

Diminazene aceturate: an antibacterial agent for Shiga-toxin-producing *Escherichia coli* O157:H7

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Abstract: The aim of this study was to investigate the bacteriostatic and bactericidal effects of diminazene aceturate (DA) against five strains of pathogenic bacteria and two strains of nonpathogenic bacteria. The results showed that 5 µg/mL of DA suppressed the growth of pathogenic *Escherichia coli* by as much as 77% compared with the controls. Enterohemorrhagic *E. coli* EDL933 (an *E. coli* O157:H7 strain) was the most sensitive to DA with a minimum inhibitory concentration of 20 µg/mL. Additional investigations showed that DA induced the highest level of intracellular reactive oxygen species in EDL933. A positive correlation between the reactive oxygen species levels and DA concentration was demonstrated. DA (5 µg/mL) was also a potent uncoupler, inducing a stationary phase collapse (70%–75%) in both strains of *E. coli* O157:H7. Further investigation showed that the collapse was due to the NaCl:DA ratio in the broth and was potassium ion dependent. A protease screening assay was conducted to elucidate the underlying mechanism. It was found that at neutral pH, the hydrolysis of H-Asp-pNA increased by a factor of 2–3 in the presence of DA, implying that DA causes dysregulation of the proton motive force and a decrease in cellular pH. Finally, a commercial verotoxin test showed that DA did not significantly increase toxin production in EDL933 and was a suitable antibacterial agent for Shiga-toxin-producing *E. coli*.

Keywords: *E. coli* O157, Berenil[®], ROS, reversible, protease, enterohemorrhagic

Introduction

According to recent reports from the Center for Disease Control and Prevention, there were eleven multistate outbreaks of Shiga-toxin (Stx)-producing *Escherichia coli* (STEC) in the US from 2011 to 2014 with six of them attributed to *E. coli* O157:H7.¹ Most *E. coli* O157:H7 strains produce Stx type 2; its subunits bind to the surface of intestinal cells, and its A subunits enter and turn off protein synthesis by disrupting the large ribosomal unit.^{2–4} Bloody diarrhea is a hallmark of symptomatic cases. STEC-producing bacteria also cause hemolytic uremic syndrome (HUS). HUS is a disease characterized by hemolysis, thrombocytopenia, and acute kidney injury, although other organs may be involved. Most cases of HUS are due to infection with STEC.^{5–10} The use of antibiotics is controversial. It is recommended not to use antibiotics to treat STEC infection (*E. coli* O157) because of either no benefit or potential increase in the risk of developing HUS.^{11,12} Stx toxicity and the acute inflammatory response raised by the host determine the development of HUS. Currently, there is no specific therapy to control Stx damage. The pathogenic role of reactive oxygen species (ROS) on endothelial injury is well documented.^{13,14} Therefore, it is not surprising that antibiotics, bactericidal agents, and nanomaterials that utilize ROS-dependent mechanisms to kill pathogenic Stx bacteria increase the potential risk of HUS in certain individuals. In addition, reports from the microbiological community show that extracellular ROS

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can induce Stx production,¹⁵ which in turn induces ROS production in the host tissue.

One way to counter ROS damage to host tissue is to administer an antioxidant with an antibiotic. This was eloquently shown in the work by Kalghatgi et al¹⁶ in which *N*-acetyl-L-cysteine was used in combination with tetracycline (growth inhibitor) to inhibit the growth of *E. coli* and minimize ROS mammalian cell damage. Whether tetracycline induces Stx 1 or Stx 2 production, STEC O157 via ROS-dependent or ROS-independent pathways has not been reported. There are many old and new antioxidant/antibacterial compounds such as *N*-acetyl-L-cysteine and *S*-ethyl-L-cysteine that can be used to neutralize ROS.¹⁷ Another old antibacterial compound is diminazene aceturate (DA; active compound in Berenil), which is a potent inhibitor of trypsin-like enzymes¹⁸ and useful in the treatment of many diseases such as coronary and cerebral infarction, vascular clotting, arthritis, tumor cell invasion, and pancreatitis.^{19–23} Recent toxicity studies by Cappoen et al showed that adding DA to mammalian cell cultures did not induce ROS production.²⁴ DA is reported to selectively inhibit the growth of commensal *E. coli* by binding to the ribosomal subunits.²⁵ However, the commensal population of *E. coli* should be maintained to prevent enterohemorrhagic *E. coli* gaining a competitive foothold in the ileum and lower intestine. Moreover, there are many contradictory reports regarding the toxicity and clinical benefits of DA. For example, DA is classified as a topoisomerase II (Top II)²⁶ poison in eukaryotes but improves the remodeling of damaged cardiac tissue which is dependent on Top II β activity in muscle cells.²⁷

Given the numerous reports on STEC^{28–31} and the various factors (genetic and culturing conditions) that influence Stx induction, a potential antibacterial agent should exhibit two of the following three properties: first, bactericidal agents, antibiotics, and nanomaterials must inhibit and kill the pathogen (duality required) at several stages of the fission cycle. Second, the compound should not significantly induce either Stx 1 or Stx 2 production. Third, the compound must not induce ROS in mammalian cells.

The aim of this work was to investigate whether DA exhibited the first two properties. This was performed by studying the effects and antibacterial mechanisms of DA on the growth and viability of different strains of STEC O157:H7, commensal, and other pathogenic bacteria. The effects of DA on intracellular and extracellular ROS levels in different strains of *E. coli* O157:H7 were measured by ROS probe and peroxidase assay. Physical changes in cell morphology and membrane integrity after exposure to DA were imaged with scanning electron microscopy (SEM).

The effects of DA and common antibiotics on Stx production were measured with a commercial verotoxin (VT) test.

Materials and methods

Bacterial strains and growth cultures

E. coli O157:H7 (Korean Agricultural Culture Collection [KACC] 11598, EDL933) isolated from the 1982 outbreak of hemorrhagic colitis in Michigan was obtained from the KACC (South Korea) as were *E. coli* strains (KACC 10005 and K-12). *E. coli* O157:H7 (KACC 10765, EDL931) was donated by Professor Lee's laboratory. *Listeria monocytogenes* (ATCC 19115) and *Enterococcus faecalis* (ATCC 29212) were purchased from the American Type Culture Collection (ATCC, Manassas, VA, USA). *Staphylococcus aureus* (KACC 13252, STA) was a gift from Professor Seong Soo A An's laboratory. All of these strains were maintained in 15% glycerol stock at -80°C and reactivated in proper broth before culturing.

The effects of DA on bacterial growth patterns

For the negative controls, K-12, EDL931, and EDL933 (10^6 CFU/mL) were grown in Luria–Bertani (LB) broth (Sigma-Aldrich Co., St Louis, MO, USA). *L. monocytogenes*, *E. faecalis*, and *S. aureus* (10^6 CFU/mL) were grown in nutrient broth (NB; Sigma-Aldrich Co.) (pH 7.0) at 34°C . *L. monocytogenes*, *S. aureus*, and *E. faecalis* were grown in NB media, because they grew poorly in LB broth. *E. coli* strains were grown in LB broth, because they grew well. K-12 and *E. coli* 10005 (EC10005) were chosen as model commensal strains. The optical density (OD; 600 nm) value was recorded every 2 hours using an ultraviolet (UV)–visible spectrophotometer for 24 hours; all the measurements were performed in triplicate; and the final results were averaged. This process was repeated at 37°C and 42°C .

Minimum inhibitory concentrations

In the sample groups, DA was added to the broth at the beginning with variable concentrations at 2 $\mu\text{g/mL}$, 5 $\mu\text{g/mL}$, 10 $\mu\text{g/mL}$, 20 $\mu\text{g/mL}$, 40 $\mu\text{g/mL}$, 60 $\mu\text{g/mL}$, 80 $\mu\text{g/mL}$, 120 $\mu\text{g/mL}$, and 150 $\mu\text{g/mL}$. Sample groups were incubated at 37°C and the growth was recorded using a UV–visible spectrophotometer (600 nm) every 2 hours; all the measurements were run in duplicate.

The effect of administering DA in the log phase

The incubation method was identical to the procedure mentioned earlier. For the sample groups, a fixed concentration

of DA was added to each strain of bacteria in the mid-log phase (OD value 0.45) at 34°C. The change in the OD was subsequently recorded every 2 hours; all the measurements were performed in triplicate. This process was repeated at 37°C and 42°C.

The effects of LB broth pH on the antibacterial activity of DA

K-12, EDL931, and EDL933 were grown in acidic LB/DA broth (pH 5.0) at 34°C for 8 hours. This process was repeated using neutral LB/DA broth. After such time, the bacteria were centrifuged at 4000 rpm for 5 minutes and then resuspended in either fresh acidic or neutral LB/DA broth. Cell density was measured every 2 hours over a 24-hour period. Control was set by changing the broth with a new LB broth (pH 7.0) with 5 µg/mL DA at 8 hours.

