

Photothermal ablation of pancreatic cancer cells with hybrid iron-oxide core gold-shell nanoparticles

Yang Guo¹
Zhuoli Zhang¹
Dong-Hyun Kim^{1,5}
Weiguo Li¹
Jodi Nicolai¹
Daniel Procissi¹
Yi Huan²
Guohong Han³
Reed A Omary^{1,4,5}
Andrew C Larson^{1,4,5}

¹Department of Radiology, Northwestern University, Chicago, IL, USA; ²Department of Radiology, Xijing Hospital, Fourth Military Medical University, Xi'an, People's Republic of China; ³Department of Digestive Interventional Radiology, Xijing Hospital, Fourth Military Medical University, Xi'an, People's Republic of China; ⁴Department of Biomedical Engineering, Northwestern University, Chicago, IL, USA; ⁵Robert H Lurie Comprehensive Cancer Center, Chicago, IL, USA

Correspondence: Andrew C Larson
Departments of Radiology and
Biomedical Engineering, Northwestern
University Feinberg School of Medicine,
737 N Michigan Ave, 16th Floor,
Chicago, IL 60611, USA
Tel +1 312 926 3499
Fax +1 312 926 5991
Email a-larson@northwestern.edu

Purpose: Photothermal ablation is a minimally invasive approach, which typically involves delivery of photothermal sensitizers to targeted tissues. The purpose of our study was to demonstrate that gold nanoparticles are phagocytosed by pancreatic cancer cells, thus permitting magnetic resonance imaging (MRI) of sensitizer delivery and photothermal ablation.

Patients and methods: Iron-oxide core/gold-shell nanoparticles (GoldMag[®], 30 nm diameter; Xi'an GoldMag Biotechnology Co, Xi'an, People's Republic of China) were used. In a 96-well plate, 3×10^4 PANC-1 (human pancreatic cancer cell line) cells were placed. GoldMag (0, 25, or 50 $\mu\text{g/mL}$) was added to each well and 24 hours allowed for cellular uptake. Samples were then divided into two groups: one treated with photothermal ablation (7.9 W/cm^2) for 5 minutes, the other not treated. Photothermal ablation was performed using laser system (BWF5; B&W Tek, Inc, Newark, DE, USA). Intraprocedural temperature changes were measured using a fiber optic temperature probe (FTP-LN2; Photon Control Inc, Burnaby, BC, Canada). After 24 hours, the remaining number of viable cells was counted using trypan blue staining; cell proliferation percentage was calculated based on the total number of viable cells after treatment compared with control. MRI of GoldMag uptake was performed using a 7.0T ClinScan system (Bruker BioSpin, Ettlingen, Germany).

Results: Temperature curves demonstrated that with increased GoldMag uptake, laser irradiation produced higher temperature elevations in the corresponding samples; temperature elevations of 12.89°C, 35.16°C, and 79.51°C were achieved for 0, 25, and 50 $\mu\text{g/mL}$ GoldMag. Without photothermal ablation, the cell proliferation percentage changed from 100% to 71.3% and 47.0% for cells treated with 25 and 50 $\mu\text{g/mL}$ GoldMag. Photothermal ablation of PANC-1 cells demonstrated an effective treatment response, specifically a reduction to only 61%, 21.9%, and 2.3% cell proliferation for cells treated with 0, 25, and 50 $\mu\text{g/mL}$ GoldMag. MRI was able to visualize GoldMag uptake within PANC-1 cells.

Conclusion: Our findings suggest that photothermal ablation may be effective in the treatment of pancreatic cancer. GoldMag nanoparticles could serve as photothermal sensitizers, and MRI is feasible to quantify delivery.

Keywords: photothermal ablation therapy, hybrid nanoparticles, magnetic resonance imaging

Introduction

Pancreatic cancer is the fourth leading cause of cancer mortality in the United States with an overall 5-year survival rate less than 5%.^{1,2} Although surgical resection is the sole curative treatment for pancreatic cancer, only 5%–15% of patients with pancreatic cancer are amenable for surgery at the time of diagnosis. Even with an optimal curative resection, the median survival for pancreatic cancer patients is approximately

2 years, with a 5-year survival of 15%–20%, and for those with metastatic disease, survival is shorter than 6 months.^{3–5} Disruptive approaches are critically needed to impact this deadly disease, and photothermal ablation therapy is one such emerging treatment modality.

Photothermal ablation is a minimally invasive therapy that typically involves the application of photothermal sensitizers.⁶ These photothermal sensitizers transform the absorbed light (typically laser irradiation) into heat, thus increasing temperatures within the local vicinity of these sensitizing agents. When tissues are heated above 42°C, irreversible cellular damage can occur, eventually resulting in cell death.⁷ Long-wavelength (800–1300 nm) laser irradiation is necessary for the penetration of tissues during *in vivo* photothermal ablation procedures; thus, for these applications, the absorption band for nanoparticle sensitizers must be in the near-infrared (NIR) region.⁸ NIR resonant nanomaterials such as gold nanorods, carbon tubes, and multifunctional magnetic gold nanoshells have each demonstrated the potential to serve as sensitizers during photothermal ablation.^{6,9,10} Gold nanoparticles permit targeted heat deposition due to their strong absorption cross section and light-to-heat conversion characteristics.¹¹ To enhance tumor-specific heating and thermal deposition, a number of researchers have explored the potential of multifunctional gold nanomaterials; these have been proven as effective photothermal sensitizers for NIR laser irradiation.^{6,9,12,13}

