

Anti-inflammatory and immunomodulatory effects of *Aquaphilus dolomiae* extract on in vitro models

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Background: Atopic dermatitis (AD) is a common skin disease characterized by recurrent pruritic inflammatory skin lesions resulting from structural and immune defects of the skin barrier. Previous studies have shown the clinical efficacy of Avène thermal spring water in AD, and a new microorganism, *Aquaphilus dolomiae* was suspected to contribute to these unique properties. The present study evaluated the anti-inflammatory, antipruritic, and immunomodulatory properties of ES0, an original biological extract of *A. dolomiae*, in immune and inflammatory cell models in order to assess its potential use in the treatment of AD.

Materials and methods: An ES0 extract containing periplasmic and membrane proteins, peptides, lipopolysaccharides, and exopolysaccharides was obtained from *A. dolomiae*. The effects of the extract on pruritus and inflammatory mediators and immune mechanisms were evaluated by using various AD cell models and assays.

Results: In a keratinocyte model, ES0 inhibited the expression of the inflammatory mediators, thymic stromal lymphopoietin, interleukin (IL)-18, IL-4R, IL-8, monocyte chemoattractant protein-3, macrophage inflammatory protein-3 α , and macrophage-derived chemokine and induced the expression of involucrin, which is involved in skin barrier keratinocyte terminal differentiation. In addition, ES0 inhibited protease-activated receptor-2 activation in HaCaT human keratinocytes stimulated by stratum corneum tryptic enzyme and T helper type (Th) 1, Th2, and Th17 cytokine production in Staphylococcal enterotoxin B-stimulated CD4+ lymphocytes. Lastly, ES0 markedly activated innate immunity through toll-like receptor (TLR) 2, TLR4, and TLR5 activation (in recombinant human embryonic kidney 293 cells) and through antimicrobial peptide induction (psoriasin, human beta-defensin-2, and cathelicidin), mainly through TLR5 activation (in normal human keratinocytes).

Conclusion: Overall, these in vitro results confirm the marked regulatory activity of this *A. dolomiae* extract on inflammatory and immune responses, which may be of value by virtue of its potential as an adjunctive treatment of AD inflammatory and pruritic lesions.

Keywords: atopic dermatitis, inflammation, anti-inflammation, in vitro and in vivo activities, barrier function, microorganism

Introduction

Atopic dermatitis (AD) is a common chronic skin disease characterized by recurrent pruritic inflammatory skin lesions resulting from a compromised skin barrier caused by genetically and environmentally determined structural defects and immune dysregulation.¹⁻³ It was found that serine proteases can alter skin barrier function and contribute to inflammation and pruritus, primarily by activation of protease-activated receptor-2 (PAR-2) and intracellular calcium mobilization.⁴ Skin barrier impairment leads to greater susceptibility to cutaneous infections, particularly by *Staphylococcus aureus*,

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and increased allergen penetration. This, in turn, contributes to cutaneous inflammation, mediated by the expression of various proinflammatory factors including cytokines (thymic stromal lymphopoietin [TSLP] interleukin [IL]-18) and chemokines (IL-8, regulated upon activation, normal T cell expressed and secreted [RANTES], monocyte chemoattractant protein 3 [MCP3], and macrophage inflammatory protein [MIP]-3 α) and by the activation of keratinocytes, dendritic cells, and specific T-lymphocytes directed against exogenous or autoantigens.^{5,6}

From various studies, it was found that keratinocytes express toll-like receptors (TLRs), which are a family of evolutionary conserved pattern recognition receptors (PRRs) involved in innate immunity.^{7,8} Each TLR senses specific microbial ligands and thus activates the innate immune system. Impairment of PRRs and an insufficient quantity of antimicrobial peptides (AMPs) to combat *S. aureus* are observed in patients with AD.⁹ In particular, TLR2, which recognizes *S. aureus*, is shown to be less expressed or altered in monocytes and macrophages of patients with AD.¹⁰⁻¹² In addition, T helper type (Th) 1 and Th2 infiltration is enhanced in AD lesions.¹³ Indeed, although IL-17-producing cells (Th17) are more abundant in peripheral blood and acute AD lesions, Th2 cytokines inhibit Th17 response, leading to lower AMP expression by keratinocytes.^{3,14} In addition, Th2 cytokines also alter the barrier function by inhibiting the expression of keratinocyte terminal differentiation markers, such as involucrin.¹⁵

In recent years, several studies have shown the clinical efficacy of Avène thermal spring water in AD, in particular immunomodulation of Th1-dependent and Th2-dependent cytokine production.¹⁶⁻²¹ In vitro experiments have also demonstrated multiple effects of Avène thermal spring water on numerous mediators involved in the immune response and on membrane fluidity, in addition to antiradical and anti-inflammatory properties, and stimulatory effects on keratinocyte differentiation.²² An investigation of the deep aquifer of the Avène thermal spring water pointed out a new microorganism as a potential source of these unique properties due to its ability to modulate TLRs (data not shown). Based on its distinctive phenotypic and genotypic characteristics, this newly identified strain was assigned to a new genus, as a representative of a novel species called *Aquaphilus dolomiae*. It is a chemoorganotrophic non-spore-forming bacterium of the β -Proteobacteria class.²³

In the present study, the activity of ES0, an original biological extract of *A. dolomiae*, on various AD-related mechanisms, including its effects on pruritus and inflammatory mediators, and on immune mechanisms in different immune and inflammatory cell models was evaluated.²⁴

Materials and methods

Materials

ES0 extract

ES0 extract (I-modulia™, Pierre Fabre, Toulouse, France) was obtained from *A. dolomiae*, a novel nonpathogenic, chemoorganotrophic, non-spore-forming bacterium with a cell size of 2.3 \pm 0.3 μ m long and 1.0 \pm 0.1 μ m wide. *A. dolomiae* was isolated from aquifer water in the Cévennes mountains (France), as described previously in the study by Bourrain et al.²³ Fermentation and purification were performed at Centre d'Immunologie Pierre Fabre (Saint-Julien-en-Genevois, France) in accordance with pharmaceutical Good Manufacturing Practice as follows: *A. dolomiae* bacteria were cultivated in complex medium (ie, citric acid, magnesium sulfate, ammonium chloride, dipotassium phosphate, pyruvic acid sodium salt, yeast extract, glucose, and traces of OligoMix®) under aerobic conditions at 27°C and pH 7.0. The culture was carried out in three successive steps from a 1-L Erlenmeyer flask to a 350-L stirred fermenter. High-cell-density fermentation was carried out in fed-batch mode with a sterile glucose solution. Cell growth was monitored by using optical density (OD) measurements at 620 nm and dry cell weight determination. At the end of the fed-batch fermentation, bacterial cells were processed in an aqueous basic Tris buffer (pH 11) containing Arginine at 300 mM. After incubation under stirring for 5 h at a temperature of 4°C, the cellular debris was removed by centrifugation on a disk separator followed by depth filtration. The clarified extract was filtered on a 0.2 μ m polyethersulfone filter. This extract contains periplasmic and membrane proteins, peptides, lipopolysaccharides, and exopolysaccharides.

