

Activation of Erk and p53 regulates copper oxide nanoparticle-induced cytotoxicity in keratinocytes and fibroblasts

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Abstract: Copper oxide nanoparticles (CuONP) have attracted increasing attention due to their unique properties and have been extensively utilized in industrial and commercial applications. For example, their antimicrobial capability endows CuONP with applications in dressings and textiles against bacterial infections. Along with the wide applications, concerns about the possible effects of CuONP on humans are also increasing. It is crucial to evaluate the safety and impact of CuONP on humans, and especially the skin, prior to their practical application. The potential toxicity of CuONP to skin keratinocytes has been reported recently. However, the underlying mechanism of toxicity in skin cells has remained unclear. In the present work, we explored the possible mechanism of the cytotoxicity of CuONP in HaCaT human keratinocytes and mouse embryonic fibroblasts (MEF). CuONP exposure induced viability loss, migration inhibition, and G₂/M phase cycle arrest in both cell types. CuONP significantly induced mitogen-activated protein kinase (extracellular signal-regulated kinase [Erk], p38, and c-Jun N-terminal kinase [JNK]) activation in dose- and time-dependent manners. U0126 (an inhibitor of Erk), but not SB 239063 (an inhibitor of p38) or SP600125 (an inhibitor of JNK), enhanced CuONP-induced viability loss. CuONP also induced decreases in p53 and p-p53 levels in both cell types. Cyclic pifithrin- α , an inhibitor of p53 transcriptional activity, enhanced CuONP-induced viability loss. Nutlin-3 α , a p53 stabilizer, prevented CuONP-induced viability loss in HaCaT cells, but not in MEF cells, due to the inherent toxicity of nutlin-3 α to MEF. Moreover, the experiments on primary keratinocytes are in accordance with the conclusions acquired from HaCaT and MEF cells. These data demonstrate that the activation of Erk and p53 plays an important role in CuONP-induced cytotoxicity, and agents that preserve Erk or p53 activation may prevent CuONP-induced cytotoxicity.

Keywords: cell cycle arrest, CuONP, MAPK, nutlin-3 α , cyclic pifithrin- α

Introduction

Copper oxide nanoparticles (CuONP) have been widely applied in semiconductors, catalysts, microelectronic materials, and lithium batteries.¹ Similar to silver and ZnO nanoparticles, CuONP exhibit excellent antimicrobial efficiency.²⁻⁴ Much attention has been paid to the feasible applications of CuONP, such as antimicrobial masks⁵ and textiles.^{6,7} However, it is crucial to evaluate their potential effects on the environment and human health before their practical application. Certain reports have revealed that CuONP are toxic to the aquatic macrophyte *Lemna gibba*⁸ and *Xenopus laevis*.⁹ CuONP were also reported to be cytotoxic to human cells, such as brain,¹⁰ lung,^{11,12} liver,^{13,14} kidney,¹⁵ and skin¹⁶ cells. A test using the human lung adenocarcinoma cell line A549 implied that CuONP were more toxic than other metal oxide nanoparticles (TiO₂, CuZnFe₂O₄, Fe₃O₄, and Fe₂O₃) and carbon-based nanomaterials (carbon nanpowders and multiwalled carbon nanotubes).¹⁷ Organs such as the lung and skin were

more vulnerable to nanoparticles due to their direct contact with the external environment. Many studies have focused on CuONP-induced cytotoxicity and corresponding mechanisms in human lung epithelial cells.^{11,12,17–19} Oxidative stress and autophagy have been reported to play vital roles in CuONP-induced cytotoxicity.^{12,18} Hanagata et al reported that CuONP exposure upregulated genes involved in mitogen-activated protein kinase (MAPK) pathways while downregulating genes involved in cell cycle progression.¹¹ Semisch et al showed that compared with microscale CuO particles and copper chloride, CuONP were more cytotoxic and induced genotoxicity due to their nanoscale features.²⁰ CuONP were also reported to induce oxidative stress and apoptosis in HaCaT human keratinocytes. However, the decisive factors and the accompanying changes involved in the toxicity of CuONP are unclear.¹⁶ Moreover, the underlying mechanism of the toxicity of CuONP to skin-associated cells has not been clarified. In the present work, we examined the cytotoxicity

of CuONP to skin-associated cells, keratinocytes (HaCaT) and mouse embryonic fibroblasts (MEF), and investigated the underlying molecular mechanisms.

Materials and methods

Materials

CuONP was synthesized as Zhu et al reported,²¹ and characterized using transmission electron microscope H-7650 (Hitachi Ltd., Tokyo, Japan), a Malvern Zetasizer Nano ZS90 (Malvern Instruments, Malvern, UK), and X-ray Photoelectron Spectrometer (Thermo Fisher Scientific, Waltham, MA, USA) (Figure 1). 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyltetrazolium bromide (MTT), SB 239063, U0126, propidium iodide, SP600125, and anti-actin primary antibody were purchased from Sigma-Aldrich Co. (St Louis, MO, USA). Fetal bovine serum (FBS) was purchased from Biological Industries Israel Beit-Haemek Ltd. (Kibbutz Beit-Haemek, Israel). Dulbecco's Modified Eagle's Medium (DMEM),

