

ERCCI and XRCCI as biomarkers for lung and head and neck cancer

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Abstract: Advanced stage non-small cell lung cancer and head and neck squamous cell carcinoma are both treated with DNA damaging agents including platinum-based compounds and radiation therapy. However, at least one quarter of all tumors are resistant or refractory to these genotoxic agents. Yet the agents are extremely toxic, leading to undesirable side effects with potentially no benefit. Alternative therapies exist, but currently there are no tools to predict whether the first-line genotoxic agents will work in any given patient. To maximize therapeutic success and limit unnecessary toxicity, emerging clinical trials aim to inform personalized treatments tailored to the biology of individual tumors. Worldwide, significant resources have been invested in identifying biomarkers for guiding the treatment of lung and head and neck cancer. DNA repair proteins of the nucleotide excision repair pathway (*ERCCI*) and of the base excision repair pathway (*XRCCI*), which are instrumental in clearing DNA damage caused by platinum drugs and radiation, have been extensively studied as potential biomarkers of clinical outcomes in lung and head and neck cancers. The results are complex and contradictory. Here we summarize the current status of single nucleotide polymorphisms, mRNA, and protein expression of *ERCCI* and *XRCCI* in relation to cancer risk and patient outcomes.

Keywords: nucleotide excision repair, base excision repair, DNA damage, DNA repair, chemotherapy, NSCLC, HNSCC, single nucleotide polymorphism

Introduction

Lung cancer is the second most common cancer in the USA and is the leading cause of cancer-related death.¹ Based on the predicted response to treatment and known risk factors, lung cancers are categorized in two groups: small cell and non-small cell lung cancers (NSCLC). NSCLC are more frequent, and smoking is a risk factor. Histologically, NSCLC are composed mainly of adenocarcinoma and, to a lesser degree, of squamous cell carcinoma (SCC) and large cell carcinoma. Treatment varies based on clinical stage. Early stage NSCLC is treated with surgery, while loco-regionally advanced and metastatic cancers are treated with multidrug systemic chemotherapy, which often includes a platinum compound.²

Head and neck cancers are similar to NSCLC in many respects, although they are less common, representing the eighth most frequent type of cancer in the USA.¹ Smoking is a recognized risk factor for head and neck cancers, like for NSCLC. Pathologically, cancers of the aerodigestive tract are mostly head and neck squamous cell carcinoma (HNSCC). As for NSCLC, early stage HNSCC is successfully treated with surgery, while treatment of loco-regionally advanced tumors includes systemic therapy.²⁻⁴ Frequently, concomitant radiotherapy and chemotherapy with a platinum-based DNA

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damaging agent (cisplatin or carboplatin) is used, either as primary treatment or as adjuvant post-operative therapy. Alternative systemic treatments that do not rely upon DNA damage, such as taxanes, base analogs, and anti-metabolites can also be used.⁴ However, currently we do not have the tools to predict which patients will respond best to the various possible therapies.

To maximize treatment success of NSCLC and HNSCC, and to reduce unnecessary toxicity, there is great demand for identifying biomarkers that predict clinical outcomes prospectively. The goal is to measure validated biomarker(s) in individual tumors to probe the biology of each tumor and predict whether it is likely to be vulnerable to genotoxic agents such radiation and platinum drugs. This would enable identification of patients likely to be resistant to these modalities, allowing use of alternative therapies, preventing unnecessary toxic side-effects, and improving clinical outcomes.

Choosing a biomarker Biomarkers in DNA repair pathways

DNA repair proteins are obvious candidate biomarkers for predicting how tumors will respond to genotoxic stress. The prediction is that overexpression of DNA repair proteins in tumors could mediate resistance to genotoxic therapies and therefore poor outcomes. In turn, persons with inherited defects in DNA repair mechanisms are frequently exquisitely hypersensitive to radiation and/or genotoxic agents. This is true of patients with ataxia telangiectasia (AT), ataxia telangiectasia-like disorder, severe combined immunodeficiency, Ligase IV syndrome, Rothmund–Thompson syndrome, Seckel syndrome, Werner syndrome, Nijmegen breakage syndrome, all due to defective repair of double-strand breaks (DSBs)⁵ or stalled replication forks.⁶ It is also true of patients with Fanconi anemia caused by defective repair of DNA interstrand crosslinks (ICLs) and patients with xeroderma pigmentosum due to a defect in nucleotide excision repair (NER) of helix-distorting DNA adducts.^{7,8} Since NSCLC and HNSCC are treated with cisplatin and radiation therapy, it is logical to predict that patients with reduced DSB repair, single-strand break (SSB) repair, ICL repair, or NER due to polymorphisms affecting the expression or function of DNA repair proteins might be most responsive to DNA damaging agents.

ERCC1-XPF repair endonuclease

ERCC1 is an attractive candidate biomarker. *ERCC1* partners with XPF to form a bi-partite nuclease that is essential for NER and ICL repair, and participates in DSB repair (Figure 1).^{9–12}

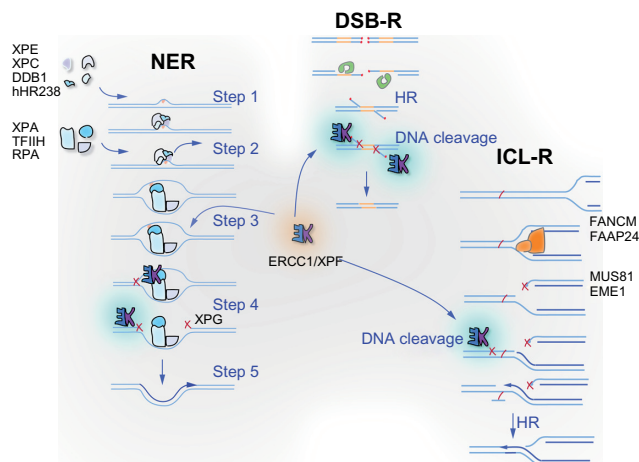


Figure 1 *ERCC1* and its obligate binding partner XPF are involved in multiple DNA repair pathways. *ERCC1*-XPF heterodimer is an endonuclease that cuts one strand of DNA at a double-strand: single-strand junction. It is critical for nucleotide excision repair (NER) of bulky chemical DNA adducts like cisplatin intrastrand crosslinks, the repair of double-strand breaks that cannot be directly ligated back together like those induced by ionizing radiation, and the repair of interstrand crosslinks (ICLs). In NER (represented on the left), adducts that cause distortion of the DNA double helix are detected by XPC-hHR23B, in some cases with the assistance of XPE-DDB1 (Step 1). These complexes recruit of TFIIH, which unwinds the DNA around the adduct and XPA and RPA, which stabilize the open complex (Step 2). XPA recruits *ERCC1*-XPF to cut the damaged strand 5' to the adduct (Step 3), while TFIIH recruits a second endonuclease XPG to cut 3' of the lesion (Step 4). The damaged base is removed as part of a single-stranded oligonucleotide. The replication machinery uses the 3'-OH created by *ERCC1*-XPF incision to prime DNA synthesis to fill the gap (Step 5). After ligation, the integrity of the DNA is fully restored. In double-strand breaks (DSB) repair (represented in the middle), two broken ends can be spliced together if they have long patches of sequence homology via homologous recombination (labeled HR) or if they have small patches of homology, known as microhomology, very close to the broken ends via alternative end-joining. In both cases, *ERCC1*-XPF is needed to remove 3' single-stranded flaps of non-homologous sequence at the ends of the breaks (labeled DNA cleavage) to allow sealing of the spliced ends by a DNA ligase. ICLs (represented on the right) are predominantly repaired during S phase of the cell cycle. ICLs are an absolute block to replication and when encountered by the replication machinery lead to the collapse of the replication fork and creation of a DSB. This DSB cannot be repaired until *ERCC1*-XPF cuts near the ICL to release it from one strand (DNA cleavage), allowing bypass of the adduct by a translesion polymerase such as REV1/Polζ.

Platinum-based chemotherapy drugs react with DNA to induce adducts that affect one strand of DNA (monoadducts and intrastrand crosslinks), which are repaired by NER, as well as adducts that affect both strands (ICLs), which are repaired by a distinct DNA repair mechanism: ICL repair.^{13–15} Because *ERCC1*-XPF is unique in being required for both NER and ICL repair pathways, it is the only enzyme required for removal of all types of DNA lesions caused by cisplatin and carboplatin. In addition, it facilitates the repair of DNA lesions caused by radiation therapy (bulky oxidative lesions and DSBs).¹⁰ Hence, it has been proposed that decreased expression of *ERCC1*-XPF might mediate increased susceptibility to chemoradiation and improved clinical outcome. It is therefore not surprising that *ERCC1* has been extensively evaluated as a biomarker in NSCLC and HNSCC, with over 90 peer-reviewed reports published on the subject.

However, it is important to emphasize that the expression level of ERCC1-XPF has not been established as rate limiting for NER, ICL, or DSB repair, therefore the influence of ERCC1-XPF protein levels on the DNA repair capacity of cells or tumors is not known.

XRCC1 scaffold protein

XRCC1 is an equally promising candidate biomarker involved in the repair of oxidative DNA damage and single-strand breaks (SSBs) (Figure 2), two types of DNA damage abundantly produced by ionizing radiation. XRCC1 does not have enzymatic activity, but it is a critical scaffold protein for base excision repair (BER) and SSB repair (reviewed in Kennedy and D'Andrea,⁸ Hoeijmakers,¹⁶ Ladiges,¹⁷ and Almeida and Sobol).¹⁸ XRCC1 interacts strongly with PARP1, which recognizes SSBs, and LIGIII that seals SSBs and BER intermediates.^{17,19} Cells lacking XRCC1 are hypersensitive to ionizing radiation, oxidative stress and alkylating agents (reviewed by Caldecott).¹⁹ It is therefore plausible that reduced expression of XRCC1 in cancer patients may lead to increased susceptibility to chemoradiation and improved

patient survival. However, like ERCC1-XPF, *XRCC1* has not been established as rate limiting for DNA repair. Thus, the impact of low expression of XRCC1 on a cell's capacity for BER and SSB is not known.

Methods to assess biomarkers and clinical endpoints

Available methods to interrogate DNA repair

Directly measuring NER, DSB repair, ICL repair, or BER would be the ideal method for predicting an individual's DNA repair capacity. However measuring DNA repair requires viable, and for some pathways, replicating cells. Thus, currently it is not possible to rapidly measure DNA repair in clinical samples because it first requires establishing a cell line from peripheral blood mononuclear cells, dermal fibroblasts, or tumors. Hence measuring DNA repair protein expression is used as a surrogate. Multiple techniques are available to measure ERCC1 and XRCC1 expression including immunohistochemistry or immunofluorescence of fixed tissue sections, quantification of mRNA expression by qRT-PCR, or quantification of protein expression by immunoblot if frozen specimens are available. It must be strongly emphasized, however, that it is not established that ERCC1 is rate limiting for NER or ICL repair, or that XRCC1 is rate limiting for BER or SSB repair. *ERCC1* and *XRCC1* can also be investigated by sequencing DNA to detect functional single nucleotide polymorphisms (SNP) affecting protein function or expression level.