Methods

Evaluation of ES0 activity on an AD keratinocyte model

Normal human keratinocytes (NHKs) were prepared from skin flaps originating from plastic surgery (breast reduction, after obtaining the patients' informed consent according to the principles of the Declaration of Helsinki). French regulations (authorization is given by the French Ministry of Research [Decree n° 2007-1220]) do not require ethical committee approval for the sampling of organ tissues for use in research studies. The cells were cultured in keratinocyte serum-free medium (KSFM; Gibco®, Life Technologies, Villebon-sur-Yvette, France) with low calcium content (0.1 mM) supplemented by 25 μ g/mL bovine pituitary extract (Gibco®) and 1.5 ng/mL epidermal growth factor (Gibco®).

Polymerase chain reaction (PCR) array analysis was conducted as previously described in a model of AD

induction in NHK.^{25,26} Briefly, NHK were stimulated for 24 h with 10 µg/mL polyinosinic/polycytidylic acid (poly I:C) and 3 ng/mL of each cytokine, tumor necrosis factor alpha (TNFα), IL-4, and IL-13 (R&D Systems, Lille, France). In order to characterize the pattern of mRNA expression in the present model of stimulated NHK, the expression of 23 mRNA, including 21 mRNA encoding various factors known to be differentially regulated in AD, such as for inflammation and keratinocyte differentiation mediators, and two housekeeping genes (*RPL13A* and *GAPDH*) were studied.²⁷ The relative quantity of mRNA expression after stimulation with poly I:C and cytokines was calculated compared with that obtained from unstimulated NHK using the formula $2^{-\Delta\Delta Ct}$ were $\Delta Ct = Ct_{\text{gene of interest}} - Ct_{\text{housekeeping gene}}$ and $\Delta\Delta Ct = \Delta Ct_{\text{treated}} - \Delta Ct_{\text{control}}$. In order to evaluate the effects of ES0, PCR analysis was conducted after a 24-h incubation of the AD cellular model with ES0 10, 30, or 60 µg/mL or with the glucocorticoid dexamethasone (DEX; Sigma-Aldrich, Lyon, France) of 2 µM (positive control), using SYBR Green probes specific to the chosen mRNA (Table S1). The relative quantity of mRNA expression after incubation of the AD model with ES0 was calculated, compared with the AD cellular model. It was also found, using a colorimetric MTT (bromure de 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium) assay, that ES0 extract was not cytotoxic at the concentrations studied.²⁸ Statistical analyses were conducted by using unpaired *t*-test on ΔCt results, in order to compare stimulated cells with unstimulated cells or treated cells with stimulated cells.

Evaluation of ES0 activity on PAR-2 assay

The spontaneously immortalized HaCaT human keratinocyte cell line was cultivated in Dulbecco's Modified Eagle's Medium (DMEM; Gibco®) with 10% fetal calf serum (FCS; Gibco®) as previously described by Boukamp et al.²⁹ PAR-2 was first activated in HaCaT keratinocytes by stratum corneum tryptic enzyme (SCTE; R&D Systems), leading to intracellular calcium flux. The inhibitory activity of ES0 on PAR-2 was assessed by measuring the intracellular calcium flux before and after the addition of ES0. HaCaT cells were incubated for 30 min with a fluorescent probe esterified by an acetoxymethyl group (Fluo-4/AM; Molecular Probes®, ThermoFisher Scientific, Villebon Sur Yvette, France; 2 µM) in a 96-well plate and then for 30 min alone or in the presence of ES0 (30 and 100 µg/mL), before the injection of SCTE 50 nM. Intracellular calcium flux was assessed for 175 s by measuring the fluorescence emitted at 535 nm by the deesterified Ca²⁺-bound form of the probe, which is the only form excitable at 485 nm. The microplate was read by Mithras LB940™ (Berthold Technologies, Thoiry, France).

Evaluation of ES0 activity on TLR assay

Human embryonic kidney (HEK; InvivoGen, Toulouse, France) 293 cells were cultivated in 4.5 g/L D-glucose + pyruvate supplemented with DMEM with 10% FCS, in the presence of Normocin™. HEK 293 cells were used as they functionally overexpress human TLR2, TLR4, or TLR5 proteins as well as a reporter gene, which is a secreted alkaline phosphatase (SEAP) expressed under the control of a nuclear factor-kappa B (NF-κB)-inducible promoter. Pam2CSK4 (TLR2 ligand) at 0.03, 0.1, 0.3, and 1 ng/mL; lipopolysaccharide (LPS) 0111 (TLR4 ligand) at 0.3, 1, 3, and 10 ng/mL; and flagellin (TLR5 ligand) at 1, 3, 10, and 30 ng/mL were used as positive controls to induce HEK 293-hTLR2, HEK 293-hTLR4, and HEK 293-hTLR5 cell lines (InvivoGen, Toulouse, France), respectively. The negative control constituted a recombinant HEK 293 cell line for the SEAP reporter gene under the control of a NF-κB-inducible promoter only.

ES0 was diluted at 3, 10, 30, and 100 ng/mL in limulus amoebocyte lysate water (Bio-Whittaker, Lonza, Levallois Perret, France), and 20 µL of each sample and the respective control were used to stimulate HEK 293 TLR2-, TLR4-, and TLR5-expressing cell lines for 20 h in 200 µL of final reaction volume. Activation of TLR2, TLR4, and TLR5 was measured by spectrophotometry, and the results were given as OD and percentage activity compared with nontreated cells for the highest ES0 concentration. All the experiments were performed in triplicate; the results were expressed as mean ± standard deviation (SD) for each treatment.

Evaluation of ES0 on AMPs expression

NHK were incubated for 24 h alone (negative control) or with IL-1β (R&D Systems) 20 ng/mL (positive control), flagellin 100 ng/mL (TLR5 ligand from InvivoGen), or ES0 6.5 µg/mL. Blocking anti-TLR2, anti-TLR4, and anti-TLR5 antibodies from InvivoGen (1 µg/mL) were preincubated for 1 h before the addition of the test products. Cells were harvested, and the QuantiGene Plex 2.0 system (Ozyme, Saint Quentin en Yvelines, France) was used to quantify the mRNA expression of AMPs (human beta-defensin-2 [hBD-2], CAMP, psoriasin) directly from cell lysates. The assay was performed in accordance with the manufacturer's instructions. Data were exported to Microsoft Excel (Microsoft Corporation, Redmond, WA, USA), where background subtraction and normalization to two housekeepers (hypoxanthine phosphoribosyltransferase and polymerase [RNA] II polypeptide A) were performed. An effect of the test product on AMP expression was considered when the relative quantity of mRNA (calculated compared with the negative control) was ≥2 (induction) or ≤0.5 (inhibition).