Reactive oxidative species assay

K-12, EDL931, and EDL933 were grown in LB broth (control) and LB-broth-containing DA (5 µg/mL) at 34°C for 6 hours and 12 hours (DA was added to LB broth in the mid-log phase). Three samples were taken of each strain, harvested by centrifugation, and then washed twice with Hanks' balanced salt solution (HBSS) as reported earlier.³² All samples were resuspended in HBSS and then diluted with the same solution until each sample contained ~10⁸ CFU/mL. A stock solution of the ROS probe, 2',7'-dichlorofluorescein diacetate (DCFHDA), was dissolved in dimethyl sulfoxide at a concentration of 1 mM and stored in dark at -20°C. In an eppendorf tube, 1 mL of suspended bacteria cells from each sample was then mixed with 1 mL of 20 mM DCFHDA/HBSS. Each tube was then wrapped in aluminum foil and placed in an automatic shaker (150 rpm) at 37°C for 2 hours. Cells were then collected by centrifugation at 5,000 rpm for 2 minutes and then washed twice with fresh medium to remove the free probe. The fluorescent intensity of the oxidized product, namely dichlorofluorescein, was detected at an emission wavelength of 535 nm (excitation wavelength [Ex] 488 nm) by a fluorescence spectrophotometer (F-4500; Hitachi Ltd., Tokyo, Japan). The fluorescent intensity of DA-treated bacteria was then compared with untreated bacteria, and the level of intracellular ROS was estimated.

Whole cell and lysate peroxidase activity

For the cell peroxidase test, K-12, EDL931, and EDL933 were grown in LB broth with and without 2% glucose at 37°C in a 130 rpm shaker for 6 hours. Then, 1 mL samples were taken of each bacterium and centrifuged (4,000 rpm for 5 minutes). In accordance with earlier reports,³³ the supernatant was

removed and the pellet was washed and resuspended with phosphate-buffered saline (PBS) solution. Cell concentrations were read at 600 nm, then diluted with PBS until an OD value of 0.1 (10⁸ CFU/mL) was achieved. Aliquots of 20 µL of cell solution were dispensed into the microtiter plate, and 3,3',5,5'-tetramethylbenzidine (TMB) solution was added. The plate was incubated at 37°C for 30 minutes, and the absorbance of the wells read at 405 nm. The procedure was repeated using cells grown in LB broth containing 2% glucose in the presence of increasing concentrations of DA (control: 0 µg/mL; samples: 1 µg/mL, 2 µg/mL, and 5 µg/mL).

For the cell lysis peroxidase assay, 1 mL of a fixed concentration of cells was removed from the broth, centrifuged, washed in deionized water, and then resuspended with PBS. Lysozyme was added to each sample (2 µg/mL); the samples were placed in an ice bath for 15 minutes and then sonicated for 2 minutes (ice bath) to help the cells to break. Samples were then washed with cold PBS and then diluted with PBS to an OD value of 0.1 at 600 nm before use. Then, 20 µL aliquots of lysate solution were added to a series of microwells and incubated with TMB solution at 37°C for 30 minutes. The reaction was stopped by H₂SO₄, and the absorbance was measured at 405 nm.

Two stock solutions of horseradish peroxidase (HRP; 20 µg/mL) were prepared in PBS (pH 7.4). A series of phosphate-buffered HRP solutions containing 0.1–5 µg/mL of enzyme were then prepared and left to equilibrate at 37°C for 1 hour. Aliquots of 20 µL of HRP solutions were then distributed into 20 µL wells (0.5 mL capacity) to which 80 µL of either TMB (positive control) or TMB and DA (5 µg/mL, well concentration) was added. The plate was then placed in a 37°C incubator and shaken at 80 rpm for 30 minutes. After incubation, 50 µL of 2 N H₂SO₄ (aq) was added to stop the reaction and the OD was measured at 450 nm using a Perkin Elmer plate reader (Victor³ spectrophotometer; PerkinElmer Inc., Waltham, MA, USA).

Examination of cell morphology by SEM

Samples of DA-treated *E. coli* O157:H7 (10⁸ CFU/mL) were transferred to fresh glass slides, and then fixed using 10% glutaraldehyde/Tris-buffered solution (0.1 M, pH 7.5) for 1 hour at room temperature. Samples were dried to critical point with CO₂, spluttered, coated, and then analyzed under field emission (FE)-SEM (SU9000 UHR FE-SEM; Hitachi High Technologies, Tokyo, Japan). Atomic force microscopy images of live and dead bacteria were obtained using an atomic force microscope (Nanoscope 111; Digital Instruments, Oyster Bay, NY, USA) in tapping mode (cantilever length: 125 µm) with Veeco RTSEP probes at a

scan rate of 1 Hz and scan size of 5 μm . Average height and surface roughness values for live and dead bacteria were collected and analyzed using Nanoscope software.

The effect of KCl on the viability of EDL933

In LB broth, 1 M stock KCl solution was prepared. To ten 50 mL of sterile polypropylene tubes, ~40 mL of EDL933 (10^6 CFU/mL) LB broth was added. The experiment consisted of two sample groups: A (tubes 1–4) and B (tubes 5–7). Tube 1 contained EDL933 broth solution (negative control). Aliquots of DA were added to tubes 2 (positive control), 3, and 4 giving a final concentration of 5 $\mu\text{g/mL}$ in each tube. To tube 3 (50 mM KCl), KCl broth solution was added (time zero). Tubes 1–4 were placed in a 37°C incubator; after 9 hours of growth, an aliquot of KCl broth solution was then added to tube 4 (50 mM KCl).

For sample group B, tube 5 was the negative control. To tube 6, a KCl aliquot was added (50 mM KCl). Tubes 5, 6, and 7 were then placed in a 37°C incubator. After 6 hours, DA (5 $\mu\text{g/mL}$) was added to tubes 6 and 7. To tube 7, an aliquot of KCl broth solution was added after 9 hours of growth. Sample groups A and B were cultured for 24 hours, and the OD was recorded every 2 hours. All the samples were performed in triplicate.

Protease screening assay

Bacterial strains, K-12, EDL933, and EDL931 were grown separately in LB broth for 6 hours at 37°C. A total of 21 para-nitroaniline (pNA) substrates were purchased from Bachem Americas, Inc. (Torrance, CA, USA) (H-Gly-Arg- β NA-HCl, H-Arg-Arg- β NA-3HCl, H-Gly-Phe-pNA, H-Lys-Ala- β NA, Ac-Glu-pNA, AC-Asp-pNA, H-Glu-Gly-Arg-pNA-acetate salt, H-Gly-Pro- β NA, H-Asp-pNA-HCl, Suc-Ala-Ala-Ala-pNA, Suc-Ala-Ala-Pro-Glu-pNA, Suc-Gly-Gly-Phe-pNA, Suc-Ala-Phe-Pro-Phe-pNA, Suc-Ala-Ala-Pro-Arg-pNA, Suc-Gly-Gly-Phe-pNA, H-Phe-pNA, Sar-Pro-Arg-pNA, H-D-Ile-Phe-Lys-pNA-salt, Z-Tyr-Lys-Arg-pNA-2TFA, Tos-Gly-Pro-Lys-pNA, MeOSuc-Ala-Ala-Pro-Met-pNA, and Z-Arg-pNA). A stock solution of each substrate (20 mM) was prepared in dimethyl sulfoxide and refrigerated until use. Dilution buffers, sodium citrate (0.1 M; pH 4 and 5), sodium phosphate (0.1 M, pH 6.0), and saline Tris buffer (0.1 M, pH 7.4) were prepared in deionized water.

From each broth, 1 mL of bacterial suspension was removed and centrifuged at 130 rpm for 5 minutes. The supernatant was removed, and the cells were diluted to 10^8 CFU/mL in pH buffer. Then, 40 μL of cells (10^8 CFU/mL), 50 μL of

buffer (pH 4), and 10 μL of pNA substrate (2 mM) were added to each well; all the samples were performed in triplicate. The reaction was left to run overnight at 37°C, and the production of pNA was read at 405 nm the next morning. The procedure was repeated using pH buffers 5, 6, and 7 and pH buffers 5, 6, 7 containing 5% glucose.

VT test

In six separate tubes (50 mL), EDL933 was grown in LB broth at 37°C until an OD value of 0.35 was obtained. To tube 1, no antibacterial agent was added (negative control), to tubes 2–6, 0.5 $\mu\text{g/mL}$ of Mitomycin, 1 $\mu\text{g/mL}$ of Ampicillin, 5 $\mu\text{g/mL}$ of DA, 5 $\mu\text{g/mL}$ of Novobiocin, and 5 $\mu\text{g/mL}$ of Pentamidine were added, respectively; all tubes were then incubated at 37°C for 20 hours. OD values of the broths were recorded on a UV spectrometer. The tubes were then centrifuged at 2,000 rpm for 10 minutes; 200 μL of the supernatant was dispensed onto the test surface in accordance with manufacturer's instructions (Duopath Verotoxins; EMD Millipore, Billerica, MA, USA). The test can detect both VT1 and VT2. Band intensities (obtained by Kodak Image Station 4000MM PRO) were converted into peak images via Free ImageJ analysis software. The amounts of VTs produced were estimated by calculating the cumulative area of the peaks (OriginLab Corporation, Northampton, MA, USA).

