

Clinical significance of TIPE expression in gastric carcinoma

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Background: TNFAIP8, also known as TIPE, is a suppressor of apoptosis. High expression of both TIPE mRNA and protein has been detected in various cancer cell lines and clinical specimens compared to healthy tissues. Many reports have shown that there is a strong correlation between TIPE overexpression and cancer progression and poor prognosis in human solid cancers.

Methods: To illustrate the functional and clinical significance of TIPE in gastric cancer, we used reverse transcription polymerase chain reaction, quantitative real-time polymerase chain reaction, and immunohistochemistry to measure TIPE expression in clinical gastric specimens. Then, TIPE expression was knocked down by using shRNA and anti-DR5ScFv, to examine different expressions of TIPE in BGC823 cell lines, while cell proliferation and apoptosis were induced.

Results: We found that there was a strong correlation between TIPE expression and TNM stage ($P=0.044$), tumor depth ($P=0.016$), lymph node metastasis ($P=0.026$), and distant metastasis ($P=0.045$). No significant correlation was found between TIPE expression with the patients' age ($P=0.062$) or sex ($P=0.459$). Anti-DR5ScFv induced TIPE depletion both in vitro and in vivo and resulted in apoptosis and suppression of proliferation.

Conclusion: Our results suggested that TIPE expression was associated with gastric cancer progression, and most importantly, suppressing TIPE expression might be an effective therapeutic strategy.

Keywords: TNFAIP8, gastric cancer, shRNA, anti-DR5ScFv, metastasis, apoptosis

Introduction

Gastric cancer is the most common digestive system tumor. Gastric cancer is lethally aggressive and malignant, so it is necessary to find suitable biomarkers of malignancy to monitor cancer progression. Many mutations and/or abnormal expression of genes are associated with cancer initiation, progression, and metastasis.¹

Higher mRNA expression of TNFAIP8, also known as TIPE, GG2-1, SCC-S2, or NDED, has been detected in many malignant cell lines and tissues.²⁻⁵ TIPE expression levels have been studied in several kinds of human cancers both in vitro and in vivo, but no study has been conducted in gastric cancer so far. From the current research, studies have shown that TIPE overexpression was found to be correlated with cancer progression and poor prognosis in patients with lung cancer. In contrast, in vitro studies showed the depletion of TIPE mRNA, by using siRNA, in lung cancer cell lines and esophageal squamous cell carcinoma significantly inhibited cell growth and invasion.^{6,7}

The benefit of using small molecules and monoclonal antibodies could be nontoxic to most of the proteins.⁸ Thus, using siRNA to target mRNA upstream appears to be the next logical strategy. The siRNA can be formed by cutting the hairpin parts of the shRNA by using the Dicer enzyme.⁹ Instead of directly making siRNA, making the

shRNA vectors beforehand showed a more stable, efficient, and long-lasting siRNA.

The aim of this study was to establish the clinical significance and functional features of TIPE expression in gastric cancer. Quantitative real-time polymerase chain reaction (qPCR) and immunohistochemistry were used to evaluate the TIPE expression, and the correlation between TIPE expression and cancer progression was evaluated following the guidelines of REMARK.¹⁰ TIPE expression was also knocked down in gastric cancer cell lines (BGC823) by using shRNA, to study the effects of TIPE on cell proliferation and apoptosis.

Materials and methods

Patients and clinical tissue samples

Surgical specimens were obtained from 46 gastric cancer patients (39 males and seven females) who underwent endoscopic biopsy or potentially curative surgery at Zhongshan Hospital, from 2012 to 2014. The patients' age ranged from 43 to 88 (mean 68) years. Tumor stage and disease grade were classified according to the 6th edition of the tumor-node-metastasis (TNM) classification of the International Union against Cancer. The tumor differentiation evaluation was based on the histological criteria, outlined by the Chinese Society for gastric cancer Disease. None of the patients had received irradiation or chemotherapy before surgery. Patients who had undergone noncurative surgery and/or who had received inadequate follow-up were excluded from the study. Tumor tissues, from which mRNA and protein were isolated, were fresh specimens collected from resection surgery. All samples were obtained after receiving written informed consent from all patients, and the study was approved by the Committee on Medical Ethics of Zhongshan Hospital.