Evaluation of ES0 activity on adaptive immunity

Peripheral blood mononuclear cells were obtained from three donors (French Blood Service, Pyrénées-Méditerranée, France). This was obtained under agreement number with French Blood Service 21/PLER/TOU/PF01/2013-0023. Written informed consent was also obtained. The cells were isolated from buffy-coat blood samples. Human CD4+ lymphocytes were purified by positive magnetic immunoselection (Miltenyi Biotec SAS, Paris, France). Lymphocytes were cultured in Roswell Park Memorial Institute 1640 medium with GlutaMax™ I and 5% FCS at 37°C in 5% CO₂ atmosphere. After incubation of CD4+ cells with Staphylococcal enterotoxin B (SEB) of 300 ng/mL and ES0 of 30, 60, and 100 µg/mL (n=3) for 20 h, culture supernatants were collected, centrifuged (3000 rpm, 15 min, 4°C), and frozen at -20°C before proceeding to Th1 (IL-2, IFN-γ, IL-12, IL-1β, and TNFα), Th2 (IL-4 and IL-10), and Th17 (IL-17) cytokine immunoassay by the Luminex™ method using the Bio-Plex Pro® Human Cytokine group 8-PLEX multiplex, in accordance with the supplier's instructions (Bio-Rad, Hercules, CA, USA). For each parameter, the results were expressed as mean ± SD.

Statistical analysis

Statistical evaluation of between-group comparisons was performed by one-way analysis of variance and Dunnett's test using Graph Pad Prism. For all the analyses, a difference with $P < 0.05$ was considered significant.

Results

Poly I:C-, TNFα-, IL-4-, and IL-13-stimulated NHK: an AD keratinocyte model

PCR array analysis of the expression of 21 mRNA, known to be differentially regulated in AD, after 24 h stimulation with poly I:C, TNFα, IL-4, and IL-13 demonstrated similarities with the regulation of expression observed in AD in this *in vitro* model. Indeed, a markedly increased expression of eight chemokines, five cytokines, and one cytokine receptor normally upregulated in AD and a markedly lower expression of three downregulated keratinocyte differentiation markers were observed. In particular, the two cytokines, IL-18 and TSLP, were increased by twofold and 12-fold, respectively, and the chemokines, MCP3/CC chemokine ligand (CCL) 7, macrophage-derived chemokine (MDC)/CCL22, MIP-3α/CCL20, and IL-8 by eightfold and more than 30-, 200-, and 700-fold, respectively (Table 1).

Anti-inflammatory effects of ES0 in stimulated NHK

The effect of ES0 on the regulation of mRNA involved in inflammation was assessed by comparing the effect of incubating stimulated NHK with ES0 or the positive control, DEX. First, it was confirmed that there was no cytotoxicity of the ES0 extract at the concentrations studied with MTT assay analysis (Table 2). Compared with control, NHK were completely viable (104%–105%) in the presence of ES0 1.1–100 µg/mL; however, cell viability was reduced to 16% with ES0 300 µg/mL and was lost (2%) at ES0 900 µg/mL.

Compared with DEX, which regulated nine mediators, ES0 regulated 14 AD-related targets with a dose–effect relationship. Among the 21 mRNA considered, three were upregulated by ES0 and 11 were downregulated (Table 1). In particular, ES0 induced the involucrin keratinocyte differentiation marker by up to 11-fold at a concentration of 60 µg/mL versus 1.1-fold for DEX of 2 µM; ES0 also induced RNASE7 and psoriasin AMPs by up to fivefold and 21-fold, respectively, versus 0.30- and 0.46-fold for DEX of 2 µM (Table 1). On the other hand, the concentration of 60 µg/mL of ES0 reduced the expression of the two inflammatory cytokines TSLP and IL-18 by 0.08-fold and 0.35-fold and that of the four chemokines MCP3/CCL7, MDC/CCL22, MIP-3α/CCL20, and IL-8 by 0.12-, 0.42-, 0.20-, and 0.12-fold compared with the stimulated NHK model, respectively (Table 1).

PAR-2 inhibition by ES0

Another cell model was used to evaluate the inhibitory effect of ES0 on the pruritus mediator, PAR-2. First, PAR-2 was activated in HaCaT keratinocyte cells by SCTE, leading to intracellular calcium flux. Then, the inhibition of PAR-2 by ES0 was assessed by measuring the intracellular calcium flux before and after the addition of ES0. ES0 dose-dependently inhibited SCTE-activated PAR-2 receptors (Figure 1). A significant effect was observed from a concentration of 30 µg/mL ($P \leq 0.05$ vs SCTE only), and inhibition was greater at 100 µg/mL, with a 50% decrease in SCTE-induced stimulation ($P \leq 0.001$).

Immunomodulatory activity of ES0

Induction of TLR activation

Evaluation of the immunomodulatory effect of ES0 on TLR activity was performed by TLR assay in stably transfected HEK 293 cell lines that overexpressed human TLR2, TLR4, or TLR5 proteins. Pam2CSK4 (TLR2 ligand), LPS 0111 (TLR4 ligand), and flagellin (TLR5 ligand) were used as positive controls to induce the hTLR2, hTLR4, and hTLR5

Table 1 PCR array analysis of the expression of 21 genes

Biological markers	Regulation of gene expression in 24-h-activated NHK	Effect of ES0 on gene expression of 24-h-activated NHK			Effect of DEX on gene expression of 24-h-activated NHK
		10 µg/mL	30 µg/mL	60 µg/mL	2 µM
Thymic stromal lymphopoietin (TSLP)	12.6±1.4 ***	0.44±0.04***	0.25±0.02***	0.08±0.02***	0.08±0.02***
Interleukin 1, alpha (IL1A)	35±1.4***	0.65±0.02**	0.54±0.04**	0.44±0.09**	0.42±0.08***
Interleukin 18 (IL18)	2±0.1 ***	0.73±0.06*	0.57±0.03**	0.37±0.09**	0.51±0.1**
Interferon, alpha 2 (IFNA2)	2.5±0.4***	1.1±0.05	1.16±0.31	1.1±0.3	0.57±0.21*
Interferon, beta 1 (IFNB1)	24.3±1.5***	0.35±0.08***	0.18±0.01***	0.1±0.02***	0.46±0.09**
Interleukin 8 (IL8)	732±155***	0.65±0.17*	0.47±0.08**	0.13±0.08***	0.24±0.08***
MIP-1α (CCL3)	5759±2977***	0.9±0.06	0.57±0.04**	0.27±0.06***	0.21±0.04***
RANTES (CCL5)	15591±1441***	0.85±0.03	0.57±0.04**	0.37±0.07**	0.8±0.15
MCP-3 (CCL7)	7.8±2.4***	0.61±0.18*	0.40±0.13**	0.14±0.07***	1.11±0.22
TARC (CCL17)	21.5±6.2***	0.44±0.08**	0.37±0.03**	0.67±0.17	1.11±0.21
MIP-3α CCL20	212±17***	0.6±0.04**	0.4±0.03***	0.21±0.04***	0.54±0.11*
MDC (CCL22)	33.1±4.2***	0.84±0.03	0.56±0.04**	0.45±0.09**	0.5±0.09**
Skinkine (CCL27)	57.8±0.7***	0.72±0.03*	0.68±0.05*	0.65±0.13	0.37±0.07***
Interleukin 4 receptor (IL4R)	6.3±0.6***	0.7±0.03*	0.56±0.04**	0.32±0.07**	0.23±0.05***
Cathelicidin antimicrobial peptide (CAMP)	5.7±0.8***	1.1±0.1	1.3±0.2	1.8±0.4*	0.42±0.08**
Ribonuclease, RNase A family, 7 (RNASE7)	7.7±0.1 ***	2.4±0.1***	3.5±0.2***	5.3±1.1***	0.27±0.05***
S100 calcium binding protein A11 (S100A11)	3±0.1 ***	0.88±0.03	0.88±0.06	0.84±0.18	0.6±0.11*
Psoriasis (S100A7)	16.6±0.2***	4.6±0.2***	11.4±1***	22.5±5.1***	0.42±0.08**
Ceramide synthase 6 (CERS6)	0.26±0.0***	0.94±0.04	0.95±0.16	0.93±0.23	1.3±0.2
Involucrin (IVL)	0.18±0.0***	1.6±0.1**	4.2±0.34***	11.8±2.4***	0.98±0.18
Filaggrin (FLG)	0.24±0.01***	1.02±0.04	0.83±0.05	0.74±0.15	1.47±0.27**

