammation.¹ The liver cannot replace damaged liver cells, which leads to the development of serious problems like hepatic encephalopathy and hepatorenal syndrome, as well as symptoms including jaundice and poor blood coagulation.² This, in turn, can result in cirrhosis, liver failure, and, potentially, liver cancer, with a mortality rate ranging from 50% to 90%.³ Current therapies for many liver diseases, however, are limited. In the case of advanced liver disease, especially cirrhosis and liver failure, the only possible treatment is a liver transplant performed in the patient's body.⁴

However, the limited availability of liver donors, exorbitant expenses, complications after surgery, and the risk of organ rejection are significant deterrents to transplantation.⁵ As a result, researchers from all around the world have been exploring alternate types of therapy that can reverse fibrosis, decrease liver inflammation and necrosis, and promote the regeneration of liver cells.^{6,7}

MSCs are found in various tissues and are not restricted to mesodermal organs such as the bone marrow, adipose tissue, muscle, or bone.^{8,9} They can also be extracted from the brain, spleen, liver, kidneys, lungs, thymus, and pancreas.^{10–12} Owing to their remarkable ability to regenerate, in recent times, MSCs have undergone clinical testing for a broader spectrum of disease indications,¹³ and the utilization of MSCs in clinical trials has seen tremendous growth.^{14,15} MSCs are easily extracted from adult tissues and have a strong *in vitro* expansion capacity.^{16,17}

Despite being found in a variety of tissues, MSCs share comparable phenotypic characteristics, and some may even have extra features that are indicative of the source tissues.^{17–19} MSCs can transform into a minimum of three different cell types, namely, adipocytes, chondrocytes, and osteocytes.^{19,20} Because MSCs are able to distinguish, their use in clinical trials has expanded substantially.²¹ MSCs are now well recognized as a type of precursor cells that support the stroma and have the ability to differentiate into stroma-supporting cells. They also produce pro-stromal factors and diverse cell growth factors.^{22–25}

MSCs²⁶ play a role in maintaining the hematopoietic stem cell balance within the matrix and also contribute to the maintenance of microenvironmental homeostasis.²⁷ Self-renewal and differentiation maintain equilibrium between vital and static hematopoietic stem cells within this microenvironment.²⁸ MSCs are becoming increasingly important in the study of liver diseases due to their capacity for self-renewal and multi-directional differentiation. These cells can be obtained from different tissues, including bone marrow, umbilical cord, and adipose tissue. In addition, they have immunomodulatory, apoptosis-inhibition, cell-regeneration, and anti-fibrotic properties.^{13,19,29–33} The liver, being a crucial organ in regulating immunity and metabolism, contains numerous immune cells, including myeloid and lymphoid cells. Disruption of hepatic immune homeostasis is often linked to various liver diseases.^{34,35} Recent clinical investigations have shown that therapies utilizing MSCs can reduce liver damage, improve liver function, and promote the regeneration of liver tissue.^{36,37}

However, with the growing use and study of MSCs, concerns have emerged regarding their theoretical basis related to their ability to differentiate. Multiple studies have shown that although MSC therapy leads to the restoration of function, only a small number of cells achieve differentiation into appropriate tissues after MSC transplantation.^{38,39} Moreover, the therapeutic effect of transplanted MSCs does not depend on the closeness of the transplantation location to the injury site.⁴⁰ Currently, there is a prevailing opinion that the immunomodulatory function of MSCs is predominantly accomplished through the paracrine pathway. This pathway leads to the reduction of damage and promotes the process of healing.^{41,42} Extracellular vesicles (EVs) are released by phospholipid bilayer-containing cells and play a crucial role in facilitating cell-to-cell communication by carrying membrane and cytoplasmic proteins, lipids, and RNA.^{24,40,43–45} MSC-derived EVs (MSC-EVs) are significantly smaller and more readily obtainable and storable compared to MSCs.⁴⁶ Following *in vivo* intravenous administration, MSC-EVs primarily accumulate in the liver. The occurrence of liver diseases is intimately associated with an imbalance in immunological homeostasis, which is closely related to the function of MSC-EVs.⁴⁷ Hence, researchers are investigating the utilization of MSC-EVs as a substitute for MSCs in liver ailments.⁴⁸ This review examines the advancements in the utilization of MSC-EVs in treating various chronic liver ailments.

Overview of MSCs

MSCs are pluripotent cells present in nearly all types of postnatal organs and tissues.⁴⁹ Moreover, MSCs possess a robust capacity for migration.¹¹ MSCs possess strong immunomodulatory properties.⁵⁰ MSCs are being increasingly utilized in clinical trials that focus on tissue repair and regeneration owing to their immunomodulatory and cell survival-enhancing characteristics.⁵¹ Nevertheless, in contrast to expectations, numerous preclinical investigations have reported that MSCs do not integrate effectively into tissues in significant quantities, and the duration of integration is inadequate.¹²

MSCs possess strong immunomodulatory properties, effectively suppressing T cells, B cells, and natural killer (NK) cells in addition to their ability to differentiate in multiple directions.⁵² Presently, there is substantial evidence backing the therapeutic and immunomodulatory roles of MSCs, primarily relying on paracrine effects and the release of secretory components. Therefore, many scholars are showing interest in the paracrine functions of MSCs, focusing specifically on paracrine secretions such as EVs.^{53,54} According to their findings, MSCs are capable of producing and releasing numerous growth factors, chemokines, and cytokines, which, in turn, have an impact on the neighboring cells.²⁰ In 2005, Gnechhi et al proposed that MSCs can release protective substances.⁵⁵ They intramyocardially injected the culture supernatants from MSCs with high Akt gene expression, as well as MSCs alone, into an animal model of acute infarction. Both animal groups showed

a decrease in the myocardial infarction area to a similar extent, thereby validating their hypothesis.⁵⁶ Furthermore, it has been reported that MSCs enhance microcirculation and produce various anti-apoptotic proteins, including Bcl-2 and Akt, thereby averting cell death.^{57,58} Numerous studies have confirmed the likelihood of the hypothesis that the healing benefits of MSCs are attributable to their paracrine effects rather than their differentiation effects.

Recently, studies have shown that under appropriate conditions, MSCs can be differentiated into hepatocyte-like cells (HLCs), providing new ideas for the treatment of liver diseases. Liver-specific miR-122 can be efficiently transfected into MSCs and activate their differentiation into HLCs. Transplantation of these engineered MSCs can treat acute liver failure by replenishing hepatocyte function.⁵⁹ Various liver diseases can lead to hepatic failure, and obtaining a sufficient number of functional hepatocytes is the key to liver regeneration.⁶⁰ Since the functions of primary hepatocytes are unstable, directing the differentiation of MSCs and other stem cells into HLCs, as well as developing and utilizing more stable and safer extracellular vesicle products, are the directions that researchers are paying attention to. These studies have provided a theoretical basis for the application of MSCs in the treatment of liver diseases.

Overview of MSC-EVs

Research on MSCs' release of extracellular vesicles has advanced significantly to date, and it has been shown that MSCs secrete a variety of extracellular vesicles, including exosomes, microvesicles, and microparticles.⁶¹ Studies indicate that MSC-EVs have a significant impact on the transfer and regulation of intercellular information.^{47,62} Scientists initially conducted thorough investigations into the mechanism of MSC-EVs generation. The study determined that the generation of MSC-EVs predominantly takes place through the endoplasmic reticulum and multivesicular body pathways.⁶³ The endoplasmic reticulum pathway involves the formation of vesicles on the endoplasmic reticulum, which bind to membrane transport proteins (eg, MVB) via fusion and are ultimately released outside the cell.^{64,65} The multivesicular body pathway includes the discharge of MSC-EVs by shedding vesicles directly from the cellular membrane in reaction to the equilibrium of the plasma membrane.⁶⁶ Gaining knowledge about these pathways provides a basis for further investigation into the biological functions of MSC-EVs.

In addition, a comprehensive examination of the components of MSC-EVs was carried out. A multitude of bioactive constituents, including proteins, nucleic acids, and lipids, have been identified inside MSC-EVs.⁴⁸ Proteomics analysis revealed that MSC-EVs contain a diverse range of operational proteins, such as cell adhesion agents, enzymes that degrade the extracellular matrix, and factors that modulate the immune system.⁶⁷ Furthermore, MSC-EVs contain miRNAs, mRNAs, and various nucleic acids in abundance, which have the potential to influence gene expression and cellular functions when transferred to recipient cells.⁶⁸ MSC-EVs demonstrate various biological capabilities and potential uses in medical practice.⁶⁹ Based on empirical evidence, MSC-EVs have been demonstrated to exhibit anti-inflammatory, anti-fibrotic, angiogenesis-promoting, and immune-modulating properties.⁷⁰

Furthermore, extensive research has been conducted on the use of MSC-EVs to manage diverse ailments, including cardiovascular disorders, neurological conditions, and immune-related diseases.^{71–73} The potential applications of MSC-EVs in disease treatment and tissue regeneration are being investigated through preclinical and clinical trials, which have shown promising results.

Currently, exosomes are the most characterized extracellular vesicles as they have more biologically and biochemically defined parameters that can be detected in conventional laboratories.⁷⁴ Exosomes range from 40–100 nm in size, have a density of 1.10–1.18 g/mL on a sucrose gradient, and are linked to marker proteins such as Alix and Tsg101, as well as tetrameric transmembrane proteins including CD9, CD63, and CD81.^{75,76} Exosomes, derived from introns,⁷⁶ are the sole extracellular vesicles that are currently recognized. Introns form vesicles with several layers and a large number of luminal vesicles by invaginating the endosomal membrane.⁷⁶ When they fuse with the cell membrane, luminal vesicles are released to form exosomes.⁷⁷ This distinguishes the process of exosome formation from that of other extracellular vesicle types. Through extensive examination of MSC-EVs, it has been established that these minuscule extracellular vesicles have a significant impact on the communication and regulation between cells.⁷⁶ Additional investigation will reveal the precise molecular mechanism of MSC-EVs, improve their composition, and broaden their potential uses in a therapeutic context. This has the potential to provide innovative strategies and techniques for enhancing the management of chronic liver disease and promoting tissue regeneration.⁷⁸

Functions of MSC-EVs

MSC-EVs have received extensive research attention as a novel extracellular messaging medium.⁷⁹ The primary role of exosomes is to transmit cellular elements from secretory cells to receptor cells in one direction, thereby controlling the activity of the latter.⁸⁰ Similarly, MSC-EVs are responsible for intercellular communication.⁸¹ Exosomes derived from MSCs can control the activity and role of immune cells through the transportation of diverse regulatory proteins, miRNAs, and other biologically active substances.⁸² Clinical investigations demonstrate that MSC-EVs can perform an immunomodulatory effect by suppressing T cell activation, regulating macrophage polarization, and modifying B cell activity.⁸³ Treatment of autoimmune liver diseases⁸⁴ relies on this immunomodulatory property.

