

Circular RNA Circ_0006282 Promotes Cell Proliferation and Metastasis in Gastric Cancer by Regulating MicroRNA-144-5p/Tyrosine 3-Monooxygenase/Tryptophan 5-Monooxygenase Activation Protein β Axis

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Background: Circular RNAs (circRNAs) are a class of non-coding RNAs which function as novel regulators in human cancers. In this study, we aimed to investigate the functional roles and related molecular mechanisms of circ_0006282 in gastric cancer (GC) progression.

Methods: Fifty-five GC patients were enrolled in this study. GC cells (AGS and HGC-27) and normal cells (GES-1) were cultured in RPMI1640 added with 10% FBS and 1% penicillin-streptomycin. Quantitative real-time polymerase chain reaction (qRT-PCR) assay was used to determine the expression levels of circ_0006282, transcription elongation factor B subunit 1 (TCEB1) mRNA, miR-144-5p and tyrosine 3-monooxygenase/tryptophan 5-monooxygenase activation protein β (YWHAB; also known as 14-3-3 β). RNase R assay was used to determine the characteristic of circ_0006282. Cell Counting Kit-8 (CCK-8) assay and colony formation assay were employed for cell proliferation. Transwell assay was conducted for cell migration and invasion. Western blot assay was carried out to measure the protein levels of Cyclin D1, matrix metalloprotein 9 (MMP9) and YWHAB. Dual-luciferase reporter assay, RNA pull-down assay and RIP assay were adopted to analyze the interaction between miR-144-5p and circ_0006282 or YWHAB. Murine xenograft model assay was performed to explore the function of circ_0006282 in vivo.

Results: Circ_0006282 level was increased in GC tissues and cells compared to normal tissues and cells. Silencing of circ_0006282 restrained GC cell proliferation, migration and invasion. For mechanism analysis, circ_0006282 was identified to function as the sponge for miR-144-5p to positively regulate YWHAB expression in GC cells. Moreover, miR-144-5p inhibition or YWHAB overexpression effectively reversed the impacts of circ_0006282 knockdown on GC cell growth and motility. Additionally, circ_0006282 knockdown blocked tumor growth of GC in vivo.

Conclusion: Circ_0006282 facilitated the malignant behaviors of GC cells through circ_0006282/miR-144-5p/YWHAB axis.

Keywords: gastric cancer, circ_0006282, miR-144-5p, YWHAB

Introduction

Gastric cancer (GC) is a usual deadly cancer in the world, which responsible for a large proportion of cancer-associated mortalities.^{1,2} Tumor metastasis and

recurrence are the main obstacles for GC therapy.^{3,4} Although the methods of surgery, chemotherapy and radiotherapy can alleviate the symptoms and improve the survival of GC patients, the prognosis of GC patients remains unfavorable.⁵ Therefore, exploring the pathological mechanisms of GC is crucial to search for effective therapeutic targets for this lethal disease.

Circular RNAs (circRNAs) are newly discovered endogenous, circular transcripts with neither 5'-3' polarities nor polyadenylated tails.^{6,7} CircRNAs can adsorb microRNAs (miRNAs), a sort of non-coding RNAs with ~22 nucleotides, to modulate gene expression.^{8,9} Up to date, emerging evidence has reported that circRNAs are widely expressed and participate in the development of human malignancies, including GC.¹⁰ For instance, circ-SFMBT2 aggravated the malignancy of GC by decreasing miR-182-5p and increasing CREB1.¹¹ Circ_0008035 contributed to GC cell viability and invasion via sponging miR-375 to enhance YBX1 expression.¹² Circ_0006282 originated from TCEB1 gene and played an oncogenic role in GC by modulation of miR-155/FBXO22 axis.¹³ Even so, the molecular mechanism by which circ_0006282 mediating GC progression remains largely unclear. In this study, the functional roles and regulatory mechanisms of circ_0006282 in GC cell growth and metastasis were investigated.

The aberrant expression of miRNAs exerts an outstanding role in regulating the biological roles of tumor progression.¹⁴ In GC, multiple miRNAs have been claimed to serve as a tumor inhibitor through targeting the 3' untranslated regions (3'UTR) of target genes.^{15,16} For example, miR-12129 impeded cell growth and cell cycle process in GC by targeting SIRT1.¹⁷ MiR-338-3p knockdown facilitated GC cell migration and prevented apoptosis through interaction with PTP1B.¹⁸ MiR-144-5p was found to be lowly expressed in GC and linked to the overall survival of GC patients.¹⁹ Nevertheless, the exact roles of miR-144-5p and its related mechanisms still need further investigation.

Tyrosine 3-monooxygenase/tryptophan 5-monooxygenase activation protein β (YWHAB; also termed as 14-3-3 β) is a form of 14-3-3 proteins, which can modulate cell proliferation, survival and motility.²⁰ It has been documented that YWHAB plays a crucial role in the carcinogenesis of diverse human cancers, such as papillary thyroid carcinomas,²¹ glioma²² and hepatocellular carcinoma.²³ However, the effects of YWHAB on GC are unknown.

Here, we found that miR-144-5p contained the potential binding sites of circ_0006282 and YWHAB. However, the relationships among circ_0006282, miR-144-5p and YWHAB in GC development remain unclear. Thus, we deciphered the functions of circ_0006282, miR-144-5p and YWHAB in GC and their relationships in regulating GC cell malignant behaviors.

Materials and Methods

Tissues Collection

Fifty-five GC patients were recruited in this study. After the work was allowed by the Ethics Committee of Baotou Cancer Hospital (NeiMengGu, China) and written informed consents were obtained from the patients, the tumor tissue specimens and adjacent non-tumor tissue specimens were harvested from the patients at Baotou Cancer Hospital during April 2015 and January 2019. The inclusion criteria were as follows: (1) The patients were pathologically diagnosed with GC; (2) The patients did not receive any chemotherapy or radiotherapy before surgery. The exclusion criteria were as follows: (1) The patients <18 years old or >80 years old; (2) pregnant or lactating patients; (3) The patients had autoimmune diseases or other malignancies. The harvested specimens were stored at -80°C before use.

Cell Culture

Human normal gastric mucosal epithelial cells (GES-1) and GC cells (AGS and HGC-27) were selected based on the previous literatures^{24,25} and purchased from Procell (Wuhan, China). These cells were maintained in Roswell Park Memorial Institute 1640 Medium (RPMI1640; Procell) added with 10% fetal bovine serum (FBS; Procell) and 1% penicillin-streptomycin (Procell) at 37°C in a humid incubator with 5% CO₂.