Results

The effects of DA on the growth of pathogenic and nonpathogenic bacteria

Figure 1A–D shows the effect of DA (5 $\mu\text{g/mL}$) in LB broth on the growth of different strains of bacteria at a fixed temperature (34°C). When compared with the negative controls, DA suppressed the growth of K-12 the most (–78%), followed by EDL933 (–75%), EDL931 (–72%), and EC10005 (–35%). DA/NB broth did not significantly suppress the growth of neither *L. monocytogenes* and *S. aureus* nor *E. faecalis*. Figure 1E shows the effects of temperature on the percentage population of different strains of *E. coli* grown for 24 hours in DA/LB (5 $\mu\text{g/mL}$) broth compared with the control. An 8°C rise in temperature from 34°C to 42°C increased the percentage populations of K-12 by 46%, EDL931 and EC10005 by 9%. EDL933 samples grown for 24 hours (Table S1) in 2 $\mu\text{g/mL}$ and 20 $\mu\text{g/mL}$ of DA/LB broth revealed similar OD values of 0.182 and 0.168 after 24 hours of growth implying that DA not only inhibits EDL933 growth but also kills the bacteria during the fission cycle.

The minimum inhibitory concentrations for each strain of bacteria are shown in Table 1. EDL933 and K-12 strains of

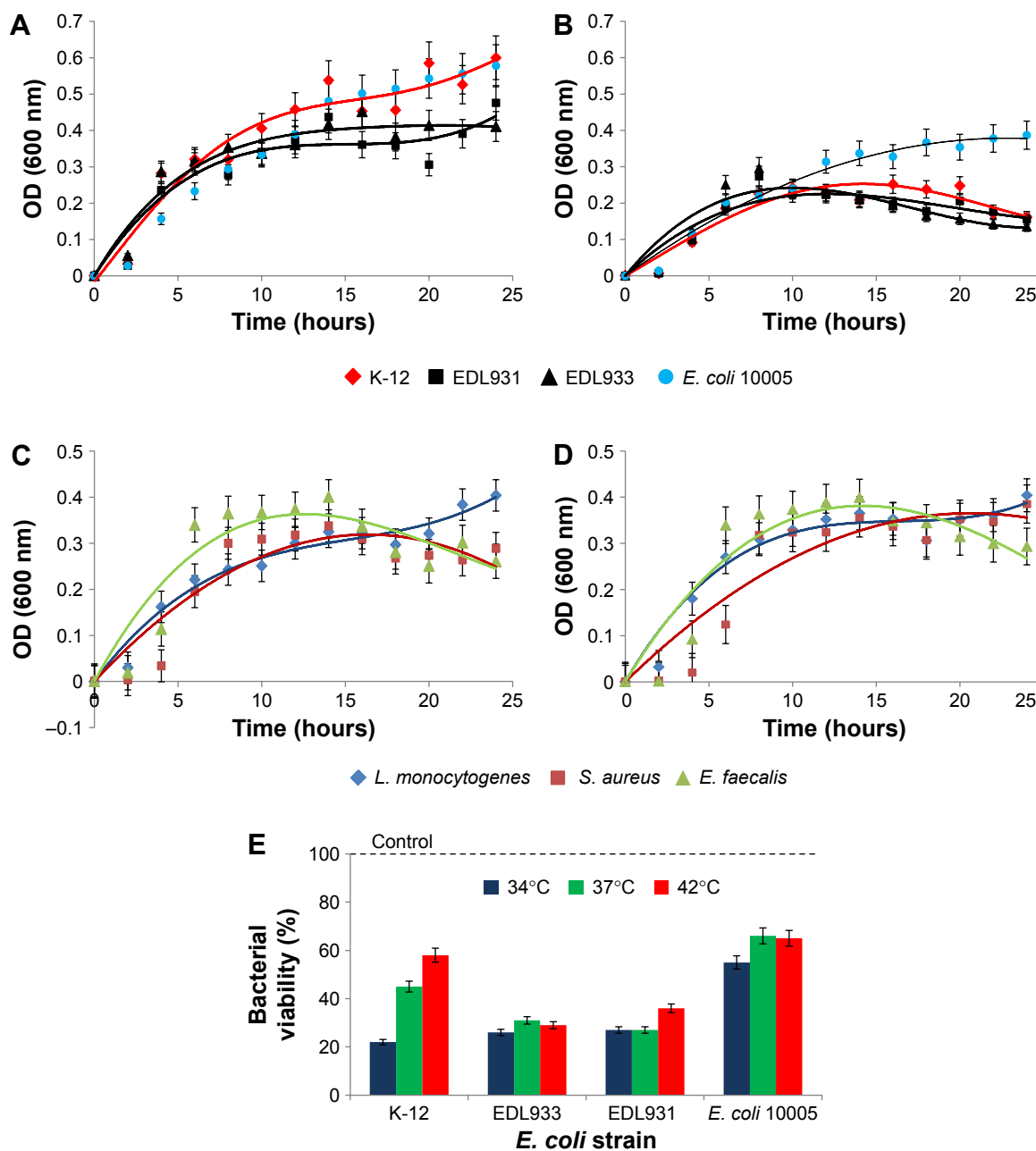


Figure 1 The effects of DA on the growth patterns of pathogenic and nonpathogenic strains of bacteria grown at 34°C. **Notes:** (A) K-12, EDL933, EDL931, and EC10005 grown with no DA. (B) K-12, EDL933, EDL931, and *E. coli* 10005 grown with DA. (C) *L. monocytogenes*, *S. aureus*, and *E. faecalis* grown with no DA. (D) *L. monocytogenes*, *S. aureus*, and *E. faecalis* grown with DA. (E) The effects of temperature on the population of different strains of *E. coli* grown for 24 hours in DA (5 mg/L)/LB broth. Bars represent the standard error of the mean of three independent experiments ($P < 0.01$). **Abbreviations:** DA, diminazene aceturate; *E. coli*, *Escherichia coli*; *E. faecalis*, *Enterococcus faecalis*; *L. monocytogenes*, *Listeria monocytogenes*; OD, optical density; *S. aureus*, *Staphylococcus aureus*.

Table 1 DA concentrations required to inhibit the growth for a minimum of 14 hours at 37°C (n=3)

Bacteria	K-12	EDL931	EDL933	<i>E. coli</i> 10005	<i>L. monocytogenes</i>	<i>S. aureus</i>	<i>E. faecalis</i>
MIC (µg/mL)	125	40	20	70	120	150	150

Abbreviations: DA, diminazene aceturate; *E. coli*, *Escherichia coli*; *E. faecalis*, *Enterococcus faecalis*; *L. monocytogenes*, *Listeria monocytogenes*; MIC, minimum inhibitory concentration; *S. aureus*, *Staphylococcus aureus*.

E. coli were the most and least sensitive to DA exhibiting values of 20 $\mu\text{g}/\text{mL}$ and 120 $\mu\text{g}/\text{mL}$, respectively. *L. monocytogenes*, *S. aureus*, and *E. faecalis* were highly resistant to DA exhibiting minimum inhibitory concentrations $>120 \mu\text{g}/\text{mL}$. Such concentrations are toxic to humans; therefore, these strains were excluded from further testing.^{34,35}

The effect of administering DA in the log phase

In the next stage of this study, we examined the effect of adding DA (5 $\mu\text{g}/\text{mL}$) to growing bacteria (mid-log phase) to see whether DA could kill as well as inhibit the growth of different strains of *E. coli* O157 in accordance with the first part of the criteria. Figure 2A shows the impact of adding DA in mid-log phase on the growth patterns of K-12 and both strains of *E. coli* O157:H7. The growth pattern of K-12 is unaffected by the addition of DA in the mid-log phase. However, 6–8 hours after the addition of DA, the growth curves for both strains of *E. coli* O157:H7 collapse with mortality rates ranging from 70% to 75%. The experiment was then repeated at 37°C and 42°C, and the results are shown in Figure 2B. An 8°C rise in temperature from 34°C to 42°C improved the population viability of K-12 by 23.4%, EDL931 by 26.6%, and EDL933 by 2.8%. The effects of broth pH on the antibacterial activity of DA are shown in Figure S1. The next part of the investigation aims to elucidate the mechanisms by which DA inhibits the growth and induces the collapse in populations of EDL933 and EDL931.

DA inhibits the growth of EDL933

As mentioned earlier, ROS are known to enhance VT production. The level of ROS generated in *E. coli* strains, K-12,

EDL931, and EDL933, grown in DA (5 $\mu\text{g}/\text{mL}$) in LB broth was detected using the fluorescent probe DCFHDA. The probe is hydrolyzed via an esterase to form DCFH in the cytoplasm of the bacteria and converted into a fluorescent product (DCF) only in the presence of ROS. Figure 3A–C shows that the amount of ROS generated in the controls was negligible. Samples (Figure 3D–F) showed that ROS production was strain dependent with EDL933 producing approximately ten times (735 AU) more ROS than K-12 (68 AU). We also recorded the fluorescence signal when DA was added in mid-log phase (samples, Figure 3G–I), but no significant amount of ROS was detected in any of the samples compared with the control. It must be noted that the oxidative stresses experienced by these cells in aerobic culture may not occur when the cells are cultured under anaerobic conditions. ROS causes the induction of Stx-converting prophages, but the relationship between extracellular ROS and its effects on extracellular peroxidase activity has not been reported.