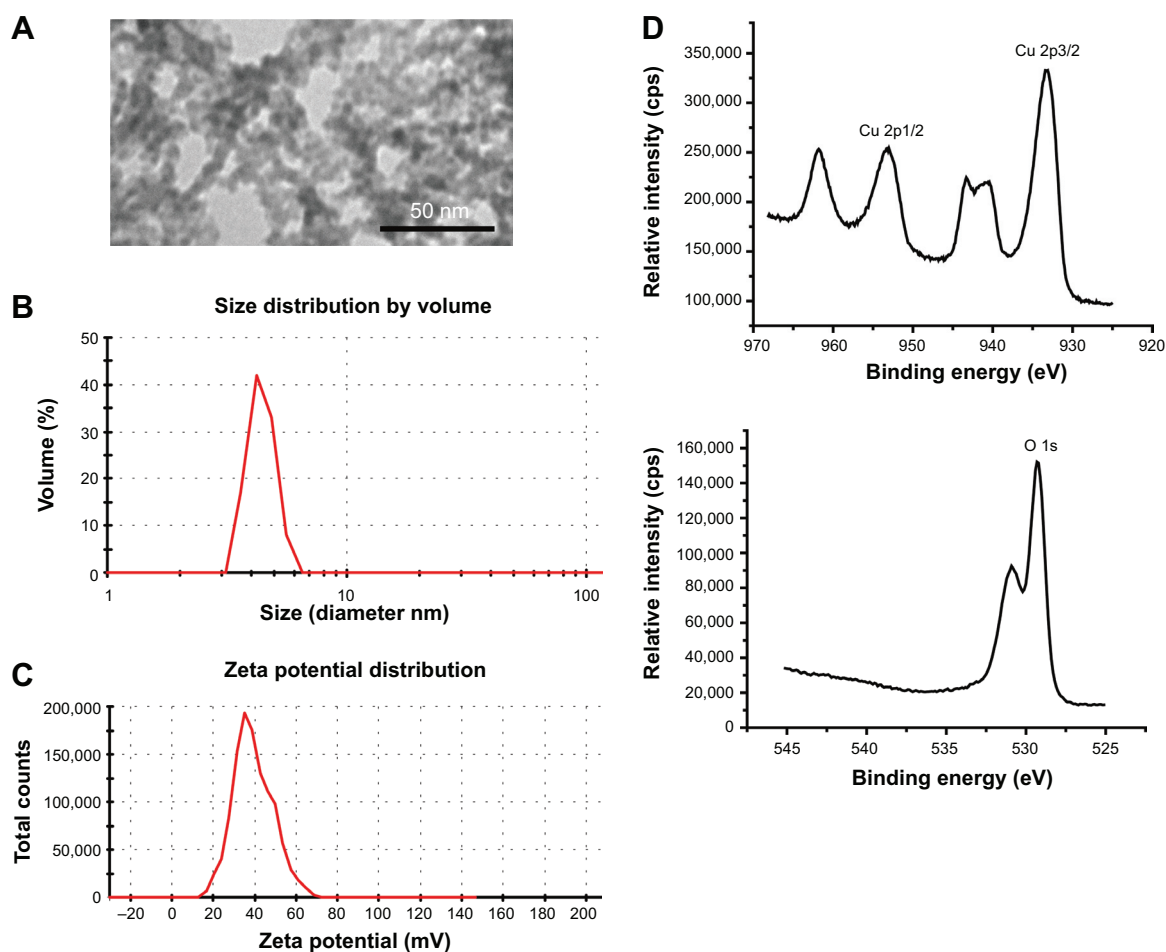


Figure 1 Physicochemical characterization of the CuONP.

Notes: (A) Transmission electron microscope image. (B) Hydrodynamic diameter. (C) Zeta potential. (D) Chemical composition by X-ray photoelectron spectrometer analysis.

Abbreviations: CuONP, copper oxide nanoparticles; cps, counts per second.

penicillin, streptomycin, and trypsin were purchased from Thermo Fisher Scientific. Primary antibodies against p-p53 and phosphorylated extracellular signal-regulated kinase (Erk) were obtained from Cell Signaling Technology, Inc. (Beverly, MA, USA). Primary antibodies against cyclin A, cyclin B1, p53, phosphorylated c-Jun N-terminal kinase (JNK), JNK, p-p38, p38, Erk, and cyclic pifithrin- α (PFT- α) were purchased from Santa Cruz Biotechnology Inc. (Dallas, TX, USA). Horseradish peroxidase-conjugated secondary antibodies were purchased from Jackson ImmunoResearch Laboratories, Inc. (West Grove, PA, USA). The bicinchoninic acid assay kit and enhanced chemiluminescence Western blotting substrate were obtained from Thermo Fisher Scientific. Cell lysis buffer was purchased from Beyotime Biotechnology (Haimen, Jiangsu, People's Republic of China). Other chemicals were obtained from local suppliers. Deionized water was used in all experiments.

Cell culture and treatment

Human keratinocytes HaCaT and mouse embryonic fibroblast cells were cultured in DMEM supplemented with 10% FBS and penicillin (100 U/mL)/streptomycin (100 μ g/mL) in a 5% CO₂ humidified incubator at 37°C (Thermo Fisher Scientific). In a dose-dependent manner, HaCaT and MEF cells were treated with the indicated concentrations of CuONP for 24 hours. In a time-dependent manner, HaCaT or MEF cells were treated with 60 or 80 μ mol/L CuONP, respectively.

For the MAPK-inhibitor treatments, HaCaT cells were pretreated with 5 μ mol/L U0126 for 2 hours and then treated with 20 or 40 μ g/mL CuONP for 24 hours; HaCaT cells were pretreated with 20 μ mol/L SB 239063 or 20 μ mol/L SP600125 for 2 hours and then treated with 20 or 60 μ g/mL CuONP for 24 hours; MEF cells were pretreated with 5 μ mol/L U0126 for 2 hours and then treated with 40 or 60 μ g/mL CuONP for 24 hours; MEF cells were pretreated with 20 μ mol/L SB 239063 or 20 μ mol/L SP600125 for 2 hours and then treated with 40 or 80 μ g/mL CuONP for 24 hours.