Measuring protein expression

Immunohistochemistry (IHC) and immunofluorescence are semi-quantitative methods that permit estimation of protein expression level in clinical samples. The intensity of the histochemical reaction or fluorescent signal varies with the expression level of the protein of interest and can be scored as positive versus negative or on a graded scale. These methods are advantageous since they employ paraffin embedded tissue specimens, which are readily available. However, several caveats must be considered while interpreting data from immunohistochemical methods. Protein expression within a given tumor may vary from one area to another.^{20,21} Therefore expression measured on a biopsy specimen or in a tissue core in an array, which represent only a small fraction of a tumor, may not reflect overall expression. In one patient cohort, however, it was established that ERCC1 expression in biopsies correlated with expression measured in tumor sections.²² Another important technical consideration is the

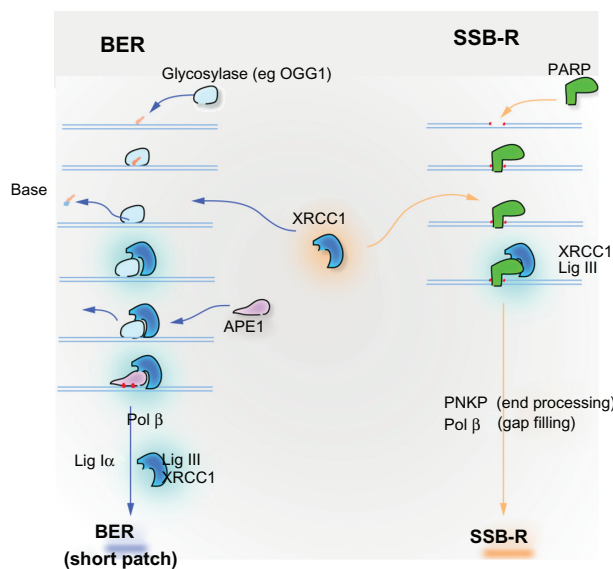


Figure 2 XRCC1 is instrumental in base excision repair (BER) of small oxidative lesions and a related mechanism for the repair of single-strand breaks (SSB-R), both caused by ionizing radiation. Oxidative damage and alkylation leads to small alterations of bases that are principally repaired through BER pathway. Damaged bases are recognized and excised by glycosylases, such as OGG1, which removes the abundant oxidative lesion 8-oxodeoxyguanosine. Excision of the damaged base leaves an abasic (AP) site. The DNA backbone adjacent to the AP site is incised by APE1 endonuclease to create a single-strand break (SSB). XRCC1 has no enzymatic activity, but is critical as a scaffolding protein in BER. It is recruited to the site of damage by the glycosylase or by PARP1, which binds the newly created SSB. XRCC1 forms a tight complex with LIG3, the ligase that seals the SSB repair intermediate to complete BER. Primary SSBs, a common consequence of ionizing radiation, are directly recognized by PARP1, which recruits XRCC1-LIG3 to repair the broken strand. PNKP removes 3' phosphate groups that block DNA ligation by LIG3. Polβ may be required to replace missing nucleotides at the site of the break.

fact that tissue collection method, handling, storage, fixation, processing, and analysis influence the biomarker readout, and causes inter-study variability.²³ This has led to the publication of guidelines for evaluation of biomarkers, in an attempt to unify methods of biomarker analysis.²⁴

Equally important, immunodetection methods are by definition indirect measures of protein expression, dependent upon the sensitivity and specificity of the antibody used. The specificity of the commercially available antibodies is rarely rigorously tested. ERCC1 protein expression was erroneously quantified in virtually all oncology studies prior to 2010 due to the implementation of an antibody raised against ERCC1 that lacks specificity.²⁵ Finally, methods for quantifying and scoring biomarker expression vary from study to study, and are somewhat subjective. For instance, biomarker positivity can be defined as the presence of any staining detected by a pathologist, calculated as an H-score based on the staining intensity and number of positive cells, or quantified by an automated system to minimize subjectivity. Thus, while immunohistochemical methods are potentially useful for quantifying biomarker protein expression, multiple factors can introduce intra- or inter-study variability.

Measuring mRNA expression

mRNA expression is often used as a surrogate marker for protein expression. Typically this is done by quantitative RT-PCR, using primers specific for the target biomarker. The advantages of quantifying mRNA are that the method is very sensitive, highly specific, and can be applied to fixed specimens. However, quantitative methods to measure mRNA levels are not readily available outside of biomedical research facilities. Importantly, mRNA and protein expression do not always correlate.^{26,27} Translational regulation, post-translational modification and protein stability alter protein levels independently of mRNA.²⁸ So while mRNA levels can be a useful biomarker to predict clinical outcomes, mRNA levels do not necessarily reflect protein levels. Therefore, changes in mRNA levels should not be used to infer changes in biological activity in the absence of experimental evidence.

Genomic approaches

Base changes in a gene can lead to reduced expression of the encoded protein if they affect the promoter, 5' or 3' untranslated sequence, regulatory miRNA binding sites, splice sites, or the coding sequence if the change leads to protein misfolding or destabilization, or utilization of a less abundant tRNA during translation. Missense mutations in the coding sequence can also alter protein function by affecting

protein:protein interactions or catalytic activity. Single nucleotide polymorphisms (SNPs) are defined as single base changes that occur in more than 1% of the population. They occur every 360 bases in the human genome, and, thus, affect all genes (reviewed by Kim and Misra).²⁹ The National Center for Biotechnology Information (www.ncbi.nlm.nih.gov/projects/SNP) reports 246 SNPs in *ERCC1*, and 550 SNPs in *XRCC1*. In silico, in vitro, or epidemiological studies can be used to identify SNPs with the highest likelihood of being a useful biomarker. This includes SNPs with a known impact on mRNA level or protein expression, or activity. Fourteen SNPs in *ERCC1* and eleven for *XRCC1* have been investigated in NSCLC and/or HNSCC. The advantages of analyzing SNPs as biomarkers are that multiple SNPs can be evaluated in one sample using an array and DNA hybridization method and require only DNA extracted from a simple blood draw.^{29,30} However, it is important to remember that the genotype of a tumor may differ from the germline genotype found in the rest of the body, as tumors are inherently genomically unstable and accumulate DNA mutations. Therefore SNPs identified in a patient's blood sample may not reflect a patient's tumor's genotype.³¹ Furthermore, because SNPs are much more abundant than recombination events in the human genome, they are inherited in clusters, referred to as haplotypes. Thus, a SNP in *ERCC1* or *XRCC1* could be a useful biomarker for predicting outcomes in cancer without having any impact on DNA repair.

Clinical endpoints

In oncology, clinical outcomes for which it would be desirable to have biomarkers include: (1) risk of cancer, (2) prognosis in untreated patients, (3) tumor response to therapy, (4) severity of treatment-related toxicities, (5) progression-free survival, and (6) overall survival. DNA repair-related endpoints could logically contribute to any of these endpoints, in particular when genotoxic chemotherapeutics or radiation is the therapy of choice.

One of the most widely recognized risk factors for NSCLC and HNSCC is smoking. The pathogenesis of these tumors involves tobacco-related DNA damage. It is rational to hypothesize that persons with low expression of ERCC1 or XRCC1 may have impaired ability to remove tobacco-induced DNA damage and therefore are more likely to develop smoking-related cancers. The best way to test this hypothesis is with well-powered prospective risk analysis. But these types of studies are difficult to conduct because they necessitate large cohorts and long follow-up times. For instance, >520,000 patients would have to be followed for

10 years to find 116 lung cancer and 82 HNSCC.³² Thus, most published studies evaluating cancer risk associated with *ERCC1* and *XRCC1* are retrospective case-control studies, which have their inherent limitations.

Since DNA repair-related biomarkers could have value for multiple clinical endpoints, they could potentially have prognostic or predictive value. *Prognostic* biomarkers estimate progression-free or overall survival in an untreated patient population. It gives information on the natural course of the disease.³³ In contrast, *predictive* biomarkers estimate how likely a given treatment is expected to work (efficacy). Predictive value is determined in prospective randomized trial settings with treatment and control arms. Both prognostic and predictive biomarkers are useful but they require different study designs. Once identifying a biomarker of interest, validation is essential and ultimately the greatest barrier to implementation of the biomarker in clinic practice.³⁴ Validation includes establishing that a biomarker of interest (expression, genotype) consistently predicts a particular clinical outcome (response rate, progression free survival, overall survival). Thus, validation requires multiple clinical studies conducted by multiple independent groups. With these considerations in mind, we now critically review the literature on *ERCC1* and *XRCC1* SNPs as biomarkers in NSCLC and HNSCC.

ERCC1 as biomarker for NSCLC and HNSCC

ERCC1 as a biomarker for cancer risk

Two SNPs, Asn118Asn and C8092A, have been described as potentially affecting *ERCC1* expression. Asn118Asn involves a synonymous polymorphism at codon 118, where AAC is changed to AAT. While the amino acid sequence does not change, the variant (T) allele is associated with lower mRNA and protein levels in ovarian cancer cells.^{35,36} C8092 is in the 3'-UTR of *ERCC1*. The 3'-UTR is implicated in translational repression of *ERCC1* mRNA.²⁸ However the impact of the polymorphism on *ERCC1* protein expression has not been critically evaluated to date. In patients, the C8092A polymorphism correlates neither with mRNA,³⁷ nor with protein levels.³⁸ Numerous other SNPs in *ERCC1* have been studied, but like C8092, their functional impact on *ERCC1* expression or activity has not been clearly established.

Studies evaluating *ERCC1* as a potential biomarker to predict the risk of developing NSCLC or HNSCC rest principally on SNP analysis. There are ten studies examining *ERCC1* SNPs in relation to NSCLC.^{32,39-47} In these studies,

only 14 of 246 reported SNPs in *ERCC1* were evaluated, with just six SNPs analyzed in greater than one study (Table 1). Most report retrospective case-controlled studies focused on Asn118, C8092, and IVS3. While case-control studies are important for identifying new biomarkers, they have inherent biases that can limit the generalization of the results. For instance, if the biomarker is not robust, confounding factors in the cohort may lead to erroneous conclusions. In most of the retrospective studies, SNPs in *ERCC1* were not significantly associated with susceptibility of developing NSCLC.^{32,39-42,46-48} However, there was not good concordance between studies.⁴²⁻⁴⁵ To clarify the role of SNPs in *ERCC1* as risk factor for NSCLC, meta-analyses were done. When patients from the diverse studies were combined into large data pools, none of the four SNPs in *ERCC1* meeting study inclusion criteria reached statistical significance as a risk factor for NSCLC.⁴⁸⁻⁵⁰ Furthermore, mRNA levels in blood samples were not identified as a risk factor for lung cancer.⁵¹ In summary, our review of the literature suggests that neither SNPs in *ERCC1* studied to date by more than one group, nor peripheral mRNA levels, constitute a risk factor for NSCLC.

Head and neck cancers are less common than lung cancer. Hence clinical studies to identify biomarkers that predict the risk of developing HNSCC are less frequent and smaller. We identified six studies evaluating whether polymorphisms in *ERCC1* are a risk factor for HNSCC (Table 1).^{32,47,52-55} Only four SNPs were assessed more than once: (Asn118Asn), (C8092A), 119216 C > G, and 4855 C > T. None showed statistically significant association with risk of HNSCC, with the exception of one large case control study in which 4855 C > T appeared to be protective.⁵⁴ One small retrospective case-controlled study suggested that low *ERCC1* mRNA in peripheral blood might be a risk factor for HNSCC,⁵⁶ but the findings could not be confirmed by others after multivariate analysis.³⁷ Therefore, we conclude that none of the SNPs in *ERCC1* tested thus far, nor peripheral *ERCC1* mRNA levels are definitive risk factors for HNSCC. However, 4855 C > T deserves close attention in future studies. Further, we cannot exclude the possibility that these or other *ERCC1* SNPs may be useful biomarkers in selected subpopulations for predicting cancer risk.