Cell lines and plasmids

The gastric cancer cell lines BGC823 were used. The cell lines were maintained in our laboratory. All cell lines were cultured in Dulbecco's Modified Eagle's Medium (Gibco, Thermo Fisher Scientific, Waltham, MA, USA), containing 10% fetal bovine serum and 1% penicillin–streptomycin in a humidified 5% CO₂ incubator at 37°C. TIPE shRNA (Top Strand 5'-gatccGCGTTCAGGCACAAAAGAAGTTCAAGAGACTTCTTTTGTGCTGAACGTTTTTTACGCGTGg-3', Bottom Strand 3'-gCGCAAGTCCGTGTTTTCTTCAA GTTCTCTGAAGAAAACACGGACTTGCAAAAAAATGC GCActaa-5') were synthesized in our laboratory. BGC823 were infected with TIPE-shRNA-pSIREN-RetroQ/PVSVG recombinant lentiviral vectors and pSIREN-RetroQ/PVSVG as the negative control lentiviral vectors. Culture medium was

replaced 24 hours after infection, and after an additional 48 hours the resistant cell clones were selected with puromycin, the expression of TIPE level was verified by Western blot and reverse transcription PCR (RT-PCR).

Protein extraction and Western blot

All cells were harvested at approximately 80% confluence, and the protein was extracted. The protein concentration was determined using the BCA Protein Assay Kit (Pierce, Thermo Fisher Scientific). The protein (30 µg) samples were run on 12% Tris-Tricine Ready Gel (Bio-Rad Laboratories Inc., Hercules, CA, USA) and then electrotransferred to a Hybond-enhanced chemiluminescence membrane (Amersham). The membranes were incubated overnight at 4°C with rabbit polyclonal antibody against TIPE (1:4,500; Abcam plc, Cambridge, UK) and mouse monoclonal antibody against β-actin (1:5,000; Sigma-Aldrich Co., St Louis, MO, USA). The bands were detected by using an enhanced chemiluminescence detection system, where β-actin served as the control. Scanning densitometry was performed using Adobe Photoshop (Apple, Inc, Cupertino, CA, USA).

RNA extraction and RT-PCR

Total RNA was extracted from the tissue using the TRIzol total RNA extraction reagent (CW BIO, Beijing, People's Republic of China). RNA quality and concentration were measured using a NanoDrop 2000 spectrophotometer (Thermo Fisher Scientific). Reverse transcription and PCR were performed using Takara PrimeScript™ RT-PCR Kit (Takara-Dalian, Dalian, People's Republic of China) and 2× EasyTaq® PCR super mix (TransGen, Beijing, People's Republic of China), template cDNA was synthesized from 2.0 µg of total RNA. RT-PCR was performed using the 96-well Biometra PCR System in a total volume of 20 µL at 94°C for 3 minutes followed by 30 cycles of 94°C for 30 seconds, 56°C for 30 seconds, 72°C for 1 minute, and 72°C for 5 minutes. The sequences of the primer pairs were as follows: TIPE forward, 5'-CCCAGGGAAGTGGCTACAGA-3', TIPE reverse, 5'-GCCTCCTTCTTGTCTGGGT-3' and β-actin forward, 5'-AGCCATGTACGTAGCCATCC-3', β-actin reverse, 5'-ACCCTCATAGATGGGCACAG-3'. RT-PCR imaging results, application of gel analysis, and the gray value against the corresponding β-actin were used as the reference.

qRT-PCR

cDNA synthesis and qPCR were performed using ReverTra Ace® qPCR RT Kit and SYBR® Green Realtime PCR Master Mix Plus (TOYOBO, Osaka-shi, Japan), respectively.

The StepOne Real-Time PCR System (ABI, Cambridge, UK) was used to perform the reaction, where β -actin was used to normalize data. The data of the qPCR were calculated using the $2^{-\Delta\Delta C(t)}$ method.

Expression and purification of human anti-DR5ScFv (ZF1)

The obtained bacterial recombination was selected and cultured at 37°C in 3 mL Luria–Bertani broth (LB medium) for 8–12 hours, later it was added to 300 mL of fresh LB medium at 37°C/250 rpm. After 7 hours, when the cell density reached between 0.6–0.8 (OD_{590}) isopropyl- β -D-thiogalactopyranoside was added to LB medium to induce expression. *Escherichia coli* Rosetta-gami™ (EMD Millipore, Billerica, MA, USA) cells from shaker flasks were disintegrated with ultrasonication (300 W, 30 seconds per/30-second intervals for 20 minutes). The suspension was precipitated by centrifugation (12,000 rpm, 20 minutes). The target protein was mainly present in inclusion bodies. The inclusion bodies were washed at 4°C for 7–8 hours in a washing fluid containing 2 M urea. After washing, the inclusion bodies were dissolved in a solution containing 8 M urea. The solution was centrifuged, and the supernatant was transferred to a Ni-nitrilotriacetic acid affinity column (GE Healthcare Life Sciences, Piscataway, NJ, USA) to purify the protein. After loading and eluting, the targeted fractions were pooled and identified by sodium dodecyl sulfate polyacrylamide gel electrophoresis.