Quantitative Real-Time Polymerase Chain Reaction (qRT-PCR)

The extraction of total RNA was done utilizing TRIzol (Beyotime, Shanghai, China). The RNA was quantified with the NanoDrop 2000c spectrophotometer (Thermo Fisher Scientific, Waltham, MA, USA). For RNase treatment, 20 μ g RNA was treated with or without RNase R (3U/ μ g; Epicenter Biotechnologies, Madison, WI, USA) for 20 min at 37°C. Next, for circ_0006282 and mRNA, PrimeScript™ RT reagent Kit (Takara, Dalian, China) and either Oligo (dt)₁₈ primers or Random primers were used for cDNA synthesis. For miR-144-

5p, TaqMan microRNA Assay kit (Applied Biosystems, Foster City, CA, USA) was utilized for cDNA synthesis. Afterwards, qRT-PCR reaction was executed with BeyoFast™ SYBR Green qPCR Mix (Beyotime) and relevant primers (GeneCopoeia, Guangzhou, China) on the StepOnePlus Real-Time PCR System (Applied Biosystems). The primers were: circ_0006282: (F: 5'-AGGCACGATAAAAGCCATGT-3' and R: 5'-GGTCCTTCACAGCCACCATA-3'); TCEB1: (F: 5'-TTATGCATGGGAAAGAAGC-3' and R: 5'-CTCAA CTCCTGGCCTCAAG-3'); miR-144-5p: (F: 5'-GCGCGAA TTCGAGATCTTAACAGACCCTAGCTC-3' and R: 5'-GCGCGGATCCGTGCCCTGGCAGTCAGTAGG-3'); YWHAB: (F: 5'-GTCACCAGGTCTCCCAAGTG-3' and R: 5'-TTCTCCCCACTGCAGTGTTCC-3'); glyceraldehyde 3-phosphate dehydrogenase (GAPDH): (F: 5'-ACCCACTCCTCC ACCTTTGAC-3' and R: 5'-TGTTGCTGTAGCCAAAT TCGTT-3'); U6: (F: 5'-CTCGCTTCGGCAGCACACA-3' and R: 5'-AACGCTTCACGAATTTGCGT-3'). The expression was computed via the $2^{-\Delta\Delta Ct}$ strategy with GAPDH or U6 as a control.

Subcellular Fraction Assay

The cytoplasmic and nuclear fractions in AGS and HGC-27 cells were separated through the usage of PARIS Kit (Invitrogen, Carlsbad, CA, USA) in line with the manufacturer's instructions. Then the aforementioned qRT-PCR analysis was adopted for the levels of circ_0006282, U6 and GAPDH. U6 and GAPDH were employed as the controls of nuclear transcript and cytoplasmic transcript, respectively.

Cell Transfection

Circ_0006282 small interfering RNA (si-circ_0006282) and scramble siRNA (si-NC), miR-144-5p mimics (miR-144-5p) and mimics NC (miR-NC), miR-144-5p inhibitors (anti-miR-144-5p) and inhibitors NC (anti-NC), the over-expression vector of YWHAB (YWHAB) and its control (Vector), circ_0006282 short hairpin RNA (sh-circ_0006282) and sh-NC were all bought by GeneCopoeia. AGS and HGC-27 cells were seeded into 12-well plates with 1.0×10^4 cells/well and then transfected with 50 nM synthetic oligonucleotides or 2 μ g vectors using Lipofectamine 2000 (Invitrogen) based on the protocols. Forty-eight-hour later, the transfected cells were harvested for subsequent experiments.

Cell Counting Kit-8 (CCK-8) Assay

The proliferation capacity of AGS and HGC-27 cells was tested by CCK-8 experiment. In brief, the transfected cells

(1×10^4 cells/well) were plated into 96-well plates. After cultivating overnight, 10 μ L CCK-8 (Boster, Wuhan, China) was supplemented into each well at 24 h, 48 h and 72 h followed by incubation for a further 4 h. Finally, the optical density (OD) value was examined with a microplate reader (Bio-Rad Laboratories, Hercules, CA, USA) at 450 nm. The experiment was run in triplicate.

Colony Formation Assay

Following 48 h of transfection, AGS and HGC-27 cells (500 cells/well) were collected and plated onto 6-well plates in complete medium and cultivated for 2 weeks. The medium was changed every 4 days. Thereafter, the colonies were fixed with 4% paraformaldehyde (Sangon, Shanghai, China) for 20 min and dyed with 0.1% crystal violet (Sangon) for 15 min. Finally, the colonies were photographed and counted.

Transwell Assay

The transwell chambers (BD Bioscience, San Jose, CA, USA) were employed for cell migration and invasion as previously described.²⁶ Briefly, transfected AGS and HGC-27 cells (1×10^5) were harvested, resuspended in 200 μ L serum-free medium and then plated into the top compartment coated with or without Matrigel (BD Bioscience) to assess cell invasion or migration. Six hundred microliter medium added with 10% FBS (Procell) was filled in the lower compartment. Twenty-four-hour later, the cells passed through the membranes were fixed with 4% paraformaldehyde (Sangon) and dyed with 0.1% crystal violet (Sangon). An inverted microscope (Olympus, Tokyo, Japan) was employed to count the invaded/migrated cells at the magnification of 100 \times .

Western Blot Assay

The protein isolation was finished using Radio Immunoprecipitation Assay (RIPA) buffer (Beyotime) and examined using a BCA Protein Quantification Kit (Vazyme, Nanjing, China). Then the protein samples were electrophoresed on 10% sodium dodecyl sulfonate-polyacrylamide gel (SDS-PAGE; Solarbio, Beijing, China) and blotted on polyvinylidene difluoride membranes (PVDF; Pall Corporation, New York, NYC, USA). Next, the membranes were blocked in 5% defatted milk for 1 h at room temperature. After that, the membranes were incubated overnight with primary antibodies against GAPDH (bs-10900R; Bioss, Beijing, China), Cyclin D1 (bs-20596R; Bioss), matrix

metalloprotein 9 (MMP9; bs-4593R; Bioss) or YWHAB (bs-12420; Bioss) at 4°C and then interacted with relevant secondary antibody (bs-0924M; Bioss) for 2 h at room temperature. At last, the protein bands were visualized with an ECL reagent (Vazyme).

Dual-Luciferase Reporter Assay

The potential binding sites between miR-144-5p and circ_0006282 or YWHAB 3'UTR were predicted by Starbase V3.0 and TargetScan, respectively. To verify the prediction, the sequences of circ_0006282 (or YWHAB 3'UTR) consisting of the mutant (MUT) or wild-type (WT) binding sites of miR-144-5p were cloned into pmirGLO vector (Promega, Fitchburg, WI, USA), constructing circ_0006282-WT, circ_0006282-MUT, YWHAB-3'UTR-WT and YWHAB-3'UTR-MUT. Then AGS and HGC-27 cells were seeded into 24-well plated at a density of 5.0×10^4 cells/well. Next, the constructs (100 ng) were co-transfected into in combination with miR-144-5p or miR-NC (50 nM) using Lipofectamine 2000 (Invitrogen). After 48 h of transfection, the Renilla and firefly luciferase activities were examined through the usage of Dual-Luciferase Reporter Assay Kit (Promega). The experiment was done referring to previous research.¹¹

RNA Pull-Down Assay

The Pierce Magnetic RNA-Protein Pull-Down Kit (Thermo Fisher Scientific) was used for RNA pull-down

assay.²⁷ In brief, 3'-biotinylated miR-144-5p (Bio-miR-144-5p) or Bio-miR-NC was transfected into AGS and HGC-27 cells. After 48 h, the cells were lysed and incubated with streptavidin-coupled beads (Invitrogen). Next, the biotin-coupled RNA complex was pulled down. Finally, the enrichment of circ_0006282 and YWHAB in bound fractions was detected by qRT-PCR.