ERCC1 SNPs as biomarkers for clinical outcome

Polymorphisms in *ERCC1* could affect tumor sensitivity to treatment, and hence influence patient outcomes. Patients with a

Table 1 Association between SNPs in *ERCC1* and cancer risk

Cancer	rs	SNPs	Alternate names	Reference	n (case-control)	Risk ^a
NSCLC	rs11615	Asn118 Asn	C118T; 354 C > T; T19007C; C19007T; 3525 C > T	Zhou et al ³⁹	1752–1358	0
				Matullo et al ^{32,#}	116–> 520,000	0
				Yin et al ⁴⁰	151–143	0
				Hung et al ⁴¹	4460–5217	0
				Yu et al ⁴²	988–986	0
				Deng et al ⁴³	315–315	1
				Zienolddiny et al ⁴⁴	343–413	1
				Zhou et al ³⁹	1752–1358	0; 1 in heavy smokers
				Zienolddiny et al ⁴⁴	343–413	0
				Yu et al ⁴²	988–986	0
	rs3212986	C8092A	14443 C > A	Hung et al ⁴¹	4688–4546	0
				Shen et al ⁴⁶	122–122	0
				Jones et al ¹⁶⁷	452–790	0
				Zienolddiny et al ⁴⁴	343–413	0
	rs3212948	19716 C > G	IVS3 174G > C	Ma et al ⁴⁵	1010–1011	2
				Ma et al ⁴⁵	1010–1011	0
				Yu et al ⁴²	988–986	1
	rs3212930	(-)433 T > C		Shen et al ⁴⁶	122–122	0
				Yu et al ⁴²	1000–1000	0
	rs3212961	4855 C > T	IVS5 + 33 C > A; 17677 C > A	Zienolddiny et al ⁴⁴	343–413	0
Ma et al ⁴⁵				1010–1011	0	
rs3212955			Jones et al ¹⁶⁷	452–790	0	
			Ma et al ⁴⁵	1010–1011	0	
rs3212981			Ma et al ⁴⁵	1010–1011	0	
rs16979802	15310 C > G		Zienolddiny et al ⁴⁴	343–413	1	
rs3212951			Ma et al ⁴⁵	1010–1011	0	
rs1007616			Ma et al ⁴⁵	1010–1011	2	
rs1319052			Jones et al ¹⁶⁷	452–790	0	
rs735482			Jones et al ¹⁶⁷	452–790	0	
rs2298881	262 G > T		Yu et al ⁴²	988–986	0; (1) in smokers	
unnamed			Ma et al ⁴⁵	1010–1011	0	
HNSCC	rs11615	Asn118 Asn	354 T > C; 19007 T > C; 3525 C > T	Abbasi et al ⁵³	257–769	0
				Canova et al ⁵⁴	1511–1457	0
				Matullo et al ³²	82–> 520,000	0
	rs3212986	C8092A	14443 C > A	Abbasi et al ⁵³	257–769	0
				Sugimura et al ⁵²	122–244	(1); 1 in smoker
	rs3212948	19716 C > G	IVS3 + 74C > G	Sturgis et al ⁵⁵	313–313	(2)
				Canova et al ⁵⁴	1511–1457	0
	rs3212961	4855 C > T	IVS5 + 33C > A	Jones et al ¹⁶⁷	175–790	0
				Abbasi et al ⁵³	257–769	0
	rs1319052			Canova et al ⁵⁴	1511–1457	2
rs735482			Jones et al ¹⁶⁷	175–790	0	
rs3212955			Jones et al ¹⁶⁷	175–790	0	

Notes: ^aRisk for variable allele, 0 = non significant, (1) = trend to increased, 1 = increased, (2) = trend to protective, 2 = protective; #retrospective analysis of prospective study.

Abbreviations: HNSCC, head and neck squamous cell carcinoma; NSCLC, non-small cell lung cancers; rs, reference SNP; SNPs, single nucleotide polymorphisms.

polymorphic variant of *ERCC1*, which results in impaired NER and/or ICL repair capacity, may be exquisitely sensitive to chemotherapy with genotoxic agents or radiation. This could mean their tumors respond better to chemoradiation therapy and outcomes are improved. Alternatively, the host may be hypersensitive to genotoxic stress leading to exaggerated side effects of therapy and poor outcomes.

In NSCLC, we identified sixteen studies testing whether *ERCC1* polymorphisms influence clinical outcome,^{38,57–71} including five prospective studies (Table 2).^{58,62,69,70} The only two SNPs tested were Asn118 and C8092. The results are inconsistent, weakening the generalizability of the conclusions. When more than 500 patients from multiple studies were pooled into a single meta-analysis, Asn118 Asn

Table 2 Association between SNPs in *ERCC1* and clinical outcome

Cancer	rs	SNPs	Alternate names	Reference	n	Outcome ^a
NSCLC	rs11615	Asn118 Asn	C118T; 354 T > C; 19007 T > C; 3525 C > T	Zhou et al ⁶³	128	0
				Gandara et al (2005) ^b	526	0
				Suk et al ⁵⁹	214	0 (toxicity)
				De Las Penas et al ^{71,b}	135	0
				Tibaldi et al ⁶¹	65	0
				Takenaka et al ⁷³	122	0
				Vinolas et al ^{62,b}	94	0
				Park et al ⁶⁴	178	(1); 1 for stage III
				Ryu et al ⁶⁵	109	1
				Isla et al ⁶⁸	62	1
	Su et al ⁶⁶	230	1			
	Kalikaki et al ⁵⁷	119	1			
	Okuda et al ³⁸	90	1			
	Yin et al ⁶⁷	257	1			
	Li et al ^{70,b}	115	2			
	Zhou et al ^{69,b}	130	2			
	Zhou et al ⁶³	128	1			
	Suk et al ⁵⁹	214	1 (toxicity)			
	Park et al ⁶⁴	178	0			
	Okuda et al ³⁸	90	1			
Takenaka et al ⁷³	122	1				
Kalikaki et al ⁵⁷	119	2				
Li et al ^{70,b}	115	2				
HNSCC	rs3212986	C8092A	14443 C > A	Quintela-Fandino et al ⁷⁴	103	-1
	rs735482	Lys259Thr	1264 A > C	Grau et al ^{75,b}	47	0
				Carles et al ⁷⁶	108	1 (but only 4% of carrier)

Notes: ^aOutcome for variable allele, 0 = non significant, (1) = trend to worse, 1 = worse, (2) = trend to better, 2 = better; ^bprospective study.

Abbreviations: HNSCC, head and neck squamous cell carcinoma; NSCLC, non-small cell lung cancers; rs, reference SNP; SNPs, single nucleotide polymorphisms.

was predictive of tumor response to chemotherapy.⁷² As expected, the variant allele (C→T), which presumably causes lower *ERCC1* expression, correlated with a higher response rate.⁷² However, this meta-analysis excluded one important report, a large phase Phase III study (n = 526) in which Asn118 did not predict clinical outcome, including response to treatment.⁵⁸ These conflicting results, derived from equally large studies, suggest that this *ERCC1* SNP is not a robust predictive biomarker in an unselected population. To our knowledge, C8092 has not been evaluated in a large prospective study or in a meta-analysis as a predictor of clinical outcomes in NSCLC. In retrospective cohorts, C8092 showed mixed results as predictive biomarker. The general tendency was slightly weighed toward the variant allele (C→A) predicting worse outcomes.^{38,59,63,73} In summary, none of the SNPs in *ERCC1* tested have been identified as strongly predictive biomarkers for outcomes in NSCLC, but C8092 emerges as a potentially promising candidate.

In HNSCC, we identified only three studies evaluating the predictive value of SNPs in *ERCC1* (Table 2).⁷⁴⁻⁷⁶ Like

NSCLC, in HNSCC, there was a trend towards an association between the variant allele of C8092 (C→A) with poor response to chemoradiation, and no correlation with survival.⁷⁴ A new SNP (rs735482) located in the 3'UTR of *ERCC1* was evaluated for predictive value of clinical outcome in two separate cohorts, but results were mixed.^{75,76} Therefore, we conclude that there is currently no strong evidence that SNPs in *ERCC1* can predict clinical outcome in HNSCC.

ERCC1 protein expression as a biomarker of patient outcomes in NSCLC

While SNPs are often used as a crude estimate of ERCC1 expression or activity, immunodetection approaches permit a more direct quantification of ERCC1 protein level in tumor samples. We identified 17 studies addressing whether quantification of ERCC1 expression in NSCLC tumors by immunohistochemistry has prognostic or predictive value (Table 3).^{27,38,60,73,77-91} In a seminal retrospective analysis of a phase III trial, more than 780 patients with fully resected early stage NSCLC were randomized to observation versus

Table 3 Association between ERCC1 protein expression and clinical outcome

Cancer	Reference	n	Outcome ^a
NSCLC	Planchard et al ⁹⁰	188	0
	Koh et al ⁸⁹	130	0
	Zheng et al ²⁷	187	1
	Kang et al ⁸⁸	82	1
	Okuda et al ³⁸	55	(2)
	Okuda et al ⁹¹	90	2
	Olausson et al ⁸¹	783	2
	Azuma et al ⁶⁴	67	2
	Fujii et al ⁸³	35	2
	Lee et al ⁸⁷	130	2
	Holm et al ⁸⁶	163	2; men $P = 0.005$, women $P = 0.7$
	Azuma et al ⁸⁵	34	2
	Lee et al ⁸²	50	2
	Ota et al ⁸⁰	156	2
	Reynolds et al ^{79,b}	69	2
	Vilmar et al ^{78,b}	264	2
	Wang et al ⁷⁷	214	2
	Taillade et al ²²	34	Biopsy vs tumor correlation
	Gomez-Roca et al (2009)	49	Primary vs metastasis
Kang et al ⁶⁴	82	Primary vs metastasis	
Papay et al (2009)	17	Change after chemotherapy	
Besse et al (2010) ^c	761	Brain metastasis	
HNSCC	Fountzilas et al ³¹	37	0
	Koh et al ⁸⁹	80	0
	Handra-Luca et al ⁹⁷	96	2
	Jun et al ⁹⁸	45	2
	Fountzilas et al ^{31,b}	26	2

Notes: ^aOutcome for low ERCC1 expression, 0 = non significant changes, (1) = trend to worse, 1 = worse, (2) = trend to better, 2 = better; ^bprospective study; ^cretrospective analysis of prospective study.

Abbreviations: HNSCC, head and neck squamous cell carcinoma; NSCLC, non-small cell lung cancers.

multidrug chemotherapy.⁸¹ The results suggested that tumoral ERCC1 protein expression was a biomarker with a complex profile. High ERCC1 levels correlated with good prognosis for untreated cases. But patients with low ERCC1 levels did significantly better when treated with multidrug chemotherapy. These results are consistent with the prediction that decreased expression of ERCC1 could promote sensitivity to genotoxic chemotherapy. Most studies agree that low ERCC1 protein expression is a marker for better clinical outcome after genotoxic therapy in NSCLC. Thirteen of 17 studies reported that low ERCC1 correlated with better clinical outcome (total $n = 1815$),^{77–85,87,91,92} or had a statistical trend towards better outcome (total $n = 218$).³⁸ Two studies showed no correlation between ERCC1 level and outcome ($n = 218$),^{89,90} while two studies showed a significantly worse outcome (total $n = 269$).^{27,88}

in patients with tumors expressing low levels of ERCC1. A recent meta-analysis evaluated NSCLC patients treated with platinum compounds.⁹³ Low expression of ERCC1 in tumors quantified by immunohistochemistry was associated with a better clinical response to cisplatin, which translated into better survival.⁹³ Despite some variability between individual studies, ERCC1 appears to emerge as a good candidate biomarker predictive of clinical outcome in NSCLC. An important point, however, is that in all 18 of the studies the monoclonal antibody, 8F1 was used to measure ERCC1 expression, and this antibody is not specific for ERCC1.²⁵ Therefore, the claim that low ERCC1 expression correlates with better outcome is inaccurate. The more precise conclusion is that low 8F1 signal correlates with better outcome. More recent studies comparing 8F1 and another antibody specific for ERCC1 reveal that they have different predictive capacities with relation to clinical outcomes in cervical cancer.⁹⁴

In HNSCC, only five studies (total $n = 285$) evaluated whether ERCC1 protein expression in tumors correlated with clinical outcome (Table 3).^{31,95–98} The 8F1 antibody was used in all of the studies. Low 8F1 signal was associated with better outcome in three studies (total $n = 168$),^{95,97,98} while no significant association was found in the other two ($n = 117$).^{31,96}

ERCC1 transcript levels as a biomarker in NSCLC and HNSCC

As a surrogate marker of ERCC1 expression, ERCC1 mRNA was measured in NSCLCs in cell lines,⁹⁹ and in six retrospective^{68,100–104} and six prospective studies.^{105–110} The results were mixed, but most studies showed an association between low ERCC1 mRNA and better clinical outcome, either significantly (seven studies)^{100,102–105,108,109} or with a statistical trend (three studies).^{68,105,110} In a meta-analysis, both low tumoral mRNA and protein levels correlated with a better response rate to chemoradiation and overall patient survival.⁹³ While assays used to measure mRNA levels in tumors are not yet readily available for clinical use in all cancer centers, ERCC1 mRNA may prove to be a reasonable predictive biomarker of outcome in NSCLC patients treated with platinum-based chemotherapy.⁹³ Interestingly, ERCC1 mRNA and protein levels were found to be not correlated in NSCLC²⁷ and inversely correlated in ovarian cancer.¹¹¹ Furthermore, mRNA levels were not correlated with chemosensitivity in NSCLC cell lines⁹⁹ nor with response to chemotherapy in HNSCC.³¹ Thus, the relationship between ERCC1 mRNA and DNA repair capacity is not direct and remains to be clarified.