For the p53-transcriptional-inhibitor (cyclic PFT- α) treatment, HaCaT cells were pretreated with 1 or 10 μ mol/L cyclic PFT- α for 2 hours and then treated with 20 μ g/mL CuONP for 24 hours or with 60 μ g/mL CuONP for 6 hours, respectively. MEF cells were pretreated with 1 or 10 μ mol/L cyclic PFT- α for 2 hours and then treated with 40 μ g/mL CuONP for 24 hours or with 80 μ g/mL CuONP for 3 hours, respectively. For p53-stabilizer (nutlin-3 α) treatment, HaCaT or MEF cells were pretreated with 30 μ mol/L nutlin-3 α for 2 hours and then treated with 60 μ g/mL or 80 μ g/mL CuONP for 24 hours, respectively.

Cellular viability assessment

The cells were washed with FBS-free DMEM once and incubated with 0.5 mg/mL MTT in FBS-free DMEM for 1 hour at 37°C in a CO₂ incubator. The formazan generated was dissolved in DMSO and recorded at 490 nm with a microplate reader (Molecular Devices LLC, Sunnyvale, CA, USA).²²

Migratory assay

The cells were scratched with a 200 μ L pipette tip across the center of a plate to create a wound. The detached debris was washed off with FBS-free DMEM. The cells were treated with the indicated concentrations of CuONP. Photographs were taken at 0 and 24 hours after the treatment.²³

Cell cycle analysis

The cells were trypsinized and washed with PBS once. The cells were then resuspended in PBS, followed by fixation in ice-cold 70% ethanol and storage at -20°C for at least 24 hours. The fixed cells were collected by centrifugation at 1,000 rpm for 10 minutes and washed with PBS once, followed by incubation with a propidium iodide working solution (0.1% Triton-100, 200 μ g/mL RNase A, and 50 μ g/mL stock solution) for 15 minutes in the dark and analysis by flow cytometry.²⁴

Western blot analysis

The cells were lysed and centrifuged at 13,000 rpm for 15 minutes at 4°C to collect the supernatant. The protein content in the lysate's supernatant was quantified with a bicinchoninic acid assay kit. The proteins were separated by sodium dodecyl sulfate polyacrylamide gel electrophoresis and transferred to nitrocellulose membranes. The membranes were blocked with 5% non-fat milk in Tris-buffered saline containing 0.1% Tween 20 (TBST) for 1 hour at room temperature, probed with primary antibodies against the target proteins overnight at 4°C, and then washed with TBST to remove unbound antibodies. Next, the membranes were incubated with horseradish peroxidase-conjugated secondary antibodies for 1 hour at room temperature and washed with TBST again. Specific signals on the probed membranes were detected using enhanced chemiluminescence and an X-ray-film exposure system.

Statistical analysis

A one-way analysis of variance, followed by Fisher's least significant difference test, was used to evaluate the significant differences among the treatments. The data are presented as the mean \pm standard deviation ($n > 3$). Statistical significance was indicated as follows: * $P < 0.05$ and ** $P < 0.01$.

Results

CuONP induce cytotoxicity in HaCaT and MEF cells

CuONP significantly suppressed the proliferation of HaCaT and MEF cells in a dose-dependent manner in 24 hours' treatment (Figure 2A and B). Cellular migration capacities were also inhibited in both CuONP-treated cells in scratch-wound assay (Figure 2C and D).

Cell cycle arrest may promote growth and migration inhibition in CuONP-treated HaCaT and MEF cells. To verify this assumption, CuONP-treated cells were subjected to cell cycle analysis with flow cytometry. There was a significant increase in the population of cells in G₂/M phase (from 3.9% to 42.8% in HaCaT, or from 1.2% to 32.8% in MEF), with corresponding decreases in the G₁ and S phases after 24 hours' CuONP treatment, which implied that the cell cycle was arrested in G₂/M phase in a dose-dependent manner in CuONP-treated cells (Figure 3A and B). There was a dose- and time-dependent decrease in the levels of cyclin A and cyclin B1 in CuONP-treated HaCaT cells (Figure 3C and D) and MEF cells (Figure 3E and F). The downregulation of those G₂/M transition-related proteins further revealed that CuONP induced G₂/M phase arrest.

Erk activation may prevent CuONP-induced growth inhibition in HaCaT and MEF cells

The activation of Erk, p38, and JNK was significantly induced in dose- and time-dependent manners in CuONP-treated HaCaT cells (Figure 4A and B) and MEF cells (Figure 4C and D). The inhibition of Erk activation by pretreatment with U0126 significantly promoted growth inhibition relative to treatment with CuONP alone in HaCaT cells (Figure 4E) and MEF cells (Figure 4F). However, pretreatment with SB 239063 (an inhibitor of p38 activation) and SP600125 (an inhibitor of JNK activation) exhibited little or no effect on the viabilities of CuONP-treated HaCaT cells (Figure 4G) and MEF cells (Figure 4H). These data implied that Erk plays a vital role in CuONP-induced cytotoxicity, whereas the changes in p38 and JNK were incidental.

Loss of p53 accelerates CuONP-induced growth inhibition in HaCaT and MEF cells

There was a remarkable decrease in the protein levels of p53 and p-p53 in a dose-dependent manner in CuONP-treated HaCaT cells (Figure 5A) and MEF cells (Figure 5B).