The effect of DA and glucose on extracellular peroxidase activity

In this section, we investigated whether glucose or DA can render EDL933 more susceptible to external ROS attack when it is grown in saturated glucose LB broth and saturated glucose LB-containing DA using a simple TMB assay. Figure S2 shows that glucose (2%, w/v) produces a negligible drop in the peroxidase activity of K-12 and EDL931, while the peroxidase activity of EDL933 decreases $>65\%$. Figure 4A also shows that the extracellular peroxidase activity of live EDL933 and its lysate is inhibited in a concentration-dependent manner by DA. Overall, the highest decrease (-80%) in extracellular peroxidase activity was

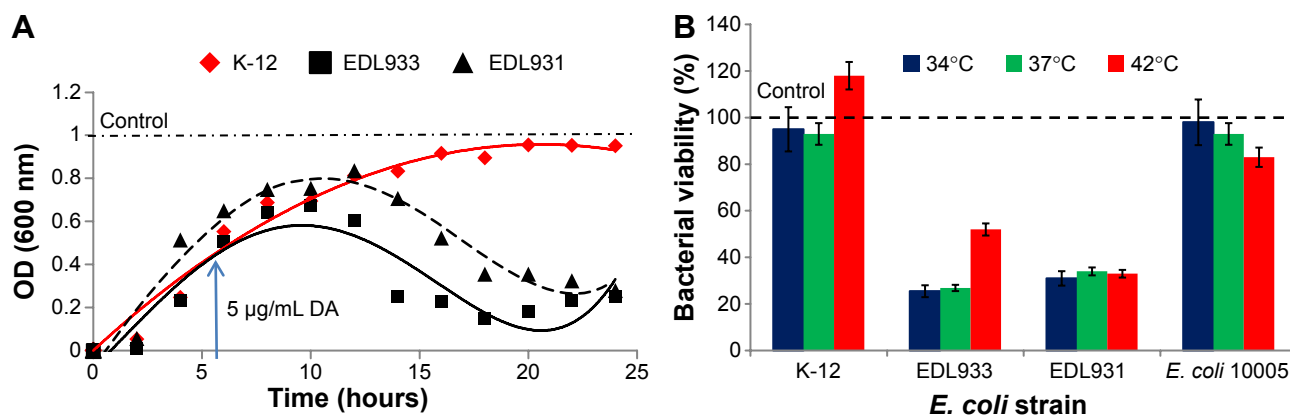


Figure 2 (A) The effects of administering DA (5 mg/L) in mid-log phase on the growth patterns of K-12 (model) and both strains of *E. coli* O157:H7. The blue arrow indicates the approximate time DA was added. **(B)** Inhibitory effects of DA administered in mid-log phase (+6 hours) on the viability of K-12, EDL933, EDL931, and *E. coli* 10005 grown at different temperatures. Bars in **(B)** represent the standard error of the mean of three independent experiments ($P < 0.01$).

Abbreviations: DA, diminazene aceturate; *E. coli*, *Escherichia coli*; OD, optical density.

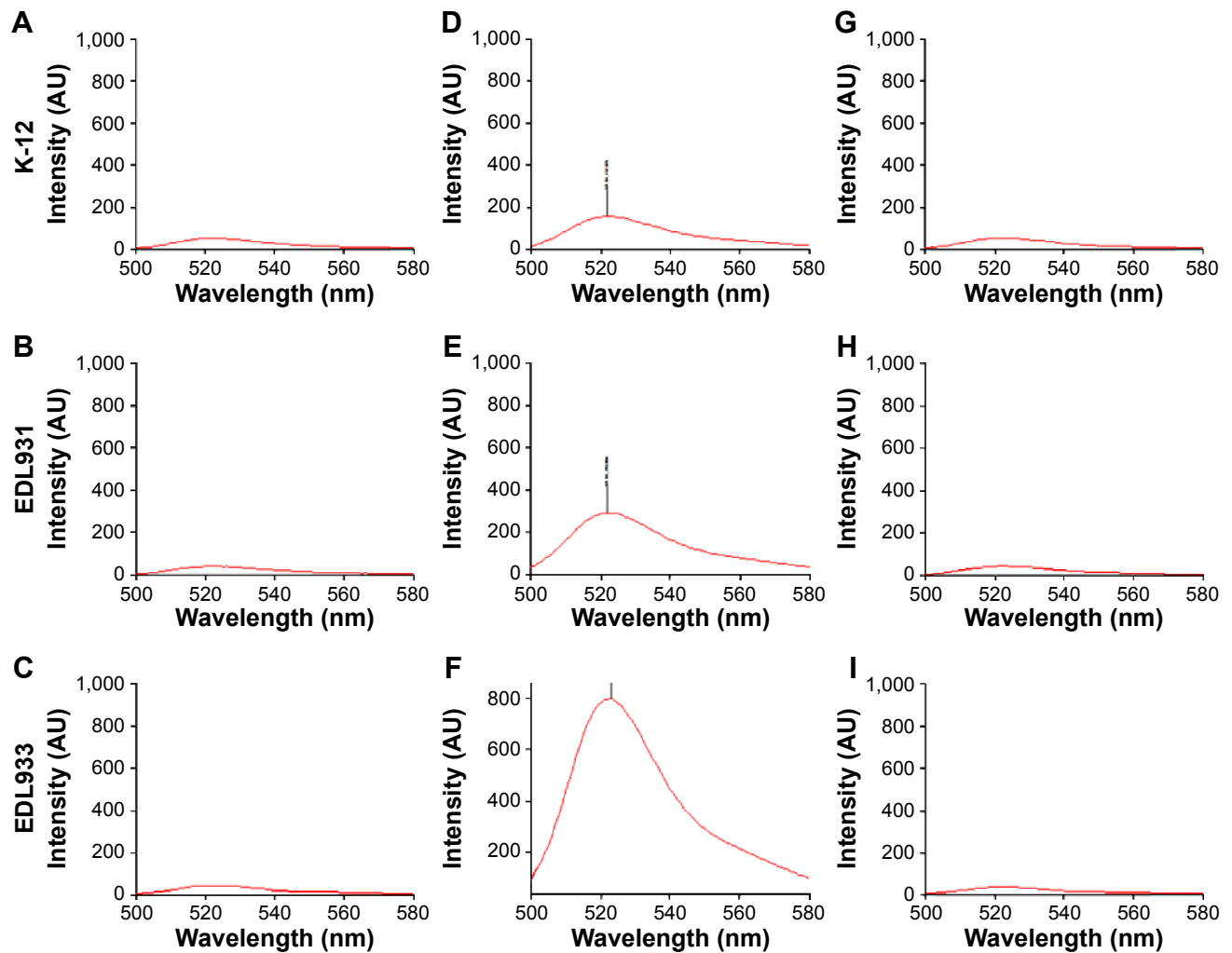


Figure 3 Internal ROS activity of K-12, EDL931, and EDL933 grown for 6 hours in LB broth (A–C), (D–F) LB/DA (5 mg/L) broth, (G–I) addition of DA in mid-log phase cells grown for further 6 hours ($\Delta T=12$ hours).

Abbreviations: DA, diminazene aceturate; LB, Luria–Bertani; ROS, reactive oxygen species.

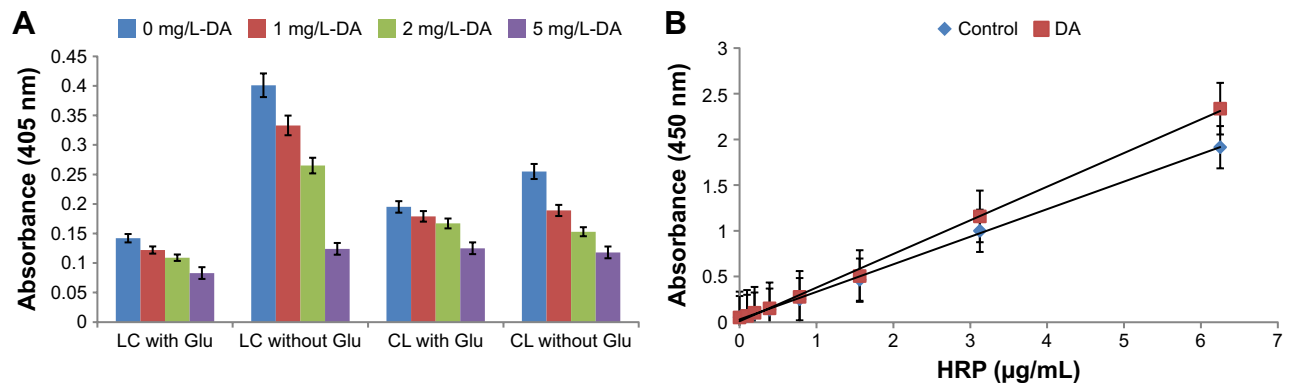


Figure 4 (A) Concentration-dependent effect of DA on the peroxidase activity of EDL933 LC and CL grown in LB broth with 2% glucose and without. (B) The effect of DA (5 mg/L) on HRP–TMB catalysis. Bars in (A) represent the standard error of the mean of three independent experiments ($P<0.01$).

