

Antibody-induced secondary treatment failure in a patient treated with botulinum toxin type A for glabellar frown lines

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Abstract: Botulinum toxin type A (BTX-A) preparations are widely used nonsurgical treatments for facial wrinkles. Higher doses of BTX-A are also used for therapeutic purposes in the treatment of conditions involving increased muscle tone, such as cervical dystonia. The phenomenon of antibody-induced treatment failure is well known in the therapeutic setting, but reports are also emerging following cosmetic use of BTX-A. We describe the case of a 41-year-old female nurse who developed secondary treatment failure during 6 years of BTX-A treatment for glabellar lines. After a good response to the first BTX-A injection, the intensity and duration of effect decreased after subsequent treatments. Antibody tests revealed a high titer of neutralizing anti-BTX-A antibodies. This case shows secondary treatment failure due to the production of neutralizing antibodies following administration of BTX-A formulations for cosmetic purposes and demonstrates that immunogenicity of BTX-A preparations is an important consideration, even in the cosmetic setting.

Keywords: botulinum toxin type A, neutralizing antibodies, antibody-induced treatment failure

Introduction

Botulinum toxin type A (BTX-A) preparations are commonly used in therapeutic and cosmetic applications with great success. However, after initial good responses, therapy can subsequently fail either partially or completely (secondary therapy failure) due to a number of causes, including inadequate dosage, injection of inappropriate muscles, and development of BTX-A neutralizing antibodies.^{1,2} Antibody-induced treatment failure following treatment with BTX-A for therapeutic purposes has been reported to range from 4% to 10% of patients treated^{3,4} and to decrease to 1%–6% after the foreign protein load of the preparation used is reduced.^{5,6} The risk of developing antibody-induced treatment failure has been shown to increase with short injection intervals and high injected doses.^{2,7} Despite lower BTX-A dosages being used in cosmetic applications compared with therapeutics, there are now emerging reports of antibody-induced treatment failure in facial esthetics.^{8,9}

Case report

Here we report the case of a 41-year-old Caucasian woman who had been receiving BTX-A preparations for the treatment of glabellar lines for 6 years (Table 1). She was initially treated in 2004 with a commercially available BTX-A preparation, abobotulinumtoxinA (Dysport[®], Ipsen Ltd, Slough, UK). The effects of treatment lasted for 3–4 months. However, following her next treatment with

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Table 1 Treatment history

Date	BTX-A preparation	Dosage	Area treated	Duration of effect	Approximate treatment interval
2004	AbobotulinumtoxinA	N/A	Glabellar	First treatment: 12–16 weeks; Second treatment: maximum 8 weeks	6 months
2005	AbobotulinumtoxinA	N/A	Glabellar	4–8 months	6 months
2006	AbobotulinumtoxinA	N/A	Glabellar	3–4 weeks	6 months
2007	AbobotulinumtoxinA	N/A	Glabellar	3–4 weeks	6 months
2008	AbobotulinumtoxinA	N/A	Glabellar	3–4 weeks	6 months
February 11, 2009	OnabotulinumtoxinA	28 U	Glabellar	Patient complained of incomplete treatment	N/A
February 25, 2009	OnabotulinumtoxinA	9 U	Glabellar, periorbital	2–3 weeks	1.5 months
May 28, 2009	OnabotulinumtoxinA	10 U	Glabellar, periorbital	2–3 weeks	3 months
August 26, 2009	IncobotulinumtoxinA	20 U	Glabellar, periorbital	3–4 weeks	3 months
December 24, 2009	IncobotulinumtoxinA	22 U	Glabellar, periorbital	3–4 weeks	4 months
January 19, 2010	IncobotulinumtoxinA	44 U	Glabellar, periorbital	3–4 weeks	1 month

Abbreviations: N/A, not available; BTX-A, botulinum toxin type A.

abobotulinumtoxinA in the glabellar region, the duration of effect was reduced to 8 weeks. From 2005 to 2008, prior to presentation at our clinic, the patient received further injections of abobotulinumtoxinA in the glabellar area twice yearly and reported that the duration of effect subsequently diminished to a maximum effect of 3–4 weeks' duration. From the beginning of 2009, we treated this patient with other BTX-A preparations, first with onabotulinumtoxinA (Botox[®]/Vistabel[®], Allergan, Irvine, CA), and more recently with incobotulinumtoxinA (Xeomin[®]/Bocouture[®], Merz Pharmaceuticals GmbH, Frankfurt, Germany).