Furthermore, MSC-EVs can secrete bioactive compounds that consist of anti-inflammatory proteins and miRNAs, thereby controlling inflammatory reactions and preventing the release of inflammatory mediators.⁸⁵ Studies show that MSC-EVs can mitigate the inflammatory pathological process, thereby mitigating the progression of inflammatory liver condition.⁸⁶ Moreover, MSC-EVs can transport numerous biologically active substances that enhance the growth of cells and the formation of new blood vessels, ultimately aiding in tissue healing and renewal.⁸⁷ Several studies have indicated the role of MSC-EVs in the regeneration of cardiovascular, neural, hepatic, skeletal, and various other tissues.^{84,88–90} Tissue regeneration function could be a novel approach to treating liver tissue injury. In addition, MSC-EVs, as a natural nanoparticle, have potential applications in drug delivery.⁹¹ Researchers have modified the composition and internal structure of exosomes to enable the precise delivery of drugs to specific cells and tissues.⁹² Targeted therapy and personalized medicine may be based on this drug delivery mechanism to treat liver diseases. MSC-EVs generally have a wide range of activities, such as immunomodulation, inflammation reduction, tissue regeneration promotion, and drug delivery facilitation^{79,91,93,94} (Figure 1). More studies on MSC-EVs can lead to a better

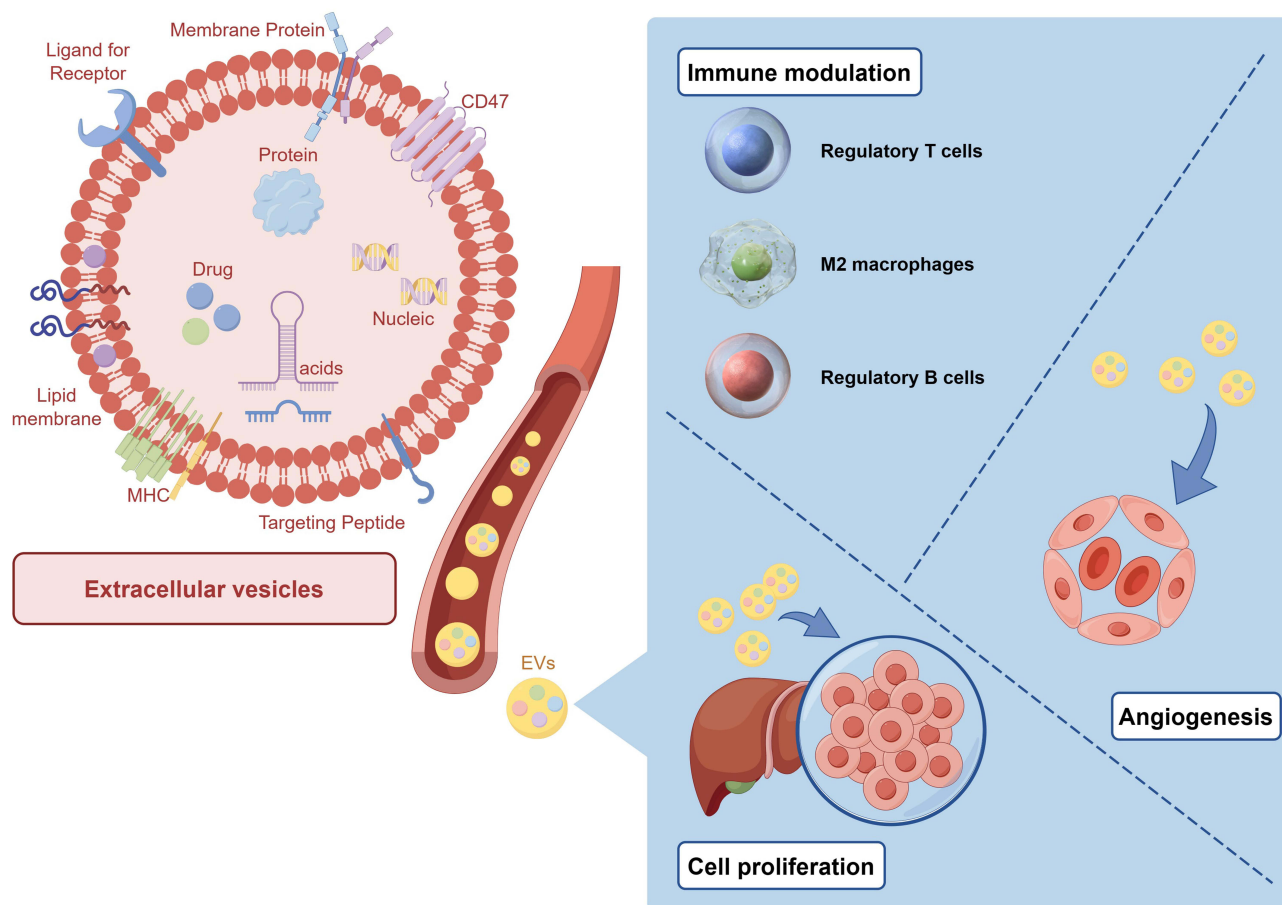


Figure 1 Functions of MSC-EVs. MSC-EVs can transport a variety of bioactive substances, promote cell growth and the formation of new blood vessels, and control the activity and role of immune cells. In addition, MSC-EV, as a natural nanoparticle, has potential applications in drug delivery.

Note: This figure is originally drawn by Figdraw platform (www.figdraw.com).

understanding of their molecular mechanisms, which will facilitate the development of more effective clinical implementation strategies and provide new therapeutic prospects for the treatment of disease and tissue regeneration.

MSC-EVs in the Treatment of Liver Diseases

Lately, a growing body of research has demonstrated the profound promise of MSC-EVs made from MSCs as a medicinal strategy for treating liver conditions.⁹⁵ In the human body, the liver is the primary organ responsible for metabolism and immune function.⁹⁶ The majority of the blood that enters the liver is from the portal vein instead of the hepatic artery.⁹⁷ The liver parenchyma receives blood primarily through the periportal vessels and is then drained from the liver parenchyma by the central hepatic vein through the intricate system of hepatic sinusoids.⁹⁸ Hepatic sinusoids serve as anatomical locations that maintain immune homeostasis.⁹⁹ Lymphocyte extravasation is promoted by the prolonged contact between lymphocytes and antigen-presenting cells due to the sluggish circulation in the hepatic sinusoids.¹⁰⁰ Besides hepatocytes and cholangiocytes, there are nonparenchymal cells such as hepatic stellate cells (HSCs) and hepatic sinusoidal endothelial cells, as well as numerous immune cells such as T cells, macrophages, and dendritic cells in the liver.^{101,102} Liver diseases are typically linked to an imbalance in immune equilibrium, and this immune control includes the varied functionality of macrophages and dendritic cells, the proportion of various T-cell subsets (such as T helper [Th]17 and regulatory T [Treg] cells), the equilibrium between inflammatory and anti-inflammatory cytokines, and other factors related to immunity, which align perfectly with the role of MSC-EVs.^{99,103,104} MSC-EVs primarily accumulate in the liver when administered intravenously in vivo.⁶⁸ Through immunomodulation, the regulation of MSC-EVs can lead to a decrease in the release of interleukin (IL)-6 and IL-1 β by macrophages, and a decrease in the expression of CD154 by CD4⁺ T cells. This reduction in cytokine release and CD154 expression could potentially alleviate hepatic inflammation and provide relief in various liver diseases.^{63,69,105}

Role of MSC-EVs in Liver Diseases

This section provides an overview of preclinical investigations on MSC-EVs as a treatment modality for liver diseases such as liver failure, steatohepatitis linked to metabolism, autoimmune hepatitis (AIH), hepatic fibrosis, and hepatic ischemia reperfusion injury (HIRI). [Table 1](#) summarizes the specific role of MSC-EVs in different liver diseases. The emphasis is on understanding the molecular mechanisms and regulation of MSC-EVs, as discussed in relevant studies.^{48,106,107}

MSC-EVs and Acute Liver Injury

The two main pathophysiological features of liver failure that impact the organism as a whole and the liver specifically are immunological dysfunction and inflammation.¹³⁸ The rapid occurrence of apoptosis and necrosis in a large number of liver cells due to severe immune damage is followed by the development of ischemia, hypoxia, and endotoxemia. This sequence of events drives and speeds up the advancement of liver failure, commonly referred to as the “triple whammy” theory of liver failure.^{138–140} Membrane permeability increases in the injured hepatocytes in the early stages of liver injury. This results in the release of molecules linked to damage-associated intracellular patterns. These molecules then bind to Toll-like receptors, triggering the activation of downstream signaling pathways such as c-jun amino terminal kinase, mitogen-activated protein kinase (MAPK), nuclear factor- κ B (NF- κ B), and signal transducer and activator of transcription 3.

Consequently, inflammatory cells are induced to infiltrate the affected area, further exacerbating liver failure through the promotion of tumor necrosis factor (TNF)- α , IL-1 β , IL-6, IL-8, and other inflammatory factors, as well as cascade reactions.¹⁴¹ Exosomes derived from various MSCs have been used recently by researchers to treat various liver failure models, proving that they can protect against liver damage.¹⁴² Yan et al¹⁴³ administered human umbilical cord MSC-derived exosomes (ucMSC-Ex) at low (8 mg/kg), medium (16 mg/kg), and high (32 mg/kg) doses to mice with carbon tetrachloride (CCl₄)-induced liver failure. Large areas of steatosis and hepatocellular necrosis were found in the liver tissues of mice administered the low dose, whereas the liver tissue lesions in mice in the medium- and high-dose groups were significantly alleviated. EVs promoted recovery from hepatic oxidative injury by delivering GPX1. Besides, several animal experiments have confirmed that the protective role of MSC-Ex in various models of liver failure included inhibition of inflammation, anti-apoptosis effects, and attenuation of oxidative stress.¹⁴⁴

Table 1 Is a Summary of the Specific Role of MSC-EVs in Different Diseases

Disease	Species Sex	Damage	Cell Source	Diameter (nm)	EV Treatment Group (Method/Dose)	Therapy Time	Therapy Mechanism	Ref.
AIH	C57BL/6 mice male	S100	mBMSC	30–100	i.p./20 µg/mL	Day 21/28/35 after injure	Regulate NLRP3 and caspase-1 by miR-223	[108]
AIH	C57BL/6 mice male	S100	Mouce-BMSC	40–100	i.v./2 µg/g (200 µL)	Day 21/35 after injure	Regulate macrophages by miR-223-3p	[109]
AIH	C57B6 mice male	Con-A induce AIH	Mouse-BMSC	135	i.v./10 µg (0.1 mL)	Once and three times after injure	Anti-apoptosis, increase Ki-67 d and Treg	[110]
AIH	BALB/c mice	Con-A induce AIH	Mouse-BMSC	120	i.v./5 mg/kg (100 µL)	Single after injure	Anti-inflammation	[111]
ALF	C57BL/6 mice male	D-GalN/LPS	MenSC	30–100	i.v./1 µg/µL	Single before injure	Anti-apoptosis	[112]
ALF	C57Bl/6 mice male	TNF- α /D-GalN	hBMSC/mouse-BMSC	116 \pm 46	i.p./2 \times 1010 particles	Single after injure	Anti-apoptosis	[113]
ALF	C57BL/6 mice male	LPS/D-GalN	hUCMSC	100	i.v./100 µg (250 µL)	Single 1 h after injure	Anti-NLRP3 inflammasome	[114]
ALF	C57BL/6j mice	LPS/D-GalN or TNF- α /D-GalN	Mouse-ADSC	40–100	i.v./400 µg (300 µL)	Single after injure	Anti-TXNIP/NLRP3 inflammasome	[115]
ALF	C57BL/6 mice male	CCL4	hESC	55–65	i.s./0.4 µg (100 µL)	24 h after injure	Activate proliferation and regeneration, anti-apoptosis	[116]
ALF	C57BL/6 mice male	LPS + D-GalN	hUCMSC	30–150	i.v./100 mg	1 h after injure	Anti-NLRP3	[117]
I/RI	Wistar rats male	IRI or CCL4	rat-BMSC	165 \pm 3	i.v./50 µg	Single after injure	Antioxidant	[118]
I/RI	C57BL/6 mice male	I/RI	Mouse-BMSC	115 \pm 48	i.v./2 \times 1010 particles	Single before injure	Anti-apoptosis, anti-inflammation	[119]
I/RI	Sprague-Dawley rats male	I/RI	hUCMSC	178 \pm 64	i.v./10 mg/kg	Single after injure	Antioxidant, anti-neutrophil inflammatory response	[120]
I/RI	C57BL/6 mice Male	THS	Mouse-BMSC	90–142	Femoral artery/20 µg	Single after resuscitation	Regulate Kupff cells by IL-10	[121]
I/RI	C57BL/6	I/RI	hUCMSC	0–200	i.v./2.5 \times 1012	Single after injure	Regulate GSK3 β /Wnt/ β -catenin pathway by miR-1246	[122]
I/RI	C57BL/6 mice male	I/RI	hUCMSC	30–150	i.v./100 µg/100 µL	Single after injure	Regulate CD4+ T cells by Ca2+-calcineurin-NFAT1 signaling pathway	[123]
I/RI	C57BL/6 mice male	I/RI	hAMSC	120–200	i.v./1 \times 109 particles (200µL)	Single after injure	Anti-inflammation, increase Ki67	[124]
Liver fibrosis	FVB.129P2-Abcb4tm1Bor mice male	PSC	hBMSC	45–372	i.p./9.1 \times 109 particles/mL (100 µL)	Once a week for 3 weeks after injure	Reduce granulocytes and T cells, increase VCAM-1	[125]
Liver fibrosis	Swiss albino mice female	CCL4	hADSC/WJMSC	40–120	i.v./250 µg	Single after injure	Anti-inflammatory, anti-fibrosis	[84]
Liver fibrosis	C57BL/6 mice male	TAA	hADSC	94.2 \pm 4.7	i.v./200 µL (1 \times 107; 1 \times 108)	Single after injure	Anti-fibrosis	[126]
Liver fibrosis	C57BL/6 mice Male	CCL4	hTMSC	50–290	i.v./150 mg (100 µg/mL)	Once a week for 3 weeks after injure	Inactivate hedgehog signaling by miR-486	[127]
Liver fibrosis	Kunmingbai strain mice	CCL4	hUCMSC	40–100	Liver directly injected/250 µg (330 µL)	Single after injure	Inhibit EMT	[128]
Liver fibrosis	Wistar rats male	TAA	hES-MSC	190.8 \pm 18	i.p./350 µg (400 µL)	Single after injure	Anti-fibrosis, anti-inflammation, anti-apoptosis, promote regeneration	[129]
Liver fibrosis	Sprague-Dawley rats male	CCL4	hAMSC	80–110	i.v./15 µg/kg and 20 µg/kg (200 µL)	Single after injure	Anti-fibrosis, reduce Kupfer cells	[130]
Liver fibrosis	Sprague-Dawley rats female	CCL4	hBMSC	30–100	i.v./250 mg (500 µL)	Single after injure	Alleviate liver fibrosis through the Wnt/ β -catenin	[131]
Liver fibrosis	Sprague-Dawley albino rats male	CCL4	rat-BMSC	113.7	i.v./80 µg	Single after injure	Anti-fibrosis	[132]

