

Improved LFIA for highly sensitive detection of BNP at point-of-care

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Abstract: Heart failure (HF) has become a major cause of morbidity and mortality with a significant global economic burden. Although well-established clinical tests could provide early diagnosis, access to these tests is limited in developing countries, where a relatively higher incidence of HF is present. This has prompted an urgent need for developing a cost-effective, rapid and robust diagnostic tool for point-of-care (POC) detection of HF. Lateral flow immunoassay (LFIA) has found widespread applications in POC diagnostics. However, the low sensitivity of LFIA limits its ability to detect important HF biomarkers (e.g., brain natriuretic peptide [BNP]) that are normally present in low concentration in blood. To address this issue, we developed an improved LFIA by optimizing the gold nanoparticle (GNP)-antibody conjugate conditions (e.g., the conjugate pH and the amount of added antibody), the diameter of GNP and the concentration of antibody embedded on the test line and modifying the structure of test strip. Through these improvements, the proposed test strip enabled the detection of BNP down to 0.1 ng/mL within 10–15 min, presenting ~15-fold sensitivity enhancement over conventional lateral flow assay. We also successfully applied our LFIA in the analysis of BNP in human serum samples, highlighting its potential use for clinical assessment of HF. The developed LFIA for BNP could rapidly rule out HF with the naked eye, offering tremendous potential for POC test and personalized medicine.

Keywords: heart failure, biomarker, colorimetric assays, personalized medicine

Introduction

Heart failure (HF) is prevalent all over the world, which has become a major cause of mortality and morbidity.¹⁻³ According to a report from the American Heart Association, there were ~5.1 million people diagnosed with HF annually in the United States, of which ~50% of them die within 5 years of diagnosis.⁴ The prevalence is predicted to increase by 25% in 2030,^{5,6} and the financial burden is estimated to rise from \$32 billion in 2013 to \$70 billion in 2030.⁷ Generally, HF is classified into four stages based on the progression of the disease, where effective interventions can be taken at early stages to prevent or attenuate its progression.⁸ Therefore, early diagnosis of HF is of paramount importance. The existing clinical diagnosis of HF is mainly based on the electrocardiogram (ECG), electrocardiography, imaging tests (e.g., chest X-ray) and laboratory tests (blood and urine tests), which are equipment dependent, time-consuming and expensive. Thus, they are less applicable in resource-limited settings,⁹ especially in developing countries where access to health care is limited while higher incidence of HF is present. Therefore, a more feasible tool for early and rapid diagnosis of HF in resource-limited areas is vital to release the local economic pressure and reduce the threat of this disease.

To address the issue, point-of-care (POC) testing of cardiac biomarkers, which is a promising method for improving disease management,¹⁰ holds a significant potential.¹¹

Ideal biomarkers for early diagnosis of HF should be highly specific, sensitive and easily accessible. Various biomarkers have been identified for HF, such as renin, norepinephrine, endothelin-1 and natriuretic peptides.^{12,13} Among these, brain natriuretic peptide (BNP) and the N-terminal fragment of BNP precursor (NT-proBNP) in serum have been known as the gold standard to identify HF and widely utilized in clinical laboratories.^{14–16} BNP and NT-proBNP are the products of proteolytic processing of the precursor proBNP (pre-proBNP). The removal of signal peptide from pre-proBNP results in proBNP molecule, which is further split into BNP (biologically active molecule) and NT-proBNP (inactive amino terminal fragment). Since small soluble molecules are preferred for early diagnosis due to their fast release kinetics and rapid diffusion from injured tissue to blood,¹⁷ BNP is a more desirable cardiac biomarker for detecting HF compared to NT-proBNP.¹⁴ Furthermore, BNP possessing a defined cutoff (0.1 ng/mL)⁹ is easier to make a judgment than NT-proBNP, whose cutoff is susceptible to patient's age. Therefore, BNP represents a vital blood cardiac biomarker in patients with acute myocardial infarction and advanced HF.¹²

With advances in POC testing,^{18–22} lateral flow assays (LFAs) hold tremendous potential to address the above-mentioned challenges^{23,24} due to the simple, affordable and user-friendly features.²⁵ However, the existing LFAs are associated with the limitation of poor sensitivity.^{11,26} Several methods have been developed to enhance the sensitivity of LFAs, such as sample concentration,^{27–30} thermal contrast,³¹ fluidic control,^{32–34} temperature–humidity technique,³⁵ probe-based signal enhancement,^{23,36} enzyme-based signal amplification^{37,38} and electrochemical devices-based enhancement.³⁹ However, these approaches require either external equipment,³⁵ high-cost reagents^{23,37} or complicated fabrication with multistep procedure,³² limiting their application for POC testing. To date, a low-cost, convenient and equipment-free sensitivity enhancement method has not yet been explored. Hence, there is still an unmet need to develop a simple and sensitive lateral flow immunoassay (LFIA) for the detection of BNP.

In this study, we introduced a highly sensitive and specific LFIA for POC detection of HF through improving the gold nanoparticle (GNP)–antibody conjugate conditions (e.g., the conjugate pH and the amount of added antibody), optimizing the diameter of GNP and the concentration of capture antibody at the test line, and modifying the structure of lateral flow test strip. Through these improvements, we achieved highly sensitive detection of BNP down to 0.1 ng/mL within 10–15 min with a ~15-fold increase in the sensitivity over

conventional LFA,^{30,35,40} as further quantified by using a smartphone. We also successfully applied our LFIA in the analysis of BNP in human serum samples to diagnose HF, demonstrating the potential of the proposed simple sensitivity enhancement strategy for clinical assessment. The prototype would enable rapid, cost-effective and highly sensitive BNP detection, offering great potential for POC and personalized diagnostics especially in the developing world.