The first treatment in our clinic was 28 U of onabotulinumtoxinA in the glabellar area, but the treatment was sub-optimal and the patient returned approximately 2 weeks later, when she received an additional 9 U of onabotulinumtoxinA. For this second treatment and subsequent treatments, BTX-A was injected in the periorbital region as well as the glabellar region at the patient's request. The patient reported the duration of effect to be 2–3 weeks. Three months later, the patient received one further treatment in the glabellar and periorbital areas, with 10 U onabotulinumtoxinA, a lower dose than usual, as requested by the patient. However, the patient was still dissatisfied with the treatment outcome and duration of effect. Therefore, we changed to administration of incobotulinumtoxinA at a higher dose (20 U) into the glabellar and periorbital regions, but the duration of effect was only 3–4 weeks. Indeed, two subsequent injections of incobotulinumtoxinA at higher doses (22 U and 44 U) also failed to elicit a response of longer duration. The clinical photograph taken approximately 1 month after the final injection shows no remaining effect of neurotoxin (Figure 1C).

Therefore, we considered the possibility that the patient had neutralizing anti-BTX-A antibodies. This seemed likely since neutralizing anti-BTX-A antibodies would not be overcome by switching to another BTX-A formulation, and high antibody titers could prevent a response even to larger doses.

In December 2009, the patient's serum was tested for the presence of neutralizing anti-BTX-A antibodies at a specialized laboratory (Toxogen GmbH, Hannover, Germany) using an *in vitro* mouse hemidiaphragm assay.¹⁰ The patient was positive for a high titer of neutralizing antibodies, suggesting that the cause of the secondary therapy failure experienced by this patient was neutralizing anti-BTX-A antibodies.

In this example, following treatment with a complexing protein-containing BTX-A formulation, the patient developed neutralizing antibodies and subsequently did not respond to any of the BTX-A formulations tested. Resistance that develops following the cosmetic use of BTX-A may impact on the success of any subsequent therapeutic BTX-A treatment (eg, for poststroke spasticity) that the patient may need in the future. It also limits further esthetic use of BTX-A. Some clostridial complexing proteins have been found to enhance antibody production,¹¹ and may therefore increase the risk of neutralizing anti-BTX-A antibodies. Indeed, reducing the foreign protein load of the early preparation of onabotulinumtoxinA decreased the risk of neutralizing anti-BTX-A antibodies.⁴ A further reduction in the protein load of BTX-A injections, for instance by using highly purified formulations,¹² may be beneficial in patients receiving several cycles of BTX-A injections.

For this patient, switching to a botulinum toxin type B (BTX-B) preparation was not considered because the data show that BTX-B has a greater diffusion potential¹³ and a



Figure 1 Clinical photographs taken at maximum frown. **(A)** Patient prior to injection with incobotulinumtoxinA on December 24, 2009, after developing nonresponsiveness to preparations containing botulinum toxin complex. **(B)** Patient following injection with incobotulinumtoxinA on January 19, 2010, after the patient developed nonresponsiveness to preparations containing botulinum toxin complex. **(C)** Patient on February 25, 2010, about 1 month after the final botulinum toxin type A injection on January 19, 2010.

highly-acidic pH,¹² and therefore is associated with more side effects.¹² Also, the duration of effect seen with BTX-B is not maintained for as long as that observed with BTX-A.¹⁴ Currently, there is no BTX-B preparation that is approved for an esthetic indication and, consequently, it is not commonly used for this purpose.

Although initial reports stated that less than 1% of patients develop neutralizing anti-BTX-A antibodies in esthetics,^{15–17} this case, and others like it,^{8,9} highlight the fact that immunogenicity of BTX-As and neutralizing anti-BTX-A antibodies are still important issues. Indeed, recent reports monitored BTX-A neutralizing antibodies in patients receiving 1–10 cycles of BTX-A injections, with few patients achieving the maximum number of cycles.^{15–17} Therefore, the long-term effects of BTX-A on neutralizing anti-BTX-A antibody development still need to be investigated to estimate accurately the incidence and importance of neutralizing

anti-BTX-A antibodies in esthetics. This is of particular significance given the widespread offlabel use of BTX-A for cosmetic indications,^{18,19} and the administration of BTX-A preparations by nonmedically trained individuals.