In vivo approaches for noninvasive quantification of gold nanoparticle delivery may be critically needed to optimize these laser irradiation procedures. Super-paramagnetic iron oxide and/or ultra-small superparamagnetic iron oxide nanoparticles have been used to label cells, microcapsules, and embolic particles for *in vivo* tracking with magnetic resonance imaging (MRI) following systemic administration.^{14–17} Magnetic susceptibility differences between the iron oxide particles and surrounding tissues leads to local magnetic field gradients with proportional signal losses via R2 and R2* relaxation mechanisms. Thus, the quantification of R2 and R2* can be used to estimate intratumoral concentrations and the biodistribution of particles containing iron-oxide nanoparticles.¹⁸

In current study, we investigated the use of gold shell–iron oxide core hybrid nanoparticles (Fe₃O₄/Au) for both imaging and laser irradiation. The aim of our study was to demonstrate that these hybrid nanoparticles are phagocytosed by pancreatic cancer cells, permit quantitative MRI, and elicit a photothermal response with NIR laser irradiation.

Material and methods

Hybrid iron oxide core gold-shell nanoparticles

Hybrid iron oxide core gold-shell nanoparticles (GoldMag[®], Xi'an GoldMag Biotechnology Co, Xi'an, People's Republic of China) were purchased and used as the photosensitizing agent for the current study.^{19,20} Briefly, the composite particle was prepared by the reduction of Au³⁺ with hydroxylamine in the presence of Fe₃O₄ particles as seeds. First, the seeds were co-precipitated Fe (II) and Fe (III) ions in alkaline medium, followed by rinsing with water several times until the pH reached 6–7. Second, Fe₃O₄ particles were dispersed in 0.1 mol/L HAuCl₄·4H₂O solution in a beaker, and slowly mixed in a shaking incubator to allow adsorption of Au³⁺ onto the Fe₃O₄ surface. NH₂OH solution of 80 mmol/L was then added to the system. The mixture was incubated with shaking for 1 hour. The core/shell structure of Fe₃O₄/Au was formed and then washed with 1 M HCl solution. Finally, the particles were washed with water until the pH was 6.7.

Transmission electron microscopy (TEM) images of these GoldMag nanoparticles were digitized with a high-magnification (FEI Tecnai Spirit G2; FEI Company, Hillsboro, OR, USA). Absorption spectra of the GoldMag nanoparticles (Fe₃O₄/Au core/shell nanoparticles) were measured and compared with bare Fe₃O₄ nanoparticles.

Cell culture

A human pancreatic cancer cell line (PANC-1) was purchased from American Type Culture Collection (ATCC, Manassas, VA, USA) and cultured using DMEM (Dulbecco's Modified Eagle's Medium; Life Technologies, Carlsbad, CA, USA) with 10% fetal bovine serum (Gemini Bio-Products, West Sacramento, CA, USA) and 1% Pen-Strep (Sigma-Aldrich, St Louis, MO, USA). Cells were maintained in culture flasks at 37°C in a humidified atmosphere containing 5% CO₂. The viability of the cells was evaluated with trypan blue staining (confirming >90% cell viability when plating the cells).

Temperature measurements

Representative samples with 0 µg/mL, 25 µg/mL, and 50 µg/mL of the GoldMag nanoparticles in 200 µL DMEM culture medium were plated in a 96-well cell culture plate (well bottom area = 0.32 cm²). Samples were irradiated using an NIR laser system (BWF5; B&W Tek, Inc, Newark, DE, USA) with diode fiber laser and continuous wave at wavelength of 808 nm; four separate samples for each nanoparticle concentration were irradiated with

power densities = 3.2 W/cm² (1.0 W), 4.7 W/cm² (1.5 W), 6.3 W/cm² (2.0 W), and 7.9 W/cm² (2.5 W), respectively. The intraprocedural temperature changes were recorded using a fiber optic temperature probe (FTP-LN2; Photon Control Inc, Burnaby, BC, Canada) with measurements recorded at 15-second intervals. The fiber probe was vertically placed in the center of each sample, approximately 3–5 mm deep inside the culture media. Baseline temperature was recorded for first 30 seconds before application of the NIR laser irradiation; each sample was then irradiated for 5 minutes and temperature curve plotted.

Cellular uptake of nanoparticles

TEM (FEI Tecnai Spirit G2) was performed to evaluate the cellular uptake of nanoparticles. Three samples of PANC-1 pancreatic cancer cells were treated with nanoparticles at concentrations of 0, 25, and 50 µg/mL, respectively. After allowing 24 hours incubation in complete culture media at 37°C, the cells were washed three times with phosphate-buffered saline to remove free particles in the medium and rapidly placed into glutaraldehyde and fixed for at least 1 hour at room temperature. The cell samples were then placed in osmium tetroxide, embedded in agar, and processed. The samples were dehydrated in graded concentrations of ethanol and propylene oxide, and embedded in Spurr's plastic. Semi-thin (5 µm) sections were cut from blocks, and the blocks were selected for thinning with thin sections (1.5 µm) cut and placed on copper grids. Each sample was digitized to visualize the subcellular location of the nanoparticles within the PANC-1 cells.