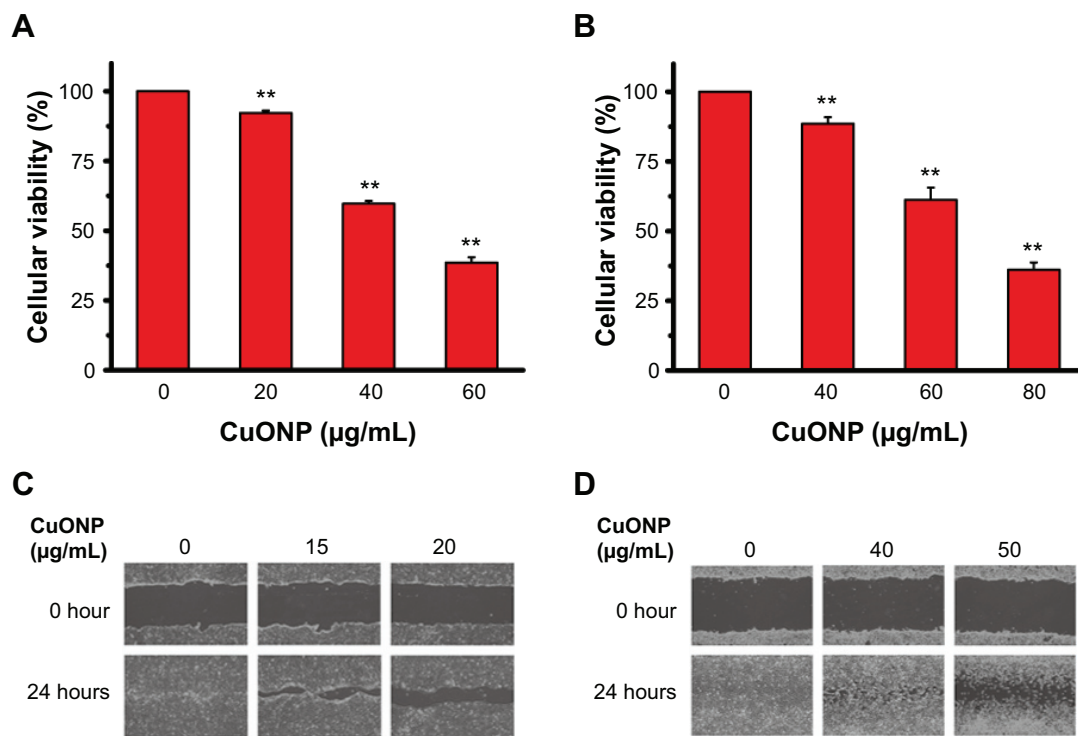


Figure 2 CuONP-induced cytotoxicity in HaCaT and MEF cells.

Notes: Viability loss in HaCaT (A) and MEF (B) cells with CuONP treatment for 24 hours. Migration inhibition in CuONP-treated HaCaT (C) and MEF (D) cells. The data are presented as the mean \pm standard deviation ($n > 3$). ** $P < 0.01$ versus vehicle control.

Abbreviations: CuONP, copper oxide nanoparticles; MEF, mouse embryonic fibroblasts.