Notes: NHKs were incubated with 10 µg/mL poly I:C + 3 ng/mL of each cytokine (TNFα + IL-4 + IL-13). NHKs were exposed to three different doses of ES0 (10, 30, and 60 µg/mL) for 24 h compared with DEX of 2 µM. Relative quantity ± SE of mRNA expression compared to control cells (for 24 h-activated NHK) or to 2 h-activated NHKs (for the effect of ES0 or DEX). Data presented as mean ± standard deviation. *P<0.05; **P<0.01; ***P<0.001.

Abbreviations: DEX, dexamethasone; ES0, extract from *Aquaphilus dolomiae*; IL, interleukin; NHK, normal human keratinocyte; PCR, polymerase chain reaction; SE, standard error; TNFα, tumor necrosis factor alpha.

Table 2 Effect of ES0 on normal human keratinocyte cell viability: MTT assay

	Treatment (24 h; µg/mL)	Cell viability (percentage of control ± SE)
Control		100±2
ES0	1.1	104±3
	3.3	105±4
	10	106±4
	30	107±4
	60	105±5
	100	105±4
	300	16±1
	900	2±0

Abbreviations: ES0, extract from *Aquaphilus dolomiae*; SE, standard error.

cell lines, respectively. Following the incubation of the three HEK 293 TLR2-, TLR4-, and TLR5-expressing cell lines with various dilutions of ES0, measurement of TLR2, TLR4, and TLR5 activation by spectrophotometry showed

that ES0 statistically significantly and dose-dependently (10–100 ng/mL) activated TLR2 ($P<0.001$ vs control; +351% [OD =2.45] at 100 ng/mL) and TLR5 (at 3, 10, and 100 ng/mL; $P<0.001$ vs control; +317% [OD =1.04] at 100 ng/mL; Figure 2). ES0 also significantly ($P<0.001$ vs control) activated TLR4 at 3 and 10 ng/mL (+75% [OD =0.8]; Figure 2).

Induction of antimicrobial peptides expression

Compared with 24 h incubation of NHK alone (negative control), incubation with IL-1β 20 ng/mL (positive control) significantly induced the expression of the AMPs, hBD-2 ($P<0.05$), cathelicidin CAMP (LL-37; $P<0.01$), and psoriasis ($P<0.001$; Figure 3). Similarly, incubation with ES0 of 6.5 mg/mL significantly induced the expression of hBD-2 ($P<0.05$), CAMP ($P<0.01$), and psoriasis ($P<0.001$), whereas flagellin of 100 ng/mL (TLR5 ligand)

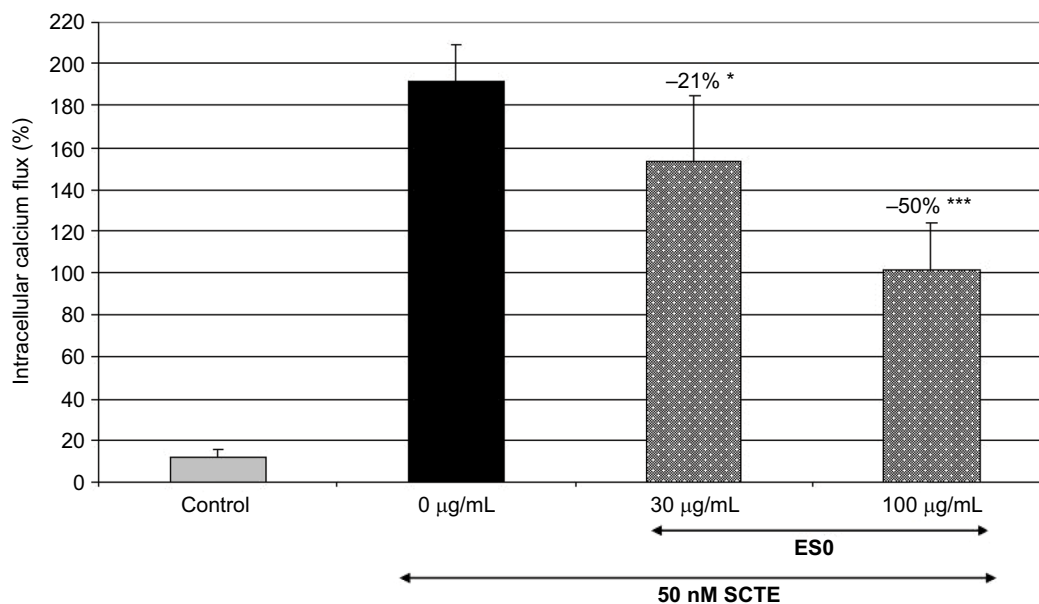


Figure 1 Stimulation of PAR-2 receptors by SCTE in HaCaT keratinocyte cells.

Notes: Dose-dependent inhibition by ES0 (30 and 100 µg/mL). * $P < 0.05$; *** $P < 0.001$ versus SCTE; data are representative of three independent experiments.

Abbreviations: ES0, extract from *A. dolomia*; PAR-2, protease-activated receptor-2; SCTE, stratum corneum tryptic enzyme.

markedly (hBD-2) or significantly (CAMP and psoriasin) induced these AMPs.

In addition, in order to test whether activation was mediated by TLR2, TLR4, or TLR5, the cells were preincubated with anti-TLR2, anti-TLR4, and anti-TLR5 antibodies (1 µg/mL) before the addition of the test products. The effect of flagellin was completely inhibited by the anti-TLR5 antibody and partially reduced by anti-TLR2, whereas the effect of ES0 was partially inhibited by the anti-TLR5 antibody but was not affected by anti-TLR2 or anti-TLR4 antibodies. These results suggest that ES0 activated AMP expression in keratinocytes through TLR5 receptor activation.