MRI after nanoparticle uptake

To evaluate the MRI properties of these hybrid nanoparticles macroscopically after cellular uptake, first we constructed three phantoms, each containing 10×10^6 PANC-1 pancreatic cancer cells. These cells were exposed to 0, 25, and 50 µg/mL dose of nanoparticles for 24 hours, respectively. After washing the cells three times using phosphate-buffered saline, the pancreatic cancer cells were gently mixed with 10 mL of 0.8% agarose and placed within 15 mL centrifuge tubes. MRI studies were performed using a 7.0T ClinScan MRI scanner (Bruker BioSpin, Ettlingen, Germany) with Siemens gradient system/pulse sequences and a surface coil. Quantitative R2 measurements were performed using a Carr–Purcell–Meiboom–Gill sequence with the following parameters: repetition time (TR) = 800 ms, echo time (TE) = 11.5, 23, 34.5, and 46 ms, slice thickness = 0.9 mm, 10 slices, field of

view (FOV) = 20 mm × 35 mm, matrix = 114 × 192, and bandwidth = 150 Hz/pixel. R2* was measured using a multiple gradient-recalled echo sequence with the following parameters: TR = 700 ms, TE = 2.47, 6.66, 9.65, and 13.24 ms, slice thickness = 0.9 mm, 10 slices, FOV = 20 mm × 35 mm, matrix = 114 × 192, and bandwidth = 500 Hz/pixel. MRI post-processing was performed offline using Matlab software (The Math Works Inc, Natick, MA, USA). R2 and R2* maps were calculated on a voxel-wise basis employing the nonlinear Levenberg–Marquardt algorithm to fit a mono-exponential function $S(TE_i) = S(0) \cdot \exp(-R2^* \cdot TE_i)$. A region of interest (ROI) was manually drawn within each resulting R2 and R2* map to record the mean R2 and R2* values within these ROI for each slice within each phantom.

To demonstrate the potential to visualize single cells after nanoparticle uptake, high-resolution three-dimensional multiple gradient-recalled echo scans were performed with the following parameters: TR = 80 ms, TE = 3.22, 6.81, 10.4, and 13.99 ms, slice thickness = 0.15 mm, FOV = 29 mm × 29 mm, matrix = 384 × 384, and bandwidth = 305 Hz/pixel. PANC-1 pancreatic cancer cells treated with 25 µg/mL nanoparticles for 24 hours were prepared in an MRI compatible dish (diameter = 1 inch). After placing these PANC-1 cells at the bottom of the dish, a thin film of agarose was added atop the cells. MRI images were collected using a surface coil with the diameter of 3 cm.

Photothermal ablation

In a 96-well plate, 3×10^4 PANC-1 cells were placed in each well and exposed to a 0, 25, or 50 µg/mL dose of nanoparticles. After allowing 24 hours for cellular uptake, the samples exposed to these three different nanoparticle concentrations were divided into two groups, one exposed to laser irradiation at a power density of 7.9 W/cm² (2.5 W/well) for 5 minutes, the other group left untreated thus serving as controls. After exposure, the cells were returned to a 37°C incubator for another 24 hours before cell viability tests. These tests were performed in triplicate.

Next, we investigated the efficacy of this photothermal approach when using different power densities during irradiation. Twenty-four hours after PANC-1 cells were exposed to a 25 µg/mL solution of nanoparticles, the cells were irradiated with power densities of 3.2 W/cm² (1.0 W), 4.7 W/cm² (1.5 W), 6.3 W/cm² (2.0 W), or 7.9 W/cm² (2.5 W), respectively, for 5 minutes. After these NIR laser procedures, the PANC-1 cells were incubated for another 24 hours before cell viability tests.

To evaluate the cell proliferation percentage for each of the aforementioned samples, cells were stained using trypan blue, and the total viable cells were counted three times using an automated cell counter (Countess; Invitrogen, San Diego, CA, USA). The cell proliferation percentage was calculated based on the total viable PANC-1 cells after treatment compared with the control group. Each sample was counted in triplicate.

Statistical analysis

All statistics were performed using the SPSS statistical software package (SPSS, Chicago, IL, USA), version 17. R2 and R2* values for three groups (control, 25 $\mu\text{g}/\text{mL}$, and 50 $\mu\text{g}/\text{mL}$) were compared by one-way analysis of variance test followed by post hoc tests. Paired *t*-test was performed to compare the cell proliferation between non-ablated cells (GoldMag treated without laser irradiation) and ablated cells (GoldMag treated with laser irradiation) for each GoldMag concentration. Test was considered statistically significant with a *P*-value < 0.05.

Results

Heating properties of hybrid nanoparticles

Representative TEM image for GoldMag nanoparticles is shown in Figure 1A, and absorption spectra of GoldMag nanoparticles ($\text{Fe}_3\text{O}_4/\text{Au}$ core/shell nanoparticles) and bare Fe_3O_4 nanoparticles is shown in Figure 1B. A surface plasmon resonance band was observed at 590 nm for the GoldMag nanoparticles. The surface plasmon resonance band for the $\text{Fe}_3\text{O}_4/\text{Au}$ core/shell bimetallic nanoparticles showed

a red-shift and broadening of the peak, which is commonly observed in other Au bimetallic systems.^{21,22} When we utilized an 808 nm laser, a corresponding absorption value at 808 nm for the $\text{Fe}_3\text{O}_4/\text{Au}$ core/shell nanoparticles was directly converted to heat energy that resulted in our heating profiles (Figure 2). However, bare Fe_3O_4 nanoparticles did not show absorption peaks around 808 nm, and no heating resulted following 808 nm laser irradiation.