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Disclosures

EKB is a consultant for Merz Pharmaceuticals GmbH. GS has no conflicts of interest to disclose in this work.

References

- Jankovic J, Schwartz K. Response and immunoresistance to botulinum toxin injections. *Neurology*. 1995;45(9):1743–1746.
- Dressler D. Clinical presentation and management of antibody-induced failure of botulinum toxin therapy. *Mov Disord*. 2004;19 Suppl 8: S92–S100.
- Greene P, Fahn S, Diamond B. Development of resistance to botulinum toxin type A in patients with torticollis. *Mov Disord*. 1994;9(2): 213–217.
- Jankovic J, Vuong KD, Ahsan J. Comparison of efficacy and immunogenicity of original versus current botulinum toxin in cervical dystonia. *Neurology*. 2003;60(7):1186–1188.
- Muller K, Mix E, Adib Saberi F, Dressler D, Benecke R. Prevalence of neutralizing antibodies in patients treated with botulinum toxin type A for spasticity. *J Neural Transm*. 2009;116(5):579–585.
- Brin MF, Comella CL, Jankovic J, Lai F, Naumann M. Long-term treatment with botulinum toxin type A in cervical dystonia has low immunogenicity by mouse protection assay. *Mov Disord*. 2008;23(10): 1353–1360.
- Lange O, Bigalke H, Dengler R, Wegner F, deGroot M, Wohlfarth K. Neutralizing antibodies and secondary therapy failure after treatment with botulinum toxin type A: Much ado about nothing? *Clin Neuropharmacol*. 2009;32(4):213–218.
- Borodic G. Immunologic resistance after repeated botulinum toxin type A injections for facial rhytides. *Ophthalm Plast Reconstr Surg*. 2006;22(3):239–240.
- Lee S-K. Antibody-induced failure of botulinum toxin type A therapy in a patient with masseteric hypertrophy. *Dermatol Surg*. 2007; 33(1 Spec No.):S105–S110.
- Göschel H, Wohlfarth K, Frevert J, Dengler R, Bigalke H. Botulinum A toxin therapy: Neutralizing and nonneutralizing antibodies – therapeutic consequences. *Exp Neurol*. 1997;147(1):96–102.
- Lee JC, Yokota K, Arimitsu H, et al. Production of anti-neurotoxin antibody is enhanced by two subcomponents, HA1 and HA3b, of Clostridium botulinum type B 16S toxin-haemagglutinin. *Microbiology*. 2005;151(Pt 11):3739–3747.
- Carruthers A, Carruthers J. Botulinum toxin products overview. *Skin Therapy Lett*. 2008;13(6):1–4.
- Flynn TC, Clark RE 2nd. Botulinum toxin type B (Myobloc) versus botulinum toxin type A (Botox) frontalis study: Rate of onset and radius of diffusion. *Dermatol Surg*. 2003;29(5):519–522.
- Flynn TC. Botulinum toxin: Examining duration of effect in facial aesthetic applications. *Am J Clin Dermatol*. 2010;11(3):183–199.
- Kawashima M, Harii K. An open-label, randomized, 64-week study repeating 10- and 20-U doses of botulinum toxin type A for treatment of glabellar lines in Japanese subjects. *Int J Dermatol*. 2009;48(7):768–776.

16. Lawrence I, Moy R. An evaluation of neutralizing antibody induction during treatment of glabellar lines with a new US formulation of botulinum neurotoxin type A. *Aesthet Surg J*. 2009;29(6 Suppl):S66–S71.
17. Monheit G, Carruthers A, Brandt F, Rand R. A randomized, double-blind, placebo-controlled study of botulinum toxin type A for the treatment of glabellar lines: Determination of optimal dose. *Dermatol Surg*. 2007;33(1 Spec No.):S51–S59.
18. Cheng CM, Chen JS, Patel RP. Unlabeled uses of botulinum toxins: A review, Part 1. *Am J Health Syst Pharm*. 2006;63(2):145–152.
19. Cheng CM, Chen JS, Patel RP. Unlabeled uses of botulinum toxins: A review, Part 2. *Am J Health Syst Pharm*. 2006;63(3):225–232.

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