Inhibition of lymphocyte cytokines production

In order to assess the effect of ES0 on cytokine production, fresh human CD4+ lymphocytes were incubated with SEB (300 ng/mL) to induce Th1, Th2, and Th17 cytokine production. Various doses of ES0 were added to the culture before proceeding with the cytokine immunoassay. ES0 dose-dependently inhibited Th1, Th2, and Th17 cytokines produced by CD4+ lymphocytes. ES0 inhibited four of eight targets at 30 µg/mL (IL-2, IL-12, IL-10, and IL-17) and all at 100 µg/mL (IL-2, IFN- γ , IL-12, IL-1 β , TNF α , IL-4, IL-10, and IL-17; Figure 4). The percentage inhibition with ES0 of 100 µg/mL ranged from -42% for IL-4 to -95% for IL-12.

Discussion

The active ingredients present in ES0, an *A. dolomia* extract, have not been characterized further; however, different batches of this extract exhibited reproducible activity in each of the cell models used in this study. Our results showed that this extract exerted marked regulatory activity on different mechanisms involved in the pathophysiology of AD, including keratinocyte inflammation, skin barrier impairment, and innate and adaptive lymphocyte immune responses.

In the present study, it was first established that NHK stimulated for 24 h with poly I:C and cytokines (TNF α , IL-4, and IL-13) constituted a model mimicking an “AD-like” condition.²⁶ Indeed, keratinocyte differentiation markers such as involucrin, which were decreased in the present study, are important for skin barrier formation and integrity. Involucrin expression is significantly decreased in both acute and nonlesional skin of subjects with AD compared with skin from healthy subjects.¹⁵ Furthermore, all the inflammatory markers induced in the present in vitro model of stimulated NHK have been shown to be upregulated in AD, for example, TSLP and IL-18 cytokines.^{25,30}

TSLP was associated with the recruitment and activation of Langerhans cells in situ, suggesting its involvement in the early steps of allergic inflammation in the skin, and recently, it was found that TSLP plays a key role in the “atopic march” leading from AD to allergic asthma.^{31,32} IL-18, expressed in various cell types including keratinocytes and macrophages, was also shown to be involved in pruritus and could play a

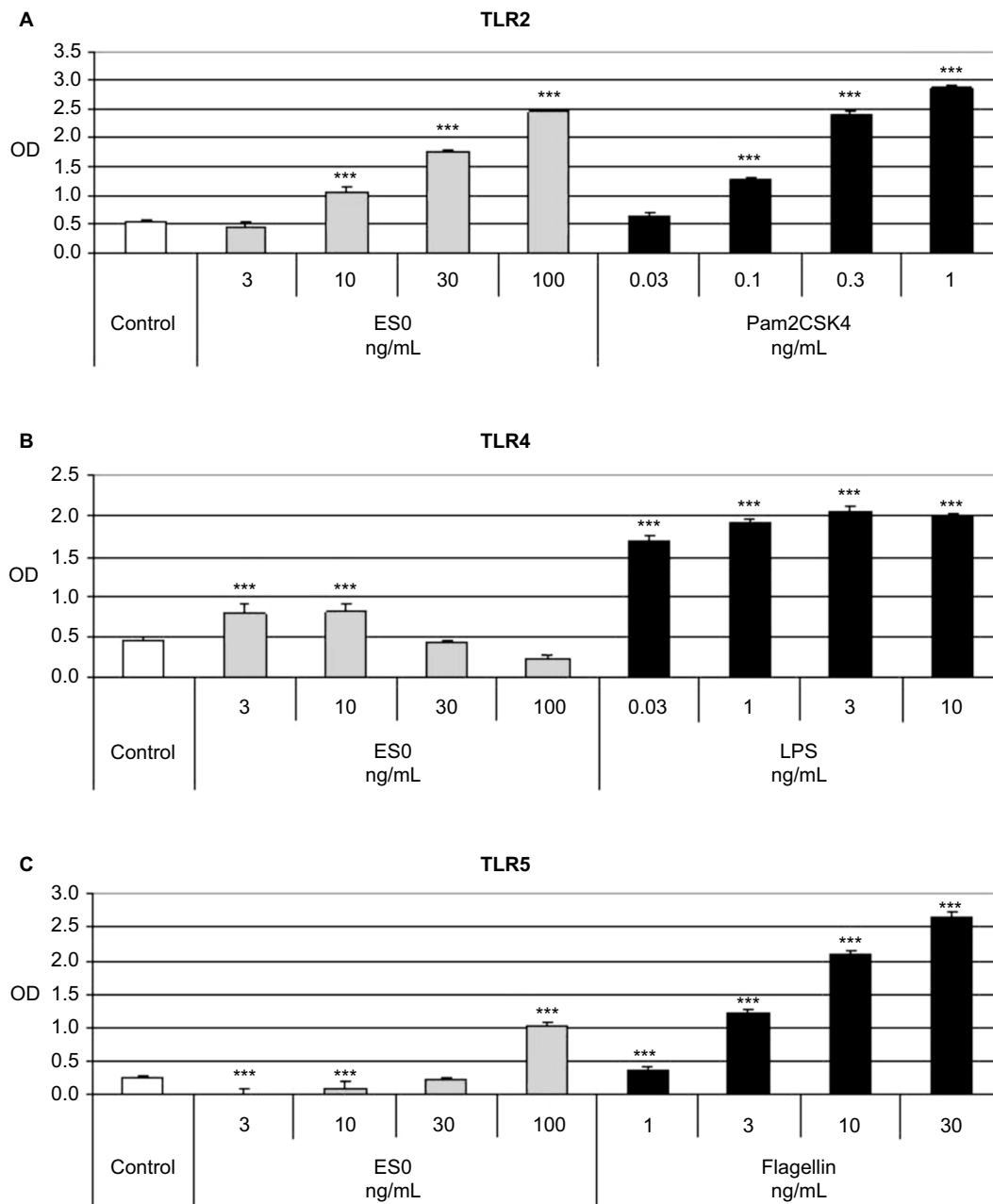


Figure 2 Spectrophotometric measurement of TLR2, TLR4, and TLR5 activation in HEK 293 cell lines.

Notes: HEK were stimulated with ES0 (3–100 ng/mL) or positive controls: Pam2CSK4 = TLR2 ligand (A), LPS = TLR4 ligand (B), and flagellin = TLR5 ligand (C). *** $P < 0.001$ versus control; data are representative of three independent experiments.