For identical laser irradiation power densities of 7.9 W/cm^2 , samples containing nanoparticles achieved significantly higher temperatures compared with those samples without the nanoparticles. With an increased concentration of nanoparticles, laser irradiation produced increasingly higher temperature elevations to 13°C, 35°C, and 80°C, respectively, at concentrations of 0, 25, and 50 $\mu\text{g}/\text{mL}$ nanoparticles. For an identical nanoparticle concentration of 25 $\mu\text{g}/\text{mL}$, increasing irradiation power density increased the magnitude of the resulting temperature elevations from 7°C at a power density of 3.2 W/cm^2 to 14°C, 26°C, and 35°C for power densities of 4.7, 6.3, and 7.9 W/cm^2 , respectively. Dynamic temperature curves during laser irradiation for each concentration and power density are shown in Figure 2.

Cellular uptake

TEM images showing cellular uptake of nanoparticles after a 24-hour exposure period are shown in Figure 3. These TEM images demonstrated that the particles were phagocytosed into the pancreatic cancer cells (arrow). Nanoparticle uptake increased within the 50 $\mu\text{g}/\text{mL}$ exposure group (more nanoparticles observed within 50 $\mu\text{g}/\text{mL}$ exposure group sample compared with both 25 $\mu\text{g}/\text{mL}$ exposure group and controls).

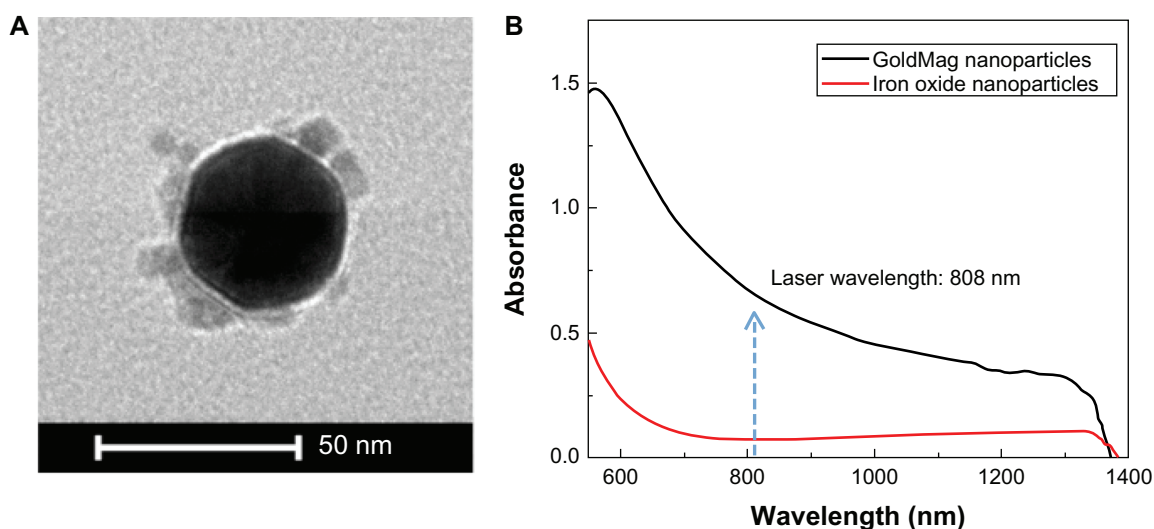


Figure 1 Transmission electron microscopy image of GoldMag nanoparticles (A), and ultraviolet-visible spectra of GoldMag ($\text{Fe}_3\text{O}_4/\text{Au}$ core/shell nanoparticles) and Fe_3O_4 in water (arrow) a wavelength of the laser (B).

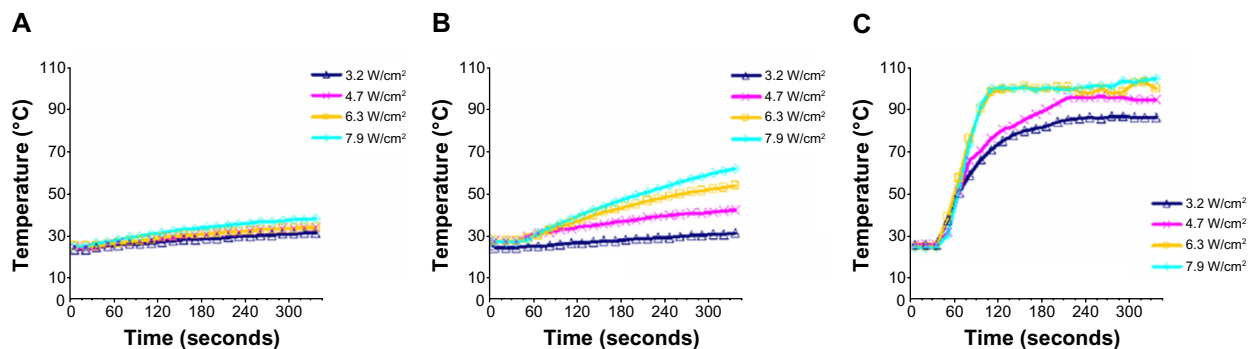


Figure 2 Photothermal response of 0 (A), 25 (B), and 50 (C) µg/mL nanoparticle samples at 3.2, 4.7, 6.3, and 7.9 W/cm² laser irradiation power densities.

MRI measurements

Voxel-wise R2 and R2* maps for cell suspension phantoms are shown in Figure 4A. Mean R2 and R2* for these cell suspension phantoms increased significantly for those samples exposed to a higher concentration of nanoparticles ($P = 0.000$ for R2, and $P = 0.000$ for R2*). A difference in mean R2 and R2* values were found between each of the three phantoms. For control sample, mean R2 \pm standard deviation (SD) = 4.77 ± 1.02 (ms⁻¹), mean R2* \pm SD = 9.71 ± 0.06 (ms⁻¹); for those cells exposed to 25 µg/mL nanoparticle dose, mean R2 \pm SD = 7.68 ± 1.75 (ms⁻¹), mean R2* \pm SD = 104.25 ± 1.83 (ms⁻¹); and for cells exposed to 50 µg/mL nanoparticle

dose, mean R2 \pm SD = 13.03 ± 0.64 (ms⁻¹), mean R2* \pm SD = 200.86 ± 4.75 (ms⁻¹) (Figure 4B).