Liver fibrosis	ICR mice Male	CCL4	hESC	120–140	i.v./NA	Twice a week for 4 weeks after injure	Anti-fibrosis by miR-6766-3p	[133]
Liver fibrosis	C57BL/6 mice male	TAA	Mouse-ADSC	117 ± 7	i.v./(1×10^7 particles), or (1×10^8 particles)	Single or three times after injure	Anti-fibrosis	[134]
Liver fibrosis	Sprague-Dawley rats male	CCL4	Mouse-ADSC	30–150	i.v./0.4 µg/µL, 100 µL	Twice a week for 8 weeks	Inhibit CXCL1 by miR-150-5p	[135]
MAFLD		MAFLD	hUCMSC	96	i.v./100 µg (500 µL)	Once a week for 2 months	miR-627-5p inhibit FTO, improve glycolipid metabolism	[136]
MAFLD	C57BL/6 J, female	MAFLD	Mouse-ADSC	95.8 ± 1.2	i.v./100 µg/100 µL	Twice a week starting from the second week of diet	Anti-fibrotic by miR-223-3p/E2F1	[137]

Abbreviations: EV, extracellular vesicle; MSC, mesenchymal stem cell; NA, not available; ALI, acute liver injure; NAFLD, nonalcoholic fatty liver disease; AIH, autoimmune hepatitis; IRI, ischemia-reperfusion injure; PSC, primary biliary cirrhosis; BMSC, bone marrow mesenchymal stem cell; UCMSC, umbilical cord mesenchymal stem cell; ADSC, adipose-derived mesenchymal stem cell; ESC, embryonic stem cell; AMSC, amnion-derived mesenchymal stromal cell; TSC, tonsil-derived mesenchymal stromal cell; MenSC, menstrual blood-derived mesenchymal stem cell; i.p., intraperitoneal; i.s., intrasplenic; i.v., intravenous injection; CCl4:carbon tetrachloride; TAA; thioacetamide; D-GalN/LPS; D-galactosamine (D-GalN) and lipopolysaccharide (LPS); DEN, diethylnitrosamine.

Human endometrial MSC-EVs have been found to alleviate liver function and reduce hepatocyte apoptosis in a mouse model of ALI, potentially by migrating to the injured liver and suppressing inflammatory responses.¹¹² Bone-marrow-derived MSC-EVs could also accumulate in the injured liver tissue and attenuate damage after systemic administration in a lethal mouse model of acute liver failure, with improved survival rates compared to controls.¹¹³ The therapeutic effects were partly mediated by a highly enriched Y RNA fragment in MSC-EVs.

Adipose tissue-derived MSC-EVs (AMSC-Exos) could protect against inflammasome-induced inflammation in acute liver failure by shuttling miR-17 into hepatic macrophages and suppressing NLRP3 activation via downregulating TXNIP.¹¹⁵ This highlights the involvement of exosomal microRNAs as important therapeutic agents. AMSC-Exos may suppress the production of inflammatory cytokines and restore liver function by targeting NLRP3-regulated inflammation. Another study found human umbilical cord MSC-EVs attenuated NLRP3 inflammasome activation and reduced liver injury markers in a mouse model of acute liver failure, likely via dampening inflammatory responses of macrophages.¹¹⁴ The effects were associated with decreased expression of NLRP3, Caspase-1, IL-1 β and IL-6. Furthermore, systemic administration of BMSC-EVs activated regenerative mechanisms and inhibited apoptosis to protect against toxicant-induced acute liver injury in mice.¹¹⁶ The hepatoprotective effects involved upregulation of proliferation and cell cycle-related genes.

In summary, accumulating evidence has characterized the therapeutic potentials of MSC-EVs in various models of acute liver damage, which are exerted through suppression of inflammasome-mediated inflammation responses, activation of hepatocyte regeneration, and inhibition of cell death pathways. Further research is warranted to understand better the responsible bioactive molecules shuttled by MSC-EVs and optimize treatment strategies.

MSC-EVs and Metabolism-Associated Fatty Liver Disease (MAFLD)

MAFLD is a condition where there is an accumulation of fat in liver tissues even in the absence of significant alcohol consumption or related metabolic disorders.¹⁴⁵ Presently, MAFLD poses a major challenge. The latest data suggest that its occurrence is on the rise worldwide, reaching 24% and resulting in a substantial financial burden.¹⁴⁶ A study found that human umbilical cord MSC-EVs containing miR-627-5p could improve insulin tolerance, alleviate liver injury, modulate glucose/lipid metabolism and reduce lipid deposition in an MAFLD rat model.¹³⁶ The effects were associated with miR-627-5p targeting and suppressing Fat Mass and Obesity associated (FTO) gene expression. In vitro, miR-627-5p overexpressing EVs also promoted cell viability, inhibited palmitic acid-induced apoptosis of LO-2 hepatocytes, and regulated a panel of metabolism-related genes. The data support miR-627-5p enriched MSC-EVs as a promising approach to improve MAFLD progression by enhancing metabolic profiles and reducing liver damage.

Another study reported that adipose-derived MSC-EVs delivering anti-fibrotic miR-223-3p could attenuate lipid accumulation and liver fibrosis by targeting E2F1, which plays a key role in hepatic stellate cell activation.¹³⁷ The anti-fibrotic effects were validated in a mouse model of diet-induced MAFLD. This highlights the therapeutic potential of functional MSC-EVs achieved by manipulating exosomal microRNA cargo. MSC-EVs can also regulate lipid metabolism. In addition, some studies show that MSC-EVs can enhance the process of fatty acid oxidation and hinder the synthesis of fatty acids, ultimately leading to decreased fat accumulation.¹⁴⁷

Furthermore, MSC-EVs affect the differentiation and viability of adipocytes and decrease the inflammatory reaction in adipose tissues.¹⁴⁸ The advancement of MAFLD is known to be frequently accompanied by hepatic fibrosis, and MSC-EVs can impede the progression of liver fibrosis.¹³⁷ According to studies, MSC-EVs can regulate the activity of matrix metalloproteinases and reduce the formation and accumulation of collagen in the liver, which decreases the severity of fibrosis.¹⁴⁹

In conclusion, MSC-EVs have emerged as a promising therapeutic agent for MAFLD, with a range of beneficial activities demonstrated in preclinical studies. These activities include resolving metabolic disorders, suppressing inflammation, inhibiting fibrosis progression, and promoting liver regeneration. Further research is required to understand the underlying mechanisms better and facilitate the translation of findings into clinical practice.

MSC-EVs and AIH

AIH is a chronic, progressive liver disease caused by autoimmune disorders. It is characterized by liver inflammation, infiltration of lymphocytes and plasma cells, and elevated ALT, AST, and immunoglobulin G (IgG) levels in the blood.¹⁵⁰ The clinical diagnosis of AIH relies on liver biopsy to evaluate for interfacial hepatitis, rosette nodules, and lymphocytic infiltration.¹⁵¹ The current initial therapy for AIH involves the use of corticosteroids with or without azathioprine.¹⁵² Although the precise origin of AIH is unknown, scientists believe that a mix of environmental variables, genetic susceptibility, and molecular mimicry may be responsible for lowering autoimmune tolerance.^{153,154}

The primary method of treating AIH at the moment is either hormone therapy alone or in conjunction with immunosuppressants. It has significant drawbacks and limitations, such as prolonged treatment duration, adverse responses, insufficient patient adherence, and failure to stop the advancement of liver fibrosis and cirrhosis, even though it may, to some extent, increase patients' chances of survival.¹⁵² Therefore, the treatment of AIH has always been a clinical focus owing to these challenges.

MSC-EVs has emerged as a promising cell-free therapy for autoimmune hepatitis. Studies have shown that MSC-exosomes can attenuate liver injury and inflammation in mouse models of autoimmune hepatitis induced by concanavalin A or liver antigen S100.^{110,111} The anti-inflammatory and hepatoprotective effects of MSC-exosomes are comparable to mesenchymal stem cell transplantation.¹¹⁰ Tracking experiments revealed that MSC-exosomes could target the injured liver after injection. MSC-exosomes can transfer microRNAs, such as miR-223-3p, to hepatocytes and immune cells like macrophages in the liver, regulating cell death, inflammation and immune responses.¹⁰⁹ It also inhibited the activation of NLRP3 inflammasome and caspase-1 activation, thus reducing IL-1 β production and pyroptosis.¹⁰⁸ MSC-exosomes modulate the polarization of macrophages from pro-inflammatory M1 phenotype to anti-inflammatory M2 phenotypes and increase the number and function of regulatory T cells in the liver through immunosuppressive molecules like TGF- β .

Furthermore, therapeutic molecules like dexamethasone can be loaded into MSC exosomes to enhance the treatment outcome. The drug-loaded exosomes exhibit better liver distribution and anti-inflammatory effects.¹¹¹ MSC-exosomes show promise as a cell-free therapy for AIH through multiple mechanisms of action. Further investigations are warranted to promote the clinical translation of MSC-exosomes.

MSC-EVs and Liver Fibrosis

Liver fibrosis is a degenerative condition that occurs as a result of long-term liver damage. It is a persistent inflammatory response characterized by the replacement of normal liver tissue with fibrous tissue. Many chronic liver diseases, such as hepatitis B and hepatitis C, fatty liver, and alcoholic liver disease, often lead to liver fibrosis. Additionally, the presence of extensive fibrosis may result in cirrhosis and liver insufficiency.¹⁵⁵ HSCs that are activated in the liver are the primary fibroblasts and have a crucial function in the development of liver fibrosis. Normally, HSCs stay dormant and store vitamin A and bilirubin. However, when the liver is damaged, HSCs become active and transform into cell types linked to fibrosis. HSCs that have been activated release collagen, fibronectin, and other substances that stimulate the production of collagen fibers, resulting in the development of liver tissue fibrosis.¹⁵⁶

Angiogenesis plays a crucial role in the development of liver fibrosis, resulting in the development of new blood vessels with a supply of oxygen and vital nutrients.¹⁵⁷ Activation of vascular endothelial cells by vascular endothelial growth factors (eg, vascular endothelial growth factor-A) and fibronectin-inducing factors (eg, transforming growth factor β) leads to the stimulation of abnormal vasculogenesis and perivascular inflammatory reactions.¹⁵⁸

However, the primary cause of liver fibrosis¹⁵⁷ is the persistent inflammatory reaction resulting from long-term liver disease. Inflammation-causing agents include cytokines (TNF- α , IL-1, IL-6) and chemokines (alcohol, compounds from oxidative stress). These substances induce hepatocellular damage and apoptosis by stimulating the development of liver fibrosis.⁴⁰ Furthermore, the accumulation of collagen and other matrix components in a fibrosed liver modifies the organization and performance of the liver. Activated HSCs release matrix metalloproteinase inhibitors, which inhibit matrix metalloproteinase activity, leading to matrix accumulation and deposition and promoting the progression of fibrosis.¹³²

In addition to the mechanisms mentioned above, several other molecular pathways, including the TGF- β /Smad pathway, Wnt/ β -catenin pathway, and NF- κ B pathway, are also considered significant in the development of liver fibrosis. Numerous preclinical investigations have recently reported the anti-fibrotic properties of MSC-EVs in the liver,¹⁵⁹ lungs,¹⁶⁰ kidneys,¹⁶¹ and heart.¹⁶² As stated previously, the progression of liver fibrosis is intricately linked to a persistent inflammatory reaction, and MSC-EVs exert anti-inflammatory effects. MSC-EVs can suppress the synthesis of inflammatory cytokines, including TNF- α and IL-1 β , leading to a decrease in inflammation.¹⁴¹ The transformation of epithelial cells to mesenchymal cells can be inhibited by MSC-EVs, leading to the amelioration of CCl₄-induced hepatic fibrosis.¹²⁸

Furthermore, the activation of HSCs can be significantly inhibited by MSC-EVs. The primary feature of hepatic fibrosis is the accumulation of collagen and other matrix components. MSC-EVs can reduce the progression of fibrosis by suppressing collagen production and regulating matrix metalloproteinase activity.¹³² MSC-EVs can directly act on hepatic stellate cells to inhibit their activation and proliferation, thereby reducing the expression of collagen and other fibrosis-related genes.^{127,128,133} Suppressing the excessive proliferation of hepatic stellate cells and the over-deposition of collagen is an important mechanism for the anti-fibrotic effects of MSC-EVs. MSC-EVs can also reduce the accumulation and activation of inflammatory cells, such as macrophages in the liver,^{125,130} since inflammatory responses and immune cell activation are closely related to the process of liver fibrosis. By inhibiting inflammatory signaling pathways such as NF- κ B,^{125,131,133} MSC-EVs can exert inhibitory effects on fibrosis. In addition, MSC-EVs can inhibit EMT processes and protect hepatocytes from apoptosis.^{128,129}

The occurrence of EMT can lead to an increase in fibroblasts, while hepatocyte apoptosis is also a feature of liver fibrosis. MSC-EVs reduced EMT by upregulating markers such as E-cadherin. Some studies have indicated that the therapeutic effects of MSC-EVs are superior to MSC themselves^{131,132} because EVs can avoid some of the problems of cell therapy, such as the risk of embolism. Surface-engineered EVs can improve targeting,¹³⁴ while sustained-release formulations can prolong the action time of EVs in target organs.¹²⁹ Furthermore, miRNAs enriched in MSC-EVs, such as miR-223 and miR-486,^{109,127,133} are also involved in their anti-fibrotic mechanisms. These miRNAs can downregulate hepatic stellate cell activation-related genes to exert effects. miR-150-5p can inhibit hepatic stellate cell activation by targeting CXCL1.¹³⁵

Therefore, MSCs and their secreted EVs exert synergistic effects through multiple pathways, inhibiting activation of hepatic stellate cells and inflammatory cells, reducing EMT, protecting hepatocytes and other mechanisms to treat liver fibrosis.