Materials and methods

Reagents and instruments

Two kinds of anti-BNP antibodies, 24C5 and 50E1, (Abcam, Cambridge, UK) were used as the capture and conjugate antibodies, respectively. The standard BNP antigen was also obtained from Abcam. Goat anti-mouse IgG polyclonal antibody was obtained from Fapon Biotech Inc. (Shenzhen, China). $\text{HAuCl}_4 \cdot 4\text{H}_2\text{O}$ was supplied by Sinopharm Chemical Reagent Co., Ltd. (Shanghai, China). Trisodium citrate was purchased from Amresco, LLC (Solon, OH, USA). Bovine serum albumin (BSA), trehalose, casein and phosphate-buffered saline (PBS; pH7.4, 0.01 M) were obtained from MP Biomaterials, LLC (Solon, OH, USA). Sucrose was obtained from Aladdin Reagent Co., Ltd. (Shanghai, China). All the components for fabricating lateral flow test paper, including a polyvinyl chloride (PVC) backing pad, conjugate pad, absorbent pad, sample pad and nitrocellulose (NC) membrane (HFB13502S25; EMD Millipore, Billerica, MA, USA), were obtained from Jiening Biotech Co., Ltd. (Shanghai, China). All chemicals employed in this study were analytical reagent grade. Other solutions were prepared with ultrapure water (>18 M Ω) from the Barnstead Nanopure ultrapure water purification system (Thermo Fisher Scientific, Waltham, MA, USA).

Preparation of GNPs

GNPs with average diameter of 13 ± 3 and 35 ± 3 nm were prepared according to the previously reported protocol.^{41,42} Briefly, 4.5 or 1 mL of 1% trisodium citrate and 1 mL of 1% chloroauric acid (HAuCl_4) were successively added to three-neck round-bottom flask filled with 100 mL boiled ultrapure water. Then, the color of the solution immediately turned from yellow to purple and finally remained wine red. The prepared GNPs⁴³ were characterized by using transmission electron microscopy (TEM; JEM-2100F) and ultraviolet–visible (UV/vis) spectrophotometry (Figure S1).

Modification of GNPs

To make a better combination of GNPs and conjugating antibody, we optimized the pH of GNP–antibody conjugation

and the amount of the added antibody. Briefly, certain volumes of potassium carbonate (0, 2, 4, 6, 8 and 10 μL) were separately added into the as-prepared 1 mL GNP solution to adjust the pH value; then, the excess antibody was added into the abovementioned solution for 10 min incubation at room temperature (RT). The sodium chloride was subsequently added into each solution, and the optimum pH was determined based on the color change and absorbance value of each solution (Figure S2A). Similarly, different volumes of conjugating antibody (0, 1, 2, 3, 4 and 5 μL) were added into the GNP solution with the optimum pH to optimize their volume based on the color change and absorbance value of each solution (Figure S2B). Then, 10% BSA was added to the abovementioned solution and incubated for 10 min to block nonspecific sites to obtain a final concentration of 0.1%. The supernatant solution was removed after the centrifugation at $12,000\times g$ for 10 min, and the red pellets were suspended in 0.22 mL eluent buffer including 1% BSA, 0.85% Tris, 5% trehalose and 20% sucrose.

Fabrication of lateral flow test strips

Control and capture antibodies of BNP were diluted with coating buffer (2% trehalose in 0.01 M PBS) to a certain concentration, which were then dispensed on NC membrane as control line and test line and incubated at 37°C for 2 h to immobilize the antibodies. The GNP–Ab conjugates were dispensed on the conjugate pad, and the sample pad was soaked in the treating fluid for half an hour and dried in oven for 2 h. Then, these elements were consecutively mounted on a PVC adhesive backing pad with 2 mm overlap between each two adjacent pads (Figure 1A). The as-assembled pads were cut into strips (2.5 mm) by Rapid Test Cutter ZQ2000 (Shanghai Kinbio Tech Co., Ltd, Shanghai, China).

BNP detection using LFIAs

A typical LFIA was performed as follows: the sample solution was dispensed onto the sample pad by using a pipette. The color changes at both test and control lines were observed after 10 min. For quantitative analysis, images of the test zones were captured with a smartphone (iPhone 6s) and their optical densities were determined using the Image-Pro Plus software.

Clinical analysis

To prove the sensitivity and specificity of the improved LFIA for BNP detection in clinical serum samples, we tested the LFIA with different concentrations of BNP and four kinds of cardiac biomarkers in spiked human serum. A series of spiked BNP sera (i.e., 1, 0.5, 0.2, 0.1 and 0.01 ng/mL)

and other cardiac biomarkers sera were prepared by diluting their antigen stock solution with normal human sera, respectively. The spiked concentration of four interfering cardiac biomarkers was 10 times of their clinical cutoff (i.e., 100 $\mu\text{g}/\text{mL}$ of C-reactive protein [CRP], 1 $\mu\text{g}/\text{mL}$ of myoglobin [Myo], 30 ng/mL of troponin I [cTnI] and 3 ng/mL of NT-proBNP).^{44–47} After LFIAs were finished, the results were captured with iPhone 6s and the optical densities of test zones were determined using the Image-Pro Plus software for further quantitative analysis.

Statistical analysis

Student's *t*-test was used to compare the data among different groups in optimization assays. Data were expressed as mean \pm standard error of the mean (SEM) of three independent experiments ($n=3$). $P<0.05$ was reported as statistically significant.

Results and discussion

The work principle of LFIA

The principle of the improved LFIA is based on an on-strip sandwich Ag–Ab reaction, and the protocol is illustrated in Figure 1B. In the presence of target BNP, the target would bind to the GNP–Ab, forming GNP–Ab target complexes, which would in turn interact with the immobilized Ab at the test line to produce a red signal detectable by the naked eye. The excess GNP–Ab complexes would then bind to the immobilized secondary Ab at the control line. To achieve rapid POC analysis, qualitative analysis was performed by observing the color changes, whereas quantitative analysis was achieved with a smartphone (iPhone 6s) based on the optical density of the test zone.