High-resolution T2*-weighted images permitted depiction of individual PANC-1 cell positions after uptake of the iron oxide core gold-shell nanoparticles (after 24 hour exposure period). A representative T2*-weighted image is shown in Figure 5 along with corresponding microscopy confirmation image.

Photothermal response

Representative microscopy images showing the PANC-1 cells exposed to different concentrations of nanoparticles are

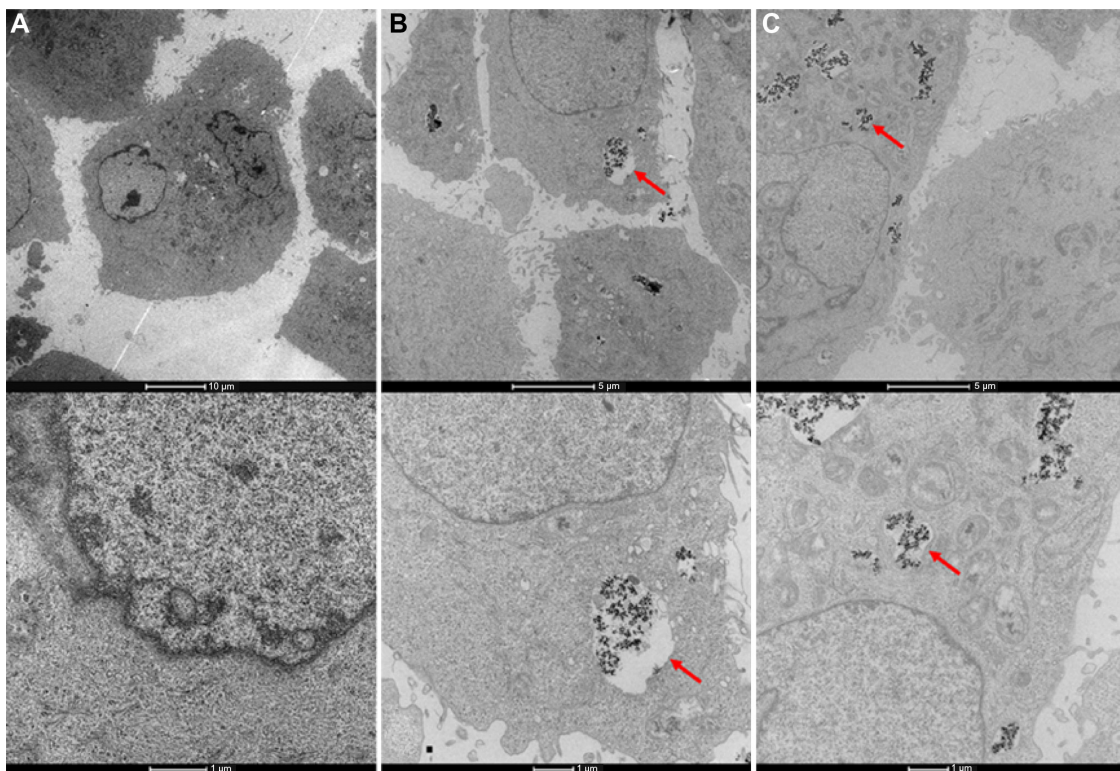


Figure 3 Transmission electron microscopy images of PANC-1 cells exposed to 0 (A), 25 (B), and 50 (C) µg/mL dose of iron oxide and gold hybrid nanoparticles (arrow).