(Figure 2). This provides a theoretical basis for MSC-EVs to become a new extracellular therapy for liver fibrosis, but further research and clinical validation are still needed.

MSC-EVs and HIRI

HIRI is a tissue injury and inflammatory response caused by hepatic ischemia (insufficient blood supply) and reperfusion (blood resupply). Liver surgery, liver transplantation, and shock often result in HIRI. Severe HIRI can result in liver failure and organ damage.¹⁶³ The intricate process of HIRI entails interactions between several cellular types and chemical communication channels. Ischemia impairs cellular activity and disturbs energy metabolism in the liver by depriving it of oxygen and nutrients.¹⁶⁴ Ischemia also leads to a decrease in intracellular ATP levels, resulting in elevated intracellular calcium ion levels, compromised mitochondrial function, and the development of oxidative stress.¹⁶⁵

When blood is being resupplied to the liver during reperfusion, several negative effects are associated with this process. Reperfusion worsens oxidative stress, which generates significant amounts of reactive oxygen radicals and inflammatory agents. These changes cause apoptosis, necrosis, and inflammatory responses, all of which contribute to tissue damage.¹⁶⁴ Oxygen radicals and inflammatory mediators cause lipid peroxidation of cell membranes, protein oxidation, and DNA damage, which worsen cellular damage and inflammatory responses.¹⁶⁵

Studies show that CD4⁺ T lymphocytes have a crucial function in initiating hepatic inflammatory reactions in response to HIRI. Following reperfusion, a notable increase is observed in the proliferation, infiltration, and aggregation of CD4⁺ T cells that have specificity for antigens in ischemia-affected tissues. CD154, a cell-surface protein weighing 32–39 kDa and belonging to the TNF superfamily, is abundantly expressed by activated CD4⁺ T cells. This protein can

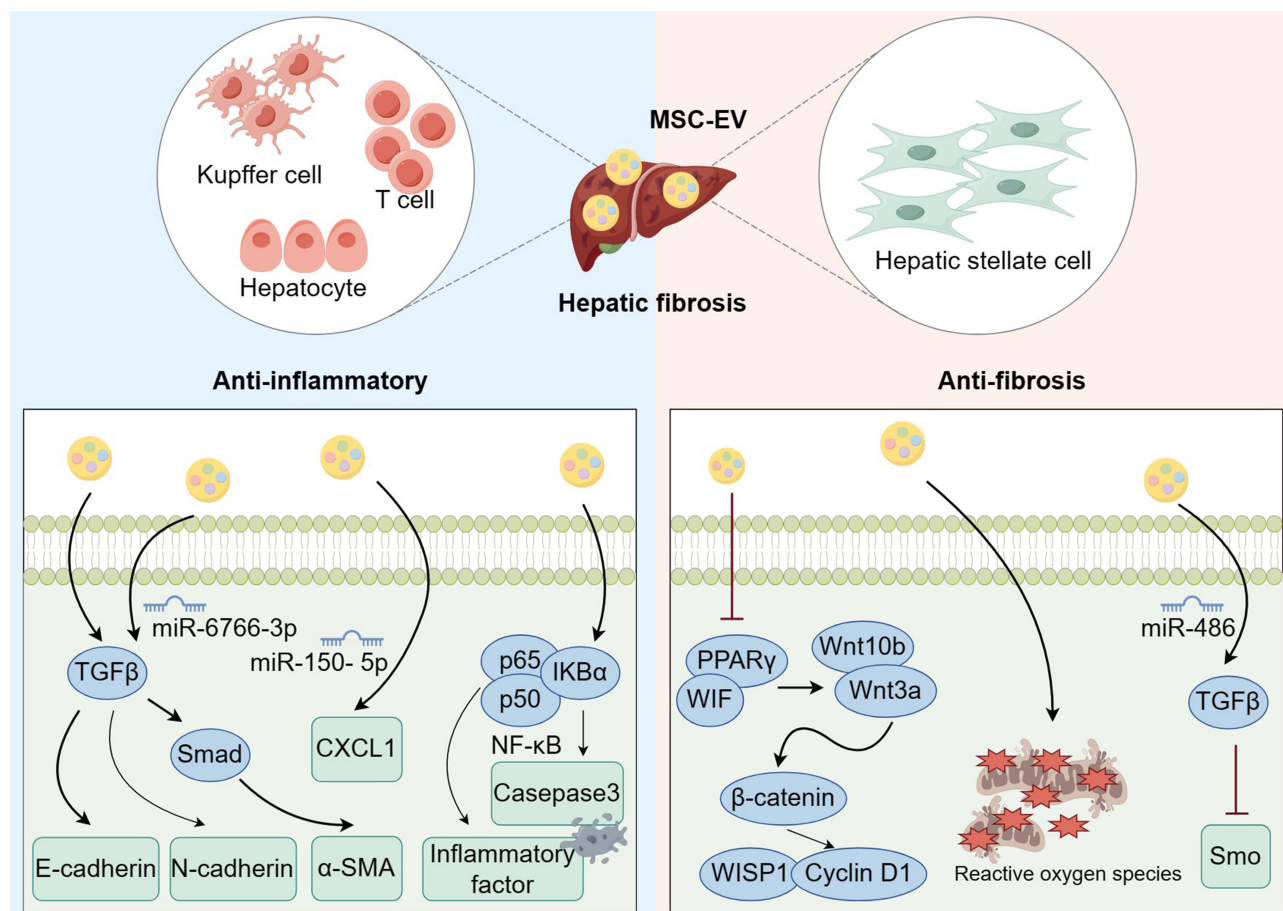


Figure 2 Mechanisms of mesenchymal stem cells in treating liver fibrosis. MSC-EVs can inhibit liver fibrosis by reducing inflammatory response, suppressing hepatic stellate cell activation and proliferation, and promoting hepatocyte regeneration.

Note: This figure is originally drawn by Figdraw platform (www.figdraw.com).

enhance the immune response, promote platelet production, and worsen hepatocellular injury by interacting with CD40/CD154. Low levels of CD154 are typically expressed during normal circumstances. CD4⁺ T cell activation leads to the continuous synthesis and expression of a significant level of CD154 on the cell surface. CD154 binds to CD40 on B cells, natural killer (NK) cells, dendritic cells, macrophages, basophils, and eosinophils, triggering the activation of downstream transcription factors that stimulate cytokine production.

Furthermore, CD154 is a protein that can undergo rapid degradation over a short period.¹⁰⁵ Hence, early therapeutic strategies are essential to regulate the manifestation of CD154 on hepatic CD4⁺ T cells to alleviate HIRI and decrease the complications and mortality in OLT. Thus, the hepatoprotective effects of MSC-EVs are attributed to the suppression of CD154 expression on CD4⁺ T cells in the liver. Additional mechanistic investigations revealed that chaperone proteins that include the TCP1 subunit 2 in MSC-EVs were moved to CD4⁺ T cells. These cells are responsible for controlling the calcium influx/NFAT1 signaling pathway, affecting the synthesis and expression of CD154.¹²³

Moreover, MSC-EVs can reduce serum aminotransferase levels, alleviate tissue necrosis, increase the number of Ki-67-positive hepatocytes, and inhibit the transcription of inflammation-related genes, demonstrating regenerative repair potential.^{118,124} Compared to unfractionated conditioned medium, the fractionated EVs secreted by MSCs have greater cytoprotective and restorative capacities.¹¹⁸ In mouse IRI models, MSC-EVs can reduce apoptotic and caspase-3 positive cells, decrease inflammatory factor mRNA levels, increase hepatocyte viability, and inhibit oxidative stress and NF-κB activation, thereby suppressing inflammatory responses and cell apoptosis.^{119,120} This is related to the antioxidant enzyme MnSOD carried and delivered by MSC-EVs.¹²⁰ Exogenous MSC-EVs can transport IL-10 into the liver, be

taken up by Kupffer cells, induce PTPN22 expression, polarize Kupffer cells towards an anti-inflammatory phenotype, and alleviate inflammation and liver injury.¹²¹ Studies have demonstrated that miR-1246 originating from MSC-EVs interacts with GSK 3 β in liver cells, significantly suppressing Wnt1, Wnt 3a, and β -catenin expression. This interaction activates the Wnt/ β -catenin signaling pathway and reduces the production of HIRI-induced TNF- α , IL-6, and IL-1 β .

Consequently, this reduction in inflammation alleviates HIRI.¹²² The exploration of employing MSC-EVs to enhance HIRI is presently in its preliminary stages, thus raising several unresolved inquiries. Further research is required to conduct comprehensive investigations into the mechanisms and lasting impacts of MSC-EVs in the treatment of HIRI.

Limitations of MSC-EVs and Improvements in Clinical Applications

Pluripotent stem cells can be employed to acquire MSC-EVs, which exhibit potential for clinical application. However, within a clinical setting, MSC-EVs are linked to specific disadvantages that require attention. Moreover, the methods for manufacturing MSC-EVs have not yet been well standardized. The exploitation of these resources in a clinical setting is impeded by the obstacles presented by their diversity and limited length of use. Different techniques of preparation produce various batches of MSC-EVs, each possessing unique features and functions. Therefore, it is crucial to create standardized preparation processes in order to ensure consistency and reliability when utilized for clinical purposes.

Significant challenges also lie in the concentration and purification of MSC-EVs. The commonly used method of isolating extracellular vesicles (EVs), ultracentrifugation, is ineffective for characterizing individual EVs. To address this issue, a number of innovations in EV isolation techniques are currently being employed. These include techniques such as fluorescent marking and subsequent analysis using high-resolution flow cytometry,¹⁶⁶ specialized flow cytometry,¹⁶⁷ and the use of laser tweezers and Raman spectroscopy¹⁶⁸ for quantitative and qualitative assessment. The purity and functionality of MSC-EVs may be affected by the presence of other cell types or impurities during the current techniques used for isolation and enrichment. Hence, it is imperative to improve the efficacies of the separation and enrichment techniques to enhance the purity of MSC-EVs.

Additionally, the long-term viability of MSC-EVs is also a matter of concern. Possible alterations in the functionalities of MSC-EVs may occur as a consequence of several conditions, including temperature, pH, and oxygen levels, in both *in vitro* and *in vivo* settings. These changes have the potential to lead to a reduction or complete cessation of their functions. Recent studies focus on delivering EVs locally using tissue-engineered substances, which could be a potential approach to enhancing their practical use. Polymer networks in hydrogels have a three-dimensional arrangement that enables them to absorb substantial quantities of water or biofluids.¹⁶⁹ Hydrogels exhibit biocompatibility and remarkable mechanical characteristics.¹⁷⁰

Furthermore, hydrogels can maintain EV release, boost their stability, and enhance their efficacy.⁴⁸ Thus, the identification of a suitable modifier may enhance the durability of MSC-EVs. Nevertheless, our understanding of the processes by which MSC-EVs exert their effects and their biological activities is limited. Notwithstanding this limitation, MSC-EVs are recognized for their anti-inflammatory, anti-fibrotic, and regeneration-promoting properties. Therefore, further investigation is necessary to further understand the roles of MSC-EVs in a therapeutic context.