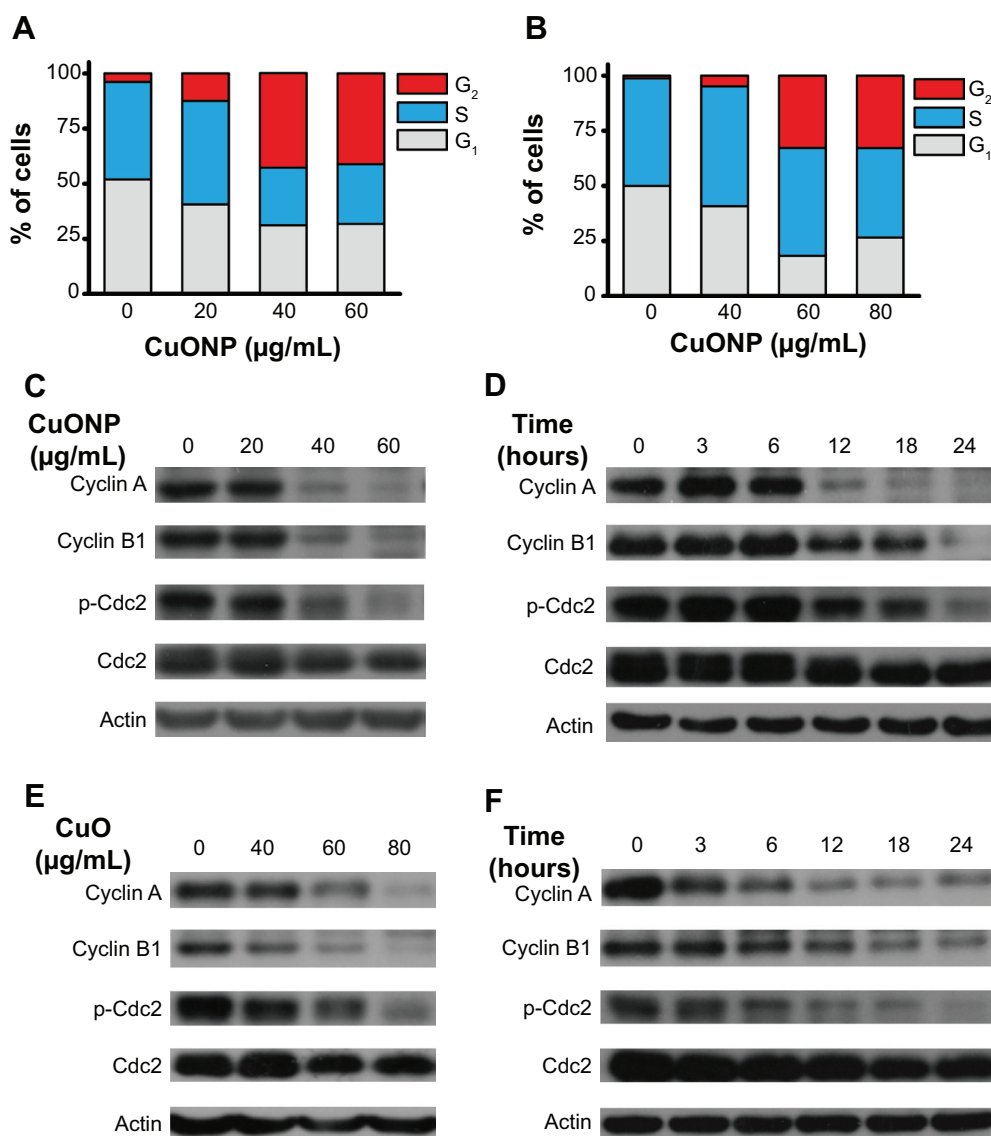


Figure 3 CuONP-induced cell cycle arrest in HaCaT and MEF cells.

Notes: Cell cycle analysis by flow cytometry for CuONP-treated HaCaT (A) and MEF (B) cells. Western blot analysis of the proteins associated with the G₂/M transition in CuONP-treated HaCaT (C, D) and MEF (E, F) cells.

Abbreviations: CuONP, copper oxide nanoparticles; MEF, mouse embryonic fibroblasts.

Pretreatment with cyclic PFT- α , a p53 transcriptional inhibitor, promoted growth inhibition relative to treatment with CuONP alone in HaCaT cells (Figure 5C and D) and MEF cells (Figure 5E and F). Briefly, pretreatment with 1 μ mol/L cyclic PFT- α promoted growth inhibition in both cells with low doses of CuONP for 24 hours' treatment (Figure 5C and E), and pretreatment with 10 μ mol/L cyclic PFT- α promoted growth inhibition in high doses of CuONP for 6 hours' (for HaCaT) or 3 hours' (for MEF) treatment (Figure 5D and F). Moreover, pretreatment with nutlin-3 α , a murine double minute-2 (MDM2)/p53 inhibitor that stabilizes p53, prevented high dose of CuONP-induced viability loss in HaCaT cells (Figure 5G). Nutlin-3 α itself was toxic

to MEF cells, so no inhibition of the viability loss induced by CuONP was observed (Figure 5H).

CuONP-induced cytotoxicity in primary keratinocytes

Primary rat keratinocytes were isolated from newborn rat skins according to a report from Albarenque and Doi.²⁵ CuONP induced dose-dependent viability loss in primary keratinocytes (Figure 6A). Erk inhibitor (U0126) and p53 transcriptional inhibitor (cyclic PFT- α) promoted CuONP-induced viability loss (Figure 6B). Nutlin-3 α was toxic to primary keratinocytes, which is the same as the MEF; no inhibition of the viability loss induced by CuONP was observed.