Abbreviations: CL, cell lysate; DA, diminazene aceturate; HRP, horseradish peroxidase; LB, Luria–Bertani; LC, live cells; TMB, 3,3',5,5'-tetramethylbenzidine.

recorded for EDL933 grown in 2% glucose/DA (5 µg/mL) broth. The drop in peroxidase activity could be a simple case of enzyme inhibition. Figure 4B shows that there is no statistical difference between the standard positive control and DA samples; therefore, DA does not directly affect TMB catalysis.

The effect of potassium on EDL933 viability

In addition to ROS, there are other factors such as pH, temperature, hydrostatic pressure, chelating agents, and UV irradiation that enhance VT production. We have shown that DA can kill EDL933 and EDL931 in the absence of ROS.

Therefore, what are these shared properties that make these strains equally susceptible to DA? In this section, we examine the effects of potassium ion concentration on population collapse. If DA is acting as a proton uncoupler and thereby limiting the uptake of K⁺, saturating the medium with the potassium will ensure sufficient uptake (via chemical gradient) of the ion by EDL933 and prevent a collapse in the proton motive force.

Figure 5A shows that the addition of potassium salt to LB/DA broth at 0 hour and +9 hours does not improve the viability of *E. coli* O157:H7 compared with the controls. On the other hand, the viability of EDL933 significantly improves when DA is added to LB/KCl (50 mM) broth after 6 hours

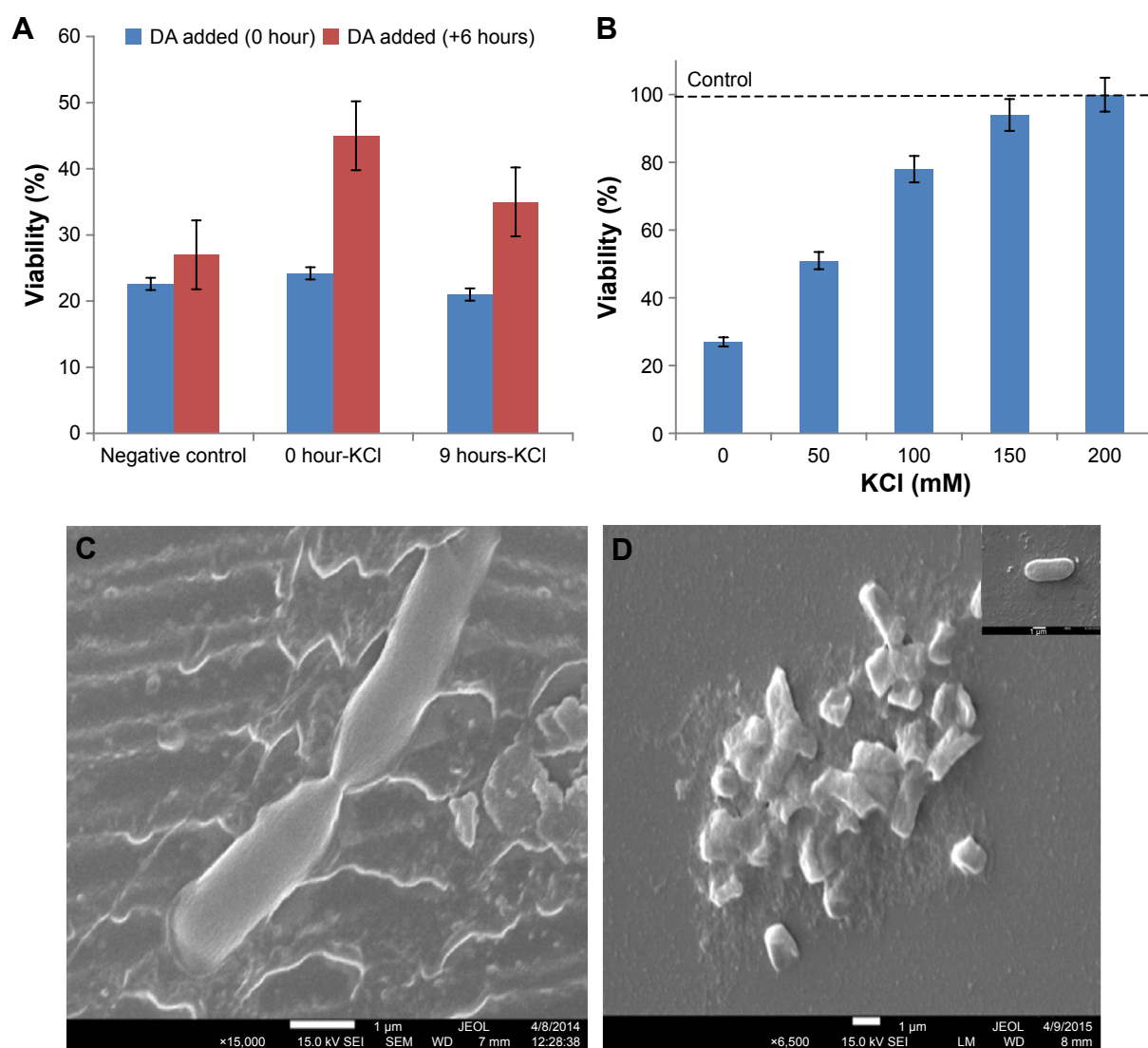


Figure 5 (A) Effects of adding KCl (50 mM) and DA (5 mg/L) to LB broth at different times of growth on the viability of EDL933. (B) Effects of increasing KCl concentration on the viability of EDL933 grown in LB-DA broth after 24 hours. (C) SEM image of dividing *E. coli* O157:H7 (inset shows the turgid state of the bacteria prior lysis). (D) SEM image of the split bacteria (inset shows membrane ghosts).

Abbreviations: DA, diminazene aceturate; *E. coli*, *Escherichia coli*; LB, Luria-Bertani; SEM, scanning electron microscopy.

of growth. We then repeated the latter experiment using LB broth containing 80 mM, 100 mM, 150 mM, and 200 mM of KCl. Figure 5B shows that the viability of EDL933 increases with increasing KCl concentration. Then, 100% viability was restored using a concentration of 200 mM KCl, which is approximately five times higher than the potassium concentration in tryptone soya broth.

Limited potassium ion uptake may be the cause of the collapse, but it does not tell us how DA induces the collapse, or why both strains of *E. coli* O157:H7 are particularly vulnerable to collapse in the log phase. SEM images of EDL933 before the addition of DA to LB broth and an hour prior population collapse are shown in Figure 5C. The image provides a unique insight into the effects of cell division on the shape of the bacteria, elasticity of the membrane, and the surface area of the poles. The number of creases/folds in the membrane at the poles is significantly higher compared with the main body of the bacteria. Further examination of the surface of main body of bacteria shows a terrace-like structure along its longitudinal axis. However, the bacteria in the insert appear swollen, distorted, and elevated from the surface, with no discernable creases in the membrane. Further SEM analysis (Figure 5D) shows that the dead bacteria are split into two or appear as membrane ghosts. The internal force required to split a bacteria in half is significantly greater than the force required to perforate one. The type of debris in Figure 5D is often seen in osmotically lysed cells.

We then investigated the effects of increasing DA concentrations on the viability of EDL933. Surprisingly, 10 $\mu\text{g/L}$ and 20 $\mu\text{g/L}$ of DA induced a smaller population collapse than 5 $\mu\text{g/L}$, suggesting that the concentration of broth constituents may influence the antibacterial activity of DA. In the final part of this section, we measured the effects of increasing NaCl concentrations (5 g/L, 6 g/L, and 7 g/L NaCl) in LB broth on the viability of *E. coli* O157:H7 using a fixed concentration of 20 $\mu\text{g/L}$ of DA (data not shown). At 7 g/L of NaCl, the viability of bacteria decreases to 20%; thus, the speed and severity of the collapse seen in both strains of *E. coli* O157:H7 are due to the concentration ratio of sodium to DA and not DA alone.

The effects of DA on protease activity of *E. coli*

The effect of buffer pH and glucose (Figure S3) on the proteolytic activity of K-12 is shown in Figure 6A. In the absence of glucose, cellular protease activity was highest at pH 5 and lowest at pH 6, H-Phe was identified as a potential marker for K-12 (pH 5). When excess glucose was present

in the assay medium (S4), cellular protease activity was lowest at pH 5 and highest at pH7, and H-Asp-pNA and Ac-Asp-pNA were identified as potential markers for K-12 (pH 7). Across the pH range (5–7), the sum of protease/proteolytic activity of K-12 measured in saturated glucose buffer was 6% lower compared with the negative control (K-12 in buffer only).

Figure 6B and C shows the influence of glucose on the protease/proteolytic activity of EDL 933. In the absence of glucose, activity is broad, weak, and nonspecific, but when the media is saturated with glucose the profile changes completely. This trend is also observed in the activity profile of EDL931 (Figure 6D). Across the pH range (5–7), the sum of protease/proteolytic activity of EDL933 measured in glucose buffer was 48% lower compared with the negative control (EDL933 buffer alone). Substrates H-Gly-Arg- β NA·HCl, H-Arg-Arg- β NA·3HCl, H-Asp-pNA·HCl, and H-D-Ile-Phe-Lys-pNA-salt were identified as potential markers for *E. coli* O157:H7.