RNA Immunoprecipitation (RIP) Assay

The Magna RNA-binding protein immunoprecipitation kit (Millipore, Billerica, MA, USA) was employed to conduct RIP assay.²⁸ Briefly, the cells were lysed in RIP buffer. Then the collected RIP buffer was incubated with magnetic beads conjugated with anti-Argonaute2 (anti-Ago2) and anti-immunoglobulin G (anti-IgG). The anti-IgG and Input were used as controls. The levels of circ_0006282 and miR-144-5p in the beads were detected by qRT-PCR assay.

Murine Xenograft Model

The 4–6 weeks old nude mice were obtained from Beijing Vital River Laboratory Animal Technology Co., Ltd. (Beijing, China). Then the mice were divided into 2 groups (n=6/group). AGS cells (1×10^7) transfected with sh-NC or sh-circ_0006282 were injected into the flank of the nude mice. The length and width of the xenograft tumor were measured weekly and tumor volume was estimated by the equation: $(\text{Length} \times \text{Width}^2)/2$. The mice were

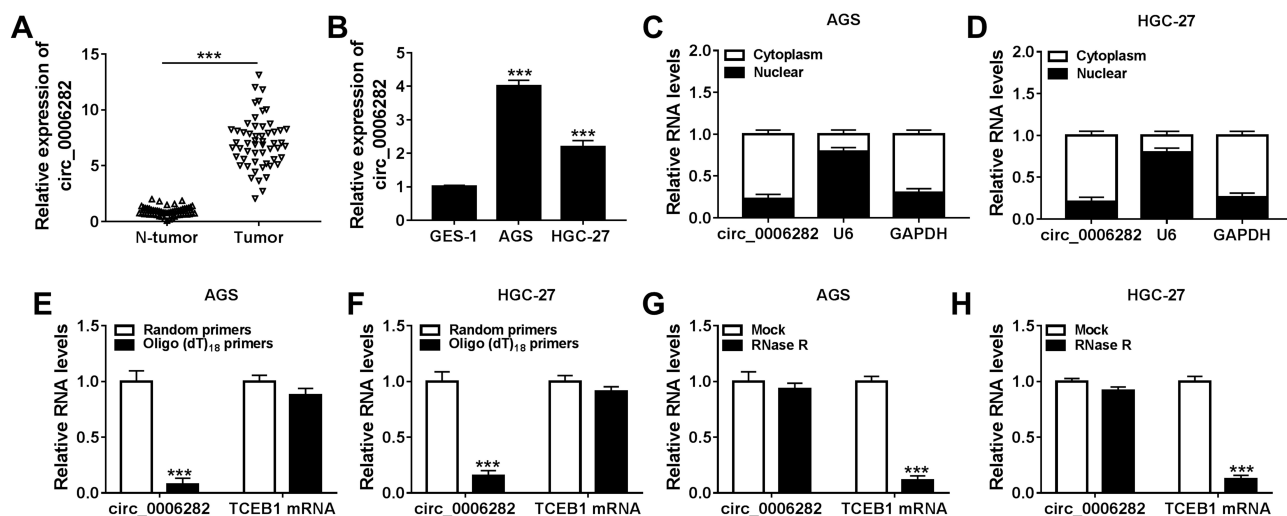


Figure 1 Circ_0006282 was upregulated in GC tissues and cells. (A and B) QRT-PCR assay was used for the expression level of circ_0006282 in GC tissues, cells and corresponding normal tissues and cells. (C and D) The expression level of circ_0006282 in the nuclear and cytosolic fractions of AGS and HGC-27 cells was determined. (E and F) Following the RNA in AGS and HGC-27 cells was reversely transcribed into cDNA with random primers or Oligo (dt)₁₈ primers, the levels of circ_0006282 and TCEB1 mRNA were measured. (G and H) The levels of circ_0006282 and TCEB1 mRNA in AGS and HGC-27 cells treated with or without RNase R were measured. ****p*<0.001 vs N-tumor, ****p*<0.001 vs GES-1, ****p*<0.001 vs Random primers, ****p*<0.001 vs Mock.

ethanized after 5 weeks and the neoplasms were harvested and weighted. The neoplasms were preserved at -80°C for further qRT-PCR and Western blot assays. The animal research was permitted by the Ethics Committee of Animal Research of Baotou Cancer Hospital. Animal studies were performed in compliance with the ARRIVE guidelines and the Basel Declaration. All animals received humane care according to the National Institutes of Health (USA) guidelines. The experiment was done according to previous study.²⁹

Statistical Analysis

The experiments were run in triple times and the collected data were presented as mean \pm standard deviation, processed by using GraphPad Prism 7 software. The differences were estimated by Student's *t*-test and one-way

analysis of variance. $P < 0.05$ was deemed as a significant difference.

Results

Circ_0006282 Was Upregulated in GC Tissues and Cell Lines

To begin with, we determined the expression level of circ_0006282 in GC tissues ($n=55$) and adjacent non-tumor tissues ($n=55$) with qRT-PCR assay. The data showed that compared to non-tumor tissues, circ_0006282 was highly expressed in tumor tissues (Figure 1A). We also found that circ_0006282 level was conspicuously elevated in AGS and HGC-27 cells compared to GES-1 cells (Figure 1B). Subcellular fraction assay showed that circ_0006282 was mainly enriched in the cytoplasm of AGS and HGC-27 cells (Figure 1C and