Immunohistochemistry

Tumor tissues were fixed and stained following the methods used in Yang et al's work.¹¹ And the concentration of affinity-purified rabbit TIPE antibody (Ab) was 1.67 g/mL.

Apoptosis assay by flow cytometry

To assess apoptosis, cells were plated at a concentration of 5×10^5 cell/L in the six-well plate, incubated for 24 hours, then treated with ZF1 (0.1 mg/mL) for 8 hours. The control groups were incubated in Dulbecco's Modified Eagle's Medium only. After incubation, the cells were harvested with 0.25% trypsin (without ethylenediaminetetraacetic acid), then washed with phosphate-buffered saline and further suspended in a binding buffer (1 \times). An aliquot of 100 μ L cell suspension was incubated with 5 μ L of Annexin V-APC and 5 μ L of propidium iodide (PI) for 15 minutes in the dark, then 1 mL binding buffer (1 \times) was added to each sample. The cells were analyzed directly by a flow cytometer. The percentage of cells was calculated by FlowJo Software.

TUNEL assay

To detect apoptosis in in vivo treatments, we used the TUNEL method. The apoptotic cells are defined by the uptake of PI (red) and FITC (green) (indicating the damage to the cell membrane) and the presence of clear nuclear condensation and/or fragmentation with DNA ends of dUTPs. The paraffin-embedded sections were deparaffinized in xylene and rehydrated in a graded series of ethanol baths. The sections were treated with 20 g/mL of proteinase K in distilled water for 10 minutes at room temperature. The tumor and liver tissues were fixed in 1% paraformaldehyde for 10 minutes. To block endogenous peroxidase, the slides were incubated in methanol containing 0.3% hydrogen peroxide for 20 minutes. The remaining procedures were performed according to the instructions provided by the Roche company. For quantification of apoptosis, five microscopic fields were randomly selected at high power magnification (200 \times), and the average counts of TUNEL-positive cells were calculated.

TIPE knock-down in a nude mice tumor model

We divided the nude mice into four groups, each group consisted of six animals. All four groups received the injection of BGC823 cells at a dose of 1×10^7 cells per mouse; however, two groups received the cells which were treated with shRNA and the other two groups received nontreated cells, as the control. When the tumor nodules started to grow it was classified as day 0. On day 3, each one of the shRNA-treated and the control group received the ZF1 injections, 80 mg/kg every day. The other two groups received saline injections. Tumor length (L), width (W), and diameter were measured every 3 days; tumor volume (mm^3) was calculated using the formula $W^2 \times (L/2)$. The mice were sacrificed on day 14, and the neutral formalin-fixed tumor samples were stained with hematoxylin–eosin (H&E), and TUNEL assay was also conducted as mentioned previously. All experimental procedures were conducted in conformity with institutional guidelines for the care and use of laboratory animals, and protocols were approved by the Animal Studies Committee of Xiamen University, People's Republic of China.

H&E staining analysis

To distinguish antitumor effects of ZF1 in vivo, H&E staining was used. Tumor and liver tissues were harvested (n=6). The H&E stained tissues were used to identify necrotic cells, tumor cells or apoptotic cells. H&E stained sections were viewed under an Olympus BHT microscope (Olympus Corporation, Tokyo, Japan).

Statistical analysis

Data were presented as mean \pm standard deviation. Statistical analysis was performed for multiple comparisons using analysis of variance and Student's *t*-test. A value of $P < 0.05$ was considered to be statistically significant.

Results

Expression of TIPE protein and mRNA in tumor cell lines and clinical gastric tissue specimens

We examined TIPE protein and mRNA expression in human gastric cancer cell lines (MGC803, BGC823), human leukemia cell line (K562), human hepatoma cancer cell line (SMC7721), human colon cancer cell line (SW480), and human gastric cancer clinical specimens with different tumor grades using the RT-PCR method. The expression level of TIPE protein and mRNA was all statistically significant in all these tested samples (Figure 1A). Based on the earlier results, we used the

BGC823 cell line to perform TIPE suppression analysis. We found that the TIPE mRNA was not only expressed in gastric cancer tissues but also in the tested adjacent gastric tissues.