Summary and Future Prospects

The utilization of MSCs is critical in the management of disorders that alter tissue function, as emerging data indicates that the compounds they produce have a therapeutic impact. MSC-EVs, being the primary constituent of MSC paracrine substances, can protect against different types of chronic liver ailments. The liver offers a range of cell types that can serve as recipient cells, including hepatic macrophages for exosomes.¹⁷¹ Recently, small sample clinical studies have been conducted to explore the safety and initial effectiveness of exosomes in the treatment of cirrhosis (ChiCTR2300075676). Nevertheless, the evaluation of MSC-EVs in clinical studies remains limited, and a significant disparity exists before their utilization in a therapeutic environment. In clinical trials involving MSC-EVs, it is crucial to address the challenges of optimizing the culture conditions of MSC, establishing standardized protocols for extracting and identifying EVs, and determining the optimal disease states that could benefit from treatment.¹⁷² Coupling gene editing tools or various biomaterials to tissue-specific extracellular vesicles may enable precise modulation of target genes to improve therapeutic efficacy.¹⁷³ For example, loading CRISPR-Cas9 ribonucleoproteins into extracellular

vesicles derived from activated hepatic stellate cells enabled targeted delivery to liver tissue.¹⁷⁴ This extracellular vesicle-mediated gene editing system specifically accumulated in the liver in vivo and exhibited significant therapeutic potential in models of acute liver injury, chronic liver fibrosis and hepatocellular carcinoma.

In conclusion, MSC-EVs have great potential for treating liver disease. These problems will undoubtedly be answered as research on MSC and its EVs progresses, opening up exciting new therapeutic opportunities for the treatment of chronic liver diseases. In the future, we anticipate that MSC-EVs will be a useful therapeutic approach for treating liver disorders.

Author Contributions

All authors made a significant contribution to the work reported, whether that is in the conception, study design, execution and interpretation, or in all these areas; took part in drafting, revising or critically reviewing the article; gave final approval of the version to be published; have agreed on the journal to which the article has been submitted; and agree to be accountable for all aspects of the work.

Funding

This review was supported by Jinhua Municipal Science and Technology Bureau Key Project (No. 2023-3-091) and Jinhua Municipal Central Hospital Science and Technology Project (No. JY2022-1-04).

Disclosure

The authors declare that they have no competing interests.

References

1. Wu Y, Min J, Ge C, et al. Interleukin 22 in Liver Injury, Inflammation and Cancer. *Int J Biol Sci.* 2020;16(13):2405–2413. doi:10.7150/ijbs.38925
2. Carmona C, Claxton L, O'Brien A, Hebditch V. Cirrhosis in over 16s: assessment and management—updated summary of NICE guidance. *BMJ.* 2023;383:2598. doi:10.1136/bmj.p2598
3. Sobeh M, Hamza MS, Ashour ML, et al. A polyphenol-rich fraction from *Eugenia uniflora* exhibits antioxidant and hepatoprotective activities in vivo. *Pharmaceuticals.* 2020;13(5):84.
4. Comarmond C, Cacoub P, Saadoun D. Treatment of chronic hepatitis C-associated cryoglobulinemia vasculitis at the era of direct-acting antivirals. *Therap Adv Gastroenterol.* 2020;13:1756284820942617. doi:10.1177/1756284820942617
5. Udompap P, Kim D, Kim WR. Current and Future Burden of Chronic Nonmalignant Liver Disease. *Clin Gastroenterol Hepatol.* 2015;13(12):2031–2041. doi:10.1016/j.cgh.2015.08.015
6. Starkey Lewis P, Campana L, Aleksieva N, et al. Alternatively activated macrophages promote resolution of necrosis following acute liver injury. *J Hepatol.* 2020;73(2):349–360. doi:10.1016/j.jhep.2020.02.031
7. Caldez MJ, Bjorklund M, Kaldis P. Cell cycle regulation in NAFLD: when imbalanced metabolism limits cell division. *Hepatol Int.* 2020;14(4):463–474. doi:10.1007/s12072-020-10066-6
8. Minami T, Aoyagi K, Kawahara A, et al. Evaluation of the expression of bone marrow-derived mesenchymal stem cells and cancer-associated fibroblasts in the stroma of gastric cancer tissue. *Ann Gastroenterol Surg.* 2020;4(4):464–474. doi:10.1002/ags3.12347
9. Freedman BR, Mooney DJ. Biomaterials to Mimic and Heal Connective Tissues. *Adv Mater.* 2019;31(19):e1806695. doi:10.1002/adma.201806695
10. Gomzikova MO, Aimaletdinov AM, Bondar OV, et al. Immunosuppressive properties of cytochalasin B-induced membrane vesicles of mesenchymal stem cells: comparing with extracellular vesicles derived from mesenchymal stem cells. *Sci Rep.* 2020;10(1):10740. doi:10.1038/s41598-020-67563-9
11. Li Q, Chen X, Li J. Marrow-derived mesenchymal stem cells regulate the inflammatory response and repair alveolar type II epithelial cells in acute lung injury of rats. *J Int Med Res.* 2020;48(4):300060520909027. doi:10.1177/0300060520909027
12. Li Z, Gong X, Li D, Yang X, Shi Q, Ju X. Intratracheal Transplantation of Amnion-Derived Mesenchymal Stem Cells Ameliorates Hyperoxia-Induced Neonatal Hyperoxic Lung Injury via Aminoacyl-Peptide Hydrolase. *Int J Stem Cells.* 2020;13(2):221–236. doi:10.15283/ijsc19110
13. Ni Z, Zhou S, Li S, et al. Exosomes: roles and therapeutic potential in osteoarthritis. *Bone Res.* 2020;8:25. doi:10.1038/s41413-020-0100-9
14. Solis MA, I M, Correa R, Huang LLH. Stem cells as a potential therapy for diabetes mellitus: a call-to-action in Latin America. *Diabetol Metab Syndr.* 2019;11:20. doi:10.1186/s13098-019-0415-0
15. Takayama Y, Kusamori K, Katsurada Y, Obana S, Itakura S, Nishikawa M. Efficient delivery of mesenchymal stem/stromal cells to injured liver by surface PEGylation. *Stem Cell Res Ther.* 2023;14(1):216. doi:10.1186/s13287-023-03446-w
16. Karaahmet F, Kocaman SA. Endothelial progenitor cells and mesenchymal stem cells to overcome vascular deterioration and cytokine storm in critical patients with COVID-19. *Med Hypotheses.* 2020;144:109973. doi:10.1016/j.mehy.2020.109973
17. Seyedrazizadeh S-Z, Poosti S, Nazari A, et al. Extracellular vesicles derived from human ES-MSCs protect retinal ganglion cells and preserve retinal function in a rodent model of optic nerve injury. *Stem Cell Res Ther.* 2020;11(1):203. doi:10.1186/s13287-020-01702-x

18. Xu B, Yuan F-Z, Lin L, et al. The Higher Inherent Therapeutic Potential of Biomaterial-Based hDPSCs and hEnSCs for Pancreas Diseases. *Front Bioeng Biotechnol.* 2020;8:636. doi:10.3389/fbioe.2020.00636
19. Wang J, Fu X, Yan Y, et al. In vitro differentiation of rhesus macaque bone marrow- and adipose tissue-derived MSCs into hepatocyte-like cells. *Exp Ther Med.* 2020;20(1):251–260. doi:10.3892/etm.2020.8676
20. Mishra VK, Shih -H-H, Parveen F, et al. Identifying the Therapeutic Significance of Mesenchymal Stem Cells. *Cells.* 2020;9(5):1145.
21. Fan T, Qu R, Yu Q, et al. Bioinformatics analysis of the biological changes involved in the osteogenic differentiation of human mesenchymal stem cells. *J Cell Mol Med.* 2020;24(14):7968–7978. doi:10.1111/jcmm.15429
22. Allameh A, Ahmadi-Ashtiani HR, Maleki N. Glutathione-related inflammatory signature in hepatocytes differentiated from the progenitor mesenchymal stem cells. *Heliyon.* 2020;6(6):e04149. doi:10.1016/j.heliyon.2020.e04149
23. Yao Z, Liu H, Yang M, et al. Bone marrow mesenchymal stem cell-derived endothelial cells increase capillary density and accelerate angiogenesis in mouse hindlimb ischemia model. *Stem Cell Res Ther.* 2020;11(1):221. doi:10.1186/s13287-020-01710-x
24. Balbi C, Costa A, Barile L, Bollini S. Message in a Bottle: upgrading Cardiac Repair into Rejuvenation. *Cells.* 2020;9(3):548.
25. Zhang J, Xie B, Hashimoto K. Current status of potential therapeutic candidates for the COVID-19 crisis. *Brain Behav Immun.* 2020;87:59–73. doi:10.1016/j.bbi.2020.04.046
26. Krambs JR, Abou Ezzi G, Yao J-C, Link DC. Canonical signaling by TGF family members in mesenchymal stromal cells is dispensable for hematopoietic niche maintenance under basal and stress conditions. *PLoS One.* 2020;15(5):e0233751. doi:10.1371/journal.pone.0233751
27. Wang L, Zhang L, Liang X, et al. Adipose Tissue-Derived Stem Cells from Type 2 Diabetics Reveal Conservative Alterations in Multidimensional Characteristics. *Int J Stem Cells.* 2020;13(2):268–278. doi:10.15283/ijsc20028
28. Chen J, Li M, Liu A-Q, et al. Gli1+ Cells Couple with Type H Vessels and Are Required for Type H Vessel Formation. *Stem Cell Reports.* 2020;15(1):110–124. doi:10.1016/j.stemcr.2020.06.007
29. Parhizkar Roudsari P, Alavi-Moghadam S, Payab M, et al. Auxiliary role of mesenchymal stem cells as regenerative medicine soldiers to attenuate inflammatory processes of severe acute respiratory infections caused by COVID-19. *Cell Tissue Bank.* 2020;21(3):405–425. doi:10.1007/s10561-020-09842-3
30. Basiri A, Pazhouhnia Z, Beheshtizadeh N, Hoseinpour M, Saghadzadeh A, Rezaei N. Regenerative Medicine in COVID-19 Treatment: real Opportunities and Range of Promises. *Stem Cell Rev Rep.* 2021;17(1):163–175. doi:10.1007/s12015-020-09994-5
31. Yu C, Peall IW, Pham SH, Okolicsanyi RK, Griffiths LR, Haupt LM. Syndecan-1 Facilitates the Human Mesenchymal Stem Cell Osteo-Adipogenic Balance. *Int J Mol Sci.* 2020;21(11):145.
32. Carvalheiro T, Zimmermann M, Radstake TRDJ, Marut W. Novel insights into dendritic cells in the pathogenesis of systemic sclerosis. *Clin Exp Immunol.* 2020;201(1):25–33. doi:10.1111/cei.13417
33. Fan D, Zeng M, Xia Q, et al. Efficacy and safety of umbilical cord mesenchymal stem cells in treatment of cesarean section skin scars: a randomized clinical trial. *Stem Cell Res Ther.* 2020;11(1):244. doi:10.1186/s13287-020-01695-7
34. van Best N, Rolle-Kampczyk U, Schaap FG, et al. Bile acids drive the newborn's gut microbiota maturation. *Nat Commun.* 2020;11(1):3692. doi:10.1038/s41467-020-17183-8
35. Alqahtani SA, Schattenberg M. Liver injury in COVID-19: the current evidence. *United Eur Gastroenterol J.* 2020;8(5):509–519. doi:10.1177/2050640620924157
36. Furuta T, Furuya K, Zheng Y-W, Oda T. Novel alternative transplantation therapy for orthotopic liver transplantation in liver failure: a systematic review. *World J Transplant.* 2020;10(3):64–78. doi:10.5500/wjt.v10.i3.64
37. Petryk N, Shevchenko O. Mesenchymal Stem Cells Anti-Inflammatory Activity in Rats: proinflammatory Cytokines. *J Inflamm Res.* 2020;13:293–301. doi:10.2147/JIR.S256932
38. Lotfi M, Hamblin MR, Rezaei N. COVID-19: transmission, prevention, and potential therapeutic opportunities. *Clin Chim Acta.* 2020;508:254–266. doi:10.1016/j.cca.2020.05.044
39. Ullah M, Liu DD, Rai S, et al. Reversing Acute Kidney Injury Using Pulsed Focused Ultrasound and MSC Therapy: a Role for HSP-Mediated PI3K/AKT Signaling. *Mol Ther Methods Clin Dev.* 2020;17:683–694. doi:10.1016/j.omtm.2020.03.023
40. Cai J, Wu J, Wang J, et al. Extracellular vesicles derived from different sources of mesenchymal stem cells: therapeutic effects and translational potential. *Cell Biosci.* 2020;10:69. doi:10.1186/s13578-020-00427-x
41. Ikeda T, Nishita M, Hoshi K, Honda T, Kakeji Y, Minami Y. Mesenchymal stem cell-derived CXCL16 promotes progression of gastric cancer cells by STAT3-mediated expression of Ror1. *Cancer Sci.* 2020;111(4):1254–1265. doi:10.1111/cas.14339
42. Watanabe T, Tsuchiya A, Takeuchi S, et al. Development of a non-alcoholic steatohepatitis model with rapid accumulation of fibrosis, and its treatment using mesenchymal stem cells and their small extracellular vesicles. *Regen Ther.* 2020;14:252–261. doi:10.1016/j.reth.2020.03.012
43. Arnold P, Li W, et al. Joint Reconstituted Signaling of the IL-6 Receptor via Extracellular Vesicles. *Cells.* 2020;9(5):1307.
44. Krammer TL, Mayr M, Hackl M. microRNAs as promising biomarkers of platelet activity in antiplatelet therapy monitoring. *Int J Mol Sci.* 2020;21:254.
45. Shimizu A, Sawada K, Kimura T. Pathophysiological Role and Potential Therapeutic Exploitation of Exosomes in Ovarian Cancer. *Cells.* 2020;9(4):814.
46. Campos A, Leyton L, Quest AFG. Caveolin-1 function at the plasma membrane and in intracellular compartments in cancer. *Cancer Metastasis Rev.* 2020;39(2):435–453. doi:10.1007/s10555-020-09890-x
47. Seo Y, Kim H-S, Hong I-S. Stem Cell-Derived Extracellular Vesicles as Immunomodulatory Therapeutics. *Stem Cells Int.* 2019;2019:5126156. doi:10.1155/2019/5126156
48. Tsiapalis D, O'Driscoll L. Mesenchymal Stem Cell Derived Extracellular Vesicles for Tissue Engineering and Regenerative Medicine Applications. *Cells.* 2020;9(4):991.
49. Ryu J-S, Seo SY, Jeong E-J, et al. Ganglioside GM3 Up-Regulate Chondrogenic Differentiation by Transform Growth Factor Receptors. *Int J Mol Sci.* 2020;21(6):548.
50. Lee DB, Verstraete FJM, Arzi B. An Update on Feline Chronic Gingivostomatitis. *Vet Clin North Am Small Anim Pract.* 2020;50(5):973–982. doi:10.1016/j.cvsm.2020.04.002
51. Volleman TNE, Schol J, Morita K, Sakai D, Watanabe M. Wnt3a and wnt5a as Potential Chondrogenic Stimulators for Nucleus Pulposus Cell Induction: a Comprehensive Review. *Neurospine.* 2020;17(1):19–35. doi:10.14245/ns.2040040.020