Optimization of LFIAs

Paper material, GNP and detection elements have been shown to have significant effect on the performance of LFA.^{48,49} However, these three elements are the personal choice of the research group for developing high-sensitive LFA⁵⁰ and have not been optimized simultaneously, although personal choice may satisfy the requirement for detecting some targets. Therefore, we developed an improved LFIA for highly sensitive detection of BNP by simultaneously modifying the structure of test strip and optimizing the GNP–Ab conjugate conditions, the diameter of GNP (Figure 1C) and the concentration of antibody at the detect line (Figure 1D).

In our previous study,²³ the lateral flow test strip was constituted with NC membrane, absorbent pad and sample pad (Figure 2A[i]). However, throughout our experiment on BNP detection, we found that the complex of GNP–Ab

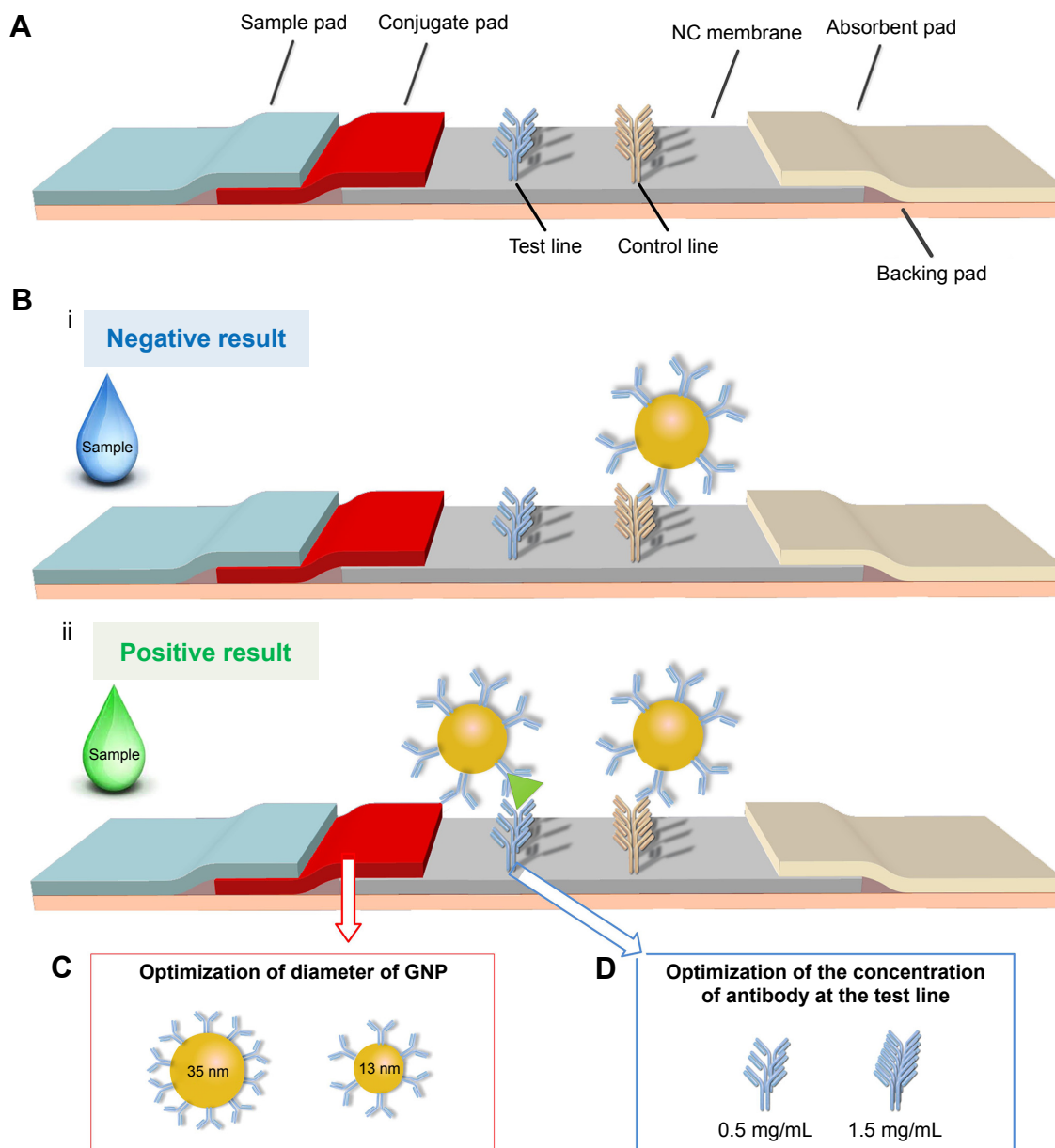


Figure 1 The schematic of the optimized LFA for BNP detection.

Notes: (A) The structure of previous LFA is adjusted by adding a conjugate pad to form a new type of LFA. (B) The negative result (i) and positive result (ii) show the principle of the LFA. (C) The diameter of GNP is adjusted from 13 ± 3 to 35 ± 3 nm. (D) The concentration of test line is adjusted from 0.5 to 1.5 mg/mL.