The expression of TIPE was investigated by using RT-PCR in 46 gastric cancer specimens. The results of TIPE mRNA expression in a gastric tumor and its adjacent tissues are shown in Figure 1B. Our results showed that TIPE mRNA expression in gastric cancer tissue was statistically higher ($P < 0.05$) than in adjacent tissues; to further confirm TIPE mRNA expression level, qPCR was used (Figure 1C). The mean expression value of TIPE mRNA level in cancer tissues (1.417 ± 0.342) was significantly higher than the value (0.404 ± 0.091) in normal gastric cancer tissues ($P < 0.05$). According to our results obtained from RT-PCR and qPCR, TIPE mRNA expression levels showed positive correlations to tumor invasion in gastric cancer.

The correlations between TIPE mRNA expression level and the clinicopathological characteristics of gastric cancer

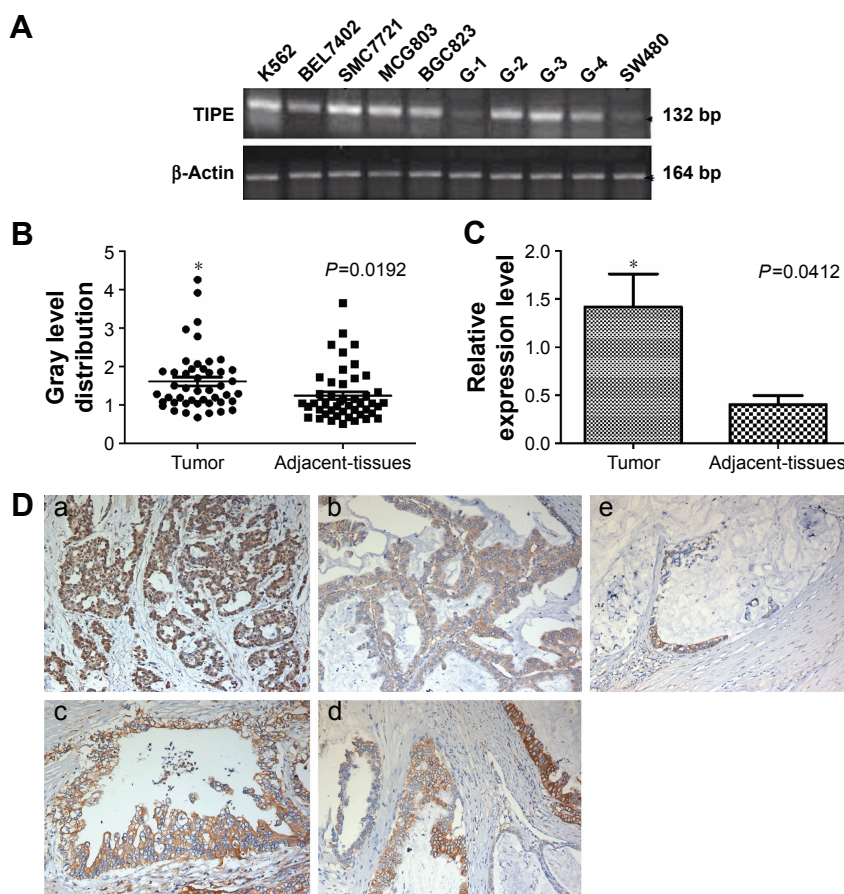


Figure 1 TIPE expression in gastric cells and gastric tissue.

Notes: (A) RT-PCR detected the expression of TIPE in K562, BEL7402, SMC7721, MCG803, BGC823, Gastric cancer I–IV stages and SW480; (B) RT-PCR analysis of TIPE mRNA expression in gastric cancer tissues and the adjacent normal tissues (normalized to β -actin); (C) Real-time PCR assay of TIPE. TIPE mRNA expression in gastric cancer tissues and in adjacent tissues ($n=10$); (D) Immunohistochemical staining of TIPE in gastric cancer tissues and adjacent normal gastric tissues. (a) The expression of TIPE in cytoplasm of gastric cancer cells a poor differentiation adenocarcinoma of gastric cancer tissues. (b) Well differentiation of gastric cancer tissues. (c) TIPE protein expression in the stage III gastric cancer tissues. (d) TIPE protein expression in the stage II gastric cancer tissues. (e) Gastric adjacent tissues. Magnification: $\times 200$. *Represents a significant difference between two groups.

Abbreviation: RT-PCR, reverse transcription polymerase chain reaction.