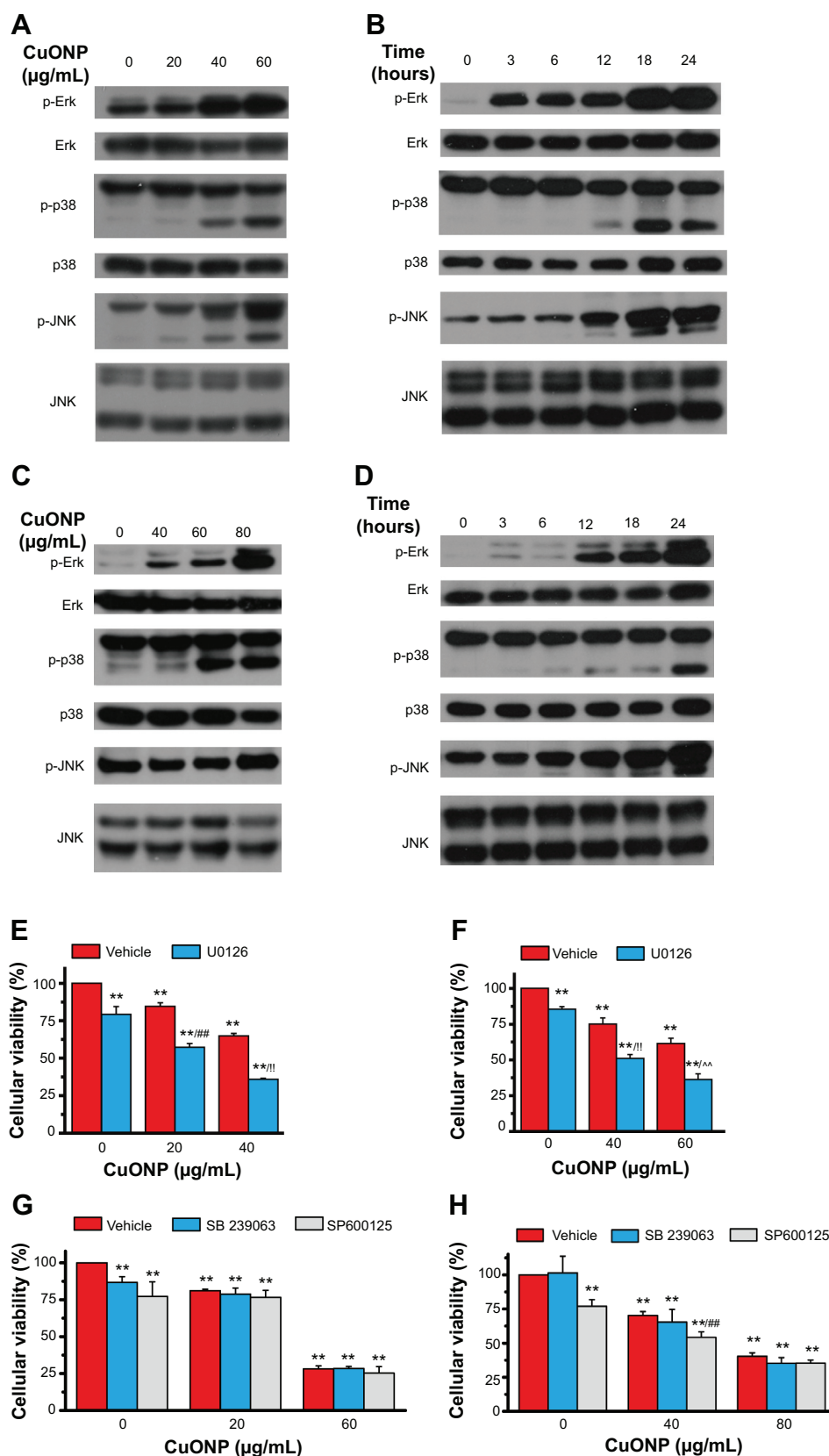


Figure 4 Erk was involved in CuONP-induced cytotoxicity in HaCaT and MEF cells.

Notes: Western blot analysis of CuONP dose-dependent MAPK activation in HaCaT (A) and MEF (C) cells; Western blot analysis of CuONP time-dependent MAPK activation in 60 $\mu\text{g/mL}$ CuONP-treated HaCaT (B) and 80 $\mu\text{g/mL}$ CuONP-treated MEF (D) cells; the effects of MAPK inhibitors (U0126, SP600125, and SB 239063) on CuONP-induced viability loss in HaCaT (E, G) and MEF (F, H) cells. The data are presented as the mean \pm standard deviation ($n > 3$). ** $P < 0.01$ versus vehicle control. ### $P < 0.01$ versus 20 $\mu\text{g/mL}$ CuONP group. # $P < 0.01$ versus 40 $\mu\text{g/mL}$ CuONP group. * $P < 0.01$ versus 60 $\mu\text{g/mL}$ CuONP group.

Abbreviations: CuONP, copper oxide nanoparticles; MEF, mouse embryonic fibroblasts; MAPK, mitogen-activated protein kinase.