Abbreviations: GNP, gold nanoparticle; BNP, brain natriuretic peptide; LFA, lateral flow assay; LFIA, lateral flow immunoassay; NC, nitrocellulose.

was unable to completely wick through the previously used sample pad, resulting in the waste and reduction of the effective tracer and degrade the detection sensitivity. This is because different paper materials are applied for different elements of LFIA, ^{50–52} where the inert glass fiber is used as the sample pad, which may not be suitable for the release of GNP–Ab. Therefore, in our modified test strip (Figure 2A[ii]), we introduced an extra surface-modified polyester layer as the conjugate pad (Figure 2B[i]), which is

much thinner than the sample pad to get a complete release of GNP–Ab. Besides, we also adjusted the composition of eluent buffer, where the trehalose was added to protect the activity of conjugate Ab⁵³ and the BSA was added to block the nonspecific binding between the GNP–Ab and NC membrane to eliminate the background signal. With the introduction of the conjugate pad and the change of composition of eluent buffer, the GNP–Ab conjugation was completely released, the background color was eliminated

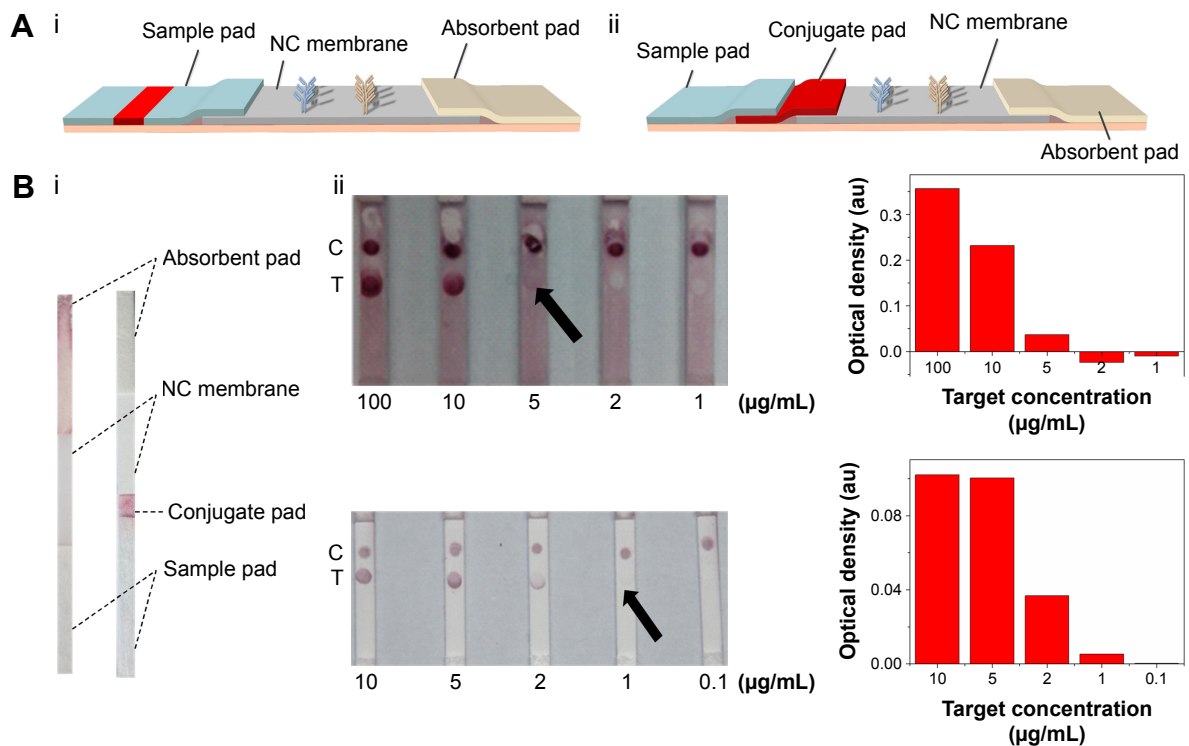


Figure 2 The effect of improved structure of test strip on LFIA performance.

Notes: (A) The schematic diagrams show the structure of test strip before (i) and after (ii) incorporating a piece of conjugate pad into the test strip. (B) The images (i) of comparison of conventional and improved LFAs and the results (ii) which show that the detection limit is improved from 5 µg/mL to 1 µg/mL (the arrows reference the detection limit of the results).

Abbreviations: LFA, lateral flow assay; LFIA, lateral flow immunoassay; NC, nitrocellulose; C, control line; T, test line.

and the signal–noise ratio and sensitivity of detection results were enhanced (Figure 2B[ii]).

Since the principle of LFIA was based on the interaction of the antibody and GNP through hydrophobic and ionic bonds, the condition of the conjugation of antibody with GNP was one of the most important parameters for this immunoassay.⁵⁴ To optimize the condition of the conjugation, we adjusted the pH value of GNP solution and the amount of conjugating antibody and found that the best condition of pH value was 8 and the amount of conjugating antibody was ~20 µg (i.e., 6.8 mg/mL). Accordingly, 4 µL of 0.2 M potassium carbonate (to make sure the pH value is 8) and 3 µL conjugating antibody (Figure S2) were successively added into the as-prepared 1 mL GNP solution for 10 min incubation at RT. The resulting GNP solution was used in the following experiments. Besides, we also adjusted the concentration of the immobilized Ab at the test line in the range of 0.5–1.5 mg/mL (Figure 3). The results indicate that the coating antibody concentration of 1.5 mg/mL showed greater signal of LFIA in BNP detection relative to 0.5 mg/mL ($n=3$, $P<0.05$). In fact, the LFIA with 0.5 mg/mL antibody can achieve a detection limit of 5 ng/mL

($n=3$, $P<0.05$), and with 1.5 mg/mL antibody can achieve a detection limit of 2 ng/mL ($n=3$, $P<0.05$), representing 2.5-fold signal enhancement. The sensitivity enhanced with the increasing concentration of test line. But concomitantly, high concentration of Ab at test line (>1.5 mg/mL) resulted in an observed false-positive result. Therefore, 1.5 mg/mL was selected as the optimized concentration.