Table 1 The expression of TIPE in gastric carcinoma and their clinicopathological relationship

Characteristics	Number of patients	TIPE value mean IOD value ($\bar{x} \pm s$)	P-value
Sex			
Male	39	1.513±0.1102	0.0618
Female	7	2.109±0.4171	
Age (years)			
<60	20	1.423±0.1525	0.4585
>60	26	1.564±0.1159	
Differentiation			
Moderate	25	1.456±0.0962	0.0224
Poor	21	1.868±0.1550	
TNM stage			
I + II	22	1.225±0.1464	0.0443
III + IV	24	1.776±0.1218	
Tumor status			
T1–3	18	1.145±0.1169	0.0161
T4	28	1.837±0.1293	
Nodal status			
N0–1	21	1.370±0.1382	0.0221
N2–3	25	1.894±0.1760	
Distant metastasis			
M0	36	1.241±0.1223	0.0451
M1	10	1.711±0.1395	

Abbreviations: TNM, tumor-node-metastasis; IOD, integrated optical density.

patients (age, sex, differentiation, TNM stage, lymph node metastasis, distant metastasis, lymphatic invasion, and venous invasion) are shown in Table 1. There were significant correlations between TIPE expression and TNM stage ($P=0.044$), tumor depth ($P=0.016$), lymph node metastasis ($P=0.026$), and distant metastasis ($P=0.045$). No significant correlations were found between TIPE expression and age ($P=0.459$) and sex ($P=0.062$). In univariate analysis, high TIPE expression was found to be a significant prognostic factor for poor survival, in addition to tumor depth and differentiation, the presence of lymph node metastasis and distant metastasis.

Overexpressed TIPE was detected in 46 gastric cancer samples, whereas negative or weak TIPE immunoreactivity was observed in adjacent tissues. TIPE was clearly localized in the cytoplasmic compartment of the tumor cells (Figure 1D). Finally, we found that the expression of TIPE in poorly differentiated gastric adenocarcinoma was higher compared with that in well-differentiated ones, a similar difference was found between adenocarcinoma stages II and III.

Depletion of TIPE expression inhibits tumor cell growth in gastric cancer cell lines

Higher expression levels of TIPE contribute to the proliferation of cancer cells. We therefore hypothesized that decreased expression of TIPE might lead to cell death.

Firstly, to test our hypothesis, we transfected BGC823 cells with a TIPE knock-down plasmid and maintained BGC823 stable transfectants (TIPE shRNA and vector) in puromycin-containing medium. Then, we confirmed the expression of endogenous TIPE (21 kDa) by using Western blotting and RT-PCR (132 bp). Western blot and RT-PCR analysis confirmed that TIPE shRNA significantly reduced the expression of TIPE (Figure 2A and B).

Apoptosis was examined by using flow cytometry after staining the cells with annexin-V and PI. As a result, BGC823 cells were treated with ZF1, 19.40% of cells underwent apoptosis, 17.98% were found in negative control cells; and 31.68% in the ZF1 treated shRNA-BGC823 cells (Figure 2C). Results showed ZF1 was able to induce more apoptosis in the TIPE treated shRNA group.

Analysis of the apoptosis pathway associated protein expression level revealed that the levels of activated caspase-8/3/9 were different in each group (Figure 2D). In comparison, the activated caspase-8/3/9 levels were similar in both negative control groups, which were all treated with ZF1, whereas activated caspase-8/3/9 levels were elevated in the TIPE shRNA treated BGC823 cells. These results, gathered from TIPE shRNA treated BGC823 cells, confer the susceptibility to ZF1 induced apoptosis in the absence of TIPE. These results indicated that TIPE might be involved with negative regulation of cell death. This inhibition could possibly be associated with caspase activity.

Knock-down TIPE impairs gastric cancer formation of BGC823 cells in vivo and enhanced anti-tumor effect of anti-DR5ScFv in tumor-bearing athymic mice

To verify the correlation of TIPE expression with experimental tumor growth and drug resistance, a xenograft tumor model was established by injecting nude mice with BGC823 cells and TIPE shRNA treated BGC823 cells. We examined the effects of inhibition of endogenous TIPE expression on the xenograft tumor growth of BGC823 cells and the susceptibility to ZF1.

Measurement of tumor size showed that at 14 days, tumors induced by TIPE shRNA treated BGC823 cells (0.019 ± 0.004 g, 232.586 ± 31.632 mm³) were significantly smaller than tumors from the control group (0.026 ± 0.003 g, 252.825 ± 34.883 mm³, $P < 0.05$). Tumors of TIPE shRNA transfected BGC823 cells treated with ZF1 (0.008 ± 0.002 g, 94.987 ± 24.287 mm³) were smaller than tumors treated with saline (0.016 ± 0.003 g, 178.387 ± 8.083 mm³, $P < 0.05$) (Figure 3A and B).