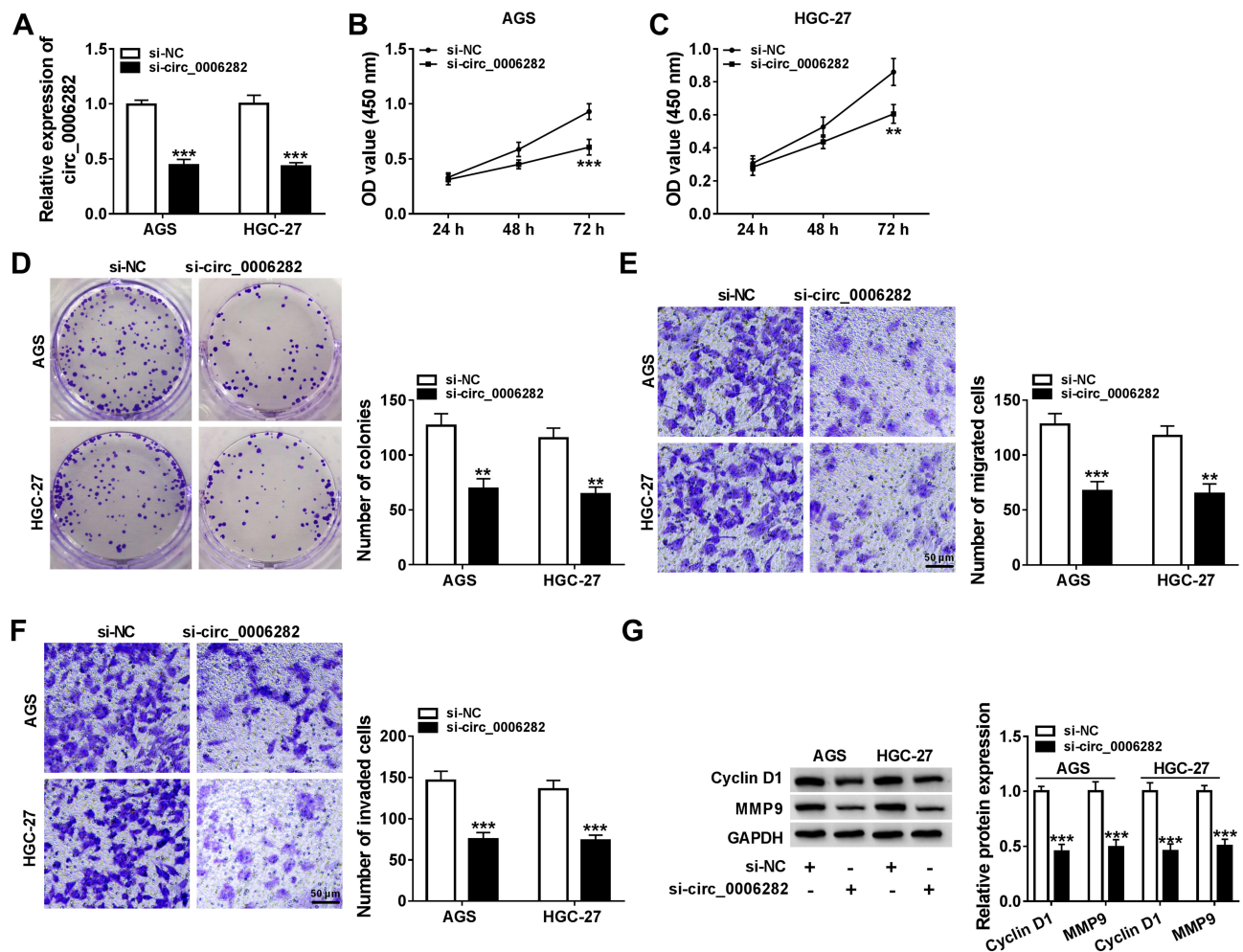


Figure 2 Knockdown of circ_0006282 suppressed GC cell growth and motility. AGS and HGC-27 cells were transfected with si-circ_0006282 or si-NC. (A) The expression level of circ_0006282 in AGS and HGC-27 cells was examined. (B and C) CCK-8 assay was performed for cell proliferation ability. (D) Colony formation assay was used to analyze cell colony formation ability. (E and F) Transwell assay was carried out for cell migration and invasion capacities. (G) Western blot assay was conducted for the protein levels of Cyclin D1 and MMP9 in AGS and HGC-27 cells. ** $P < 0.01$ vs si-NC, *** $P < 0.001$ vs si-NC.

D). Moreover, we found that Oligo (dt)₁₈ primers reversely transcribed cDNA could barely amplify circ_0006282 compared with Random primers reversely transcribed cDNA in both AGS and HGC-27 cells (Figure 1E and F). In addition, the results of RNase R digestion assay indicated that circ_0006282 was resistant to RNase R treatment, whereas TCEB1 mRNA was digested by RNase R treatment (Figure 1G and H). These findings

suggested that the dysregulation of circ_0006282 might play a role in GC progression.

Silencing of Circ_0006282 Repressed GC Cell Proliferation and Metastasis

In order to elucidate the precise roles of circ_0006282 in GC development, loss-of-function experiments were conducted by transfecting si-circ_0006282 into AGS and HGC-

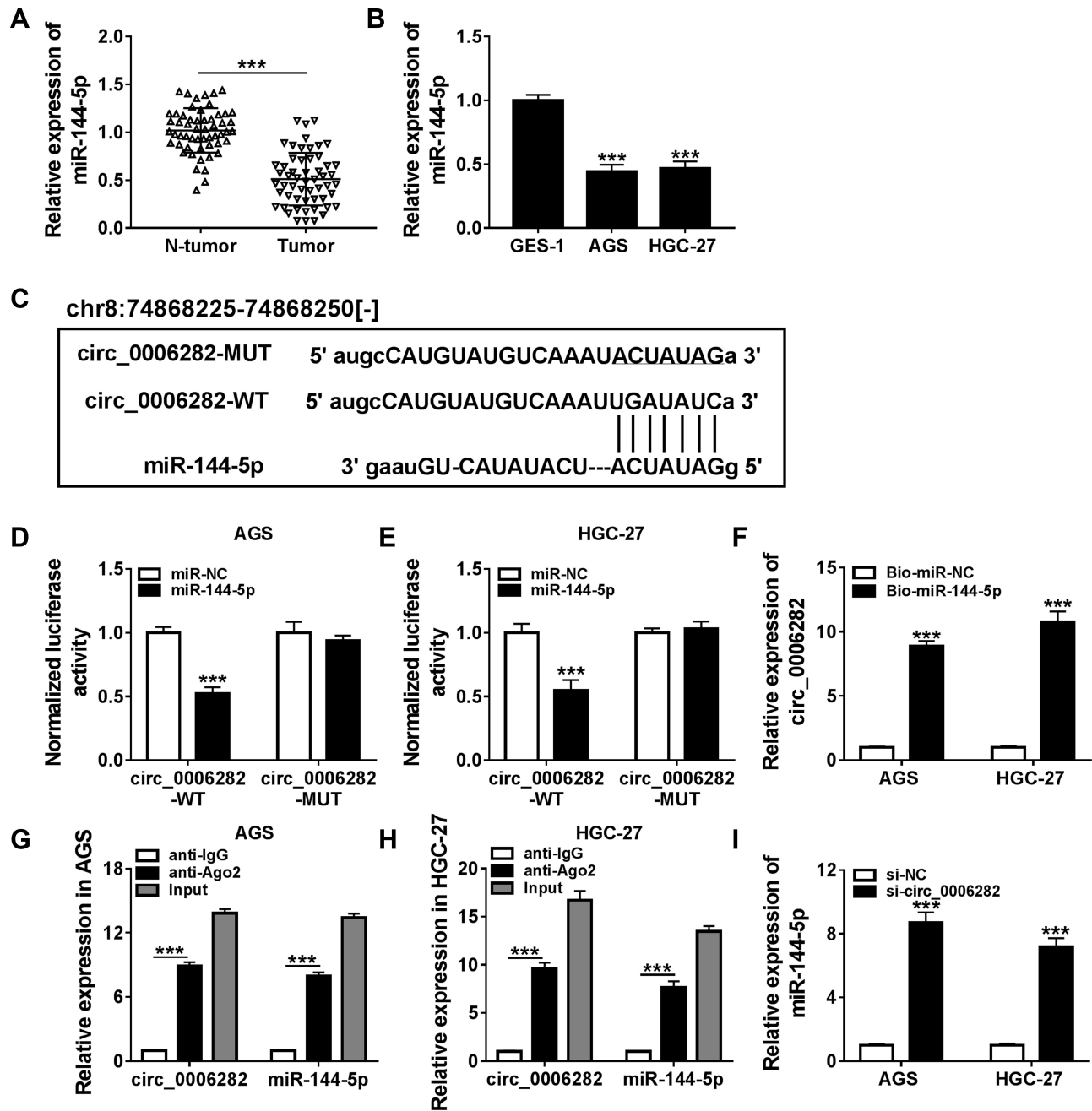


Figure 3 Circ_0006282 directly targeted miR-144-5p in GC cells. (A and B) The expression level of miR-144-5p in GC tissues, cells and corresponding normal tissues and cells was determined. (C) The complementary sequences between circ_0006282 and miR-144-5p were exhibited. (D-H) The interaction between circ_0006282 and miR-144-5p was estimated by dual-luciferase reporter assay, RNA pull-down assay and RIP assay. (I) The expression level of miR-144-5p in transfected AGS and HGC-27 cells was determined. ***P<0.001 vs N-tumor, ***P<0.001 vs GES-1, ***P<0.001 vs miR-NC, ***P<0.001 vs Bio-miR-NC, ***P<0.001 vs anti-IgG, ***P<0.001 vs si-NC.