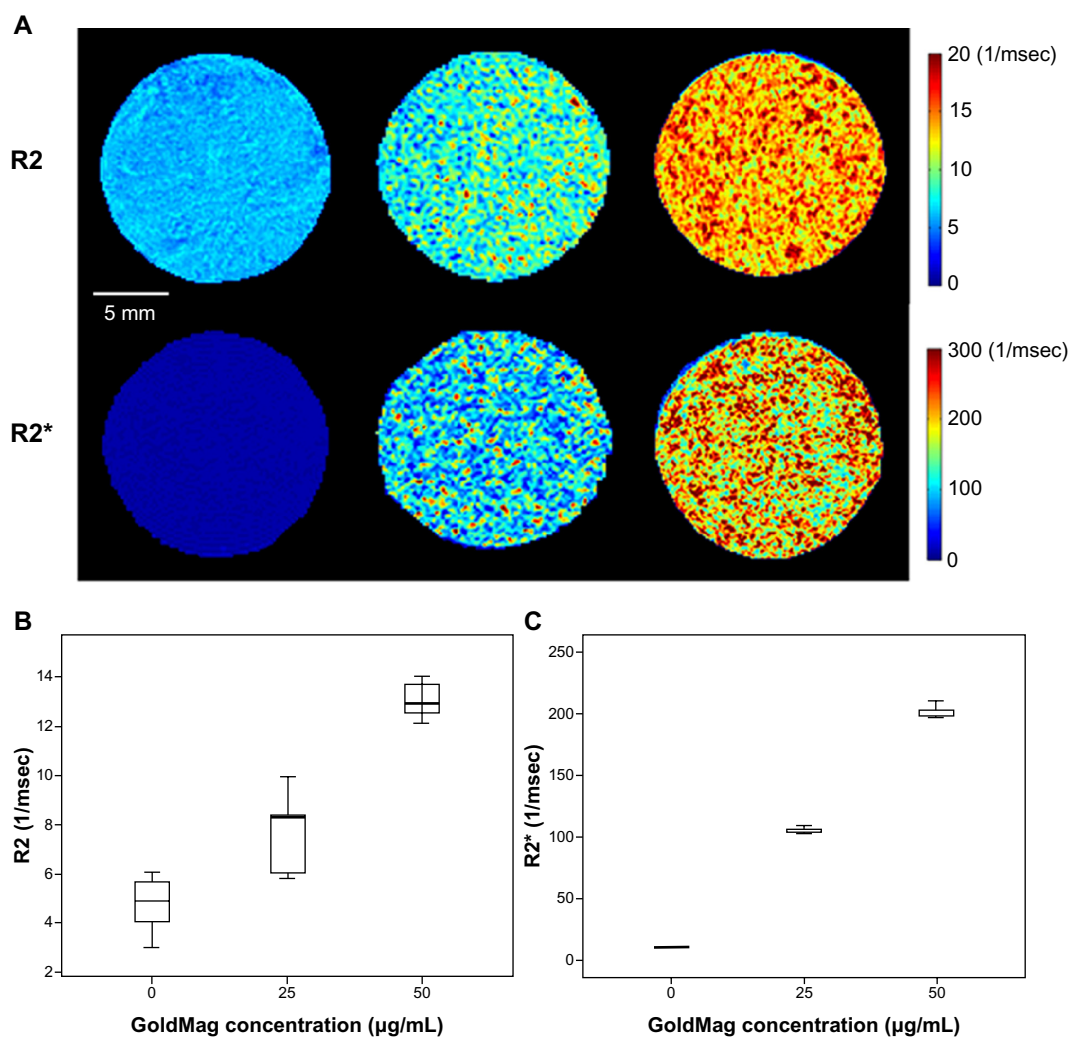


Figure 4 R2 and R2* measurements within PANC-1 cell suspensions after 24-hour exposures to nanoparticles at concentrations of 0, 25, and 50 µg/mL (each cell suspension consisting of 10×10^6 PANC-1 cells in 10 mL 0.8% agarose) (A). Mean R2 (B) and R2* values (C) \pm standard deviation for each corresponding cell suspension.

Note: $R2^* = R2 + R2$.

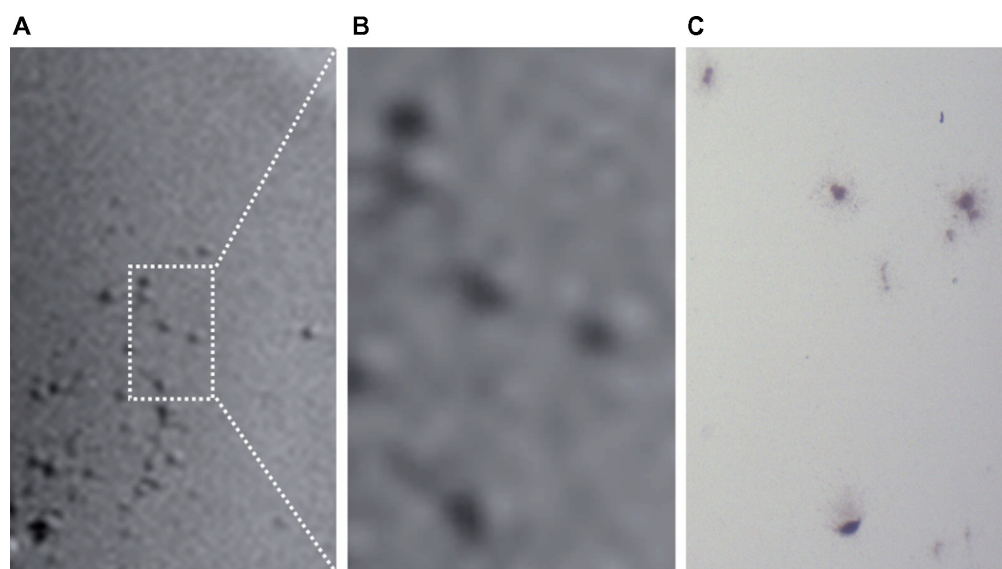


Figure 5 High resolution T2*-weighted images depicting individual PANC-1 cell positions after uptake of the iron oxide core gold-shell nanoparticles (A and B); corresponding $\times 20$ microscopy image (C).

shown in Figure 6. The top row shows the PANC-1 cells exposed to nanoparticles at concentrations of 0, 25, and 50 $\mu\text{g}/\text{mL}$ without laser irradiation; without laser irradiation, cell proliferation percentages changed from 100% to 71.3% and 47.0% with exposures to 0, 25, and 50 $\mu\text{g}/\text{mL}$ nanoparticle doses, respectively. The bottom row of images shows the pancreatic cancer cell samples treated with gold nanoparticles (0, 25, and 50 $\mu\text{g}/\text{mL}$, respectively) and laser irradiation at a power density of 7.9 W/cm^2 ; with laser irradiation, the PANC-1 cells showed significantly decreased cell proliferation (compared with corresponding samples without irradiation) to 61%, 21.9%, and 2.3%, respectively. The latter results for nanoparticle-exposed PANC-1 cells both with and without laser irradiation are summarized in Figure 7A. Difference was found between non-ablated samples and ablated samples in cell proliferation ($P = 0.000$ for each GoldMag concentration).