Since the diameter of GNP would affect the optical density of test line detected by the naked eye, we investigated the effect of different diameter of GNP (13±3 and 35±3 nm) on LFIA (Figure 4). We found that the LFIA with 35±3 nm GNP showed greater signal in BNP detection compared to that with 13±3 nm GNP ($n=3$, $P<0.05$). Indeed, the LFIA with 13±3 nm GNP can achieve a detection limit of 5 ng/mL ($n=3$, $P<0.05$), and LFIA with 35±3 nm GNP can achieve a detection limit of 1 ng/mL ($n=3$, $P<0.05$), representing five-fold signal enhancement. According to the literature,^{49,55,56} the particles reported (i.e., GNP) can be more easily seen when the size of the particles is larger, but the stability of the colloid will decrease when the diameter is >40 nm,⁵⁶ which may result in the steric hindrance and hamper the antigen–antibody interaction.⁵⁰ Thus, we used the diameter of 13±3

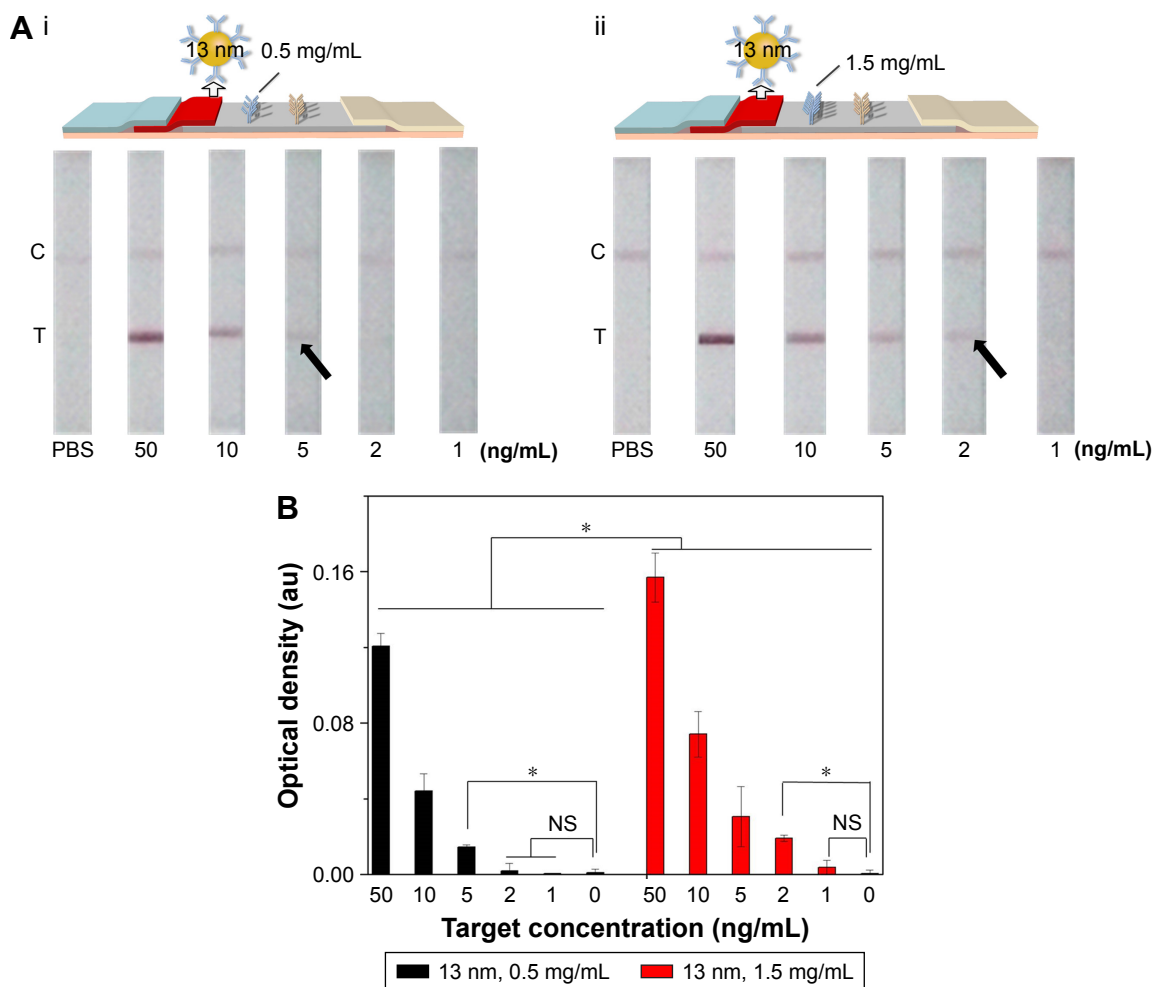


Figure 3 The effect of antibody concentration at the test line on LFI A performance.

Notes: (A) The schematic diagram and results of the LFI A with 0.5 mg/mL (i) and 1.5 mg/mL coating antibody (ii). (B) Optical density analysis of (A) ($n=3$, $*P<0.05$). (The arrows reference the detection limit of the results).

Abbreviations: LFI A, lateral flow immunoassay; NS, no significance; PBS, phosphate-buffered saline; C, control line; T, test line.

and 35 ± 3 nm in this study, and the result showed that the optimum diameter is 35 ± 3 nm.

Application of the improved LFI A

First, we used different BNP concentrations to evaluate the performance of optimized LFIA. With the optimum concentration of antibodies (1.5 mg/mL), GNP diameter (35 nm) and improved structure of test strip (with conjugate pad), the proposed test strip could sensitively detect BNP at 0.1 ng/mL ($n=3$, $P<0.05$) with ~15-fold signal enhancement over unmodified LFIA (Figure 5), which represents the clinical threshold to rule out HF. Moreover, we also assessed the limit of detection of the method. According to the concentration that shows three times of noise detection (signal to noise ratio = 0.0955),^{57,58} the estimated detection limit of our method is 95.5 pg/mL.