27 cells. The transfection efficiency was estimated by qRT-PCR assay, exhibiting that si-circ_0006282 evidently decreased the level of circ_0006282 in AGS and HGC-27 cells compared to si-NC control groups (Figure 2A). As demonstrated by CCK-8 assay, circ_0006282 silencing markedly restrained the proliferation of AGS and HGC-27 cells compared to control groups (Figure 2B and C). The colony formation assay showed that circ_0006282 knockdown suppressed AGS and HGC-27 cell colony formation compared to control groups (Figure 2D). Moreover, we conducted transwell assay to evaluate cell migration and invasion abilities, showing that circ_0006282 deficiency drastically suppressed cell migration and invasion in AGS and HGC-27 cells compared to control groups (Figure 2E and F). Additionally, the levels of proliferation-related protein (cyclin D1) and metastasis-related protein (MMP9) were measured by Western blot assay. The results showed that circ_0006282 deficiency led to marked downregulation of Cyclin D1 and MMP9 levels in AGS and HGC-27 cells compared to si-NC control groups (Figure 2G). Collectively, circ_0006282 knockdown hampered the malignant behaviors of GC cells.

Circ_0006282 Functioned as the Sponge for miR-144-5p

As shown in Figure 3A and B, miR-144-5p was weakly expressed in GC tissues and cells compared to normal tissues and cells. Through analyzing online tool Starbase V3.0, we found miR-144-5p contained the complementary sequences of circ_0006282, indicating that miR-144-5p might be a target of circ_0006282 (Figure 3C). Then dual-luciferase reporter assay exhibited that miR-144-5p transfection caused a remarkable suppression in the luciferase activity of circ_0006282-WT in AGS and HGC-27 cells, while the luciferase activity of circ_0006282-MUT was not affected, demonstrating the interaction between circ_0006282 and miR-144-5p (Figure 3D and E). The results of RNA pull-down assay showed that circ_0006282 level was abundantly pulled down by Bio-miR-144-5p RNA pull-down assay (Figure 3F). RIP assay further confirmed the interaction between circ_0006282 and miR-144-5p for the levels of circ_0006282 and miR-144-5p were enhanced by anti-Ago2 RIP compared to control anti-IgG groups (Figure 3G and H). Moreover, our results indicated that circ_0006282 silencing distinctly

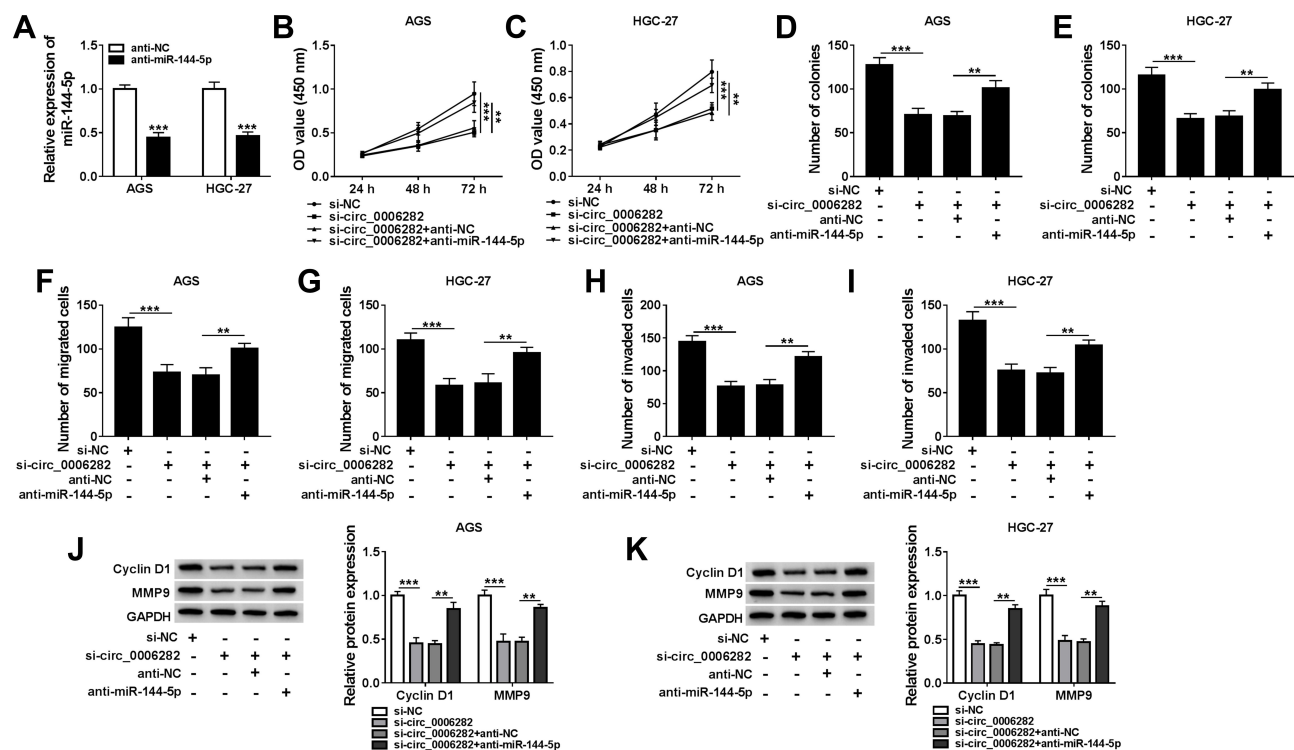


Figure 4 Inhibition of miR-144-5p ameliorated the effects of circ_0006282 knockdown on GC cell proliferation and metastasis. (A) The expression level of miR-144-5p in AGS and HGC-27 cells transfected with anti-NC or anti-miR-144-5p was determined. (B–K) AGS and HGC-27 cells were transfected with si-NC, si-circ_0006282, si-circ_0006282+anti-NC or si-circ_0006282+anti-miR-144-5p. (B–I) Cell proliferation, colony formation ability, migration and invasion of AGS and HGC-27 cells were analyzed. (J and K) The protein levels of Cyclin D1 and MMP9 were measured. ** $P < 0.01$ vs si-circ_0006282+anti-NC, *** $P < 0.001$ vs anti-NC, *** $P < 0.001$ vs si-NC.

increased miR-144-5p level in AGS and HGC-27 cells compared to si-NC groups (Figure 3I). These data indicated that circ_0006282 negatively regulated miR-144-5p expression in GC cells by sponging miR-144-5p.

Circ_0006282 Knockdown Repressed the Malignant Behaviors of GC Cells by Targeting miR-144-5p

As presented in Figure 4A, anti-miR-144-5p transfection significantly reduced miR-144-5p level in AGS and HGC-27 cells compared to anti-NC groups. To further explore the relationship between circ_0006282 and miR-144-5p in the biological behaviors of GC cells, AGS and HGC-27 cells were divided into 4 groups: si-NC, si-circ_0006282, si-circ_0006282+anti-NC and si-circ_0006282+anti-miR-144-5p. As illustrated by CCK-8 assay, colony formation assay and transwell assay, the inhibitory effects on the proliferation, colony formation, migration and invasion of AGS and HGC-27 cells mediated by circ_0006282 silencing were partially reversed by decreasing miR-144-5p (Figure 4B–I). Moreover, the results of Western blot assay showed that circ_0006282 knockdown markedly reduced the levels of Cyclin D1 and MMP9 in AGS and HGC-27 cells, while

miR-144-5p inhibition abrogated the impacts (Figure 4J and K). To sum up, circ_0006282 knockdown decelerated GC cell proliferation and metastasis by targeting miR-144-5p.