XRCCI as biomarker for NSCLC and HNSCC

XRCCI as a biomarker for cancer risk

Similar studies have sought to establish whether *XRCCI* is linked with cancer risk, prognosis, or treatment outcome. SNPs in *XRCCI* have been extensively studied in NSCLC, although only 9 SNPs out of 550 possible have been evaluated in published reports. The majority of trials focus on Arg194Trp, Arg280His, and Arg399Gln, three nonsynonymous SNPs in *XRCCI* (reviewed by Schneider et al).¹¹² Four studies, including two large ones, also analyzed a SNP in the *XRCCI* promoter ($-77T \rightarrow C$).¹¹³⁻¹¹⁶ The variant allele $-77T \rightarrow C$ alters a binding site for the zinc finger transcription factor SP1, leading to reduced transcription of *XRCCI*.¹¹³ The variant allele at position 399 (Gln) correlates with lower DNA repair capacity and increased genomic instability in multiple studies.¹¹⁷⁻¹²¹ These functional SNPs in *XRCCI* are attractive candidate biomarkers in cancer.

XRCCI SNPs as biomarkers for cancer risk

The assessment of SNPs in *XRCCI* as risk factors for developing NSCLC has focused mainly on *XRCCI* Arg194Trp, Arg280His and Arg399Gln, and to a lesser degree on $-77T \rightarrow C$ (Table 4).^{32,41,44,67,112-116,122-143} Studies failed to identify significant association between Arg194Trp, Arg280His, and Arg399Gln genotypes and NSCLC risk. However, $-77T \rightarrow C$ did emerge as a significant risk factor in two large studies.^{113,114} This is consistent with the notion that low *XRCCI* expression leads to impaired BER and SSB repair, greater mutational load and therefore increased cancer risk. A well conducted meta-analysis pooling more than 10,000 patients for the analysis of Arg194Trp, Arg280His, and Arg399Gln, and more than 1,000 patients for the analysis of Pro206Pro and $-77T \rightarrow C$ found that, in NSCLC, $-77T \rightarrow C$ was associated with cancer risk ($P < 0.0001$), while none of the other four SNPs analyzed in *XRCCI* showed association.⁵⁰ Furthermore, this meta-analysis reviewed a total of 241 associations in 16 genes, and *XRCCI* $-77T \rightarrow C$ was one of the only two associations that maintained a significant association through the most stringent analysis. Thus, there is strong epidemiological and biological credibility supporting *XRCCI* $-77T \rightarrow C$ as a risk factor for NSCLC.

In HNSCC, only five SNPs have been evaluated as cancer risk factors.^{32,54,144-154} Four of them have been evaluated

more than once: Arg194Trp, Arg280His, Arg399Gln, and Pro206Pro (Table 4). The results were mixed for all four SNPs, but primarily showed no significant association with cancer risk, except for a tendency for the homozygous variant 399Gln-Gln to be protective in Caucasians in one large pooled study.¹⁵⁴ Interestingly, when patients from individual studies were pooled for a meta-analysis, Arg194Trp emerged as a significant risk factor for HNSCC, as well as for other solid cancers (skin, esophageal, and stomach).⁵⁰ It will be interesting to follow whether future studies can validate this SNP as a biomarker for risk stratification in HNSCC.

XRCCI SNPs as biomarkers for clinical outcome

Biologically, genetic polymorphisms in *XRCCI* could potentially predict clinical outcome, because reduced *XRCCI* expression in animal models confers sensitivity to ionizing radiation. We identified eleven studies^{57,67,71,115,155-161} looking at *XRCCI* SNPs (Arg194Trp, Arg280His, Arg399Gln, and $-77T \rightarrow C$) including five prospective studies,^{71,155,157,159,160} totaling more than 1700 patients (Table 5). Results were mixed for Arg194Trp: three studies showed no association (total $n = 382$),¹⁵⁵⁻¹⁵⁷ one showed a worse prognosis for the allelic variant ($n = 229$),¹⁵⁸ and one showed a better prognosis ($n = 82$).¹⁵⁹ Results for Arg399Gln were also mixed, with significantly worse overall survival or toxicity for the allelic variant in three studies (total $n = 515$),^{57,67,156} while a better prognosis was found in two studies ($n = 238$)^{71,160} and no association was found in other studies (total $n = 559$).^{155,157-159,161} Finally, Arg280His showed no significant association with any outcome (2 studies; total $n = 428$). A meta-analysis and additional studies to examine $-77T \rightarrow C$ are needed to determine if SNPs in *XRCCI* have any value for predicting clinical outcomes in patients with NSCLC treated with chemoradiation.

In HNSCC, *XRCCI* has not been extensively studied. We identified only four reports assessing the predictive value of SNPs in *XRCCI*, focusing predominantly on Arg399Gln,^{74,76,145,162} and to a lesser extent Arg194Trp^{145,162} (Table 5). Results for Arg399Gln were mixed; two out of the four studies (total $n = 293$) showed a better outcome for the allelic variant.^{74,162} Interestingly, Arg194Trp, which was previously identified as a significant risk factor for HNSCC, did not influence treatment outcome.¹⁶² As with NSCLC, more studies and larger prospective studies are needed to evaluate whether SNPs in *XRCCI* influence response to treatment in HNSCC.

Table 4 Association between SNPs in *XRCC1* and cancer risk

Cancer	rs	SNPs	Alternate names	Reference	n (case-control)	Risk ^a			
NSCLC	rs1799782	Arg194Trp	194 C > T; 194 R > W; 194 Arg > Trp; C26304T	Butkiewicz et al ¹²⁴	96–96	0			
				Hu et al ¹¹⁴	710–710	0			
				Shen et al ⁴⁶	122–122	0			
				Matullo et al ³²	116–> 520,000	0			
				Hao et al ¹¹³	1024–1118	0			
				Zienolddiny et al ⁴⁴	343–413	0			
				Yin et al ¹³¹	247–253	0			
				Hung et al ^{41,b}	6463–6603	0			
				Improta et al ¹²⁶	940–121	0			
				Tanaka et al ¹³⁰	50–50	0			
				Ratnasinghe et al ¹²⁸	108	0; 2 in drinkers			
				David-Beabes ¹³²	332–704	0; 2 in African-Americans			
				Schneider et al ¹¹²	446–622	0; 2 in heavy smokers			
				Hung et al ^{127,b}	2188–2198	0; 2 in heavy smokers			
				Chen et al ⁵⁶	109–109	(1)			
				Pachouri et al ¹³³	103–122	(1)			
				De Ruyck et al ¹¹⁶	110–110	2			
				Yin et al ⁶⁷	55–74	2			
	rs25489	Arg280His	280 G > A; 280 R > H; 280 Arg > His	Butkiewicz et al ¹²⁴	96–96	0			
				Misra et al ^{122,b}	305–305	0			
				Vogel et al ¹²⁴	265–272	0			
				Schneider et al ¹¹²	446–622	0			
				Shen et al ⁴⁶	122–122	0			
				Hao et al ¹¹³	1024–1118	0			
				Zienolddiny et al ⁴⁴	343–413	0			
				Hung et al ⁴¹	6463–6603	0			
				Yin et al ⁶⁷	55–74	0			
				Yin et al ¹³¹	247–253	0; 2 in non-smokers			
				Hung et al ^{127,b}	2188–2198	0; 2 in heavy smokers			
				Ratnasinghe et al ¹²⁸	108	1			
				De Ruyck et al ¹¹⁶	110–110	2			
				Butkiewicz et al ¹²⁴	96–96	0			
				rs25487	Arg399Gln	G28152A; 399 G > A; 399 R > Q; 399 Arg > Gln	David-Beabes ¹³²	332–704	0
							Ratnasinghe et al ¹²⁸	108	0
							Chen et al ⁵⁶	109–109	0
							Ito et al ¹³⁵	178–449	0
Popanda et al ¹³⁷	463–460	0							
Vogel et al ¹³⁴	265–272	0							
Zhang et al ¹³⁹	1000–1000	0							
Hu et al ¹¹⁴	710–710	0							
Hung et al ^{127,b}	2188–2198	0							
Zienolddiny et al ⁴⁴	343–413	0							
Hao et al ¹¹³	1024–1118	0							
Yin et al ¹³¹	247–253	0							
Lopez-Cima et al ¹³⁶	516–533	0							
Hung et al ^{41,b}	6463–6603	0							
Improta et al ¹²⁶	940–121	0							
Yin et al ⁶⁷	55–74	0							
De Ruyck et al ¹¹⁶	110–110	0; 1 in light smokers, 2; in heavy smokers							

(Continued)

Table 4 (Continued)

Cancer	rs	SNPs	Alternate names	Reference	n (case-control)	Risk ^a	
HNSCC				Misra et al ^{122,b}	305–305	0; (2) in heavy smokers	
				Schneider et al ¹¹²	446–622	0; 2 in heavy smokers	
				Ryk et al ¹³⁸	177–153	0; 2 in non-smokers	
				Park et al ^{140,b}	192–135	(1) for SCC	
				Zhou et al ¹⁴¹	1091–1240	(1)	
				Sreeja et al ¹⁴²	171–211	1	
				Divine et al ¹⁴³	172–143	1 in Caucasian but not Hispanic	
				Shen et al ⁴⁶	122–122	(2)	
				Matullo et al ³²	116-> 520,000	2 (by stepwise regression)	
				Pachouri et al ¹³³	103–122	2	
		rs3213245	-(77) T > C		De Ruyck et al ¹¹⁶	110–110	0
					Hsieh et al ¹¹⁵	294–288	0
					Hao et al ¹¹³	1024–1118	1
					Hu et al ¹¹⁴	710–710	1
		rs915927	Pro206Pro	206 A > G; 206 pro = pro	Matullo et al ³²	116-> 520,000	0
					Yin et al ¹³¹	247–253	1
					Yin et al ⁶⁷	55–74	1
		rs17852150	Gln632Gln	632 G > A; 632 Gln = Gln	Yin et al ¹³¹	247–253	0
					Yin et al ⁶⁷	55–74	0
		rs2307191	Pro161Leu	161 Pro > Leu	Tanaka et al ¹³⁰	50	0
		rs2307177	Tyr576Ser	576 Tyr > Ser	Tanaka et al ¹³⁰	50	0
		n/a	Arg59Cys		Zienolddiny et al ⁴⁴	343–413	ND
		rs1799782	Arg194Trp	194 C > T; 194 R > W; 194 Arg > Trp; C26304T	Sturgis et al ¹⁵¹	203–424	0; 2 for oral and pharyngeal cancer
					Olshan et al ¹⁴⁸	182–202	0
					Varzim et al ¹⁶⁸	88–178	0
					Matullo et al ³²	82-> 520,000	0
					Harth et al ¹⁴⁶	312–300	0
					Applebaum et al ¹⁴⁴	722–815	0
				Csejtei et al ¹⁴⁵	108–102	0	
				Kowalski et al ¹⁴⁹	92–124	(1)	
				Tae et al ¹⁵⁰	147–168	1	
	rs25489	Arg280His	280 G > A; 280 R > H; 280 Arg > His	Tae et al ¹⁵⁰	147–168	0	
				Harth et al ¹⁴⁶	312–300	0	
				Applebaum et al ¹⁴⁴	722–815	0	
				Sturgis et al ¹⁵¹	203–424	0	
				Cho et al ¹⁵²	334–283	2	
	rs25487	Arg399Gln	G28152A; 399 G > A; 399 R > Q; 399 Arg > Gln	Varzim et al ¹⁶⁸	88–178	0	
				Cho et al ¹⁵²	334–283	0	
				Tae et al ¹⁵⁰	147–168	0	
				Huang et al ¹⁵⁴	555–792	0; 2 in Caucasian	
				Harth et al ¹⁴⁶	312–300	0	
				Canova et al ¹⁵⁴	1478–1424	0	
				Applebaum et al ¹⁴⁴	722–815	0; (1) in p16 neg smokers	