For PANC-1 cells treated with 25 $\mu\text{g}/\text{mL}$ nanoparticles, the photothermal response increased with increasing power density. PANC-1 cell proliferation percentages decreased from 100% to 75.6%, 32.9%, 20%, and 21.9% as power density increased from 3.2 to 4.7, 6.3, and 7.9 W/cm^2 , respectively (Figure 7B).

Discussion

These studies demonstrated the efficacy of using iron oxide core gold-shell nanoparticles as potent MRI-visible photosensitizers during laser irradiation of human pancreatic

cancer cells. Previous studies have also shown that other types of nanoparticles can also perform well for these types of applications.²³ The salient combination of iron oxide core gold-shell nanoparticles with the local application of NIR laser irradiation offers a promising approach for photothermal ablation therapy in the setting of pancreatic cancer. PANC-1 cell proliferation significantly decreased with exposure to increasing concentrations of the nanoparticles and increasing applied power densities. MRI studies demonstrated quantitative increases in both R2 and R2* with increasing nanoparticle uptake along with the ability to depict individual PANC-1 pancreatic cancer cells after particle phagocytosis.

Heating curves collected during NIR irradiation demonstrated that temperature increased relatively slowly from 8°C to 13°C for those PANC-1 cells that were not exposed to the iron oxide core gold-shell nanoparticles. Upon 24-hour exposure to these particles, temperature changes during irradiation increased dramatically: 7°C, 14°C, 26°C, and 35°C increases for power densities of 3.2, 4.7, 6.3, and 7.9 W/cm^2 , respectively, in 25 $\mu\text{g}/\text{mL}$ exposure group; 60°C, 68°C, 75°C, and 80°C increases for the same power densities in 50 $\mu\text{g}/\text{mL}$ exposure group. These hybrid nanoparticles significantly enhanced photothermal response during these NIR laser irradiation procedures. Similar observations have been reported for a number of prior studies using gold nanomaterials as photothermal sensitizers for ablation of human prostate cancer cells, murine renal carcinoma cells, and MDA-MB-435 cells.^{10,24}

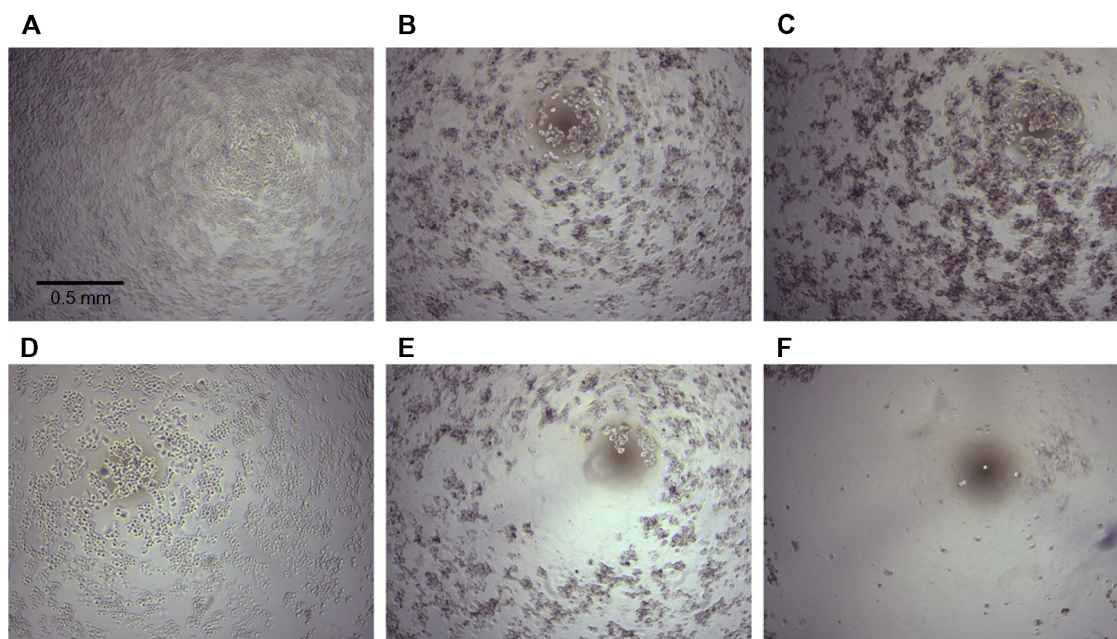


Figure 6 PANC-1 cells exposed to 0, 25, and 50 $\mu\text{g}/\text{mL}$ dose of nanoparticles, respectively, without laser irradiation (A–C) and with laser irradiation at 7.9 W/cm^2 power density (D–F).