Subsequently, to further show the potential of our proposed LFIA for HF diagnosis at the POC, we tested the

improved strip with clinical serum samples. Briefly, we spiked different concentrations of standard BNP antigen (i.e., 1, 0.5, 0.2, 0.1, 0.01 and 0 ng/mL) and four kinds of interfering cardiac biomarkers with 10-fold concentration of their clinical cutoff in normal human serum and then detected them using the proposed test strips. We found that the improved LFIA can detect BNP as low as 0.1 ng/mL ($n=3$, $P<0.05$) in spiked serum, which reaches the clinical threshold (Figure 6A). Comparing to the negative detection results of four interfering biomarkers, the test strip showed positive in BNP detection, indicating the improved LFIA with a good specificity (Figure 6B).

Besides, with the existing target concentration (0.01, 0.1, 0.2, 0.5 and 1 ng/mL) and their corresponding optical densities, we obtained a calibration curve ($y=0.00398+0.08082x$) with $R^2=0.99881$, showing an excellent linear relationship between target concentration and optical density (Figure 7). Therefore, when the proposed LFIA detected the BNP sera

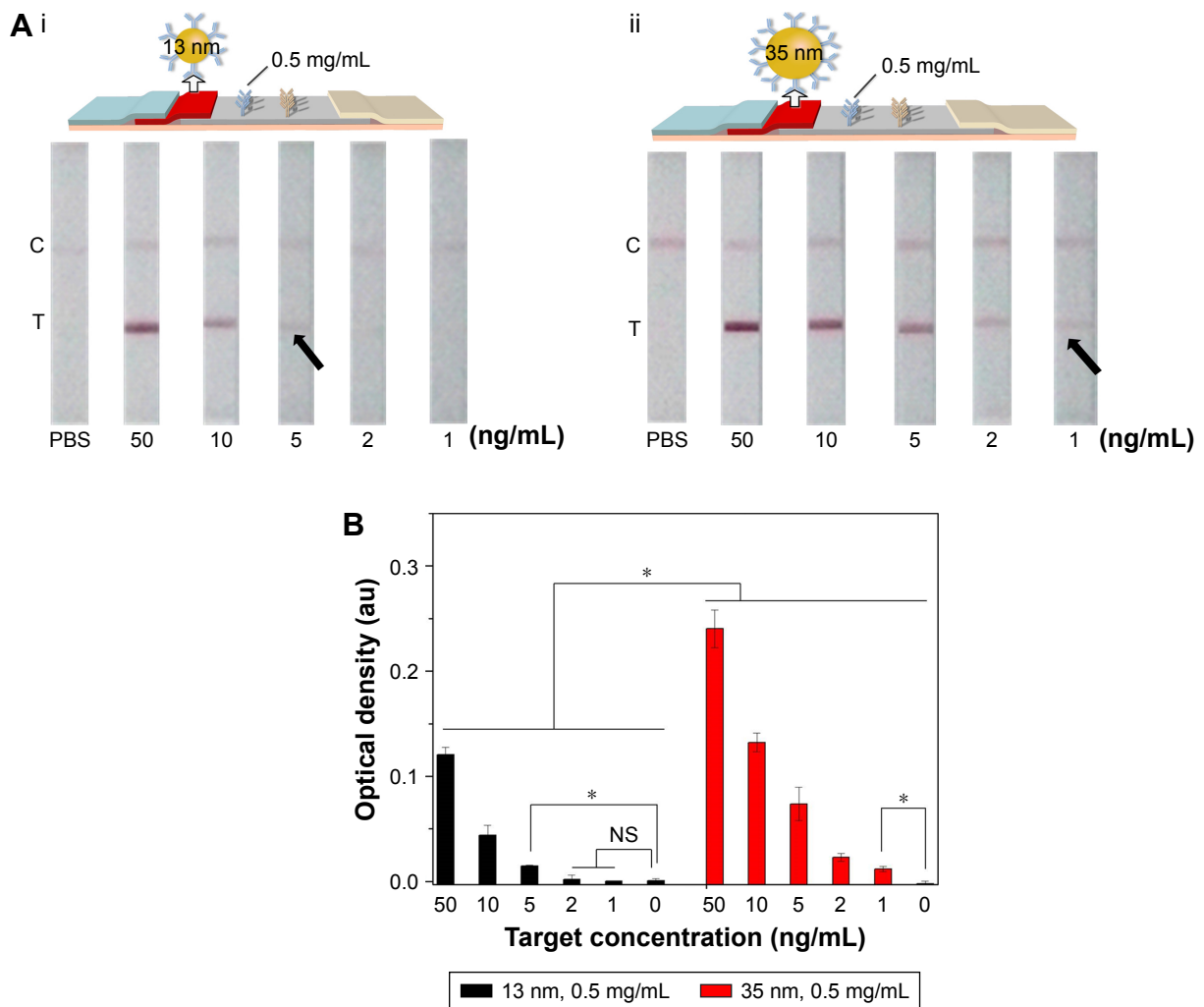


Figure 4 The effect of GNP diameter on LFIA performance. **Notes:** (A) The schematic diagram and results of the LFIA with 13±3 nm GNP (i) and 35±3 nm GNP (ii). (B) Optical density analysis of (A) (n=3, *P<0.05). (The arrows reference the detection limit of the results). **Abbreviations:** GNP, gold nanoparticle; LFIA, lateral flow immunoassay; NS, no significance; PBS, phosphate-buffered saline; C, control line; T, test line.