(Continued)

Table 4 (Continued)

Cancer	rs	SNPs	Alternate names	Reference	n (case-control)	Risk ^a
				Csejtej et al ¹⁴⁵	108–102	0
				Kowalski et al ¹⁴⁹	92–124	0
				Sturgis et al ¹⁵¹	203–424	(1)
				Olshan et al ¹⁴⁸	182–202	2
				Gal et al ¹⁵³	279	2; for overall survival only
	rs915927	Pro206Pro		Matullo et al ³²	82-> 520,000	0
				Canova et al ⁵⁴	1495–1436	0
	rs762507			Canova et al ⁵⁴	1447–1397	0

Notes: ^aRisk for variable allele, 0 = non significant, (1) = trend to increased, 1 = increased, (2) = trend to protective, 2 = protective; ND = not done; ^bretrospective analysis of prospective study.

Abbreviations: HNSCC, head and neck squamous cell carcinoma; NSCLC, non-small cell lung cancers; rs, reference SNP; SCC, squamous cell carcinoma; SNPs, single nucleotide polymorphisms.

XRCC1 expression as a biomarker of patient outcomes in cancer

There is very little data on XRCC1 expression in tumors, despite the fact that at least in NSCLC cell lines increased XRCC1 mRNA is significantly associated with cisplatin resistance.¹⁶³ There are two studies (both using the same patient cohort) reporting XRCC1 expression in NSCLC, as measured by immunohistochemistry.^{88,164} XRCC1 protein expression did not correlate with either response to treatment or survival.

Interestingly, more than half of the metastases had a stronger immunohistochemical signal than their matched primary tumor, suggesting that the level of XRCC1 may increase during cancer progression. This could have therapeutic implications if elevated expression of XRCC1 renders cells more resistant to treatment.

Only one study evaluated XRCC1 protein expression and clinical outcome in HNSCC.¹⁶⁵ High XRCC1 expression was correlated with resistance to radiotherapy. There is also a

Table 5 Association between SNPs in XRCC1 and clinical outcome

Cancer	rs	SNPs	Alternate names	Reference	n	Outcome ^a
NSCLC	rs1799782	Arg194Trp	194 C > T; 194 R > W; 194 Arg > Trp; C26304T	Petty et al ^{155,b}	49	0
				Wang et al ¹⁵⁶	139	0
				Yuan et al ^{157,b}	199	0
				Yoon et al ¹⁵⁸	229	1
				Sun et al ^{159,b}	82	2
	rs25489	Arg280His	280 G > A; 280 R > H; 280 Arg > His	Yoon et al ¹⁵⁸	229	0
				Yuan et al ^{157,b}	199	(2)
	rs25487	Arg399Gln	G28152 A; 399 G > A; 399 R > Q; 399 Arg > Gln	Yoon et al ¹⁵⁸	229	0
				Petty et al ^{155,b}	49	0
				Sun et al ^{159,b}	82	0
				Yuan et al ¹⁵⁷	199	0
				Gurubhagavatula et al ^{161,c}	103	(1)
				Kalikaki et al ⁵⁷	119	1
				Yin et al ⁶⁷	257	1
				Wang et al ¹⁵⁶	139	1 (toxicity)
rs3213245	-(77) T > C		Giachino et al ^{160,b}	203	2 (toxicity)	
			De las Penas et al ^{71,b}	135	2	
rs1799782	Arg194Trp	194 C > T; 194 R > W; C26304T	Hsieh et al ¹¹⁵	294	0	
			Geisler et al ¹⁶²	190	0	
rs25487	Arg399Gln	G28152A; 399 G > A; 399 R > Q	Csejtej et al ¹⁴⁵	108	1	
			Carles et al ⁷⁶	108	0	
			Csejtej et al ¹⁴⁵	108	0	
			Geisler et al ¹⁶²	190	2	
			Quintela-Fandino et al ⁷⁴	103	2	

Notes: ^aOutcome for variable allele, 0 = non significant, (1) = trend to worse, 1 = worse, (2) = trend to better, 2 = better; ^bprospective study; ^cretrospective analysis of prospective study.

Abbreviations: NSCLC, non-small cell lung cancers; rs, reference SNP; SNPs, single nucleotide polymorphisms.

paucity of studies on the predictive value of either peripheral or tumor *XRCC1* mRNA in cancer. In contrast to the protein data, *XRCC1* mRNA appears to be lower in early stage lung cancer compared with more advanced cancer.¹⁶⁶

Conclusion

In summary, for the past decade the biomedical community has evaluated DNA repair genes as potential biomarkers to predict cancer risk and prognosis of cancer patients treated with genotoxic agents. There has been considerable investment toward this endeavor, yet none of the candidate biomarkers, other than *BRCA1* and *BRCA2*, have yet to be translated to clinic use. *ERCC1* and *XRCC1* are two good candidate biomarkers, with robust experimental evidence demonstrating that reduced expression or activity of either protein results in increased genomic instability and sensitivity to DNA damaging agents.^{7,9–11,19} To date, investigations as to whether *ERCC1* and *XRCC1* alter cancer risk or outcomes are primarily modest-sized retrospective case controlled studies, which have yielded conflicting results. The strongest associations to date are that a CC genotype at SNP –77 of *XRCC1*, which causes reduced *XRCC1* mRNA, predicts increased risk of NSCLC. For *ERCC1*, there are numerous studies indicating that low mRNA or protein expression is associated with a better prognosis in HNSCC and NSCLC, respectively. However, it is not established that *ERCC1* expression is regulated at the transcriptional level. Furthermore, in the studies measuring protein level, a nonspecific antibody was used. Therefore these studies, while validating the utility of these biomarkers (*ERCC1* mRNA levels or 8F1 immunohistochemical signal) for predicting clinical outcomes, do not directly demonstrate that DNA repair levels are altered in tumors.

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Disclosure

The authors report no conflicts of interest in relation to this paper.

References

- Jemal A, Siegel R, Ward E, Hao Y, Xu J, Thun MJ. Cancer statistics, 2009. *CA Cancer J Clin*. 2009;59(4):225–249.
- Seiwert TY, Salama JK, Vokes EE. The concurrent chemoradiation paradigm – general principles. *Nat Clin Pract Oncol*. 2007;4(2):86–100.
- Murdoch D. Standard, and novel cytotoxic and molecular-targeted, therapies for HNSCC: an evidence-based review. *Curr Opin Oncol*. 2007;19(3):216–221.
- Seiwert TY, Salama JK, Vokes EE. The chemoradiation paradigm in head and neck cancer. *Nat Clin Pract Oncol*. 2007;4(3):156–171.
- O'Driscoll M, Jeggo PA. The role of double-strand break repair – insights from human genetics. *Nat Rev Genet*. 2006;7(1):45–54.
- Petermann E, Helleday T. Pathways of mammalian replication fork restart. *Nat Rev Mol Cell Biol*. 2010;11(10):683–687.
- Niedernhofer LJ, Lalai AS, Hoeijmakers JH. Fanconi anemia (cross) linked to DNA repair. *Cell*. 2005;123(7):1191–1198.
- Kennedy RD, D'Andrea AD. DNA repair pathways in clinical practice: lessons from pediatric cancer susceptibility syndromes. *J Clin Oncol*. 2006;24(23):3799–3808.
- Niedernhofer LJ, Garinis GA, Raams A, et al. A new progeroid syndrome reveals that genotoxic stress suppresses the somatotroph axis. *Nature*. 2006;444(7122):1038–1043.
- Ahmad A, Robinson AR, Duensing A, et al. *ERCC1*-XPF endonuclease facilitates DNA double-strand break repair. *Mol Cell Biol*. 2008;28(16):5082–5092.
- Bhagwat N, Olsen AL, Wang AT, et al. XPF-*ERCC1* participates in the fanconi anemia pathway of cross-link repair. *Mol Cell Biol*. 2009;29(24):6427–6437.
- Jaspers NG, Raams A, Silengo MC, et al. First reported patient with human *ERCC1* deficiency has cerebro-oculo-facio-skeletal syndrome with a mild defect in nucleotide excision repair and severe developmental failure. *Am J Hum Genet*. 2007;80(3):457–466.
- Zwelling LA, Anderson T, Kohn KW. DNA-protein and DNA inter-strand cross-linking by cis- and trans-platinum(II) diamminedichloride in L1210 mouse leukemia cells and relation to cytotoxicity. *Cancer Res*. 1979;39(2 Pt 1):365–369.
- McHugh PJ, Spanswick VJ, Hartley JA. Repair of DNA interstrand crosslinks: molecular mechanisms and clinical relevance. *Lancet Oncol*. 2001;2(8):483–490.
- Martin LP, Hamilton TC, Schilder RJ. Platinum resistance: the role of DNA repair pathways. *Clin Cancer Res*. 2008;14(5):1291–1295.
- Hoeijmakers JH. DNA damage, aging, and cancer. *N Engl J Med*. 2009;361(15):1475–1485.
- Ladiges WC. Mouse models of *XRCC1* DNA repair polymorphisms and cancer. *Oncogene*. 2006;25(11):1612–1619.
- Almeida KH, Sobol RW. A unified view of base excision repair: lesion-dependent protein complexes regulated by post-translational modification. *DNA Repair (Amst)*. 2007;6(6):695–711.
- Caldecott KW. *XRCC1* and DNA strand break repair. *DNA Repair (Amst)*. 2003;2(9):955–969.
- Lin Y, Hatem J, Wang J, Quinn A, Hicks D, Tang P. Tissue microarray-based immunohistochemical study can significantly underestimate the expression of HER2 and progesterone receptor in ductal carcinoma in situ of the breast. *Biotech Histochem*. 2010 Aug 12. Epub ahead of print.
- Tamaki K, Sasano H, Ishida T, et al. Comparison of core needle biopsy (CNB) and surgical specimens for accurate preoperative evaluation of ER, PgR and HER2 status of breast cancer patients. *Cancer Sci*. 2010;101(9):2074–2079.
- Taillade L, Penault-Llorca F, Boulet T, et al. Immunohistochemical expression of biomarkers: a comparative study between diagnostic bronchial biopsies and surgical specimens of non-small-cell lung cancer. *Ann Oncol*. 2007;18(6):1043–1050.
- Babic A, Loftin IR, Stanislaw S, et al. The impact of pre-analytical processing on staining quality for H&E, dual hapten, dual color in situ hybridization and fluorescent in situ hybridization assays. *Methods*. 2010;52(4):287–300.
- McShane LM, Altman DG, Sauerbrei W, Taube SE, Gion M, Clark GM. Reporting recommendations for tumor marker prognostic studies (REMARK). *J Natl Cancer Inst*. 2005;97(16):1180–1184.
- Niedernhofer LJ, Bhagwat N, Wood RD. *ERCC1* and non-small-cell lung cancer. *N Engl J Med*. 2007;356(24):2538–2540; author reply 2540–2531.