Abbreviations: ES0, extract from *A. dolomiae*; HEK, human embryonic kidney; LPS, lipopolysaccharide; OD, optical density; TLR, toll-like receptor.

key role in the pathogenesis of AD, in addition to its proinflammatory role.^{33,34} Chemokines, such as RANTES/CCL5, MCP-3/CCL7, thymus and activation-regulated chemokine (TARC)/CCL17, MIP-3a/CCL20, and MDC/CCL22, which were upregulated in the present model of AD, were also found to be increased in AD skin lesions and might support leukocyte recruitment into the skin.³⁵

By using this keratinocyte model mimicking an “AD-like” condition, it was showed that ES0 dose-dependently

inhibited the expression of all of these proinflammatory mediators, including some associated with the severity of AD lesions, such as epidermal IL-18, which contributes to the development of *S. aureus* infection-associated AD and IL-8, recently shown to be significantly higher in the stratum corneum of severe lesions than in mild or moderate AD lesions.^{36–38} ES0 almost completely inhibited IL-8 expression, which was highly expressed in the present AD model compared with normal keratinocytes. Finally, an 11-fold increase

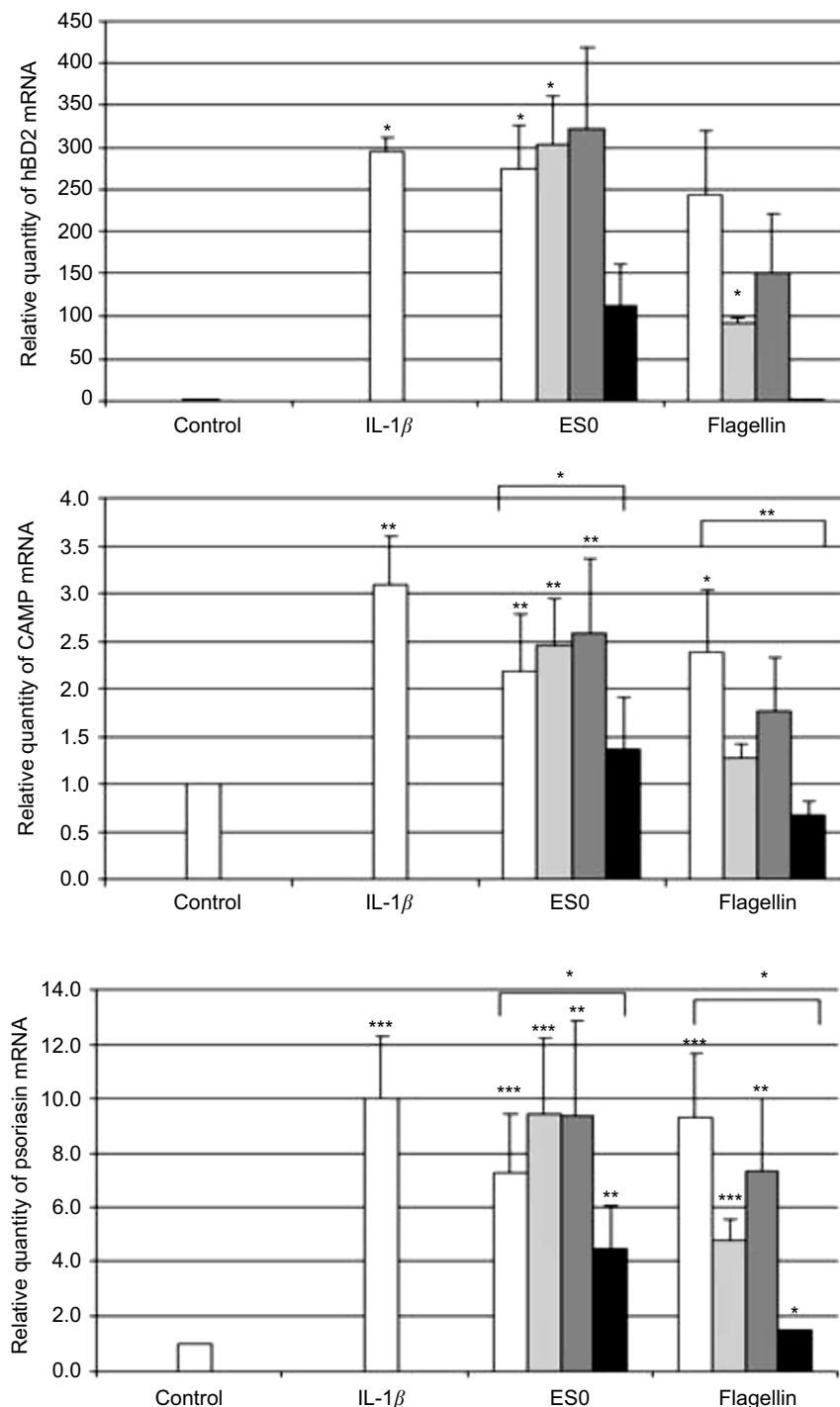


Figure 3 Relative quantity of hBD-2, CAMP, and psoriasin mRNA in NHK cells.

Notes: NHKs were stimulated by IL-1 β (20 ng/mL, positive control), ESO (6.5 μ g/mL), flagellin (100 ng/mL), or alone (negative control). Pretreatment was performed for 1 h with anti-TLR2 (□), anti-TLR4 (▨), anti-TLR5 (▩) (1 μ g/mL), or without antibody (■) to test whether stimulation was blocked in ESO- and flagellin-treated samples; data are the mean of three independent experiments. * P <0.05; ** P <0.01; *** P <0.001 versus control or comparator.

Abbreviations: CAMP, cathelicidin antimicrobial peptide; ESO, extract from *A. dolomiae*; hBD2, beta-defensin 2; NHK, normal human keratinocyte; TLR, toll-like receptor.

in involucrin expression was observed in this AD model following incubation with ESO of 60 μ g/mL, as compared with a 1.1-fold increase with DEX of 2 μ M, showing the beneficial effect of ESO on epidermal barrier integrity.

Another important feature of AD is pruritus, which is largely mediated by PAR-2 and also, as emerging evidence

has shown, TSLP.^{39,40} PAR-2 plays a key role in AD pathogenesis: besides its role in inflammation and pruritus, it also acts as a sensor for exogenous proteases from various allergens, such as house dust mites, cockroaches, pollens, molds, and bacteria.³⁹ Indeed, PAR-2 expression may be induced both by cutaneous inflammation and by these allergens. The