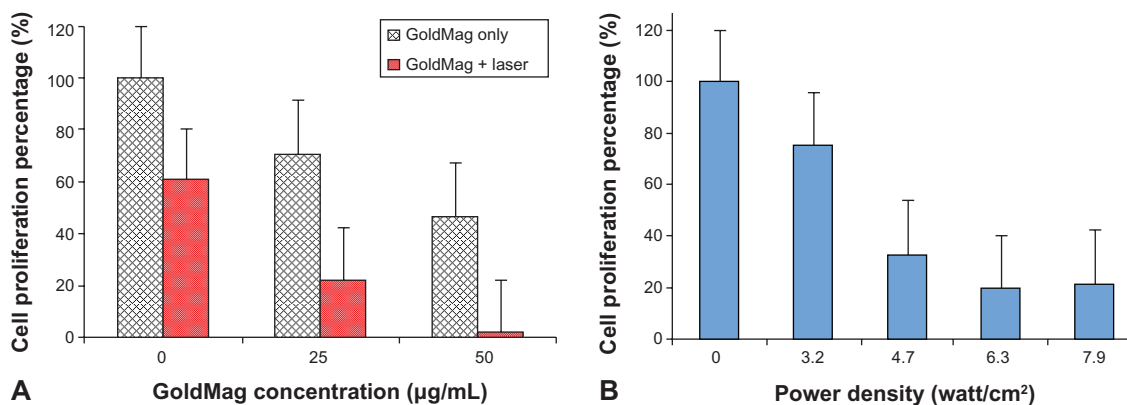


Figure 7 Cell proliferation percentage for PANC-1 cells exposed to different concentrations of nanoparticles with (red) and without (grid) laser irradiation (A); cell proliferation percentage for PANC-1 cells exposed to 25 µg/mL nanoparticle dose and increasing power densities during NIR laser irradiation (B).

Abbreviation: NIR, near-infrared.

Current clinically used thermal ablation technologies include radiofrequency ablation,^{25–27} microwave coagulation therapy,²⁸ and cryoablation therapy.²⁹ These thermal ablation approaches have a common limitation, nonspecific heating of targeted tissues, and potential injury to surrounding healthy tissues. Photothermal ablation approaches alternatively may permit improved tumor-specific heating with nanomaterials serving to photosensitize the targeted tissues. During the current study, iron oxide core gold-shell nanoparticles were readily phagocytosed into human pancreatic cancer cells. These nanoparticles permitted photothermal ablation in a controllable manner with resulting therapeutic reductions to pancreatic cell proliferation dependent upon the photosensitizer dose and power deposition during irradiation. Given the promising results of these initial in-vitro studies, additional investigations are now warranted to rigorously examine the potential therapeutic efficacy of these approaches with in vivo preclinical animal model studies. These approaches may ultimately overcome some of the limitations of current thermal ablation therapies by limiting unwanted damage to adjacent normal tissues.

Intra-procedural imaging-guidance has been widely advocated for most thermal ablation procedures with current modalities including ultrasound, computed tomography, and MRI.^{24,30,31} X-ray computed tomography, MRI, fluorescence, and optical coherence tomography techniques have each been used to quantitatively visualize gold nanoparticle distributions.^{6,32–34} However, fluorescence and optical imaging typically require cell fixation and staining; while noninvasive in vivo imaging may be feasible for rodent models, depth penetration issues limit application to the examination of only superficial tissues in large mammal and human studies. Given that lack of ionizing radiation, MRI may offer the greatest potential for clinical translation.

Cellular MRI techniques have been widely developed for in vivo visualization and quantification of single cells labeled with iron oxide nanoparticles.^{35,36} Quantitative R2 and R2* measurements can be used to map the biodistribution of iron oxide nanoparticles.^{35,36} The nanoparticles used for the current studies included an iron oxide component as a core of the particles, and subsequent MRI studies demonstrated that increased PANC-1 cell uptake yielded significant changes in both R2 and R2* relaxation. Given the noninvasive nature of these measurements, these can already be readily performed in clinical settings. Such measurements may ultimately be critical for treatment planning, allowing the physicians to potentially adjust laser irradiation parameters (power density and duration) based upon MRI assessments of nanoparticle deposition within the targeted pancreatic tumor tissues. Given the superior soft tissue contrast over competing modalities, MRI may also be ideal for real-time placement of the diode fiber laser probe (akin to current clinical procedures using MRI for the placement of radiofrequency ablation or cryo-probes as well as biopsy needles).³⁷

The primary limitation of this current in-vitro feasibility study was the lack of subsequent in vivo validation. This will be a critical next step and will allow important investigation of physiologic influences that may impact therapeutic outcomes. These influences include the resultant nanoparticle biodistribution (following intravenous systemic administration or direction percutaneous injection) as well as potential local heat losses due to perfusion of the tissue targeted for ablation (heat-sink effects).³⁸ The circulation times of nanoparticles after intravenous injection has been a challenge for targeted therapy. Particle size and surface charge are critical factors for avoiding uptake by macrophages prior to reaching the targeted tumors.

Polyethylene glycol can extend circulation times but was not used for surface modification of our nanoparticles for the current study. For future studies, the in vivo bio-availability of GoldMag nanoparticles should be further investigated. In addition, targeting moieties can be readily added to these nanoparticles to facilitate selective delivery to the targeted tumor tissues. Active targeting such as antibody targeting enhances targeting efficiency and should result in better therapeutic response than passive targeting approaches. Prostate stem cell antigen antibodies have been added to these hybrid iron oxide core gold-shell nanoparticles for selective delivery to prostate tumors.³⁹ Anti-epidermal growth factor receptor antibodies are also overexpressed in most human pancreatic cancer cell lines, offering additional potential targets to facilitate selective delivery.^{6,9} The decrease in cell proliferation after nanoparticle administration observed in our study may have resulted from the high surface charge of these nanoparticles. Future modification and improvement of these gold nanoparticles is warranted.

In conclusion, these studies demonstrated the potential to use iron oxide gold-shell nanoparticles as MRI-visible photothermal sensitizers during laser irradiation of pancreatic cancer cells. Exposure to these nanoparticles and subsequent laser irradiation led to significant reductions in pancreatic cancer cell proliferation; MRI R2 and R2* measurements correlated to nanoparticle cellular uptake levels. Future in vivo translational studies are now warranted to validate these approaches in animal models of pancreatic cancer.

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Disclosure

The authors report no conflicts of interest in this work.

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