26. Britten RA, Liu D, Tessier A, Hutchison MJ, Murray D. *ERCC1* expression as a molecular marker of cisplatin resistance in human cervical tumor cells. *Int J Cancer*. 2000;89(5):453–457.
27. Zheng Z, Chen T, Li X, Haura E, Sharma A, Bepler G. DNA synthesis and repair genes RRM1 and *ERCC1* in lung cancer. *N Engl J Med*. 2007;356(8):800–808.
28. McGurk CJ, Cummings M, Koberle B, Hartley JA, Oliver RT, Masters JR. Regulation of DNA repair gene expression in human cancer cell lines. *J Cell Biochem*. 2006;97(5):1121–1136.
29. Kim S, Misra A. SNP genotyping: technologies and biomedical applications. *Annu Rev Biomed Eng*. 2007;9:289–320.
30. Schneider J, Classen V, Philipp M, Helmig S. Rapid analysis of *XRCC1* polymorphisms using real-time polymerase chain reaction. *Mol Cell Probes*. 2006;20(3–4):259–262.
31. Fountzilias G, Kalogera-Fountzila A, Lambaki S, et al. MMP9 but not EGFR, MET, *ERCC1*, P16, and P-53 is associated with response to concomitant radiotherapy, cetuximab, and weekly cisplatin in patients with locally advanced head and neck cancer. *J Oncol*. 2009;2009:305908.
32. Matullo G, Dunning AM, Guarrera S, et al. DNA repair polymorphisms and cancer risk in non-smokers in a cohort study. *Carcinogenesis*. 2006;27(5):997–1007.
33. Ferte C, Andre F, Soria JC. Molecular circuits of solid tumors: prognostic and predictive tools for bedside use. *Nat Rev Clin Oncol*. 2010;7(7):367–380.
34. Srivastava S, Gray JW, Reid BJ, Grad O, Greenwood A, Hawk ET. Translational Research Working Group developmental pathway for biospecimen-based assessment modalities. *Clin Cancer Res*. 2008;14(18):5672–5677.
35. Ford BN, Ruttan CC, Kyle VL, Brackley ME, Glickman BW. Identification of single nucleotide polymorphisms in human DNA repair genes. *Carcinogenesis*. 2000;21(11):1977–1981.
36. Yu JJ, Mu C, Lee KB, et al. A nucleotide polymorphism in *ERCC1* in human ovarian cancer cell lines and tumor tissues. *Mutat Res*. 1997;382(1–2):13–20.
37. Yang M, Kim WH, Choi Y, et al. Effects of *ERCC1* expression in peripheral blood on the risk of head and neck cancer. *Eur J Cancer Prev*. 2006;15(3):269–273.
38. Okuda K, Sasaki H, Hikosaka Y, et al. Excision repair cross complementation group 1 polymorphisms predict overall survival after platinum-based chemotherapy for completely resected non-small-cell lung cancer. *J Surg Res*. 2009 Sep 26. Epub ahead of print.
39. Zhou W, Liu G, Park S, et al. Gene-smoking interaction associations for the *ERCC1* polymorphisms in the risk of lung cancer. *Cancer Epidemiol Biomarkers Prev*. 2005;14(2):491–496.
40. Yin J, Vogel U, Guo L, Ma Y, Wang H. Lack of association between DNA repair gene *ERCC1* polymorphism and risk of lung cancer in a Chinese population. *Cancer Genet Cytogenet*. 2006;164(1):66–70.
41. Hung RJ, Christiani DC, Risch A, et al. International Lung Cancer Consortium: pooled analysis of sequence variants in DNA repair and cell cycle pathways. *Cancer Epidemiol Biomarkers Prev*. 2008;17(11):3081–3089.
42. Yu D, Zhang X, Liu J, et al. Characterization of functional excision repair cross-complementation group 1 variants and their association with lung cancer risk and prognosis. *Clin Cancer Res*. 2008;14(9):2878–2886.
43. Deng Q, Sheng L, Su D, et al. Genetic polymorphisms in ATM, *ERCC1*, APE1 and iASPP genes and lung cancer risk in a population of southeast China. *Med Oncol*. 2010 Mar 31. Epub ahead of print.
44. Zienolddiny S, Campa D, Lind H, et al. Polymorphisms of DNA repair genes and risk of non-small cell lung cancer. *Carcinogenesis*. 2006;27(3):560–567.
45. Ma H, Xu L, Yuan J, et al. Tagging single nucleotide polymorphisms in excision repair cross-complementing group 1 (*ERCC1*) and risk of primary lung cancer in a Chinese population. *Pharmacogenet Genomics*. 2007;17(6):417–423.
46. Shen M, Berndt SI, Rothman N, et al. Polymorphisms in the DNA nucleotide excision repair genes and lung cancer risk in Xuan Wei, China. *Int J Cancer*. 2005;116(5):768–773.
47. Jones NR, Spratt TE, Berg AS, Muscat JE, Lazarus P, Gallagher CJ. Association studies of excision repair cross-complementation group 1 (*ERCC1*) haplotypes with lung and head and neck cancer risk in a Caucasian population. *Cancer Epidemiol*. 2011;35(2):175–181.
48. Li Y, Gu S, Wu Q, et al. No association of *ERCC1* C8092A and T19007C polymorphisms to cancer risk: a meta-analysis. *Eur J Hum Genet*. 2007;15(9):967–973.
49. Kiyohara C, Yoshimasu K. Genetic polymorphisms in the nucleotide excision repair pathway and lung cancer risk: a meta-analysis. *Int J Med Sci*. 2007;4(2):59–71.
50. Vineis P, Manuguerra M, Kavvoura FK, et al. A field synopsis on low-penetrance variants in DNA repair genes and cancer susceptibility. *J Natl Cancer Inst*. 2009;101(1):24–36.
51. Cheng L, Spitz MR, Hong WK, Wei Q. Reduced expression levels of nucleotide excision repair genes in lung cancer: a case-control analysis. *Carcinogenesis*. 2000;21(8):1527–1530.
52. Sugimura T, Kumimoto H, Tohna I, et al. Gene-environment interaction involved in oral carcinogenesis: molecular epidemiological study for metabolic and DNA repair gene polymorphisms. *J Oral Pathol Med*. 2006;35(1):11–18.
53. Abbasi R, Ramroth H, Becher H, Dietz A, Schmezer P, Popanda O. Laryngeal cancer risk associated with smoking and alcohol consumption is modified by genetic polymorphisms in ERCC5, ERCC6 and RAD23B but not by polymorphisms in five other nucleotide excision repair genes. *Int J Cancer*. 2009;125(6):1431–1439.
54. Canova C, Hashibe M, Simonato L, et al. Genetic associations of 115 polymorphisms with cancers of the upper aerodigestive tract across 10 European countries: the ARCAGE project. *Cancer Res*. 2009;69(7):2956–2965.
55. Sturgis EM, Dahlstrom KR, Spitz MR, Wei Q. DNA repair gene *ERCC1* and *ERCC2/XPD* polymorphisms and risk of squamous cell carcinoma of the head and neck. *Arch Otolaryngol Head Neck Surg*. 2002;128(9):1084–1088.
56. Cheng L, Sturgis EM, Eicher SA, Spitz MR, Wei Q. Expression of nucleotide excision repair genes and the risk for squamous cell carcinoma of the head and neck. *Cancer*. 2002;94(2):393–397.
57. Kalikaki A, Kanaki M, Vassalou H, et al. DNA repair gene polymorphisms predict favorable clinical outcome in advanced non-small-cell lung cancer. *Clin Lung Cancer*. 2009;10(2):118–123.
58. Gandara DR, Kawaguchi T, Crowley J, et al. Japanese-US common-arm analysis of paclitaxel plus carboplatin in advanced non-small-cell lung cancer: a model for assessing population-related pharmacogenomics. *J Clin Oncol*. 2009;27(21):3540–3546.
59. Suk R, Gurubhagavatula S, Park S, et al. Polymorphisms in *ERCC1* and grade 3 or 4 toxicity in non-small cell lung cancer patients. *Clin Cancer Res*. 2005;11(4):1534–1538.
60. Takenaka T, Yoshino I, Kouso H, et al. Combined evaluation of Rad51 and *ERCC1* expressions for sensitivity to platinum agents in non-small cell lung cancer. *Int J Cancer*. 2007;121(4):895–900.
61. Tibaldi C, Giovannetti E, Vasile E, et al. Correlation of CDA, *ERCC1*, and XPD polymorphisms with response and survival in gemcitabine/cisplatin-treated advanced non-small cell lung cancer patients. *Clin Cancer Res*. 2008;14(6):1797–1803.
62. Vinolas N, Provencio M, Reguart N, et al. Single nucleotide polymorphisms in MDR1 gen correlates with outcome in advanced non-small-cell lung cancer patients treated with cisplatin plus vinorelbine. *Lung Cancer*. 2011;71(2):191–198.
63. Zhou W, Gurubhagavatula S, Liu G, et al. Excision repair cross-complementation group 1 polymorphism predicts overall survival in advanced non-small cell lung cancer patients treated with platinum-based chemotherapy. *Clin Cancer Res*. 2004;10(15):4939–4943.
64. Park SY, Hong YC, Kim JH, et al. Effect of *ERCC1* polymorphisms and the modification by smoking on the survival of non-small cell lung cancer patients. *Med Oncol*. 2006;23(4):489–498.
65. Ryu JS, Hong YC, Han HS, et al. Association between polymorphisms of *ERCC1* and XPD and survival in non-small-cell lung cancer patients treated with cisplatin combination chemotherapy. *Lung Cancer*. 2004;44(3):311–316.