YWHAB Was the Target Gene of miR-144-5p

As shown in Figure 5A and B, the mRNA and protein levels of YWHAB were all obviously enhanced in GC tissues compared to adjacent non-tumor tissues. We also found that YWHAB protein level was elevated in AGS and HGC-27 cells in comparison with GES-1 cells (Figure 5C). Then miR-144-5p was successfully transfected into AGS and HGC-27 cells, as demonstrated by the upregulation of miR-144-5p in AGS and HGC-27 cells after miR-144-5p transfection (Figure 5D). By searching online website TargetScan, YWHAB was found to be a direct target gene of miR-144-5p and their potential binding sites were presented in Figure 5E. The dual-luciferase reporter assay was then conducted to confirm the interaction between miR-144-5p and YWHAB. The results showed that miR-144-5p transfection apparently inhibited the luciferase activity of YWHAB-3'UTR-WT in AGS and HGC-27 cells, but did not affect the luciferase activity of YWHAB-3'UTR-MUT (Figure 5F and G). RNA

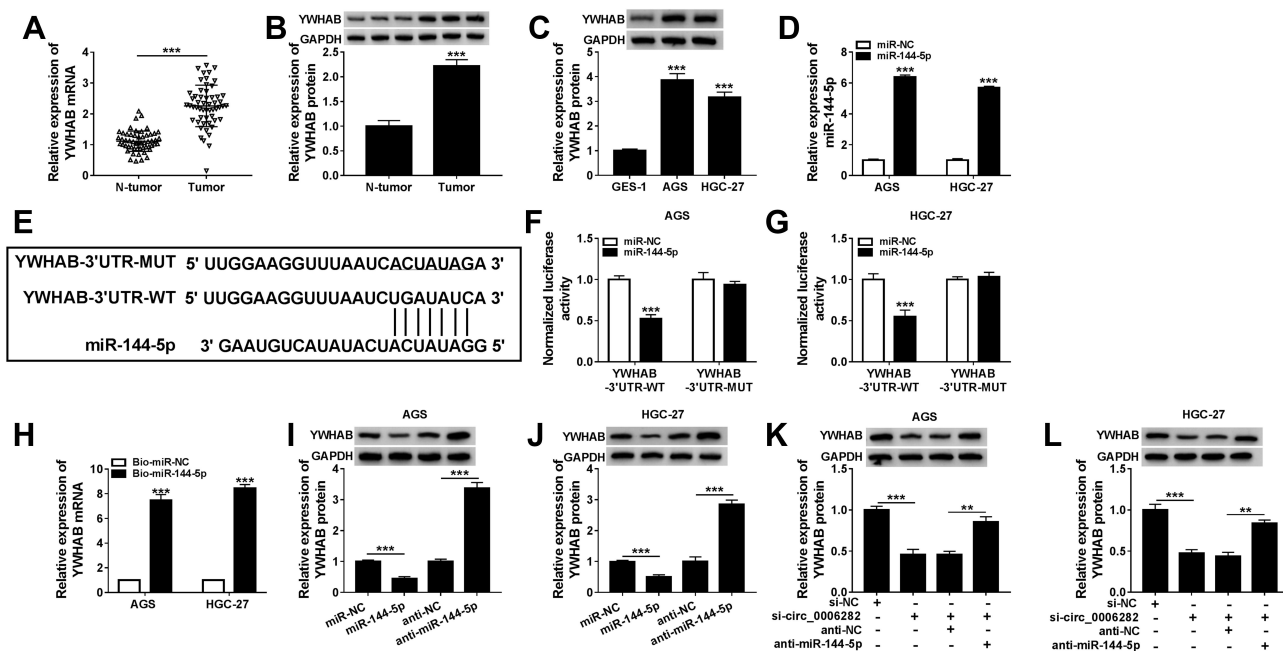


Figure 5 YWHAB was directly targeted by miR-144-5p in GC cells. (A and B) The mRNA and protein levels of YWHAB in tumor tissues and non-tumor tissues were measured. (C) The protein level of YWHAB in GES-1, AGS and HGC-27 cells was detected. (D) The expression level of miR-144-5p in AGS and HGC-27 cells transfected with miR-144-5p and miR-NC was determined. (E) The potential binding sites between miR-144-5p and YWHAB. (F–H) The combination between miR-144-5p and YWHAB was verified by dual-luciferase reporter assay and RNA pull-down assay. (I and J) The protein level of YWHAB in AGS and HGC-27 cells transfected with miR-NC, miR-144-5p, anti-NC or anti-miR-144-5p was measured. (K and L) After AGS and HGC-27 cells were transfected with si-NC, si-circ_0006282, si-circ_0006282+anti-NC or si-circ_0006282+anti-miR-144-5p, the protein level of YWHAB was measured. ** $P < 0.01$ vs si-circ_0006282+anti-NC, *** $P < 0.001$ vs N-tumor, *** $P < 0.001$ vs GES-1, *** $P < 0.001$ vs miR-NC, *** $P < 0.001$ vs Bio-miR-NC, *** $P < 0.001$ vs anti-NC, *** $P < 0.001$ vs si-NC.

pull-down assay further demonstrated the interaction between miR-144-5p and YWHAB for the level YWHAB was evidently enriched by Bio-miR-144-5p assay compared to Bio-miR-NC control groups (Figure 5H). Furthermore, we observed that miR-144-5p overexpression strikingly decreased the protein level of YWHAB in both AGS and HGC-27 cells, while miR-144-5p suppression evidently increased the protein level of YWHAB (Figure 5I and J). Additionally, our results showed that circ_0006282 knockdown led to a marked reduction of YWHAB protein level in AGS and HGC-27 cells, while miR-144-5p inhibition abolished the impact (Figure 5K and L). All these data suggested that circ_0006282 positively regulated YWHAB expression by targeting miR-144-5p.