52. Hutchings G, Janowicz K, Moncrieff L, et al. The Proliferation and Differentiation of Adipose-Derived Stem Cells in Neovascularization and Angiogenesis. *Int J Mol Sci.* 2020;21(11):991.
53. Bao X, Wang J, Zhou G, et al. Extended in vitro culture of primary human mesenchymal stem cells downregulates Brca1-related genes and impairs DNA double-strand break recognition. *FEBS Open Bio.* 2020;10(7):1238–1250. doi:10.1002/2211-5463.12867
54. Shi R, Lian W, Jin Y, et al. Role and effect of vein-transplanted human umbilical cord mesenchymal stem cells in the repair of diabetic foot ulcers in rats. *Acta Biochim Biophys Sin (Shanghai).* 2020;52(6):620–630. doi:10.1093/abbs/gmaa039
55. Gnecci M, He H, Liang OD, et al. Paracrine action accounts for marked protection of ischemic heart by Akt-modified mesenchymal stem cells. *Nat Med.* 2005;11(4):367–368.
56. Gnecci M, He H, Noiseux N, et al. Evidence supporting paracrine hypothesis for Akt-modified mesenchymal stem cell-mediated cardiac protection and functional improvement. *FASEB J.* 2006;20(6):661–669.
57. Okazaki T, Magaki T, Takeda M, et al. Intravenous administration of bone marrow stromal cells increases survivin and Bcl-2 protein expression and improves sensorimotor function following ischemia in rats. *Neurosci Lett.* 2008;430(2):109–114.
58. Wang S-P, Wang Z-H, Peng D-Y, S-M L, Wang H, Wang X-H. Therapeutic effect of mesenchymal stem cells in rats with intracerebral hemorrhage: reduced apoptosis and enhanced neuroprotection. *Mol Med Rep.* 2012;6(4):848–854. doi:10.3892/mmr.2012.997
59. Wei H, Li F, Xue T, et al. MicroRNA-122-functionalized DNA tetrahedron stimulate hepatic differentiation of human mesenchymal stem cells for acute liver failure therapy. *Bioact Mater.* 2023;28:50–60. doi:10.1016/j.bioactmat.2023.04.024
60. Hu C, Li L. In vitro culture of isolated primary hepatocytes and stem cell-derived hepatocyte-like cells for liver regeneration. *Protein Cell.* 2015;6(8):562–574. doi:10.1007/s13238-015-0180-2
61. Z-H L, Wang Y-L, Wang H-J, J-H W, Tan Y-Z. Rapamycin-Preactivated Autophagy Enhances Survival and Differentiation of Mesenchymal Stem Cells After Transplantation into Infarcted Myocardium. *Stem Cell Rev Rep.* 2020;16(2):344–356. doi:10.1007/s12015-020-09952-1
62. Zhang Y, Li Y, Li W, et al. Therapeutic Effect of Human Umbilical Cord Mesenchymal Stem Cells at Various Passages on Acute Liver Failure in Rats. *Stem Cells Int.* 2018;2018:7159465. doi:10.1155/2018/7159465
63. Zhou X, Jin N, Wang F, Chen B. Mesenchymal stem cells: a promising way in therapies of graft-versus-host disease. *Cancer Cell Int.* 2020;20:114. doi:10.1186/s12935-020-01193-z
64. Buono L, Scalabrin S, De Iulius M, et al. Mesenchymal Stem Cell-Derived Extracellular Vesicles Protect Human Corneal Endothelial Cells from Endoplasmic Reticulum Stress-Mediated Apoptosis. *Int J Mol Sci.* 2021;22(9):4930.
65. Moayedfard Z, Sani F, Alizadeh A, Bagheri Lankarani K, Zarei M, Azarpira N. The role of the immune system in the pathogenesis of NAFLD and potential therapeutic impacts of mesenchymal stem cell-derived extracellular vesicles. *Stem Cell Res Ther.* 2022;13(1):242. doi:10.1186/s13287-022-02929-6
66. Khatri M, Richardson LA, Meulia T. Mesenchymal stem cell-derived extracellular vesicles attenuate influenza virus-induced acute lung injury in a pig model. *Stem Cell Res Ther.* 2018;9(1):17. doi:10.1186/s13287-018-0774-8
67. Wu R, Fan X, Wang Y, et al. Mesenchymal Stem Cell-Derived Extracellular Vesicles in Liver Immunity and Therapy. *Front Immunol.* 2022;13:833878. doi:10.3389/fimmu.2022.833878
68. Zhao M, Liu S, Wang C, et al. Mesenchymal Stem Cell-Derived Extracellular Vesicles Attenuate Mitochondrial Damage and Inflammation by Stabilizing Mitochondrial DNA. *ACS Nano.* 2021;15(1):1519–1538. doi:10.1021/acsnano.0c08947
69. Branscome H, Paul S, Yin D, et al. Use of Stem Cell Extracellular Vesicles as a “Holistic” Approach to CNS Repair. *Front Cell Dev Biol.* 2020;8:455. doi:10.3389/fcell.2020.00455
70. Yin L, Liu X, Shi Y, et al. Therapeutic Advances of Stem Cell-Derived Extracellular Vesicles in Regenerative Medicine. *Cells.* 2020;9(3):548.
71. Li Q, Huang Z, Wang Q, et al. Targeted immunomodulation therapy for cardiac repair by platelet membrane engineering extracellular vesicles via hitching peripheral monocytes. *Biomaterials.* 2022;284:121529. doi:10.1016/j.biomaterials.2022.121529
72. Aneesh A, Liu A, Moss HE, et al. Emerging concepts in the treatment of optic neuritis: mesenchymal stem cell-derived extracellular vesicles. *Stem Cell Res Ther.* 2021;12(1):594. doi:10.1186/s13287-021-02645-7
73. Fujii S, Miura Y. Immunomodulatory and Regenerative Effects of MSC-Derived Extracellular Vesicles to Treat Acute GVHD. *Stem Cells.* 2022;40(11):977–990. doi:10.1093/stmcls/sxac057
74. Harrell CR, Jovicic N, Djonov V, Volarevic V. Therapeutic Use of Mesenchymal Stem Cell-Derived Exosomes: from Basic Science to Clinics. *Pharmaceutics.* 2020;12(5):991.
75. Qiu L, Wang J, Chen M, Chen F, Tu W. Exosomal microRNA-146a derived from mesenchymal stem cells increases the sensitivity of ovarian cancer cells to docetaxel and taxane via a LAMC2-mediated PI3K/Akt axis. *Int J Mol Med.* 2020;46(2):609–620. doi:10.3892/ijmm.2020.4634
76. Wang M, Yu F, Li P, Wang K. Emerging Function and Clinical Significance of Exosomal circRNAs in Cancer. *Mol Ther Nucleic Acids.* 2020;21:367–383. doi:10.1016/j.omtn.2020.06.008
77. Gao S, Zhu H, Zuo X, Luo H. Cathepsin G and Its Role in Inflammation and Autoimmune Diseases. *Arch Rheumatol.* 2018;33(4):498–504. doi:10.5606/ArchRheumatol.2018.6595
78. Lu T, Zhang J, Cai J, et al. Extracellular vesicles derived from mesenchymal stromal cells as nanotherapeutics for liver ischaemia-reperfusion injury by transferring mitochondria to modulate the formation of neutrophil extracellular traps. *Biomaterials.* 2022;284:121486. doi:10.1016/j.biomaterials.2022.121486
79. Pan L-F, Niu Z-Q, Ren S, et al. Could extracellular vesicles derived from mesenchymal stem cells be a potential therapy for acute pancreatitis-induced cardiac injury? *World J Stem Cells.* 2023;15(7):654–664. doi:10.4252/wjsc.v15.i7.654
80. Qian X, An N, Ren Y, Yang C, Zhang X, Li L. Immunosuppressive Effects of Mesenchymal Stem Cells-derived Exosomes. *Stem Cell Rev Rep.* 2021;17(2):411–427. doi:10.1007/s12015-020-10040-7
81. Cha K-Y, Cho W, Park S, et al. Generation of bioactive MSC-EVs for bone tissue regeneration by taurosoodeoxycholic acid treatment. *J Control Release.* 2023;354:45–56. doi:10.1016/j.jconrel.2022.12.053
82. Pu Q, Xiu G, Sun J, Liu P, Ling B. Progress on the effect of mesenchymal stem cell derived exosomes on multiple organ dysfunction in sepsis. *Zhonghua Wei Zhong Bing Ji Jiu Yi Xue.* 2021;33(6):757–760. doi:10.3760/cma.j.cn121430-20200908-00620
83. Wang J-H, Liu X-L, Sun J-M, Yang J-H, D-H X, Yan -S-S. Role of mesenchymal stem cell derived extracellular vesicles in autoimmunity: a systematic review. *World J Stem Cells.* 2020;12(8):879–896. doi:10.4252/wjsc.v12.i8.879