66. Su D, Ma S, Liu P, et al. Genetic polymorphisms and treatment response in advanced non-small cell lung cancer. *Lung Cancer*. 2007; 56(2):281–288.
67. Yin Z, Zhou B, He Q, et al. Association between polymorphisms in DNA repair genes and survival of non-smoking female patients with lung adenocarcinoma. *BMC Cancer*. 2009;9:439.
68. Isla D, Sarries C, Rosell R, et al. Single nucleotide polymorphisms and outcome in docetaxel-cisplatin-treated advanced non-small-cell lung cancer. *Ann Oncol*. 2004;15(8):1194–1203.
69. Zhou C, Ren S, Zhou S, et al. Predictive effects of ERCCI and XRCC3 SNP on efficacy of platinum-based chemotherapy in advanced NSCLC patients. *Jpn J Clin Oncol*. 2010;40(10):954–960.
70. Li F, Sun X, Sun N, et al. Association between polymorphisms of ERCCI and XPD and clinical response to platinum-based chemotherapy in advanced non-small cell lung cancer. *Am J Clin Oncol*. 2010; 33(5):489–494.
71. De las Penas R, Sanchez-Ronco M, Alberola V, et al. Polymorphisms in DNA repair genes modulate survival in cisplatin/gemcitabine-treated non-small-cell lung cancer patients. *Ann Oncol*. 2006;17(4): 668–675.
72. Wei SZ, Zhan P, Shi MQ, et al. Predictive value of ERCCI and XPD polymorphism in patients with advanced non-small cell lung cancer receiving platinum-based chemotherapy: a systematic review and meta-analysis. *Med Oncol*. 2011;28(1):315–321.
73. Takenaka T, Yano T, Kiyohara C, et al. Effects of excision repair cross-complementation group 1 (ERCCI) single nucleotide polymorphisms on the prognosis of non-small cell lung cancer patients. *Lung Cancer*. 2010;67(1):101–107.
74. Quintela-Fandino M, Hitt R, Medina PP, et al. DNA-repair gene polymorphisms predict favorable clinical outcome among patients with advanced squamous cell carcinoma of the head and neck treated with cisplatin-based induction chemotherapy. *J Clin Oncol*. 2006;24(26):4333–4339.
75. Grau JJ, Caballero M, Campayo M, et al. Gene single nucleotide polymorphism accumulation improves survival in advanced head and neck cancer patients treated with weekly paclitaxel. *Laryngoscope*. 2009; 119(8):1484–1490.
76. Carles J, Monzo M, Amat M, et al. Single-nucleotide polymorphisms in base excision repair, nucleotide excision repair, and double strand break genes as markers for response to radiotherapy in patients with Stage I to II head-and-neck cancer. *Int J Radiat Oncol Biol Phys*. 2006; 66(4):1022–1030.
77. Wang X, Zhao J, Yang L, et al. Positive expression of ERCCI predicts a poorer platinum-based treatment outcome in Chinese patients with advanced non-small-cell lung cancer. *Med Oncol*. 2010; 27(2):484–490.
78. Vilmar A, Santoni-Rugiu E, Sorensen JB. ERCCI, toxicity and quality of life in advanced NSCLC patients randomized in a large multicenter phase III trial. *Eur J Cancer*. 2010;46(9):1554–1562.
79. Reynolds C, Obasaju C, Schell MJ, et al. Randomized phase III trial of gemcitabine-based chemotherapy with in situ RRM1 and ERCCI protein levels for response prediction in non-small-cell lung cancer. *J Clin Oncol*. 2009;27(34):5808–5815.
80. Ota S, Ishii G, Goto K, et al. Immunohistochemical expression of BCRP and ERCCI in biopsy specimen predicts survival in advanced non-small-cell lung cancer treated with cisplatin-based chemotherapy. *Lung Cancer*. 2009;64(1):98–104.
81. Olausson KA, Dunant A, Fouret P, et al. DNA repair by ERCCI in non-small-cell lung cancer and cisplatin-based adjuvant chemotherapy. *N Engl J Med*. 2006;355(10):983–991.
82. Lee HW, Choi YW, Han JH, et al. Expression of excision repair cross-complementation group 1 protein predicts poor outcome in advanced non-small cell lung cancer patients treated with platinum-based doublet chemotherapy. *Lung Cancer*. 2009;65(3):377–382.
83. Fujii T, Toyooka S, Ichimura K, et al. ERCCI protein expression predicts the response of cisplatin-based neoadjuvant chemotherapy in non-small-cell lung cancer. *Lung Cancer*. 2008;59(3):377–384.
84. Azuma K, Komohara Y, Sasada T, et al. Excision repair cross-complementation group 1 predicts progression-free and overall survival in non-small cell lung cancer patients treated with platinum-based chemotherapy. *Cancer Sci*. 2007;98(9):1336–1343.
85. Azuma K, Sasada T, Kawahara A, et al. Expression of ERCCI and class III beta-tubulin in non-small cell lung cancer patients treated with a combination of cisplatin/docetaxel and concurrent thoracic irradiation. *Cancer Chemother Pharmacol*. 2009;64(3):565–573.
86. Holm B, Mellemegaard A, Skov T, Skov BG. Different impact of excision repair cross-complementation group 1 on survival in male and female patients with inoperable non-small-cell lung cancer treated with carboplatin and gemcitabine. *J Clin Oncol*. 2009;27(26):4254–4259.
87. Lee KH, Min HS, Han SW, et al. ERCCI expression by immunohistochemistry and EGFR mutations in resected non-small cell lung cancer. *Lung Cancer*. 2008;60(3):401–407.
88. Kang CH, Jang BG, Kim DW, et al. The prognostic significance of ERCCI, BRCA1, XRCCI, and betaIII-tubulin expression in patients with non-small cell lung cancer treated by platinum- and taxane-based neoadjuvant chemotherapy and surgical resection. *Lung Cancer*. 2010;68(3):478–483.
89. Koh Y, Jang B, Han SW, et al. Expression of class III beta-tubulin correlates with unfavorable survival outcome in patients with resected non-small cell lung cancer. *J Thorac Oncol*. 2010;5(3):320–325.
90. Planchard D, Domont J, Taranchon E, et al. The NER proteins are differentially expressed in ever smokers and in never smokers with lung adenocarcinoma. *Ann Oncol*. 2009;20(7):1257–1263.
91. Okuda K, Sasaki H, Dumontet C, et al. Expression of excision repair cross-complementation group 1 and class III beta-tubulin predict survival after chemotherapy for completely resected non-small cell lung cancer. *Lung Cancer*. 2008;62(1):105–112.
92. Vilmar AC, Santoni-Rugiu E, Sorensen JB. ERCCI and histopathology in advanced NSCLC patients randomized in a large multicenter phase III trial. *Ann Oncol*. 2010;21(9):1817–1824.
93. Chen S, Zhang J, Wang R, Luo X, Chen H. The platinum-based treatments for advanced non-small cell lung cancer, is low/negative ERCCI expression better than high/positive ERCCI expression? A meta-analysis. *Lung Cancer*. 2010;70(1):63–70.
94. Doll CM, Prystajek M, Eliasziw M, et al. Low ERCCI mRNA and protein expression are associated with worse survival in cervical cancer patients treated with radiation alone. *Radiation Oncol*. 2010; 97(2):352–359.
95. Fountzilas G, Bamias A, Kalogera-Fountzila A, et al. Induction chemotherapy with docetaxel and cisplatin followed by concomitant chemoradiotherapy in patients with inoperable non-nasopharyngeal carcinoma of the head and neck. *Anticancer Res*. 2009;29(2):529–538.
96. Koh Y, Kim TM, Jeon YK, et al. Class III beta-tubulin, but not ERCCI, is a strong predictive and prognostic marker in locally advanced head and neck squamous cell carcinoma. *Ann Oncol*. 2009;20(8): 1414–1419.
97. Handra-Luca A, Hernandez J, Mountzios G, et al. Excision repair cross complementation group 1 immunohistochemical expression predicts objective response and cancer-specific survival in patients treated by Cisplatin-based induction chemotherapy for locally advanced head and neck squamous cell carcinoma. *Clin Cancer Res*. 2007; 13(13):3855–3859.
98. Jun HJ, Ahn MJ, Kim HS, et al. ERCCI expression as a predictive marker of squamous cell carcinoma of the head and neck treated with cisplatin-based concurrent chemoradiation. *Br J Cancer*. 2008; 99(1):167–172.
99. Shimizu J, Horio Y, Osada H, et al. mRNA expression of RRM1, ERCCI and ERCC2 is not associated with chemosensitivity to cisplatin, carboplatin and gemcitabine in human lung cancer cell lines. *Respirology*. 2008;13(4):510–517.
100. Rosell R, Felip E, Taron M, et al. Gene expression as a predictive marker of outcome in stage IIB-IIIA-IIIB non-small cell lung cancer after induction gemcitabine-based chemotherapy followed by resectional surgery. *Clin Cancer Res*. 2004;10(12 Pt 2):4215s–4219s.

101. Simon GR, Sharma S, Cantor A, Smith P, Beppler G. *ERCC1* expression is a predictor of survival in resected patients with non-small cell lung cancer. *Chest*. 2005;127(3):978–983.
102. Ceppi P, Volante M, Novello S, et al. *ERCC1* and RRM1 gene expressions but not EGFR are predictive of shorter survival in advanced non-small-cell lung cancer treated with cisplatin and gemcitabine. *Ann Oncol*. 2006;17(12):1818–1825.
103. Lord RV, Brabender J, Gandara D, et al. Low *ERCC1* expression correlates with prolonged survival after cisplatin plus gemcitabine chemotherapy in non-small cell lung cancer. *Clin Cancer Res*. 2002; 8(7):2286–2291.
104. Ceppi P, Longo M, Volante M, et al. Excision repair cross complementing-1 and topoisomerase IIalpha gene expression in small-cell lung cancer patients treated with platinum and etoposide: a retrospective study. *J Thorac Oncol*. 2008;3(6):583–589.
105. Cobo M, Isla D, Massuti B, et al. Customizing cisplatin based on quantitative excision repair cross-complementing 1 mRNA expression: a phase III trial in non-small-cell lung cancer. *J Clin Oncol*. 2007; 25(19):2747–2754.
106. Booton R, Ward T, Ashcroft L, Morris J, Heighway J, Thatcher N. *ERCC1* mRNA expression is not associated with response and survival after platinum-based chemotherapy regimens in advanced non-small cell lung cancer. *J Thorac Oncol*. 2007;2(10):902–906.
107. Simon G, Sharma A, Li X, et al. Feasibility and efficacy of molecular analysis-directed individualized therapy in advanced non-small-cell lung cancer. *J Clin Oncol*. 2007;25(19):2741–2746.
108. Su C, Zhou S, Zhang L, et al. *ERCC1*, RRM1 and BRCA1 mRNA expression levels and clinical outcome of advanced non-small cell lung cancer. *Med Oncol*. 2010 May 14. Epub ahead of print.
109. Ren S, Zhou S, Zhang L, et al. High-level mRNA of excision repair cross-complementation group 1 gene is associated with poor outcome of platinum-based doublet chemotherapy of advanced nonsmall cell lung cancer patients. *Cancer Invest*. 2010;28(10):1078–1083.
110. Beppler G, Kusmartseva I, Sharma S, et al. RRM1 modulated in vitro and in vivo efficacy of gemcitabine and platinum in non-small-cell lung cancer. *J Clin Oncol*. 2006;24(29):4731–4737.
111. Pers comm; 13th Annual Midwest DNA Repair Meeting, Toledo, Ohio, USA. Neha Baghwat, Laura J Niedernhofer, 2011.
112. Schneider J, Classen V, Bernges U, Philipp M. *XRCC1* polymorphism and lung cancer risk in relation to tobacco smoking. *Int J Mol Med*. 2005;16(4):709–716.
113. Hao B, Miao X, Li Y, et al. A novel T-77C polymorphism in DNA repair gene *XRCC1* contributes to diminished promoter activity and increased risk of non-small cell lung cancer. *Oncogene*. 2006; 25(25):3613–3620.
114. Hu Z, Ma H, Lu D, et al. A promoter polymorphism (–77T > C) of DNA repair gene *XRCC1* is associated with risk of lung cancer in relation to tobacco smoking. *Pharmacogenet Genomics*. 2005;15(7): 457–463.
115. Hsieh WC, Cheng YW, Lin CJ, Chou MC, Chen CY, Lee H. Prognostic significance of X-ray cross-complementing group 1 T-77C polymorphism in resected non-small cell lung cancer. *Jpn J Clin Oncol*. 2009;39(2):81–85.
116. De Ruyck K, Szaumkessel M, De Rudder I, et al. Polymorphisms in base-excision repair and nucleotide-excision repair genes in relation to lung cancer risk. *Mutat Res*. 2007;631(2):101–110.
117. Ladiges W, Wiley J, MacAuley A. Polymorphisms in the DNA repair gene *XRCC1* and age-related disease. *Mech Ageing Dev*. 2003; 124(1):27–32.
118. Cheng J, Leng S, Li H, et al. Suboptimal DNA repair capacity predisposes coke-oven workers to accumulate more chromosomal damages in peripheral lymphocytes. *Cancer Epidemiol Biomarkers Prev*. 2009; 18(3):987–993.
119. Abdel-Rahman SZ, El-Zein RA. The 399Gln polymorphism in the DNA repair gene *XRCC1* modulates the genotoxic response induced in human lymphocytes by the tobacco-specific nitrosamine NNNK. *Cancer Lett*. 2000;159(1):63–71.
120. Relton CL, Daniel CP, Fisher A, Chase DS, Burn J, Tawn EJ. Polymorphisms of the DNA repair gene *XRCC1* and the frequency of somatic mutations at the glycophorin A locus in newborns. *Mutat Res*. 2002;502(1–2):61–68.
121. Lunn RM, Langlois RG, Hsieh LL, Thompson CL, Bell DA. *XRCC1* polymorphisms: effects on aflatoxin B1-DNA adducts and glycophorin A variant frequency. *Cancer Res*. 1999;59(11):2557–2561.
122. Misra RR, Ratnasinghe D, Tangrea JA, et al. Polymorphisms in the DNA repair genes XPD, *XRCC1*, *XRCC3*, and APE/ref-1, and the risk of lung cancer among male smokers in Finland. *Cancer Lett*. 2003; 191(2):171–178.
123. Yin J, Vogel U, Ma Y, Qi R, Wang H. Association of DNA repair gene *XRCC1* and lung cancer susceptibility among nonsmoking Chinese women. *Cancer Genet Cytogenet*. 2009;188(1):26–31.
124. Butkiewicz D, Rusin M, Enewold L, Shields PG, Chorazy M, Harris CC. Genetic polymorphisms in DNA repair genes and risk of lung cancer. *Carcinogenesis*. 2001;22(4):593–597.
125. Chen S, Tang D, Xue K, et al. DNA repair gene *XRCC1* and XPD polymorphisms and risk of lung cancer in a Chinese population. *Carcinogenesis*. 2002;23(8):1321–1325.
126. Improta G, Sgambato A, Bianchino G, et al. Polymorphisms of the DNA repair genes *XRCC1* and *XRCC3* and risk of lung and colorectal cancer: a case-control study in a Southern Italian population. *Anticancer Res*. 2008;28(5B):2941–2946.
127. Hung RJ, Brennan P, Canzian F, et al. Large-scale investigation of base excision repair genetic polymorphisms and lung cancer risk in a multicenter study. *J Natl Cancer Inst*. 2005;97(8):567–576.
128. Ratnasinghe D, Yao SX, Tangrea JA, et al. Polymorphisms of the DNA repair gene *XRCC1* and lung cancer risk. *Cancer Epidemiol Biomarkers Prev*. 2001;10(2):119–123.
129. Shen M, Berndt SI, Rothman N, et al. Polymorphisms in the DNA base excision repair genes APEX1 and *XRCC1* and lung cancer risk in Xuan Wei, China. *Anticancer Res*. 2005;25(1B): 537–542.
130. Tanaka Y, Maniwa Y, Bermudez VP, et al. Nonsynonymous single nucleotide polymorphisms in DNA damage repair pathways and lung cancer risk. *Cancer*. 2010;116(4):896–902.
131. Yin J, Vogel U, Ma Y, Qi R, Sun Z, Wang H. The DNA repair gene *XRCC1* and genetic susceptibility of lung cancer in a northeastern Chinese population. *Lung Cancer*. 2007;56(2):153–160.
132. David-Beabes GL, London SJ. Genetic polymorphism of *XRCC1* and lung cancer risk among African-Americans and Caucasians. *Lung Cancer*. 2001;34(3):333–339.
133. Pachouri SS, Sobti RC, Kaur P, Singh J. Contrasting impact of DNA repair gene *XRCC1* polymorphisms Arg399Gln and Arg194Trp on the risk of lung cancer in the north-Indian population. *DNA Cell Biol*. 2007;26(3):186–191.
134. Vogel U, Nexo BA, Wallin H, Overvad K, Tjonneland A, Raaschou-Nielsen O. No association between base excision repair gene polymorphisms and risk of lung cancer. *Biochem Genet*. 2004; 42(11–12):453–460.
135. Ito H, Matsuo K, Hamajima N, et al. Gene-environment interactions between the smoking habit and polymorphisms in the DNA repair genes, APE1 Asp148Glu and *XRCC1* Arg399Gln, in Japanese lung cancer risk. *Carcinogenesis*. 2004;25(8):1395–1401.
136. Lopez-Cima MF, Gonzalez-Arriaga P, Garcia-Castro L, et al. Polymorphisms in XPC, XPD, *XRCC1*, and *XRCC3* DNA repair genes and lung cancer risk in a population of northern Spain. *BMC Cancer*. 2007;7:162.
137. Popanda O, Schattenberg T, Phong CT, et al. Specific combinations of DNA repair gene variants and increased risk for non-small cell lung cancer. *Carcinogenesis*. 2004;25(12):2433–2441.
138. Ryk C, Kumar R, Thirumaran RK, Hou SM. Polymorphisms in the DNA repair genes *XRCC1*, APEX1, *XRCC3* and NBS1, and the risk for lung cancer in never- and ever-smokers. *Lung Cancer*. 2006; 54(3):285–292.