Overexpression of YWHAB Reversed the Suppressive Roles of Circ_0006282 Knockdown in GC Cell Malignant Behaviors

As presented in Figure 6A and B, YWHAB transfection distinctly elevated YWHAB protein level in AGS and HGC-27 cells compared to control groups, indicating the successful transfection of YWHAB. Subsequently, AGS and HGC-27 cells were assigned to 4 groups: si-NC, si-circ_0006282, si-circ_0006282+Vector and si-circ_0006282+YWHAB. The results of CCK-8 assay, colony formation assay and transwell assay indicated that YWHAB overexpression ameliorated the inhibitory effects

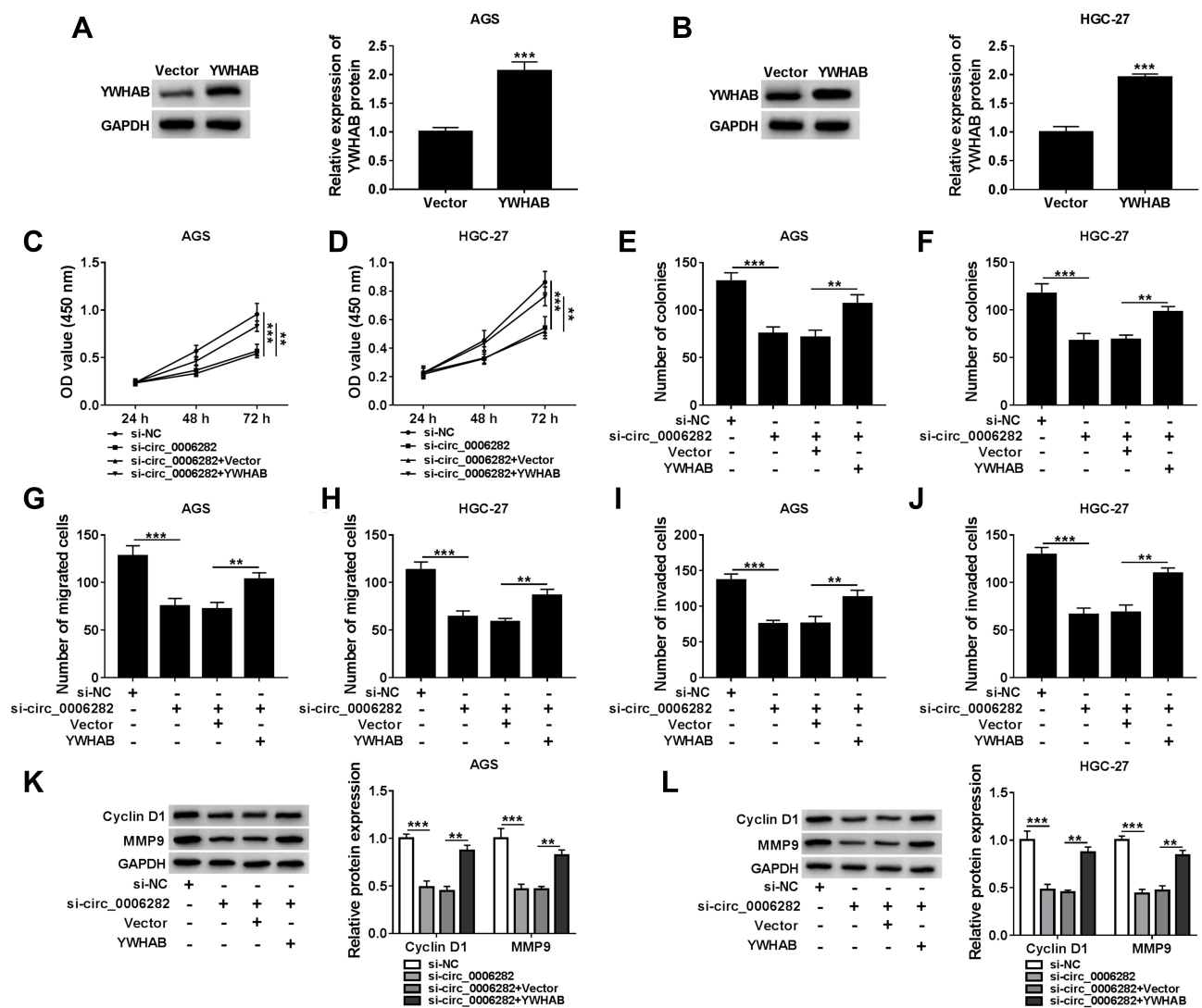


Figure 6 YWHAB overexpression abolished the impacts of circ_0006282 knockdown on GC cell proliferation and metastasis. (A and B) The protein level of YWHAB in AGS and HGC-27 cells transfected with Vector or YWHAB was measured. (C–L) AGS and HGC-27 cells were transfected with si-NC, si-circ_0006282, si-circ_0006282+Vector or si-circ_0006282+YWHAB. (C–J) The proliferation, colony formation, migration and invasion of AGS and HGC-27 cells were analyzed. (K and L) The protein levels of Cyclin D1 and MMP9 in AGS and HGC-27 cells were determined. ** $P < 0.01$ vs si-circ_0006282+Vector, *** $P < 0.001$ vs Vector, **** $P < 0.0001$ vs si-NC.

of circ_0006282 silencing on cell proliferation, colony formation, migration and invasion in AGS and HGC-27 cells (Figure 6C–J). Furthermore, the elevation of YWHAB rescued the downregulation of Cyclin D1 and MMP9 in AGS and HGC-27 cells mediated by circ_0006282 silencing (Figure 6K and L). Taken together, circ_0006282 knockdown hampered GC cell progression by downregulating YWHAB.

Circ_0006282 Knockdown Blocked Tumorigenesis of GC in vivo

At last, the functional role of circ_0006282 in GC progression in vivo was investigated by murine xenograft model assay. As a result, the mice with sh-circ_0006282 transfected AGS cells showed reduced tumor volume and tumor weight compared to control groups (Figure 7A and B). Moreover, our results showed that circ_0006282, YWHAB mRNA and YWHAB protein levels were all

declined and miR-144-5p level was raised in the tissues collected from sh-circ_0006282 groups compared to sh-NC groups (Figure 7C and D). In summary, circ_0006282 silencing repressed tumor growth of GC in vivo.

Discussion

Currently, there is a growing body of evidence exhibited that circRNAs play pivotal roles in the malignancy of GC.^{30–32} Herein, we focused on the relationship between circ_0006282 and GC cell malignant phenotypes. It was found that circ_0006282 aggravated the progression of GC. Furthermore, the regulatory network of circ_0006282/miR-144-5p/YWHAB axis was discovered in GC.

CircRNAs are key regulators implicated in GC malignant biological behaviors.^{11,12,33} Shao et al disclosed that circ_0006282 was aberrantly improved in GC tissues.³⁴ He et al reported that circ_0006282 level was closely linked to TNM stage, tumor size and lymph nodes