84. Gupta S, Pinky V, et al. Comparative Evaluation of Anti-Fibrotic Effect of Tissue Specific Mesenchymal Stem Cells Derived Extracellular Vesicles for the Amelioration of CCL₄ Induced Chronic Liver Injury. *Stem Cell Rev Rep*. 2022;18(3):1097–1112. doi:10.1007/s12015-021-10313-9
85. Angioni R, Liboni C, Herkenne S, et al. CD73+ extracellular vesicles inhibit angiogenesis through adenosine A2B receptor signalling. *J Extracell Vesicles*. 2020;9(1):1757900. doi:10.1080/20013078.2020.1757900
86. Xin D, Li T, Chu X, Ke H, Liu D, Wang Z. MSCs-extracellular vesicles attenuated neuroinflammation, synapse damage and microglial phagocytosis after hypoxia-ischemia injury by preventing osteopontin expression. *Pharmacol Res*. 2021;164:105322. doi:10.1016/j.phrs.2020.105322
87. Pu Y, Li C, Qi X, et al. Extracellular Vesicles from NMN Preconditioned Mesenchymal Stem Cells Ameliorated Myocardial Infarction via miR-210-3p Promoted Angiogenesis. *Stem Cell Rev Rep*. 2023;19(4):1051–1066. doi:10.1007/s12015-022-10499-6
88. Wu Y, Peng W, Fang M, Wu M, Wu M. MSCs-Derived Extracellular Vesicles Carrying miR-212-5p Alleviate Myocardial Infarction-Induced Cardiac Fibrosis via NLR5/VEGF/TGF- β 1/SMAD Axis. *J Cardiovasc Transl Res*. 2022;15(2):302–316. doi:10.1007/s12265-021-10156-2
89. Han M, Cao Y, Xue H, et al. Neuroprotective Effect of Mesenchymal Stromal Cell-Derived Extracellular Vesicles Against Cerebral Ischemia-Reperfusion-Induced Neural Functional Injury: a Pivotal Role for AMPK and JAK2/STAT3/NF- κ B Signaling Pathway Modulation. *Drug Des Devel Ther*. 2020;14:2865–2876. doi:10.2147/DDDT.S248892
90. Gholami L, Nooshabadi VT, Shahabi S, et al. Extracellular vesicles in bone and periodontal regeneration: current and potential therapeutic applications. *Cell Biosci*. 2021;11(1):16. doi:10.1186/s13578-020-00527-8
91. Rezaie J, Nejati V, Mahmoodi M, Ahmadi M. Mesenchymal stem cells derived extracellular vesicles: a promising nanomedicine for drug delivery system. *Biochem Pharmacol*. 2022;203:115167. doi:10.1016/j.bcp.2022.115167
92. Gentile P, Sterodimas A. Adipose Stem Cells (ASCs) and Stromal Vascular Fraction (SVF) as a Potential Therapy in Combating (COVID-19)-Disease. *Aging Dis*. 2020;11(3):465–469. doi:10.14336/AD.2020.0422
93. Abreu H, Canciani E, Raineri D, Cappellano G, Rimondini L, Chiocchetti A. Extracellular Vesicles in Musculoskeletal Regeneration: modulating the Therapy of the Future. *Cells*. 2021;11(1).
94. Liao Z, Liu C, Wang L, Sui C, Zhang H. Therapeutic Role of Mesenchymal Stem Cell-Derived Extracellular Vesicles in Female Reproductive Diseases. *Front Endocrinol (Lausanne)*. 2021;12:665645. doi:10.3389/fendo.2021.665645
95. Zhang J, Lu T, Xiao J, et al. MSC-derived extracellular vesicles as nanotherapeutics for promoting aged liver regeneration. *J Control Release*. 2023;356:402–415. doi:10.1016/j.jconrel.2023.02.032
96. Diniz AB, Antunes M, Lacerda V, et al. Imaging and immunometabolic phenotyping uncover changes in the hepatic immune response in the early phases of NAFLD. *JHEP Rep*. 2020;2(4):100117. doi:10.1016/j.jhepr.2020.100117
97. Kjærgaard K, Sandahl TD, Frisch K, et al. Intravenous and oral copper kinetics, biodistribution and dosimetry in healthy humans studied by [64Cu]copper PET/CT. *EJNMMI Radiopharm Chem*. 2020;5(1):15. doi:10.1186/s41181-020-00100-1
98. Fahey S, Dempsey E, Long A. The role of chemokines in acute and chronic hepatitis C infection. *Cell Mol Immunol*. 2014;11(1):25–40. doi:10.1038/cmi.2013.37
99. Patel SR, Lundgren TS, Spencer HT, Doering CB. The Immune Response to the fVIII Gene Therapy in Preclinical Models. *Front Immunol*. 2020;11:494. doi:10.3389/fimmu.2020.00494
100. Gottwick C, Carambia A, Herkel J. Harnessing the liver to induce antigen-specific immune tolerance. *Semin Immunopathol*. 2022;44(4):475–484. doi:10.1007/s00281-022-00942-8
101. Lin L, Gong H, Li R, et al. Nanodrug with ROS and pH Dual-Sensitivity Ameliorates Liver Fibrosis via Multicellular Regulation. *Adv Sci*. 2020;7(7):1903138. doi:10.1002/advs.201903138
102. Horng J-H, Lin W-H, C-R W, et al. HBV X protein-based therapeutic vaccine accelerates viral antigen clearance by mobilizing monocyte infiltration into the liver in HBV carrier mice. *J Biomed Sci*. 2020;27(1):70. doi:10.1186/s12929-020-00662-x
103. Van Herck M, Vonghia L, Kwanten WJ, et al. Diet Reversal and Immune Modulation Show Key Role for Liver and Adipose Tissue T Cells in Murine Nonalcoholic Steatohepatitis. *Cell Mol Gastroenterol Hepatol*. 2020;10(3):467–490. doi:10.1016/j.jcmgh.2020.04.010
104. Asadipour M, Fazeli P, Zohouri M, et al. IL-18 in Blood Serum of Hepatitis C Patients Might be of Predictive Value for Individual Outcomes. *Infect Disord Drug Targets*. 2021;21(3):389–393. doi:10.2174/187152652066200707113401
105. De Luca L, Trino S, Laurenzana I, et al. Mesenchymal Stem Cell Derived Extracellular Vesicles: a Role in Hematopoietic Transplantation? *Int J Mol Sci*. 2017;18(5):852.
106. Cai P, Mu Y, Olveda RM, Ross AG, Olveda DU, McManus DP. Serum Exosomal miRNAs for Grading Hepatic Fibrosis Due to Schistosomiasis. *Int J Mol Sci*. 2020;21:5548.
107. Morājn L, Cubero FJ. Extracellular vesicles in liver disease and beyond. *World J Gastroenterol*. 2018;24(40):4519–4526. doi:10.3748/wjg.v24.i40.4519
108. Chen L, F-B L, Chen D-Z, et al. BMSCs-derived miR-223-containing exosomes contribute to liver protection in experimental autoimmune hepatitis. *Mol Immunol*. 2018;93:38–46. doi:10.1016/j.molimm.2017.11.008
109. F-B L, Chen D-Z, Chen L, et al. Attenuation of Experimental Autoimmune Hepatitis in Mice with Bone Mesenchymal Stem Cell-Derived Exosomes Carrying MicroRNA-223-3p. *Mol Cells*. 2019;42(12):906–918. doi:10.14348/molcells.2019.2283
110. Tamura R, Uemoto S, Tabata Y. Immunosuppressive effect of mesenchymal stem cell-derived exosomes on a concanavalin A-induced liver injury model. *Inflamm Regen*. 2016;36:26. doi:10.1186/s41232-016-0030-5
111. Zhao J, Li Y, Jia R, Wang J, Shi M, Wang Y. Mesenchymal Stem Cells-Derived Exosomes as Dexamethasone Delivery Vehicles for Autoimmune Hepatitis Therapy. *Front Bioeng Biotechnol*. 2021;9:650376. doi:10.3389/fbioe.2021.650376
112. Chen L, Xiang B, Wang X, Xiang C. Exosomes derived from human menstrual blood-derived stem cells alleviate fulminant hepatic failure. *Stem Cell Res Ther*. 2017;8(1):9. doi:10.1186/s13287-016-0453-6
113. Haga H, Yan IK, Takahashi K, Matsuda A, Patel T. Extracellular Vesicles from Bone Marrow-Derived Mesenchymal Stem Cells Improve Survival from Lethal Hepatic Failure in Mice. *Stem Cells Transl Med*. 2017;6(4):1262–1272. doi:10.1002/setm.16-0226
114. Jiang L, Zhang S, Hu H, et al. Exosomes derived from human umbilical cord mesenchymal stem cells alleviate acute liver failure by reducing the activity of the NLRP3 inflammasome in macrophages. *Biochem Biophys Res Commun*. 2019;508(3):735–741. doi:10.1016/j.bbrc.2018.11.189

115. Liu Y, Lou G, Li A, et al. AMSC-derived exosomes alleviate lipopolysaccharide/d-galactosamine-induced acute liver failure by miR-17-mediated reduction of TXNIP/NLRP3 inflammasome activation in macrophages. *EBioMedicine*. 2018;36:140–150. doi:10.1016/j.ebiom.2018.08.054
116. Tan CY, Lai RC, Wong W, Dan YY, Lim S-K, Ho HK. Mesenchymal stem cell-derived exosomes promote hepatic regeneration in drug-induced liver injury models. *Stem Cell Res Ther*. 2014;5(3):76. doi:10.1186/srct465
117. Zhang S, Jiang L, Hu H, et al. Pretreatment of exosomes derived from hUCMSCs with TNF- α ameliorates acute liver failure by inhibiting the activation of NLRP3 in macrophage. *Life Sci*. 2020;246:117401. doi:10.1016/j.lfs.2020.117401
118. Damania A, Jaiman D, Teotia AK, Kumar A. Mesenchymal stromal cell-derived exosome-rich fractionated secretome confers a hepatoprotective effect in liver injury. *Stem Cell Res Ther*. 2018;9(1):31. doi:10.1186/s13287-017-0752-6
119. Haga H, Yan IK, Borrelli DA, et al. Extracellular vesicles from bone marrow-derived mesenchymal stem cells protect against murine hepatic ischemia/reperfusion injury. *Liver Transpl*. 2017;23(6):791–803. doi:10.1002/lt.24770
120. Yao J, Zheng J, Cai J, et al. Extracellular vesicles derived from human umbilical cord mesenchymal stem cells alleviate rat hepatic ischemia-reperfusion injury by suppressing oxidative stress and neutrophil inflammatory response. *FASEB J*. 2019;33(2):1695–1710. doi:10.1096/fj.201800131RR
121. Zhang Y, Zhang X, Zhang H, et al. Mesenchymal Stem Cells Derived Extracellular Vesicles Alleviate Traumatic Hemorrhagic Shock Induced Hepatic Injury via IL-10/PTPN22-Mediated M2 Kupffer Cell Polarization. *Front Immunol*. 2021;12:811164. doi:10.3389/fimmu.2021.811164
122. Xie K, Liu L, Chen J, Liu F. Exosomes derived from human umbilical cord blood mesenchymal stem cells improve hepatic ischemia reperfusion injury via delivering miR-1246. *Cell Cycle*. 2019;18(24):3491–3501. doi:10.1080/15384101.2019.1689480
123. Zheng J, Lu T, Zhou C, et al. Extracellular Vesicles Derived from Human Umbilical Cord Mesenchymal Stem Cells Protect Liver Ischemia/Reperfusion Injury by Reducing CD154 Expression on CD4+ T Cells via CCT2. *Adv Sci*. 2020;7(18):1903746. doi:10.1002/advs.201903746
124. Anger F, Camara M, Ellinger E, et al. Human Mesenchymal Stromal Cell-Derived Extracellular Vesicles Improve Liver Regeneration After Ischemia Reperfusion Injury in Mice. *Stem Cells Dev*. 2019;28(21):1451–1462. doi:10.1089/scd.2019.0085
125. Angioni R, Cal α B, Vigneswara V, et al. Administration of Human MSC-Derived Extracellular Vesicles for the Treatment of Primary Sclerosing Cholangitis: preclinical Data in MDR2 Knockout Mice. *Int J Mol Sci*. 2020;21(22):8874.
126. Han HS, Lee H, You D, et al. Human adipose stem cell-derived extracellular nanovesicles for treatment of chronic liver fibrosis. *J Control Release*. 2020;320:328–336. doi:10.1016/j.jconrel.2020.01.042
127. Kim J, Lee C, Shin Y, et al. sEVs from tonsil-derived mesenchymal stromal cells alleviate activation of hepatic stellate cells and liver fibrosis through miR-486-5p. *Mol Ther*. 2021;29(4):1471–1486. doi:10.1016/j.ymthe.2020.12.025
128. Li T, Yan Y, Wang B, et al. Exosomes derived from human umbilical cord mesenchymal stem cells alleviate liver fibrosis. *Stem Cells Dev*. 2013;22(6):845–854. doi:10.1089/scd.2012.0395
129. Mardpour S, Ghanian MH, Sadeghi-Abandansari H, et al. Hydrogel-Mediated Sustained Systemic Delivery of Mesenchymal Stem Cell-Derived Extracellular Vesicles Improves Hepatic Regeneration in Chronic Liver Failure. *ACS Appl Mater Interfaces*. 2019;11(41):37421–37433. doi:10.1021/acsami.9b10126
130. Ohara M, Ohnishi S, Hosono H, et al. Extracellular Vesicles from Amnion-Derived Mesenchymal Stem Cells Ameliorate Hepatic Inflammation and Fibrosis in Rats. *Stem Cells Int*. 2018;2018:3212643. doi:10.1155/2018/3212643
131. Rong X, Liu J, Yao X, Jiang T, Wang Y, Xie F. Human bone marrow mesenchymal stem cells-derived exosomes alleviate liver fibrosis through the Wnt/ β -catenin pathway. *Stem Cell Res Ther*. 2019;10(1):98. doi:10.1186/s13287-019-1204-2
132. Rostom DM, Attia N, Khalifa HM, Abou Nazeel MW, El Sabaawy EA. The Therapeutic Potential of Extracellular Vesicles Versus Mesenchymal Stem Cells in Liver Damage. *Tissue Eng Regen Med*. 2020;17(4):537–552. doi:10.1007/s13770-020-00267-3
133. Wang N, Li X, Zhong Z, et al. 3D hESC exosomes enriched with miR-6766-3p ameliorates liver fibrosis by attenuating activated stellate cells through targeting the TGF β RII-SMADS pathway. *J Nanobiotechnology*. 2021;19(1):437. doi:10.1186/s12951-021-01138-2
134. You DG, Oh BH, Nguyen VQ, et al. Vitamin A-coupled stem cell-derived extracellular vesicles regulate the fibrotic cascade by targeting activated hepatic stellate cells in vivo. *J Control Release*. 2021;336:285–295. doi:10.1016/j.jconrel.2021.06.031
135. Du Z, Wu T, Liu L, Luo B, Wei C. Extracellular vesicles-derived miR-150-5p secreted by adipose-derived mesenchymal stem cells inhibits CXCL1 expression to attenuate hepatic fibrosis. *J Cell Mol Med*. 2021;25(2):701–715. doi:10.1111/jcmm.16119
136. Cheng L, Yu P, Li F, et al. Human umbilical cord-derived mesenchymal stem cell-exosomal miR-627-5p ameliorates non-alcoholic fatty liver disease by repressing FTO expression. *Hum Cell*. 2021;34(6):1697–1708. doi:10.1007/s13577-021-00593-1
137. Niu Q, Wang T, Wang Z, et al. Adipose-derived mesenchymal stem cell-secreted extracellular vesicles alleviate non-alcoholic fatty liver disease via delivering miR-223-3p. *Adipocyte*. 2022;11(1):572–587. doi:10.1080/21623945.2022.2098583
138. Engelmann C, Zhang IW. Mechanisms of immunity in acutely decompensated cirrhosis and acute-on-chronic liver failure. *Liver Int*. 2023. doi:10.1111/liv.15644
139. Yu Z, Li J, Ren Z, et al. Switching from Fatty Acid Oxidation to Glycolysis Improves the Outcome of Acute-On-Chronic Liver Failure. *Adv Sci*. 2020;7(7):1902996. doi:10.1002/advs.201902996
140. Cao P, Chen Q, Shi C, Wang L, Gong Z. Fusobacterium nucleatum promotes the development of acute liver failure by inhibiting the NAD $^{+}$ salvage metabolic pathway. *Gut Pathog*. 2022;14(1):29. doi:10.1186/s13099-022-00503-2
141. Zhang S, Hou Y, Yang J, et al. Application of mesenchymal stem cell exosomes and their drug-loading systems in acute liver failure. *J Cell Mol Med*. 2020;24(13):7082–7093. doi:10.1111/jcmm.15290
142. Fang X, Gao F, Yao Q, et al. Pooled Analysis of Mesenchymal Stromal Cell-Derived Extracellular Vesicle Therapy for Liver Disease in Preclinical Models. *J Pers Med*. 2023;13(3):441.
143. Yan Y, Jiang W, Tan Y, et al. hucMSC Exosome-Derived GPX1 Is Required for the Recovery of Hepatic Oxidant Injury. *Mol Ther*. 2017;25(2):465–479. doi:10.1016/j.ymthe.2016.11.019
144. Dong X, Feng X, Liu J, et al. Characteristics of Intestinal Microecology during Mesenchymal Stem Cell-Based Therapy for Mouse Acute Liver Injury. *Stem Cells Int*. 2019;2019:2403793. doi:10.1155/2019/2403793
145. Heeren J, Scheja L. Metabolic-associated fatty liver disease and lipoprotein metabolism. *Mol Metab*. 2021;50:101238. doi:10.1016/j.molmet.2021.101238