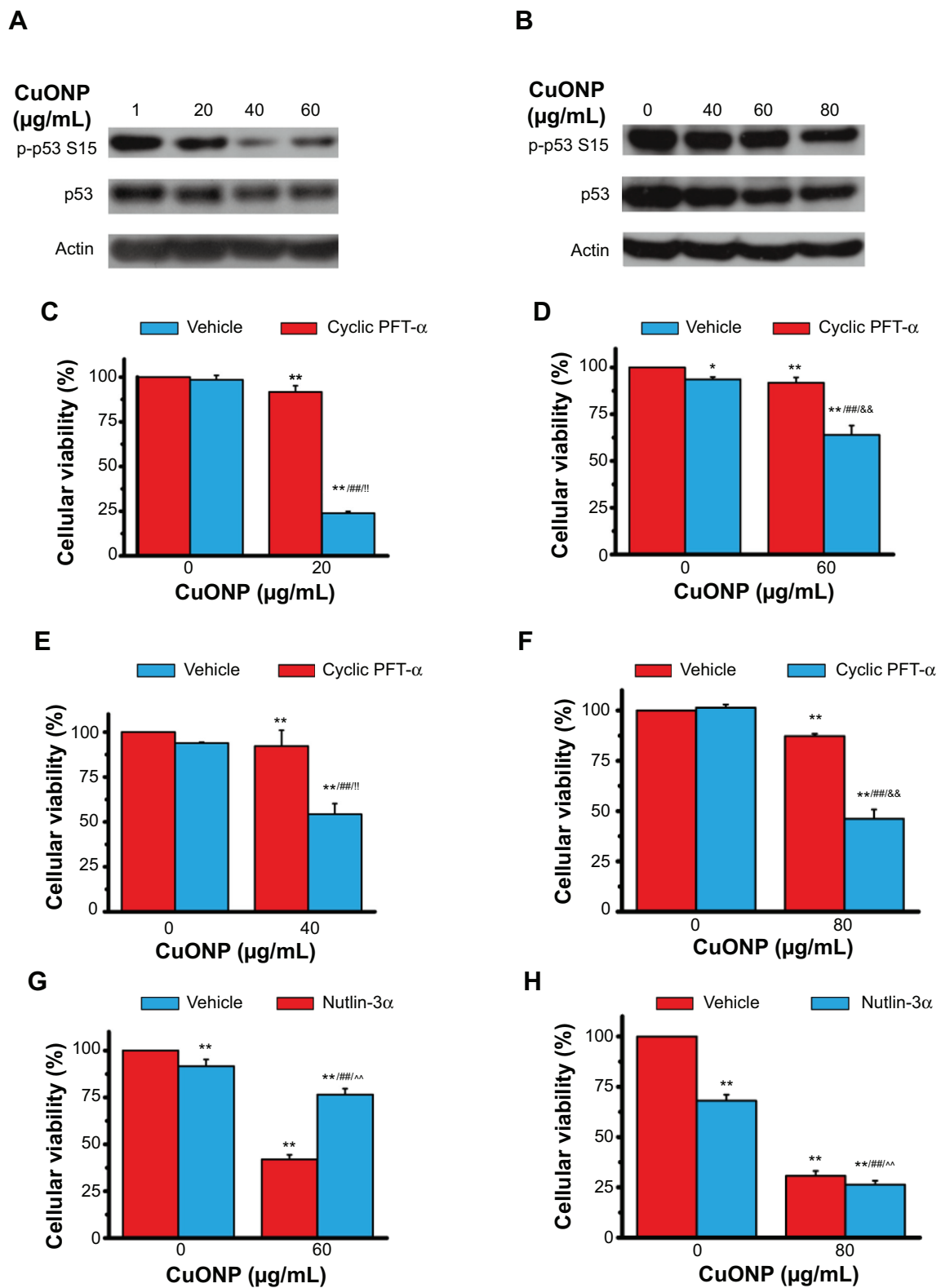


Figure 5 p53 was involved in CuONP-induced cytotoxicity in HaCaT and MEF cells.

Notes: Western blot analysis of p53 and p-p53 in CuONP-treated HaCaT (A) and MEF (B) cells. The effects of 1 $\mu\text{mol/L}$ p53 transcriptional inhibitor (cyclic PFT- α) on CuONP-induced viability loss in HaCaT (C) and MEF (E) cells with CuONP treatment for 24 hours. The effects of 10 $\mu\text{mol/L}$ cyclic PFT- α on CuONP-induced viability loss in HaCaT cells with CuONP treatment for 6 hours (D) and in MEF cells with CuONP treatment for 3 hours (F). The effects of 30 $\mu\text{mol/L}$ p53 stabilizer (nutlin-3 α) on CuONP-induced viability loss in HaCaT (G) and MEF (H) cells with CuONP treatment for 24 hours. The data are presented as the mean \pm standard deviation ($n > 3$). ** $P < 0.01$ versus vehicle control. ## $P < 0.01$ versus CuONP group. # $P < 0.01$ versus 1 $\mu\text{mol/L}$ cyclic PFT- α group. ## $P < 0.01$ versus 10 $\mu\text{mol/L}$ cyclic PFT- α group. ** $P < 0.01$ versus nutlin-3 α group.

Abbreviations: CuONP, copper oxide nanoparticles; MEF, mouse embryonic fibroblasts; PFT- α , pifithrin- α .

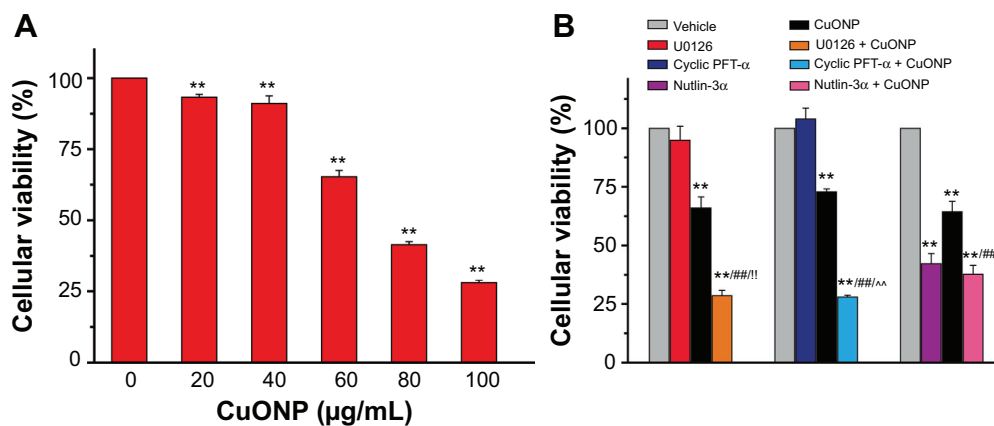


Figure 6 Cytotoxicity of CuONP in primary keratinocytes.

Notes: Viability loss in primary keratinocytes with CuONP treatment for 24 hours (A). The effects of Erk inhibitor (U0126, 5 µmol/L), p53 transcriptional inhibitor (cyclic PFT-α, 1 µmol/L), and p53 stabilizer (nutlin-3α, 30 µmol/L) on viability loss in primary keratinocytes with 60 µg/mL CuONP treatment for 24 hours (B). The data are presented as the mean ± standard deviation (n>3). **P<0.01 versus vehicle control. ###P<0.01 versus CuONP group. ^P<0.01 versus U0126 group. ^^P<0.01 versus cyclic PFT-α group.

Abbreviations: CuONP, copper oxide nanoparticles; PFT-α, pifithrin-α.

Discussion

CuONP have been applied in various industrial and medical areas, such as heat-transfer systems, gas sensors, electronic chips, and biocides.¹ The skin, respiratory tract, and gastrointestinal tract are the most susceptible to the adverse effects of hazardous materials outside the human body due to their direct exposure to the environment. The skin is the largest among these organs, which increases the risk of being affected. Moreover, potent medical applications utilizing the antibacterial activity of CuONP give rise to direct contact between CuONP and the skin.^{6,7} Gauze and bandages containing CuONP could protect wounds against microbial infection. However, there is a risk in using CuONP application as a microbicide for covering the skin surface: ie, harm to skin-associated cells while killing microorganisms. Keratinocytes and fibroblasts, cooperating to facilitate skin repair, will be affected by CuONP. The redox activities and nanoscale properties of CuONP may promote cytotoxicity to these cells. CuONP exposure has been reported to induce a significant decrease in cellular viability and glutathione levels, and an increase in oxidative stress, apoptosis, and DNA damage in HaCaT cells.¹⁶ However, the underlying mechanisms are not well clarified. The present work reveals that cell cycle arrest occurred in CuONP-treated keratinocytes and fibroblasts and that the activation of Erk and p53 protected both cell types from cytotoxicity.