A similar experiment to the one described earlier using the four substrates and EDL933 (control) was then conducted, and the results are shown in Figure 6E. EDL933 incubated with DA showed a small increase in protease/proteolytic activity to two of the four substrates; there was no change in the activity to H-D-Ile-Phe-Lys-pNA (serine protease substrate) salt, but a significant increase ($\times 2.5$) in protease activity for H-Asp-pNA (substrate for peptidase E, B and L-asparaginases). The experiment was repeated at 37°C and 42°C as well.

DA does not induce VT production in EDL933

Figure 7A shows the visual results of the commercial test. The control (LB broth) is weakly positive for VT2 and negative for VT1. Mitomycin C tests were positive for VT1 and VT2; Novobiocin and Pentamidine test was negative for VT1 and VT2, whereas DA is weakly positive for VT1 compared with the control. However, Ampicillin tested positive for VT1 and negative for VT2, which is surprising. To confirm the visual observations, samples were also imaged under UV light (Figure 7B) and subsequently analyzed (Figure 7C), and the total amount of VT was tabulated (Figure 7D). Overall, Ampicillin/LB media (1 $\mu\text{g/mL}$) contained low levels of VT2 but significantly higher levels of VT1 (57%), while LB media containing DA had significantly lower levels of VT2 and slightly higher levels of VT1 compared with the controls. In terms of total verotoxins (VT1 and VT2) in the broth,

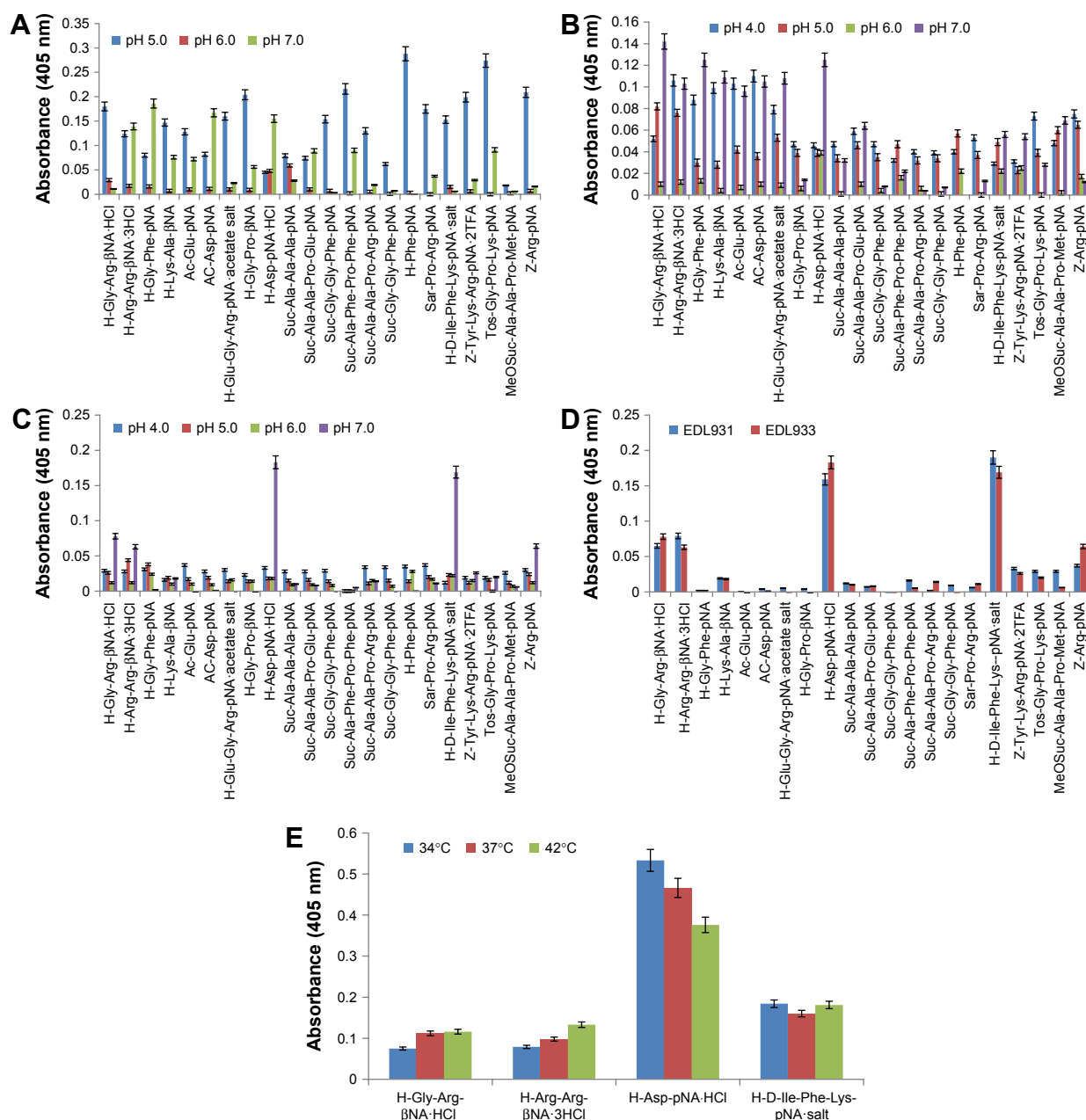


Figure 6 The effects of saturated glucose on the protease activities of K-12 and EDL933.

Notes: (A) K-12 pH buffer only. (B) EDL933 pH buffer only. (C) EDL933 pH buffer and 5% glucose. (D) The activity profile of EDL 933 and EDL931 (5% glucose, pH 7). (E) The effects of DA on the proteolytic activity of EDL933 for H-Gly-Arg-βNA-HCl, H-Arg-Arg-βNA-3HCl, H-Asp-pNA-HCl, and H-D-Ile-Phe-Lys-pNA-salt with increasing assay temperature (the data for K-12 in pH 4.0 were omitted, dead cells). Bars represent the standard error of the mean of three independent experiments ($P < 0.01$).

Abbreviations: DA, diminazene aceturate; pNA, para-nitroaniline; TFA, trifluoroacetic acid.

Mitomycin C produced the highest value of 1.21 followed by Ampicillin, LB broth, DA, Pentamidine, and Novobiocin. It should be noted that neither Pentamidine nor Novobiocin induced a collapse in the EDL933 population when administered in the mid-log phase.

Discussion

The first line of defense against any pathogenic infection is a healthy diverse commensal microbiome. *E. coli* is believed to

be an all-rounder running a thin line between commensalism and pathogenicity. Thus, it is essential to prove that DA is selective for STEC O157 over the commensal population. Within the microbiological community, K-12 is considered the model strain of *E. coli* commensalism, whereas EC10005 can act as both depending on environment stresses. It is well reported that antibiotic resistance of many strains of K-12 and wild-type *E. coli* O157 increases with temperature.³⁶ Antibiotic resistance in *E. coli* is governed by two properties