146. Zhang J-B. Therapeutic Effect of Prolyl Endopeptidase Inhibitor in High-fat Diet-induced Metabolic Dysfunction-associated Fatty Liver Disease. *J Clin Transl Hepatol.* **2023**;11(5):1035–1049. doi:10.14218/JCTH.2022.00110
147. Gao Z, Zhang C, Peng F, et al. Hypoxic mesenchymal stem cell-derived extracellular vesicles ameliorate renal fibrosis after ischemia-reperfusion injury by restoring CPT1A mediated fatty acid oxidation. *Stem Cell Res Ther.* **2022**;13(1):191. doi:10.1186/s13287-022-02861-9
148. Wei S, Li A, Zhang L, Du M. GROWTH AND DEVELOPMENT SYMPOSIUM: STEM AND PROGENITOR CELLS IN ANIMAL GROWTH: long noncoding RNAs in adipogenesis and adipose development of meat animals12. *J Anim Sci.* **2019**;97(6):2644–2657. doi:10.1093/jas/skz114
149. Grange C, Tritta S, Tapparo M, et al. Stem cell-derived extracellular vesicles inhibit and revert fibrosis progression in a mouse model of diabetic nephropathy. *Sci Rep.* **2019**;9(1):4468. doi:10.1038/s41598-019-41100-9
150. Komori A. Recent updates on the management of autoimmune hepatitis. *Clin Mol Hepatol.* **2021**;27(1):58–69. doi:10.3350/cmh.2020.0189
151. Covelli C, Sacchi D, Sarcognato S, et al. Pathology of autoimmune hepatitis. *Pathologica.* **2021**;113(3):185–193. doi:10.32074/1591-951X-241
152. Chung Y, Rahim MN, Graham JJ, Zen Y, Heneghan MA. An update on the pharmacological management of autoimmune hepatitis. *Expert Opin Pharmacother.* **2021**;22(11):1475–1488. doi:10.1080/14656566.2021.1895747
153. Martinon F, Mayor A, Tschopp J. The inflammasomes: guardians of the body. *Annu Rev Immunol.* **2009**;27:229–265. doi:10.1146/annurev.immunol.021908.132715
154. Vanaja SK, Rathinam VAK, Fitzgerald KA. Mechanisms of inflammasome activation: recent advances and novel insights. *Trends Cell Biol.* **2015**;25(5):308–315. doi:10.1016/j.tcb.2014.12.009
155. Liu X, Mi X, Wang Z, et al. Ginsenoside Rg3 promotes regression from hepatic fibrosis through reducing inflammation-mediated autophagy signaling pathway. *Cell Death Dis.* **2020**;11(6):454. doi:10.1038/s41419-020-2597-7
156. Joseph J. Serum Marker Panels for Predicting Liver Fibrosis - An Update. *Clin Biochem Rev.* **2020**;41(2):67–73. doi:10.33176/AACB-20-00002
157. Ponziani FR, Nicoletti A, Gasbarrini A, Pompili M. Diagnostic and therapeutic potential of the gut microbiota in patients with early hepatocellular carcinoma. *Ther Adv Med Oncol.* **2019**;11:1758835919848184. doi:10.1177/1758835919848184
158. Qiu B-F, Zhang G-Q. Effect of the transdifferentiation of BECs into myofibroblasts on the pathogenesis of secondary cholestatic hepatic fibrosis. *Exp Ther Med.* **2019**;17(4):2769–2776. doi:10.3892/etm.2019.7234
159. Bruno S, Pasquino C, Herrera Sanchez MB, et al. HLSC-Derived Extracellular Vesicles Attenuate Liver Fibrosis and Inflammation in a Murine Model of Non-alcoholic Steatohepatitis. *Mol Ther.* **2020**;28(2):479–489. doi:10.1016/j.ymthe.2019.10.016
160. Zhou J, Lin Y, Kang X, Liu Z, Zhang W, Xu F. microRNA-186 in extracellular vesicles from bone marrow mesenchymal stem cells alleviates idiopathic pulmonary fibrosis via interaction with SOX4 and DKK1. *Stem Cell Res Ther.* **2021**;12(1):96. doi:10.1186/s13287-020-02083-x
161. Wang B, Yao K, Huuskos BM, et al. Mesenchymal Stem Cells Deliver Exogenous MicroRNA-let7c via Exosomes to Attenuate Renal Fibrosis. *Mol Ther.* **2016**;24(7):1290–1301. doi:10.1038/mt.2016.90
162. Wang S, Li L, Liu T, Jiang W, Hu X. miR-19a/19b-loaded exosomes in combination with mesenchymal stem cell transplantation in a preclinical model of myocardial infarction. *Regener Med.* **2020**;15(6):1749–1759. doi:10.2217/rme-2019-0136
163. Guan Y, Yao W, Yi K, et al. Nanotheranostics for the Management of Hepatic Ischemia-Reperfusion Injury. *Small.* **2021**;17(23):e2007727. doi:10.1002/smll.202007727
164. Lee HM, Kim T, Choi HJ, et al. Influence of intraoperative oxygen content on early postoperative graft dysfunction in living donor liver transplantation: a STROBE-compliant retrospective observational study. *Medicine (Baltimore).* **2020**;99(21):e20339. doi:10.1097/MD.00000000000020339
165. Vald s S, Paredes SD, Garc a Carreras C, et al. S-Adenosylmethionine Decreases Bacterial Translocation, Proinflammatory Cytokines, Oxidative Stress and Apoptosis Markers in Hepatic Ischemia-Reperfusion Injury in Wistar Rats. *Antioxidants (Basel).* **2023**;12(8):4930.
166. van der Vlist EJ, Stoorvogel W, Arksteijn GJA, Wauben MHM. Fluorescent labeling of nano-sized vesicles released by cells and subsequent quantitative and qualitative analysis by high-resolution flow cytometry. *Nat Protoc.* **2012**;7(7):1311–1326. doi:10.1038/nprot.2012.065
167. Pospichalova V, Svoboda J, Dave Z, et al. Simplified protocol for flow cytometry analysis of fluorescently labeled exosomes and microvesicles using dedicated flow cytometer. *J Extracell Vesicles.* **2015**;4:25530. doi:10.3402/jev.v4.25530
168. Smith ZJ, Lee C, Rojalin T, et al. Single exosome study reveals subpopulations distributed among cell lines with variability related to membrane content. *J Extracell Vesicles.* **2015**;4:28533. doi:10.3402/jev.v4.28533
169. Hoffman AS. Hydrogels for biomedical applications. *Ann N Y Acad Sci.* **2001**;944:62–73.
170. Xu N, Xu J, Zheng X, Hui J. Preparation of Injectable Composite Hydrogels by Blending Poloxamers with Calcium Carbonate-Crosslinked Sodium Alginate. *ChemistryOpen.* **2020**;9(4):451–458. doi:10.1002/open.202000040
171. Okusha Y, Eguchi T, Tran MT, et al. Extracellular Vesicles Enriched with Moonlighting Metalloproteinase Are Highly Transmissible, Pro-Tumorigenic, and Trans-Activates Cellular Communication Network Factor (CCN2/CTGF): CRISPR against Cancer. *Cancers (Basel).* **2020**;12(4):548.
172. Kalluri R, LeBleu VS. The biology, function, and biomedical applications of exosomes. *Science.* **2020**;367:6478. doi:10.1126/science.aau6977
173. Li F, Zhang J, Yi K, et al. Delivery of Stem Cell Secretome for Therapeutic Applications. *ACS Appl Bio Mater.* **2022**;5(5):2009–2030. doi:10.1021/acsbm.1c01312
174. Wan T, Zhong J, Pan Q, Zhou T, Ping Y, Liu X. Exosome-mediated delivery of Cas9 ribonucleoprotein complexes for tissue-specific gene therapy of liver diseases. *Sci Adv.* **2022**;8(37):eabp9435. doi:10.1126/sciadv.abp9435

Journal of Inflammation Research

Dovepress

Publish your work in this journal

The Journal of Inflammation Research is an international, peer-reviewed open-access journal that welcomes laboratory and clinical findings on the molecular basis, cell biology and pharmacology of inflammation including original research, reviews, symposium reports, hypothesis formation and commentaries on: acute/chronic inflammation; mediators of inflammation; cellular processes; molecular mechanisms; pharmacology and novel anti-inflammatory drugs; clinical conditions involving inflammation. The manuscript management system is completely online and includes a very quick and fair peer-review system. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Submit your manuscript here: <https://www.dovepress.com/journal-of-inflammation-research-journal>