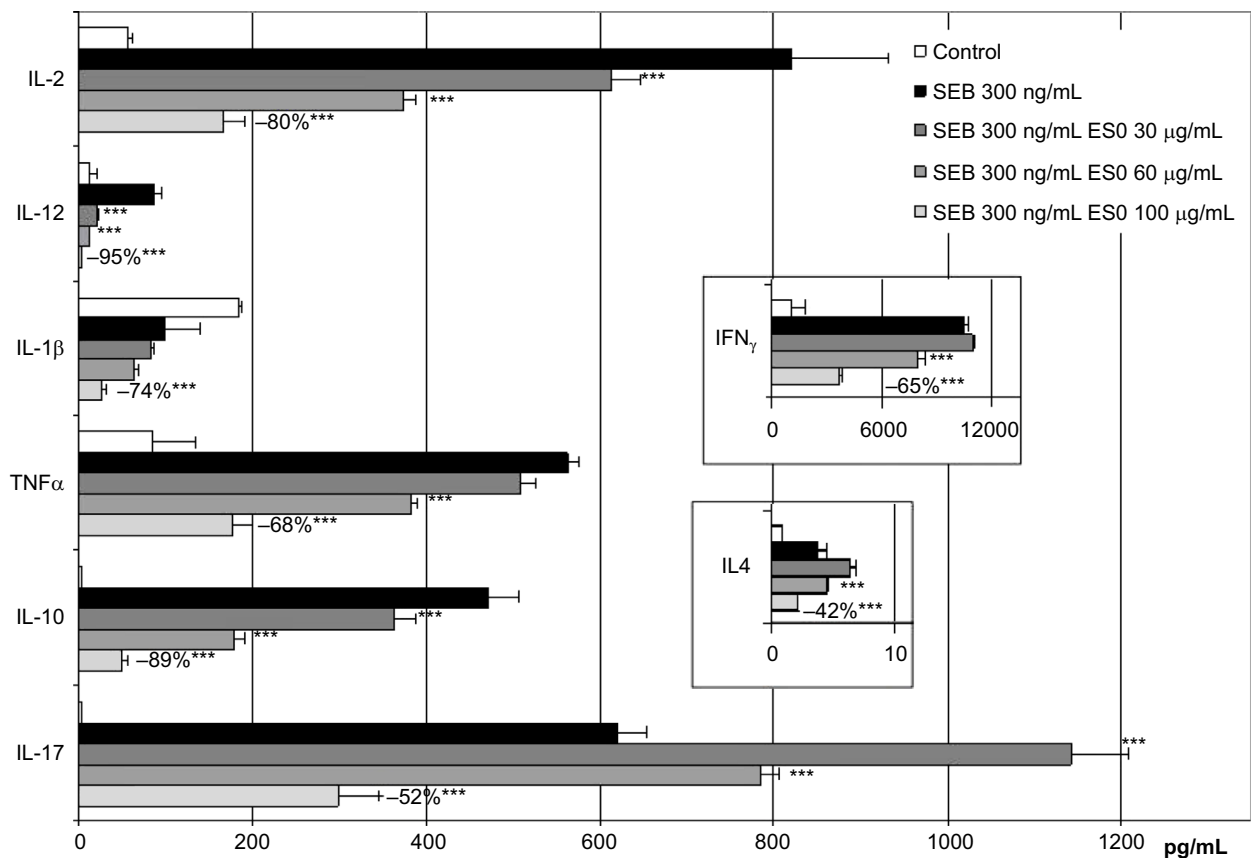


Figure 4 Lymphocyte stimulation by Staphylococcal enterotoxin B (SEB; 300 ng/mL).

Notes: Dose effect of ES0 (30, 60, and 100 µg/mL) on the production of Th1, Th2, and Th17 cytokines; data are representative of three independent experiments. *** $P < 0.001$ versus SEB-treated cells.

Abbreviations: ES0, extract from *A. dolomiae*; IL, interleukin; SEB, Staphylococcal enterotoxin B; Th, T helper cell type; TNFα, tumor necrosis factor alpha.

increased protease and PAR-2 activity in the lesional skin of patients with AD result in abnormal desquamation, alteration of the lipid barrier and antimicrobial barrier by degradation of lipid-processing enzymes and AMPs, respectively, and inflammation through activation of cytokines. As such, PAR-2 may represent a therapeutic target for the treatment of AD.³⁹ Importantly, in the present study, a marked inhibitory effect of ES0 on PAR-2 was observed, as shown by 50% inhibition of PAR-2 activation by SCTE in keratinocytes. ES0 could inhibit PAR-2 activation either by a direct inhibition on the receptor (antagonist effect) or by indirectly inhibiting SCTE (enzyme inhibition). As ES0 is also able to inhibit PAR-2 activated by its specific agonist peptide SLIGKV (data not shown), it could be suggested that ES0 has, at least, an antagonist activity on the PAR-2 receptor. Furthermore, until recently, TSLP was thought to indirectly trigger itch and AD by signaling between epithelial cells and innate immune cells. However, recently, it was shown that TSLP directly stimulates peripheral neurons to promote itch.⁴⁰ It has been shown that PAR-2 activation increases expression of TSLP in cultured epithelial cells; in the present study, it

was shown that ES0 inhibits the expression of TSLP.⁴¹ Thus, ES0 effectively modulates two targets that play a role in the genesis of pruritus in AD, TSLP, and PAR-2.

In addition, ES0 also displayed appreciable activity on innate immunity through the activation of TLRs, which are known to play a positive stimulatory role in innate immunity.^{7,8} Such an immunomodulatory effect of ES0 on TLR2, TLR4, and TLR5 activity without activation of other TLRs (TLR3, TLR7, TLR8, and TLR9; data not shown) in recombinant HEK 293 cell lines was demonstrated. These results suggest that ES0 could activate these three TLRs on normal skin cells that express these receptors.^{11,12,42,43} The combined activation of these three TLRs was shown to induce Th1 polarization synergistically via dendritic cells.⁴² This could be of interest as acute AD is characterized by a skewed balance toward Th2, leading to inhibition of the production of major terminal differentiation proteins, such as involucrin, but also loricrin and filaggrin, and suppression of the expression of hBD-2 and hBD-3.¹³ Notably, several studies have shown that patients with AD have decreased expression and/or function of TLR2 in monocytes and macrophages.^{11,12} Recently, Kuo

et al demonstrated that in cultured keratinocytes, the TLR2-deficient mouse, and a human wound model, TLR2 functions include the preservation of tight junction integrity.⁴³ This protective action would thus be reduced in patients with AD who express less TLR2, and deficient TLR2 responses would also lead to the persistence of pathogenic bacteria, such as *S. aureus*, on the skin surface and sustained inflammation. On the basis of these observations, Kuo et al concluded that strategies stimulating TLR2 expression or function might be promising in restoring epidermal integrity in AD.⁴³ In this context, the marked induction of TLR2, as well as TLR4 and TLR5, by ES0 is particularly valuable.