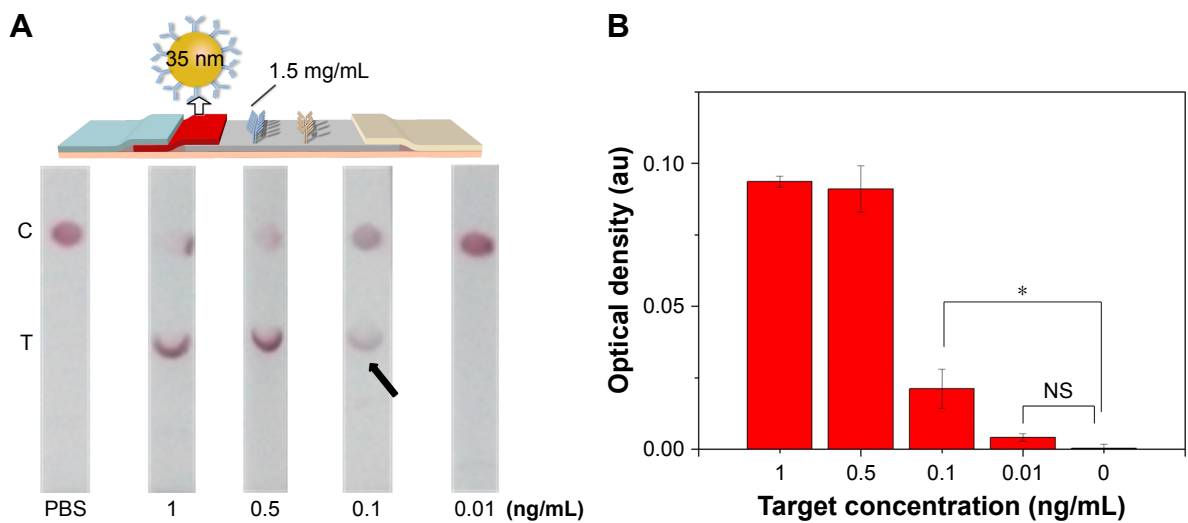


Figure 5 Sensitivity assay using the optimized LFIA. **Notes:** (A) The images showing LFIA results in the detection of different concentrations of BNP solutions. (B) Optical density analysis of (A) (n=3, *P<0.05). (The arrow references the detection limit of the results). **Abbreviations:** BNP, brain natriuretic peptide; LFIA, lateral flow immunoassay; NS, no significance; PBS, phosphate-buffered saline; C, control line; T, test line.

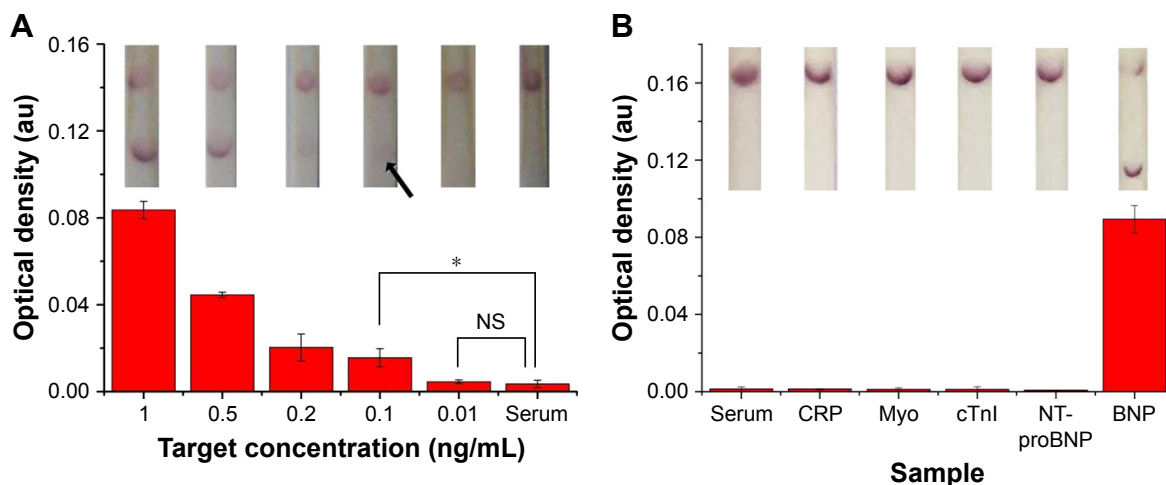


Figure 6 Clinical sample testing using the optimized LFIA.

Notes: (A) The sensitivity results of optimized LFIA detect BNP in human sera (the inset shows the images of LFA results in the detection of different concentrations of BNP solutions). (B) The specificity results of optimized LFIA detect BNP in human sera ($n=3$, $*P<0.05$) (the inset shows the images of LFA results in the detection of different samples). (The arrow references the detection limit of the results).

Abbreviations: BNP, brain natriuretic peptide; CRP, C-reactive protein; cTnI, troponin I; LFIA, lateral flow immunoassay; Myo, myoglobin; NS, no significance; NT-proBNP, N-terminal fragment of BNP precursor.

with 0.4, 0.6 and 0.8 ng/mL, their optical densities were determined using the Image-Pro Plus software. Then, we used the calibration curve to separately convert the optical density to calculate the concentration, and the recovery rates are demonstrated in Table 1, with 0.8 ng/mL associated with the largest recovery rates (0.976 ± 0.006).

In fact, various methods have been developed for the diagnosis of BNP, such as enzyme-linked immunosorbent assay (ELISA) and chemiluminescent immunoassay (CLIA).^{59–61} Although these methods have achieved quantitative detection with high sensitivity (2–10 pg/mL),⁶⁰ they are either time-consuming (~12–36 h), complicated (need tedious procedures), inconvenient (require cumbersome instruments and professional operators) or need large blood volume (~5 mL),

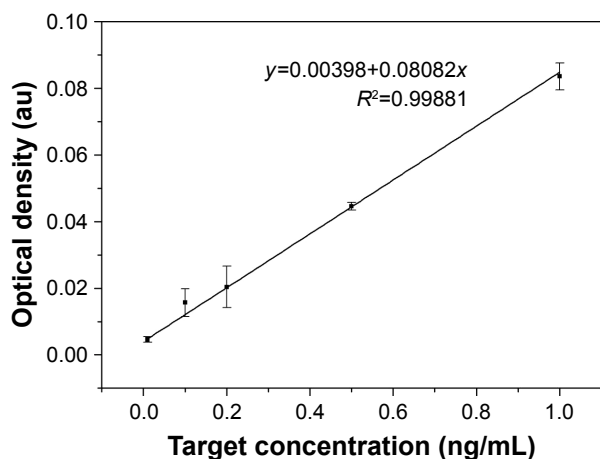


Figure 7 The calibration curve of relationship between target concentration and optical density.

which significantly limit their accessibility in resource-limited settings.⁶¹ Our proposed test strip is simple, low cost and equipment free, which can be performed by untrained users in resource-limited settings in 10–15 min using only a drop of blood, eliminating the need for large and expensive equipment or laboratory infrastructure. More importantly, compared with the existing methods, our improved LFIA can also meet the clinical demand. It has a clinically relevant BNP level detection limit of 0.1 ng/mL, which is the cutoff point to rule out the HF,⁹ making it highly suitable for early and rapid diagnosis of HF at the POC.