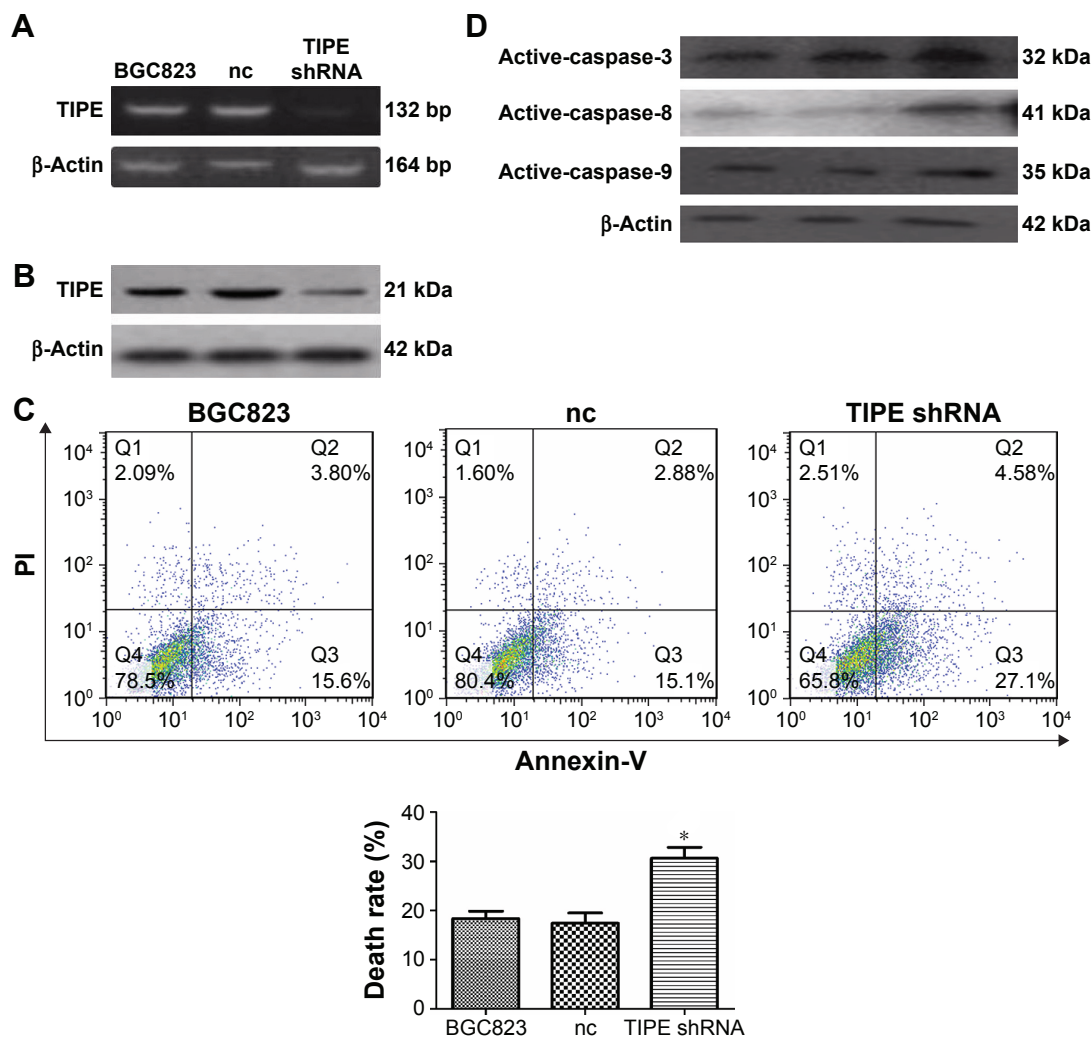


Figure 2 Suppression analysis of TIPE in the BGC823 cell line in vitro.

Notes: (A), (B) TIPE protein and mRNA levels expression in BGC823 cells that had been treated with shRNA were detected by Western blotting and RT-PCR. TIPE expression was suppressed by shRNA. (C) The rate of apoptosis for BGC823 treated with the Anti-DR5 ScFv was analyzed using flow cytometry. The apoptosis induced in the TIPE shRNA cells was enhanced compared with that induced in the negative control shRNA and the BGC823 cells. This image represents one of three sets of flow cytometry results. (D) The expression of the proteins involved in the apoptosis pathway of activated caspase-3/8/9. * $P < 0.05$.

Abbreviations: nc, negative control; PI, propidium iodide.