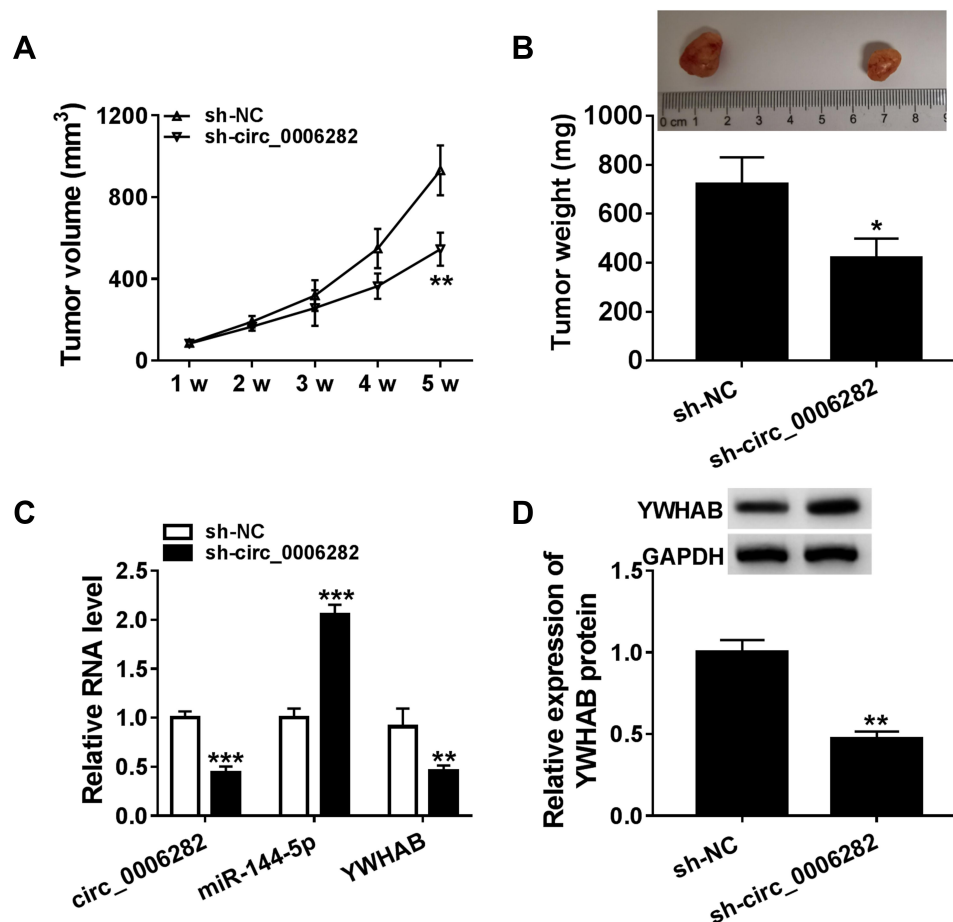


Figure 7 Circ_0006282 deficiency hampered tumor growth in vivo. (A) Tumor volume was monitored every week. (B) Tumor weight was examined after 5 weeks. (C) The levels of circ_0006282, miR-144-5p and YWHAB mRNA in the collected tumors were determined by qRT-PCR assay. (D) The protein level of YWHAB in the collected tumors was measured by Western blot assay. * $P < 0.05$ vs sh-NC, ** $P < 0.01$ vs sh-NC, *** $P < 0.001$ vs sh-NC.

motility, and promoted GC cell growth, migration and invasion by competitively binding to miR-155 and reducing FBXO22.¹³ Corresponding to these previous reports, circ_0006282 was strikingly elevated in GC tissues and cells. Moreover, interference of circ_0006282 restrained the proliferation, colony formation, migration and invasion capacities of GC cells in vitro and blocked the tumorigenicity of GC in vivo. Cyclin D1 is a critical protein in G1 phase, which can promote the rapid transformation from G1 to S phase through accelerating DNA replication and mitosis.³⁵ MMP9 can degrade extracellular matrix and basement membrane, thereby promoting tumor invasion and metastasis.^{36,37} Thus, in this study, we investigated the effects of circ_0006282 on the levels of cyclin D1 and MMP9. As a result, cyclin D1 and MMP9 levels were reduced in GC cells following the downregulation of circ_0006282. Taken together, circ_0006282 facilitated GC malignancy.

Subsequently, the regulatory mechanism of circ_0006282 in GC carcinogenesis was further investigated. As a result, circ_0006282 functioned as the sponge for miR-144-5p to positively modulate YWHAB expression in GC cells. Thereafter, our study explored the relationships among circ_0006282, miR-144-5p and YWHAB in regulating GC cell malignant behaviors for the first time. MiR-144-5p level was declined in GC. Moreover, the impacts of circ_0006282 interference on tumor cell growth and metastasis were abolished by miR-144-5p suppression, suggesting that miR-144-5p might act as a tumor inhibitor in GC. In support of our findings, Li et al declared that miR-194 level was decreased in GC, and regulated GC cell proliferation and motility by targeting ZFX.³⁸ Yao et al uncovered that miR-144 upregulation suppressed cell viability and growth, as well as induced cell apoptosis and cell cycle arrest in GC cells by binding to COX-2.³⁹ Additionally, previous reports indicated that YWHAB level was enhanced in GC and accelerated the growth, invasion and migration of GC cells.^{40,41} Consistently, we demonstrated the enhancement of YWHAB level in GC tissues and cells. Moreover, YWHAB overexpression ameliorated the suppressive roles of circ_0006282 knockdown in GC cell growth and metastasis.

However, there were still some shortcomings in our study. For example, the tumor tissue samples were not enough and the function of circ_0006282 in tumor metastasis in vivo has not been explored. We will validate our

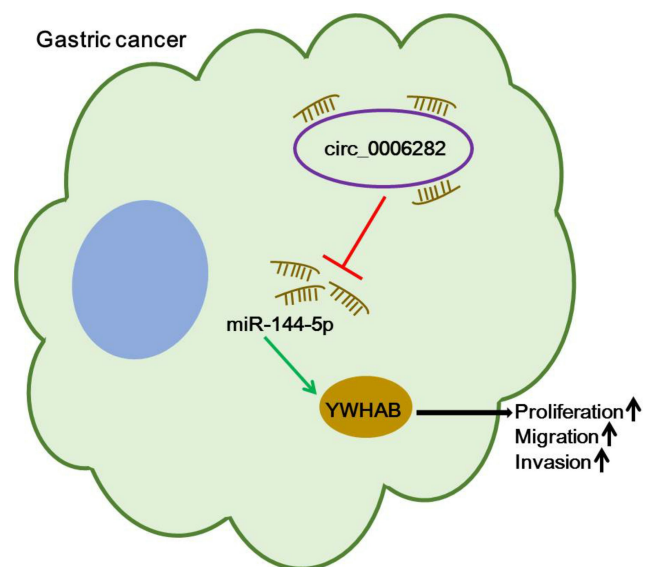


Figure 8 The schematic diagram of circ_0006282/miR-144-5p/YWHAB axis in the modulation of GC cell proliferation, migration and invasion.

results with more tumor samples and explore the role of circ_0006282 in tumor metastasis.

In summary, our present study unveiled that circ_0006282 sponged miR-144-5p to increase YWHAB expression, thereby contributing to GC cell proliferation, migration and invasion, as presented in Figure 8. This research might bring new light to the pathogenesis of GC and offer a target for GC therapy.

Abbreviations

GC, gastric cancer; circRNAs, circular RNAs; QRT-PCR, quantitative real-time polymerase chain reaction; TCEB1, transcription elongation factor B subunit 1; YWHAB, tyrosine 3-monooxygenase/tryptophan 5-monooxygenase activation protein β ; CCK-8, Cell Counting Kit-8; MMP9, matrix metalloprotein 9; RIP, RNA immunoprecipitation (RIP); MiRNAs, microRNAs; CREB1, cAMP responsive element binding protein 1; YBX1, Y-box binding protein-1; FBXO22, F-box protein 22; 3'UTR, 3' untranslated regions; SIRT1, sirtuin 1; PTP1B, protein-tyrosine phosphatase 1B; RPMI1640, Roswell Park Memorial Institute 1640 Medium; FBS, fetal bovine serum; GAPDH, glyceraldehyde 3-phosphate dehydrogenase; OD, optical density; RIPA, Radio Immunoprecipitation Assay; SDS-PAGE, sodium dodecyl sulfonate-polyacrylamide gel; PVDF, polyvinylidene difluoride; ZFX, zinc finger protein X-linked; COX-2, cyclooxygenase-2.

Disclosure

The authors declare no conflicts of interest.

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