139. Zhang X, Miao X, Liang G, et al. Polymorphisms in DNA base excision repair genes ADPRT and *XRCC1* and risk of lung cancer. *Cancer Res.* 2005;65(3):722–726.
140. Park JY, Lee SY, Jeon HS, et al. Polymorphism of the DNA repair gene *XRCC1* and risk of primary lung cancer. *Cancer Epidemiol Biomarkers Prev.* 2002;11(1):23–27.
141. Zhou W, Liu G, Miller DP, et al. Polymorphisms in the DNA repair genes *XRCC1* and *ERCC2*, smoking, and lung cancer risk. *Cancer Epidemiol Biomarkers Prev.* 2003;12(4):359–365.
142. Sreeja L, Syamala VS, Syamala V, et al. Prognostic importance of DNA repair gene polymorphisms of *XRCC1* Arg399Gln and XPD Lys751Gln in lung cancer patients from India. *J Cancer Res Clin Oncol.* 2008;134(6):645–652.
143. Divine KK, Gilliland FD, Crowell RE, et al. The *XRCC1* 399 glutamine allele is a risk factor for adenocarcinoma of the lung. *Mutat Res.* 2001;461(4):273–278.
144. Applebaum KM, McClean MD, Nelson HH, Marsit CJ, Christensen BC, Kelsey KT. Smoking modifies the relationship between *XRCC1* haplotypes and HPV16-negative head and neck squamous cell carcinoma. *Int J Cancer.* 2009;124(11):2690–2696.
145. Csejtei A, Tibold A, Koltai K, et al. Association between *XRCC1* polymorphisms and head and neck cancer in a Hungarian population. *Anticancer Res.* 2009;29(10):4169–4173.
146. Harth V, Schafer M, Abel J, et al. Head and neck squamous-cell cancer and its association with polymorphic enzymes of xenobiotic metabolism and repair. *J Toxicol Environ Health A.* 2008;71(13–14):887–897.
147. Shen H, Sturgis EM, Khan SG, et al. An intronic poly (AT) polymorphism of the DNA repair gene XPC and risk of squamous cell carcinoma of the head and neck: a case-control study. *Cancer Res.* 2001;61(8):3321–3325.
148. Olshan AF, Watson MA, Weissler MC, Bell DA. *XRCC1* polymorphisms and head and neck cancer. *Cancer Lett.* 2002;178(2):181–186.
149. Kowalski M, Przybylowska K, Rusin P, et al. Genetic polymorphisms in DNA base excision repair gene *XRCC1* and the risk of squamous cell carcinoma of the head and neck. *J Exp Clin Cancer Res.* 2009;28:37.
150. Tae K, Lee HS, Park BJ, et al. Association of DNA repair gene *XRCC1* polymorphisms with head and neck cancer in Korean population. *Int J Cancer.* 2004;111(5):805–808.
151. Sturgis EM, Castillo EJ, Li L, et al. Polymorphisms of DNA repair gene *XRCC1* in squamous cell carcinoma of the head and neck. *Carcinogenesis.* 1999;20(11):2125–2129.
152. Cho EY, Hildesheim A, Chen CJ, et al. Nasopharyngeal carcinoma and genetic polymorphisms of DNA repair enzymes *XRCC1* and hOGG1. *Cancer Epidemiol Biomarkers Prev.* 2003;12(10):1100–1104.
153. Gal TJ, Huang WY, Chen C, Hayes RB, Schwartz SM. DNA repair gene polymorphisms and risk of second primary neoplasms and mortality in oral cancer patients. *Laryngoscope.* 2005;115(12):2221–2231.
154. Huang WY, Olshan AF, Schwartz SM, et al. Selected genetic polymorphisms in *MGMT*, *XRCC1*, *XPD*, and *XRCC3* and risk of head and neck cancer: a pooled analysis. *Cancer Epidemiol Biomarkers Prev.* 2005;14(7):1747–1753.
155. Petty WJ, Knight SN, Mosley L, et al. A pharmacogenomic study of docetaxel and gemcitabine for the initial treatment of advanced non-small cell lung cancer. *J Thorac Oncol.* 2007;2(3):197–202.
156. Wang Z, Xu B, Lin D, et al. *XRCC1* polymorphisms and severe toxicity in lung cancer patients treated with cisplatin-based chemotherapy in Chinese population. *Lung Cancer.* 2008;62(1):99–104.
157. Yuan P, Liu L, Wu C, et al. No association between *XRCC1* polymorphisms and survival in non-small-cell lung cancer patients treated with platinum-based chemotherapy. *Cancer Biol Ther.* 2010;10(9).
158. Yoon SM, Hong YC, Park HJ, et al. The polymorphism and haplotypes of *XRCC1* and survival of non-small-cell lung cancer after radiotherapy. *Int J Radiat Oncol Biol Phys.* 2005;63(3):885–891.
159. Sun X, Li F, Sun N, et al. Polymorphisms in *XRCC1* and XPG and response to platinum-based chemotherapy in advanced non-small cell lung cancer patients. *Lung Cancer.* 2009;65(2):230–236.
160. Giachino DF, Ghio P, Regazzoni S, et al. Prospective assessment of XPD Lys751Gln and *XRCC1* Arg399Gln single nucleotide polymorphisms in lung cancer. *Clin Cancer Res.* 2007;13(10):2876–2881.
161. Gurubhagavatula S, Liu G, Park S, et al. XPD and *XRCC1* genetic polymorphisms are prognostic factors in advanced non-small-cell lung cancer patients treated with platinum chemotherapy. *J Clin Oncol.* 2004;22(13):2594–2601.
162. Geisler SA, Olshan AF, Cai J, Weissler M, Smith J, Bell D. Glutathione S-transferase polymorphisms and survival from head and neck cancer. *Head Neck.* 2005;27(3):232–242.
163. Weaver DA, Crawford EL, Warner KA, Elkhairi F, Khuder SA, Willey JC. *ABCC5*, *ERCC2*, *XPA* and *XRCC1* transcript abundance levels correlate with cisplatin chemoresistance in non-small cell lung cancer cell lines. *Mol Cancer.* 2005;4(1):18.
164. Kang CH, Jang BG, Kim DW, et al. Differences in the expression profiles of excision repair cross-complementation group 1, x-ray repair cross-complementation group 1, and betaIII-tubulin between primary non-small cell lung cancer and metastatic lymph nodes and the significance in mid-term survival. *J Thorac Oncol.* 2009;4(11):1307–1312.
165. Nix P, Greenman J, Stafford N, Cawkwell L. Expression of *XRCC1* and *ERCC1* proteins in radioresistant and radiosensitive laryngeal cancer. *Cancer Therapy.* 2004(2):47–53.
166. Campioni M, Ambrogi V, Pompeo E, et al. Identification of genes down-regulated during lung cancer progression: a cDNA array study. *J Exp Clin Cancer Res.* 2008;27:38.
167. Jones NR, Spratt TE, Berg AS, Muscat JE, Lazarus P, Gallagher CJ. Association studies of excision repair cross-complementation group 1 (*ERCC1*) haplotypes with lung and head and neck cancer risk in a Caucasian population. *Cancer Epidemiol.* 2010;35(2):175–81.
168. Varzim G, Monteiro E, Silva RA, Fernandes J, Lopes C. *CYP1A1* and *XRCC1* gene polymorphisms in SCC of the larynx. *Eur J Cancer Prev.* 2003;12(6):495–499.

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