There were no statistical differences in weight of mice between the ZF1 and saline groups ($P > 0.05$). Furthermore, all the mice were observed to be in a healthy condition and remained active throughout the study, suggesting that ZF1 administration had no obvious physiological or psychological side effects. H&E staining revealed that TIPE shRNA promotes more tumor cell death in vivo than tumor tissue from nude mice injected with ZF1, as well as disappearance of intracellular structures and cell sputtering (Figure 3C).

In contrast, tumor tissue from control nude mice injected with saline showed large and irregular cell shapes as expected. Results obtained from TUNEL stain showed apoptosis positive cells were more abundant in tumor tissue from TIPE shRNA mice than in tissue from the two control groups (Figure 3D and 3E).

Discussion

TIPE was originally identified in a primary human head and neck squamous cell carcinoma cell line; with its metastatic cell line established from cancer recurrence after radiation therapy.¹² Overexpression of TIPE in cancer cells enhances cell survival and proliferation in various cells.^{2,13,14} High expression of TIPE in cancer cells is correlated with enhanced cell survival and inhibition of apoptosis, including inhibition of caspase-8 and caspase-3.^{2,15}

This is the first report on the role of TIPE in gastric cancer. TIPE overexpression was significantly associated with high histologic grade, large residual tumor size, recurrence, and poor response to chemotherapy.¹⁶ Our data demonstrated that expression of TIPE protein was much higher in several human cancer cells; MGC803, BGC823, K562,