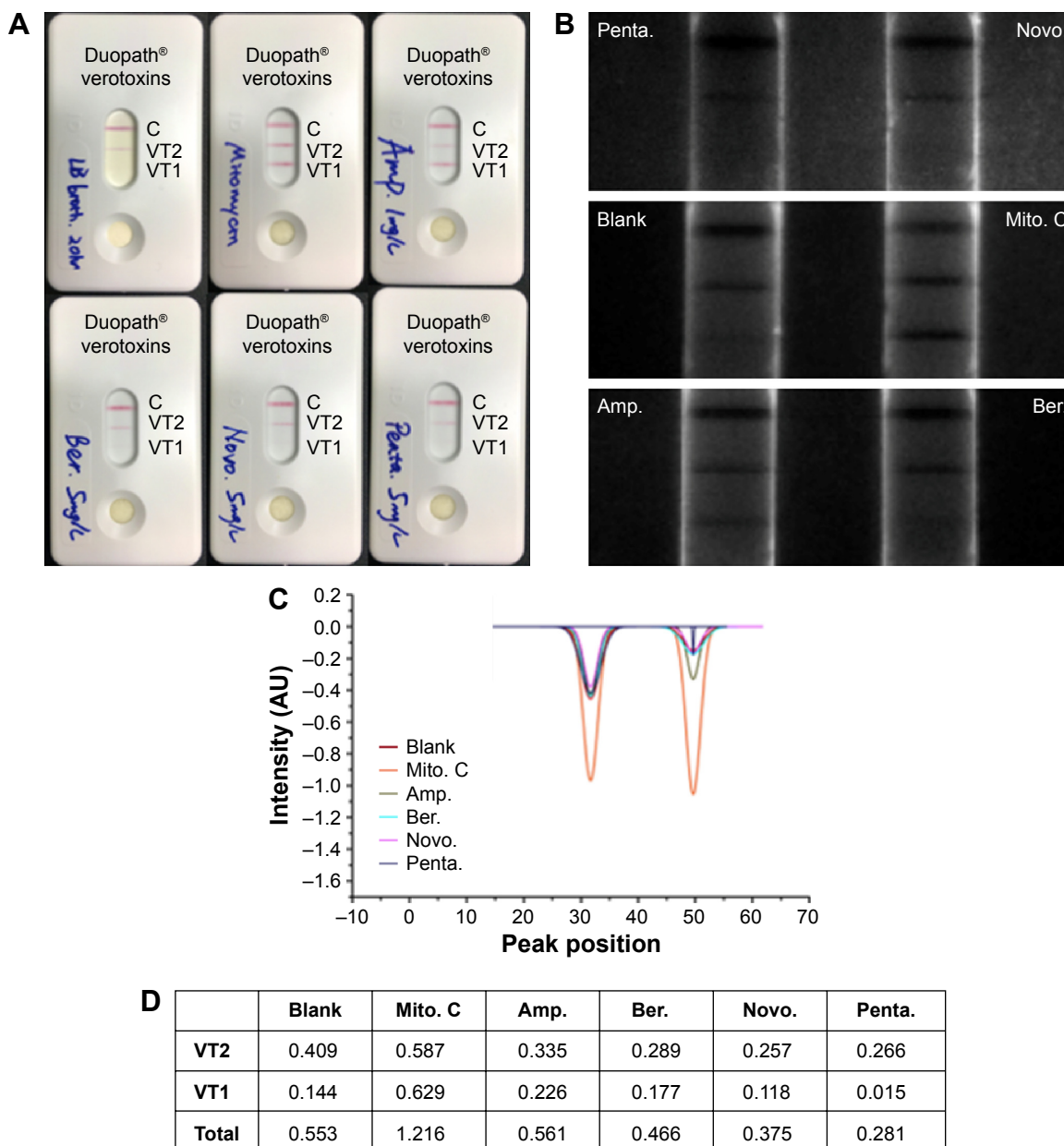


Figure 7 VT test and subsequent analysis.

Notes: (A) Images of the kit devices after incubation. (B) Images of bands of the samples from the kit devices via Kodak Image Station 4000MM PRO. (C) Standardized peak curves for VT1 and VT2 of each sample (Image) that analyzed by Origin2016. (D) The table shows the standardized cumulative area of the peaks, which indicates the amount of the production of verotoxins.

Abbreviations: Amp., Ampicillin; Ber., Berenil; Mito. C, Mitomycin C; Novo., Novobiocin; Penta., Pentamidine; VT, verotoxin.

of genes: pleiotropy and epistasis. The latter describes how the antibiotic resistance mutations affect other functions, hence their fate in other environments. Epistasis describes how well different mutations combine in their effect on resistance.³⁷ Therefore, it is necessary to know what effect temperature has on the resistance of both pathogenic and commensal *E. coli* to DA and if such effects are of benefit to the commensal population. The results in Figure 1 show that at higher temperatures DA is more selective for both strains

of *E. coli* O157 than the commensal strains. The results in Figure 2 also shows that the viability of commensal strains improves significantly when DA is administered in the log phase which could be due to a change in the membrane permeability of the commensal strains during the log phase. Taking the results from Figures 1 and 2, in addition to the pH data given in Figure S2, it is reasonable to suggest that at neutral pH DA is a selective antibacterial agent for *E. coli* O157:H7 when it is administered at 37°C.

Recent work has illustrated that phenotypes such as cell death, mutagenesis, oxidative stress, and OxyR activation can all be reduced depending on the method of culturing.³⁸ Another interesting observation was that in the presence of catalase, which can break down H₂O₂, STEC-dependent *Tetrahymena* killing in cocultures diminished.³⁹ Saturated glucose media is known to increase ROS in mammalian cells and reverse the effect of citrate and the inhibition of 933WΔtox in STEC.⁴⁰ In addition, earlier reports hinted at a possible feedback mechanism between ROS, phage induction, and membrane peroxidase activity.⁴¹ The results in Figures 3 and 4 suggest that DA inhibits the growth of EDL933 by increasing intracellular ROS levels and decreasing extracellular peroxidase activity making the host bacteria more susceptible to external ROS attack. Whether a drop in extracellular peroxidase activity indicates the bacteria has entered the lytic cycle requires further investigation, but it is reasonable to assume that the elevated ROS levels will reduce membrane integrity making lysis easier.

K⁺ transport is one of a large number of mechanisms contributing to pH homeostasis. Many bacteria primarily use net potassium uptake to compensate for proton extrusion that establishes the proton motive force (PMF), thus alkalinizing the cytoplasm at low external pH. In *E. coli*, the ΔpH of potassium-depleted cells varies with the K⁺ concentration of resuspension medium. The regulation of cytoplasmic pH during acid stress depends on ion transport and catabolic acid consumption. It is well known⁴² that K-12 has a much higher intracellular store of potassium than *E. coli* O157:H7, which may account for the absence of a population collapse in Figure 2. If DA is simply acting as a PMF uncoupler, akin to carbonylcyamide-*m*-chlorophenylhydrazine as suggested earlier, one way to test this theory is to measure the drop in the uptake of gentamicin-labeled Texas Red in O157 cells using FACS as the PMF drops. Taking the results from Figures 1 to 5 into account, it is apparent that DA can inhibit, suppress, and uncouple EDL933 by acting on multiple targets within the bacteria. In an effort to identify as many targets as possible, we then examined the effects of DA on the protease activity of EDL933 and EDL931. The assay was performed in saturated glucose to ensure that a strong PMF was maintained in EDL933 and EDL931. This resulted in both strains of O157 exhibiting near-identical proteolytic profiles (Figure 6D).

In the presence of DA, there is twofold to threefold (Figure 6E) increase in proteolytic activity for Asp at 34°C and 37°C, which indicates DA is upregulating either the *dctA* or *aspA* gene (aspartate ammonia-lyase enzyme function) in

the gluconeogenesis pathway. Further investigation should be conducted as to the effect of DA on the genes responsible for the assimilation of ethanolamine, urea, agmatine, and amino acids (Asp, Thr, Gly, Ser, and Trp) and how DA influences the gluconeogenesis pathway in EDL933 and other pathogenic strains.

As mentioned in the Introduction, a suitable antibacterial agent for STEC should not induce VT production. To show DA does not induce the phage induction cascade and is a suitable antibacterial agent for EDL933, we investigated the effects of DA on VT production. The results in Figure 7 clearly show that both Mitomycin C and Ampicillin induce the production of VTs in EDL933, whereas DA does not. Moreover, DA is a much better antibacterial agent lysing 77% of the EDL933 population compared with 30% and 20% (data not shown) for Mitomycin C and Ampicillin, respectively. The weakly VT2 positive seen in the control and DA samples is probably due to the nitrates (10 mM) in the LB media. It should be noted that the degree of lysis will affect the total VT concentration as such the peak values for Mitomycin C and Ampicillin are negatively bias (underestimated). Future work will focus on the quantification of VT and investigate the exciting possibility that DA inhibits the autocatalytic serine protease activity of the CI repressor rendering the prophage uninducible.⁴³

Conclusion

To our knowledge, no investigations regarding the ability of DA to inhibit or suppress the growth of *E. coli* O157:H7 via ROS accumulation have been reported. EDL933 demonstrated the highest peroxidase activity and the highest internal ROS levels out of all the strains investigated. In addition, we have also shown that DA is an effective uncoupler of the PMF in more than one strain of *E. coli* O157:H7 and that the underlying mechanism does not induce the production of VTs. Therefore, we conclude that DA is a suitable antibacterial agent for *E. coli* O157, because it can inhibit and kill different strains of STEC at various stages of the fission cycle using a variety of ROS-dependent and ROS-independent mechanisms without inducing a significant increase in Stx production.

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Disclosure

The authors report no conflicts of interest in this work.

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Supplementary materials

Table S1 Inhibition of EDL933 with increasing concentrations of DA

DA	0 ($\mu\text{g/mL}$)	2 ($\mu\text{g/mL}$)	5 ($\mu\text{g/mL}$)	10 ($\mu\text{g/mL}$)	20 ($\mu\text{g/mL}$)
0 hour	0.001	0.001	0.001	0.001	0.001
2 hours	0.028	0.012	0.003	0.001	0.002
4 hours	0.267	0.126	0.065	0.029	0.006
6 hours	0.377	0.292	0.158	0.096	0.004
8 hours	0.401	0.293	0.249	0.160	0.004
10 hours	0.423	0.310	0.284	0.223	0.003
12 hours	0.433	0.318	0.224	0.166	0.020
14 hours	0.546	0.325	0.210	0.158	0.016
24 hours	0.504	0.182	0.161	0.176	0.168

Note: Average values (N=3).

Abbreviation: DA, diminazene aceturate.