The proliferation and migration of keratinocytes and fibroblasts are important stages of wound repair and skin development.²⁶ A loss of skin integrity renders the body vulnerable to invasion by foreign pathogens. Viability loss restrains the proliferation and migration capacity of both cell types, which will undermine cutaneous wound healing and

lead to inflammation and infection. Cell cycle progression dysfunction is another pivotal factor affecting proliferation. Generally, one cell divides into two daughter cells when one cell cycle completes, and those progressions are under the control of checkpoints.²⁷ Combinations of cyclins and cyclin-dependent kinases, the main components of checkpoints, vary in different stages of the cell cycle and regulate cell cycle progression. The cell cycle can be blocked in certain stages due to cellular damage and stress signaling, which makes the checkpoints defective. If the protein levels of cyclin A and B do not increase after G₁ phase, cell progression cannot pass through G₂/M phase and complete mitosis.²⁸ In the present work, following CuONP treatment, the deregulation of cyclin A and B dynamics led to blockage of G₂/M transitions and further promoted viability loss (Figures 2 and 3).

Erk, JNK, and p38 MAPK compose a large family of MAPKs that regulates various cellular functions and activities, such as proliferation, differentiation, mitosis, survival, and death.²⁹ The activation of Erk, p38, and JNK was significantly induced in dose- and time-dependent manners in CuONP-treated HaCaT cells (Figure 4A and B) and MEF cells (Figure 4C and D). The activation of JNK and p38 MAPK has been reported to promote cell death.³⁰ However, neither the inhibition of p38 activation nor the inhibition of JNK activation prevented viability loss in CuONP-treated HaCaT and MEF cells (Figure 4G and H). These data imply that the activation of p38 and JNK under CuONP exposure was incidental. Strong Erk activation was induced and sustained for several hours (Figure 4A–D). There are two opposing functions of Erk activation in cell proliferation and cell death.³⁰ When Erk activation was suppressed by inhibitors (eg, U0126), cell death due to

harmful reagents was promoted or suppressed, corresponding to Erk's prosurvival and prodeath functions, respectively. Martin and Pognonec described both functions of Erk in cadmium toxicity, depending on different experimental conditions (eg, the cadmium treatment and cell type).³¹ It was necessary to inhibit Erk activation to uncover the roles that Erk played in CuONP-induced cytotoxicity. The inhibition of Erk activation promoted viability loss in cells treated with both low and high concentrations of CuONP, which implies a prosurvival function for Erk in this context. Several studies reported that CuONP induced reactive oxygen species (ROS) generation in different cells,^{10,13,18,32} including keratinocytes¹⁶ and fibroblasts.³³ For example, Alarifi et al reported that CuONP induced dose-dependent cell death and ROS generation in keratinocytes.¹⁶ Considering that ROS generation would activate Erk,^{29,34,35} the dose-dependent Erk activation in the present work might be similarly due to CuONP-induced ROS generation.

p53 is a transcription factor that regulates the expression of hundreds of genes. Those genes are involved in metabolic homeostasis, antioxidant defense, DNA repair, growth arrest, senescence, apoptosis, autophagy, and other processes. The stability of p53 protein is mainly regulated by interaction with MDM2. The p53/MDM2 complex promotes p53 degradation by ubiquitylation and blocks the transactivation domain.³⁶ Nutlin-3 α can stabilize p53 levels and the transcriptional activity by inhibiting the MDM2-p53 interaction.³⁷ The phosphorylation of p53 at serine 15 stimulates its transactivation.^{38,39} Cyclic PFT- α , a cyclic analog of PFT- α , inhibits p53-dependent gene transcription.⁴⁰ Surprisingly, in the current study, p53 and p-p53 levels decreased in CuONP-treated cells (Figure 5A and B), which implies that p53 did not have a prodeath function. Cyclic PFT- α and nutlin-3 α were used to further evaluate whether p53 had a prosurvival function. Figure 5C–H show that cyclic PFT- α significantly promoted CuONP-induced viability loss in both HaCaT and MEF cells, whereas nutlin-3 α treatment inhibited the cellular viability loss of CuONP-treated HaCaT cells. These data indicate that p53 is involved in CuONP-induced cytotoxicity and plays a prosurvival role. There was no viability-loss inhibition observed following nutlin-3 α treatment of CuONP-treated MEF cells, which may have resulted from the cytotoxicity of nutlin-3 α to MEF cells. The different responses of HaCaT and MEF cells to nutlin-3 α treatment might have been due to the different p53 statuses of these two types of cells: there are p53 mutations in HaCaT cells.⁴¹ Tovar et al reported that nutlin-3 treatment induced growth inhibition in p53-wildtype fibroblasts.⁴² However, no literature has reported similar phenomena in HaCaT cells. These reports suggest that HaCaT

cells may be more tolerant to high p53 levels than MEF cells are. Therefore, pretreatment with nutlin-3 α stabilized p53 and increased its levels in MEF cells, resulting in cytotoxicity.

Conclusion

In conclusion, we have investigated the cytotoxic effects of CuONP on skin-associated HaCaT cells, MEF cells, and primary keratinocytes, as well as the underlying mechanisms. The present study showed that CuONP is toxic to skin-associated cells and that Erk and p53 may be the key factors regulating this cytotoxicity. These results suggest that contact with CuONP may cause damage to the skin and that the activation of Erk and p53 might be a possible way to prevent the potential toxicity of CuONP to the skin.

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Disclosure

The authors report no conflicts of interest in this work.

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