Another effect of ES0 on innate immunity demonstrated in this study is the induction of expression of several AMPs (ie, CAMP, hBD-2, and psoriasin) in NHK. The effect of ES0 was markedly, but not completely inhibited by anti-TLR5, suggesting that the induction of AMPs by ES0 was mediated by activation of the TLR5 receptor. As TLR5 has been shown to be activated by bacterial flagella, the effect of ES0 on TLR5 could thus be explained by the fact that *A. Dolomia* bears a polar flagellum.⁴⁴ The effect of ES0 on AMP expression is of interest, as insufficient expression of several AMPs in AD was related to a higher susceptibility to *S. aureus* colonization, correlating with the severity of atopic lesions.^{45,46} There is evidence that AMPs are upregulated in AD; however, in particular, the expression of CAMP and hBD-2 is not sufficiently upregulated in patients with AD to kill *S. aureus*.⁴⁷ In addition to their antimicrobial function, hBD-2, LL-37 (CAMP), and psoriasin also contribute to host defenses against infections by their chemotactic activity.^{9,45} hBD-2 activates immature dendritic cells, inducing a strong Th1 response, LL-37 recruits inflammatory cells and triggers cytokine release through various mechanisms, and psoriasin also potentiates inflammation through its chemoattractant properties.^{45,48,49}

Finally, ES0 displayed an immunomodulatory effect on lymphocyte cytokines by inhibiting Th1, Th2, and Th17 cytokine production (IL-2, IFN γ , IL-12, IL-1 β , TNF α /IL-4, IL-10/IL-17, respectively) in CD4+ lymphocytes. These cytokines play complex roles in AD.⁵⁰ The relative absence of IL-17 production by Th17 in AD tissues was suspected to account for reduced AMP levels and a potential increase in the incidence of skin infections.⁵⁰ However, the proinflammatory role of IL-17 was also established, suggesting that it could be useful to downregulate its expression.⁵¹ Th1 and Th2 cytokines have been shown to be involved in the development of AD. Therefore, downplaying the expression of both Th2 and Th1 cytokines, as shown in experimental models, might be valuable in controlling AD.¹³

Conclusion

In conclusion, data from the present study show that ES0, an *A. dolomia* extract, has marked regulatory activity on keratinocyte inflammation, particularly PAR-2-dependent pruritus-associated inflammation, skin barrier impairment, and innate and adaptive immune responses. These effects on various pathophysiological mechanisms of AD suggest the potential value of this extract in topical preparations designed to regulate AD. Moreover, a recent study showed that ES0 counteracts the effects of cutaneous *S. aureus* secretome isolated from atopic children on CD4+ T-cell activation.⁵² Owing to the role of *S. aureus* colonization in driving inflammation in AD, the immunosuppressive property of ES0 might be useful to reduce disease severity. This study, together with the results, showed very interesting activities of ES0 on different parameters implicated on AD.

Moreover, clinical studies were performed in infants, children, and adults with AD in order to assess the efficacy of an ES0-based cream in vivo; these results have been recently published and confirmed that an ES0-based cream applied twice daily decreased the SCORing Atopic Dermatitis⁵³ and pruritus after 28 days of application.⁵⁴ Furthermore, this ES0-based cream protected the skin from *S. aureus* proliferation and preserved microflora biodiversity. These results confirmed the interesting activities of *A. dolomia* extract on adjunctive treatment of AD inflammatory and pruritic lesions.

Indeed, as proposed by Guttman-Yassky et al, “a combination of more than one biologic agent or a biologic agent that targets more than one immunologic factor or cell type might be the best approach to treating AD.”⁵⁰

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Disclosure

The authors are employees of Pierre Fabre Dermo-Cosmétique, France. The authors report no other conflicts of interest in this work.

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Supplementary material

Table S1 Polymerase chain reaction primer sequences and amplicon length

Gene name	Gene symbol	Genebank accession number	Sense primer sequence	Antisense primer sequence	Amplicon Length (bp)
Thymic stromal lymphopoietin	TSLP	NM_033035, NM_138551	CAAAGAAATGTTCCGCCATGA	TGGTCTGTGAAATATGACCA	233
Interleukin 1, alpha	IL1A	NM_000575	ATCAGTACCTCACGGCTGCT	AACAAGTTTGGATGGGCAAC	283
Interleukin 18	IL18	NM_001562	CCAAGGAAATCGGCCTCTAT	TCAGGAGGATTCATTTCCTTAAA	201
Interferon, alpha 2	IFNA2	NM_000605	AGGAGACTCCAATCTGGCTGTG	AGGTGAGCTGGCATAAGAAATCAATG	216
Interferon, beta 1, fibroblast	IFNB1	NM_002176	ATTGCTCCTGTGTGCTTCTCC	TGGGCTCCTCCTTCTGG	207
Interleukin 8	IL8	NM_000584	CTCTTGGAGCCTTCCCTG	TTGGGTCAGACAGAGC	218
Chemokine (C-C motif) ligand 3	CCL3	NM_002983	CCTTCTTGGCTCTGCTGACACTC	CTGGCTGCTCTCAAAGTAGTC	222
Chemokine (C-C motif) ligand 5	CCL5	NM_002985	AGCTACTCGGAGGCTAAGG	GAGGCATGCTGACTTCCCTC	263
Chemokine (C-C motif) ligand 7	CCL7	NM_006273	GGCTGAGACCAAAACCAGAAA	GGACAGTGGCTACTGGTGGT	241
Chemokine (C-C motif) ligand 17	CCL17	NM_002987	TTCAAAACCAAGGCTGCTCC	CTGCCCTGCACAGTTACAAA	279
Chemokine (C-C motif) ligand 20	CCL20	NM_004591	TGAGCTAAAACCCATGTGCTGTA	AGCATTGATGTCACAGCCTTC	199
Chemokine (C-C motif) ligand 22	CCL22	NM_002990	TGTGCCAACTCTCTGCATTC	AGGCTGGAGACAGAGATGGA	258
Chemokine (C-C motif) ligand 27	CCL27	NM_006664	CAGCTTACCAGAAAGCCACT	GCATCCAAAATTCAGCTTG	217
Interleukin 4 receptor	IL4R	NM_000418	CCTGGGAAATCGATGAGAA	CAGGTTGGCTGAGCATATTT	268
Cathelicidin antimicrobial peptide	CAMP	NM_004345	CCCAGGTCCTCAGCTACAAG	GGTCACTGTCCCAATACACC	251
Ribonuclease, RNase A family, 7	RNASE7	NM_032572	CCTCCCCAGAAAAGGACTC	GGAAATTGAAGGTCAGCCAAA	106
S100 calcium binding protein A11	S100A11	NM_005620	CTTGCCATGACTCCTTCCCTC	GCCTGCATGAGGTGGTTAGT	158
S100 calcium binding protein A7	S100A7	NM_002963	GCATGATCGACATGTTTCAAAATACAC	TGGTAGTCTGGGCTATGCTCCC	226
Ceramide synthase 6	CERS6	NM_203463	TTCGACAAAAGACGCAATCAG	AAAACGACAGCTCCAGGATG	216
Involucrin	IVL	NM_005547	CTGGGAACAGCATGAGGAAT	GGAGTCCAAACAGTTGCTCT	194
Filaggrin	FLG	NM_002016	TGATGGTATTCAGTTGGCTCA	TGTTTCTTTGGGCTCTTGG	215
Glyceralddehyde-3-phosphate dehydrogenase	GAPDH	NM_002046	GGCTCTCCAGAACATCATCCCTGC	GGGTGCTGCTGTTGAAAGTCAGAGG	269
Ribosomal protein L13a	RPL13A	NM_012423	TAAACAGTACTGCTGGGCCGGGAAAGGTG	CACGTTCTTCTCGGCCCTGTTTCCGTAGC	483

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