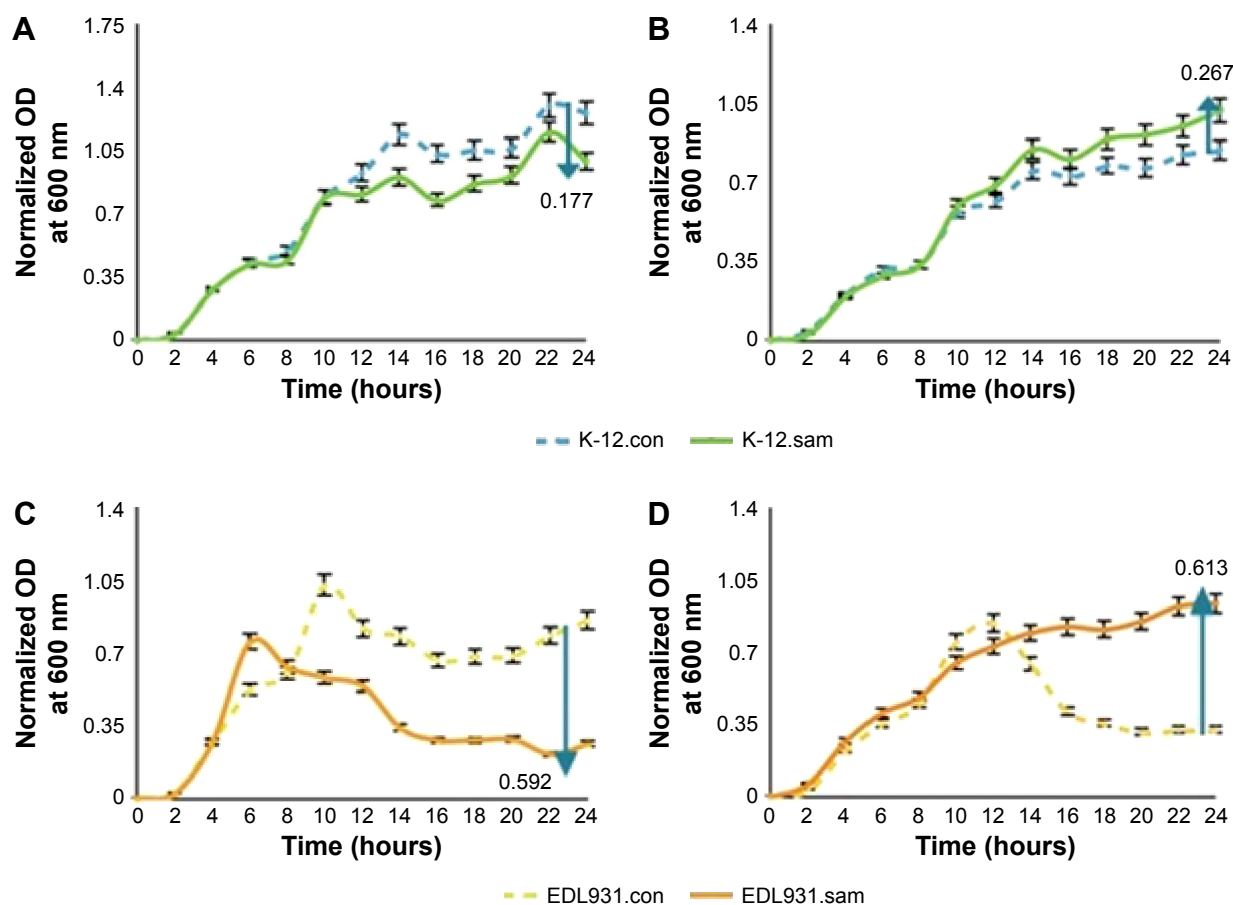


Figure S1 (Continued)

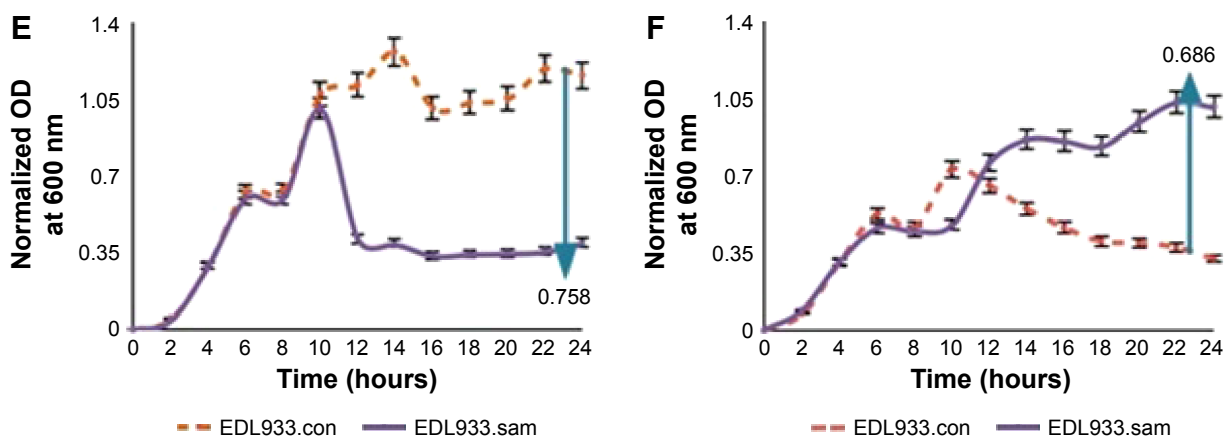


Figure S1 The effects of broth pH on the antibacterial activity of DA.

Notes: The effects of DA when broth pH was converted at 8 hours. (A) K-12, (C) EDL931, and (E) EDL933 were grown in LB broth (pH 5.0) with 5 $\mu\text{g/mL}$ DA and then converted to pH 7.0 at 8 hours; (B) K-12, (D) ATCC 35150, and (F) EDL933 were grown in LB broth (pH 7.0) with 5 $\mu\text{g/mL}$ DA and then converted to pH 5.0 at 8 hours.

Abbreviations: DA, diminazene aceturate; LB, Luria-Bertani; OD, optical density.

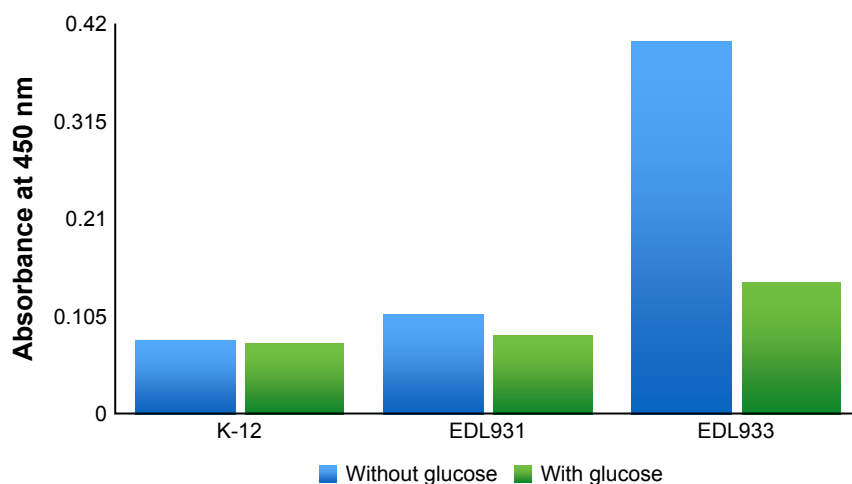


Figure S2 Cell peroxidase/TMB activity of different strains of *E. coli* (2% glucose) (N=3).

Abbreviations: *E. coli*, *Escherichia coli*; TMB, 3,3',5,5'-tetramethylbenzidine.

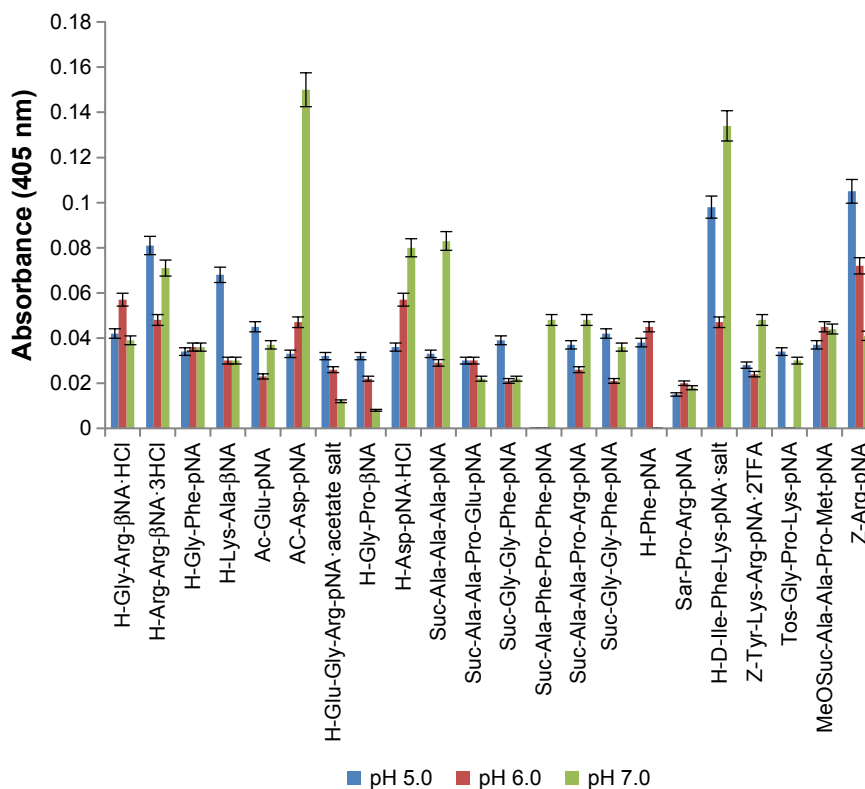


Figure S3 Protease activity of K-12 incubated in saturated glucose media at different pH.
Abbreviations: pNA, para-nitroaniline; TFA, trifluoroacetic acid.

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