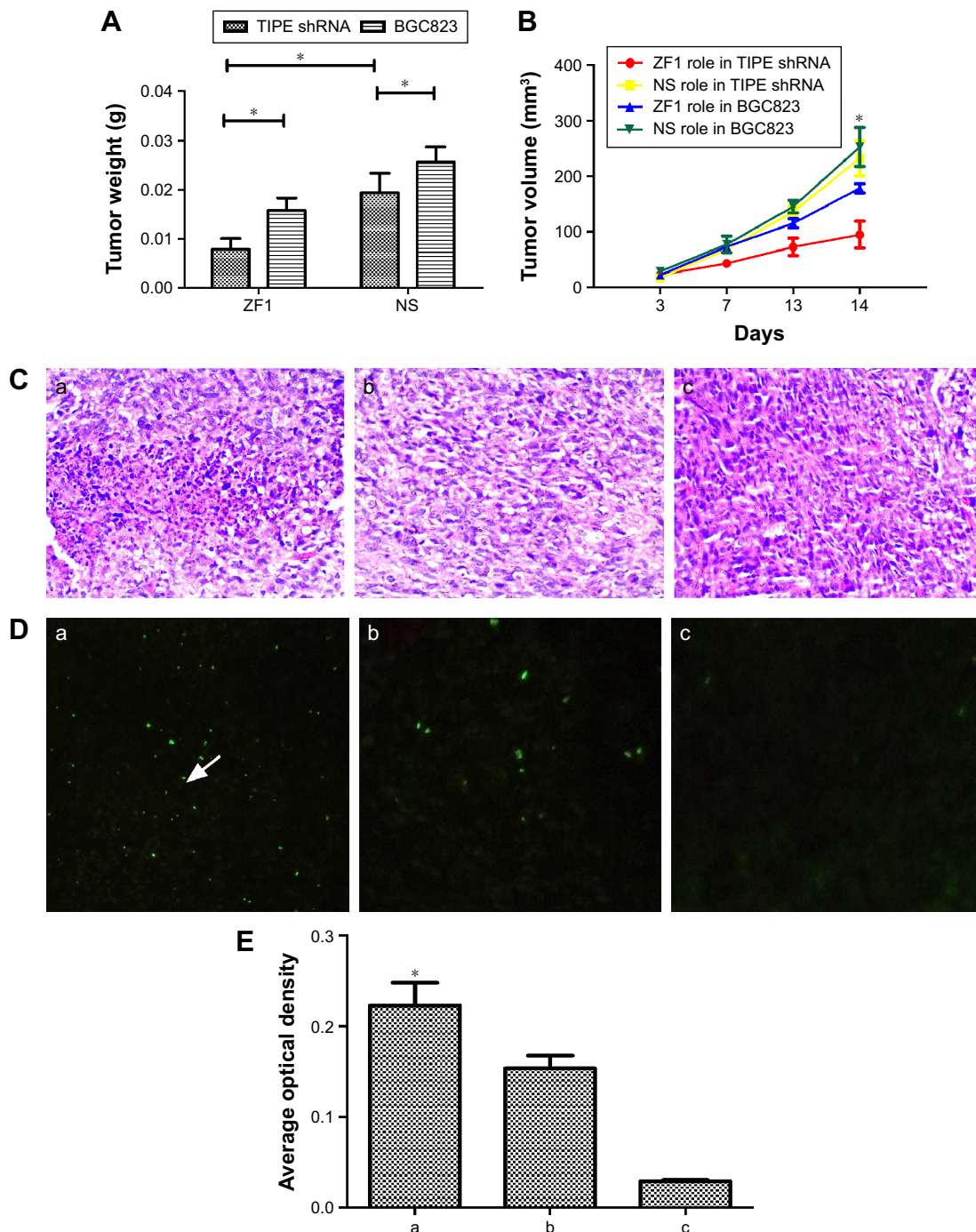


Figure 3 Downregulated TIPE expression inhibits the growth of BGC823 xenografts tumor in athymic mice.

Notes: Comparison of tumor weight and tumor volume in BGC823 tumor-bearing mice of each group. **(A)** Weight of mice tumor after 14 days treatment, $*P < 0.05$. **(B)** Volume of mice tumor treated per day, $*P < 0.05$. **(C)** Hematoxylin–eosin staining, tumor tissue from nude mice injected with Anti-DR5 ScFv showed sheet necrosis, disappearance of intracellular structures, and cell sputtering; (a) TIPE shRNA tumor treated with Anti-DR5 ScFv, (b) BGC823 tumor treated with Anti-DR5 ScFv (c) BGC823 tumor treated with normal saline (NS) (400 \times). **(D)** TUNEL staining; (a) TIPE shRNA tumor treated with Anti-DR5 ScFv; the arrow indicates the apoptosis cell; (b) BGC823 tumor treated with Anti-DR5 ScFv, (c) BGC823 tumor treated with normal saline (200 \times). **(E)** Average optical density of panels in **(D)**. *Represents a significant difference between groups a, b, and c.

SMC7721, and SW480. These results were in line with previous published data; TIPE was discovered to have higher expression levels in various human cancer cell lines, such as K562, MOLT4, A549, and SKOV-3, but lower levels in SW480.¹⁵ Moreover, TIPE expression was found to be higher in gastric carcinomas compared to the normal adjacent tissues

examined. In this study, results showed significant correlations between TIPE expression and TNM stage ($P=0.044$), tumor depth ($P=0.016$), lymph node metastasis ($P=0.026$), and distant metastasis ($P=0.045$). No significant correlations were found between TIPE expression and age ($P=0.062$) or sex ($P=0.459$). Its expression pattern is similar to human

breast carcinomas, non-small cell lung cancer, and renal cell carcinomas.^{4,6,17}

In the absence of TIPE expression in BGC823 cells, via shRNA knock-down, there was increased cell death in BGC823 cells. Our findings of decreased TIPE expression and increased cell death are consistent with the role of TIPE in cell survival and metastasis.^{13,15}

Anti-DR5mAb could induce cell apoptosis by binding to DR5. He et al's, group showed that apoptosis plays a key role in antitumor activities in colon cancer cells and xenografts through the DR5, FADD, and caspase-8 axis.¹⁸ Overexpression of DR5 and DR4 receptors can induce tumor cell apoptosis and reverse the drug resistance of tumors to TRAIL.¹⁹ Activated DR5 recruits the adaptor protein FADD, then cleaves procaspase-8 and initiates caspase-8 activation, which activates the downstream effector caspase-3 that leads to apoptosis.^{20,21}

Our laboratory successfully prepared and purified a high purity and stable Anti-DR5ScFv, which could specifically bind to eDR5 and induce cell apoptosis.¹¹ TIPE could suppress TNF- α -mediated apoptosis by inhibiting caspase-3 and caspase-8 activity.^{2,17} This inhibition might lead to its resistance to chemotherapy and antibody treatments, thereby halting tumor progression.

Conclusion

In this study, we demonstrated that after the administration of Anti-DR5ScFv, apoptosis was induced to a greater degree in the TIPE shrank group than in the control group, both in vitro and in vivo. In addition, the activity of caspase-8, caspase-3, and caspase-9 was successfully induced, and more apoptotic cells were found in the tumor in the absence of TIPE. Therefore, TIPE might play a critical role in interfering with anticancer drug resistance. In the absence of TIPE expression in BGC823, induced cell apoptosis was significantly elevated, showing a direct link between TIPE expression and tumor cell growth inhibition. These findings indicated that TIPE would be a useful tumor therapy target in the close future.

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Disclosure

The authors report no conflicts of interest in this work.

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