Conclusion

We developed a simple, cost-effective and highly sensitive LFIA for rapid BNP detection at the POC. We optimized the conjugating condition of GNP–Ab, the diameter of GNP and the concentration of immobilized Ab at test line and adjusted the structure of conventional LFA. Through these improvements, the proposed LFIA could sensitively detect BNP at 0.1 ng/mL in serum samples, representing ~15-fold

Table 1 The recovery rates of other three concentrations of BNP

Added concentration (ng/mL)	Optical density (au)	Calculated concentration (ng/mL)	Standard deviation	Recovery rates
0.4	0.067	0.374	0.008	0.937 ± 0.020
0.6	0.044	0.499	0.007	0.831 ± 0.012
0.8	0.034	0.781	0.005	0.976 ± 0.006

Abbreviation: BNP, brain natriuretic peptide.

signal improvement over conventional LFIA. We envision that the proposed test strip may show great potential to rapidly detect HF and facilitate early and appropriate health management at the POC.

Acknowledgments

This work was financially supported by the National Instrumentation Program of China (2013YQ190467). All the data generated during this study are included within the published article and its Supplementary materials.

Disclosure

The authors report no conflicts of interest in this work.

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Supplementary materials

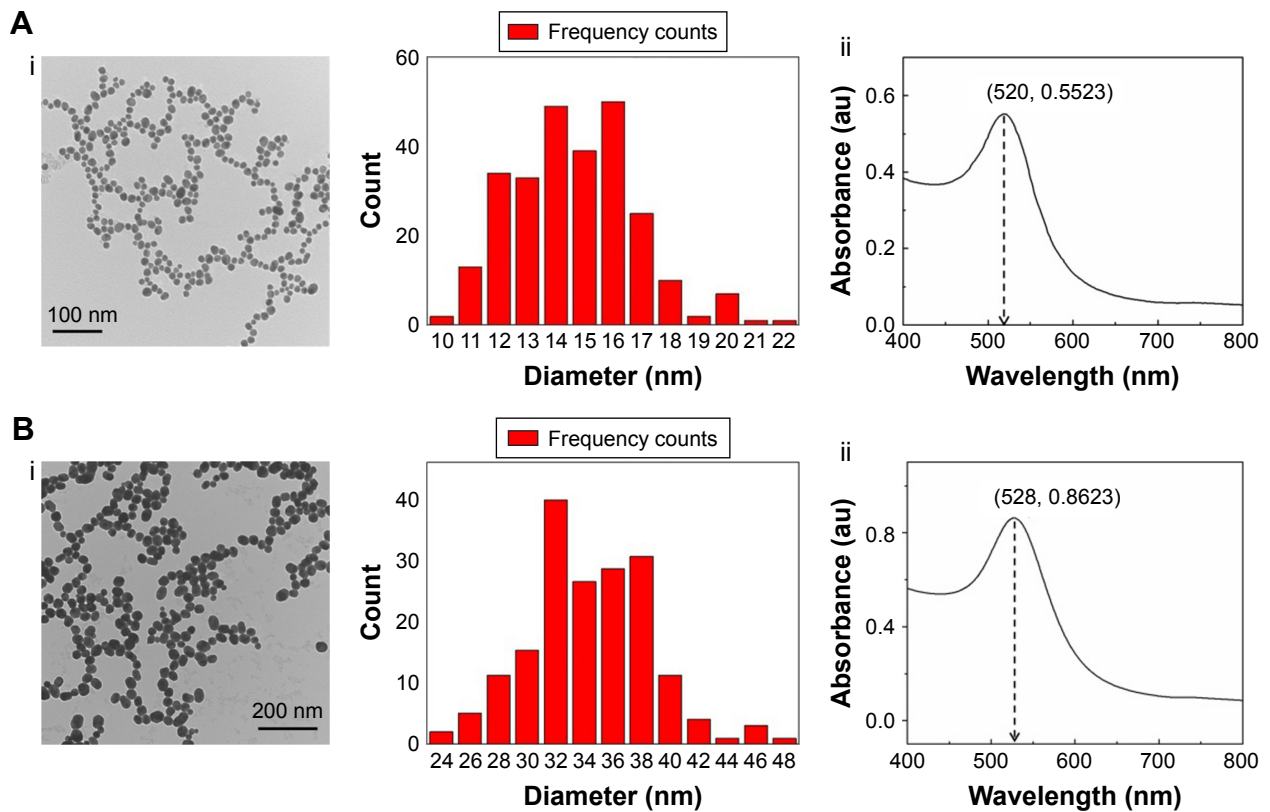


Figure S1 Characterization of GNP.

Notes: (A) Characterization of GNP with 13 nm: (i) TEM and size distribution of GNP; (ii) UV-vis absorption spectra of GNP. (B) Characterization of GNP with 35 nm: (i) TEM and size distribution of GNP; (ii) UV-vis absorption spectra of GNP.

Abbreviations: GNP, gold nanoparticle; TEM, transmission electron microscopy; UV-vis, ultraviolet-visible.

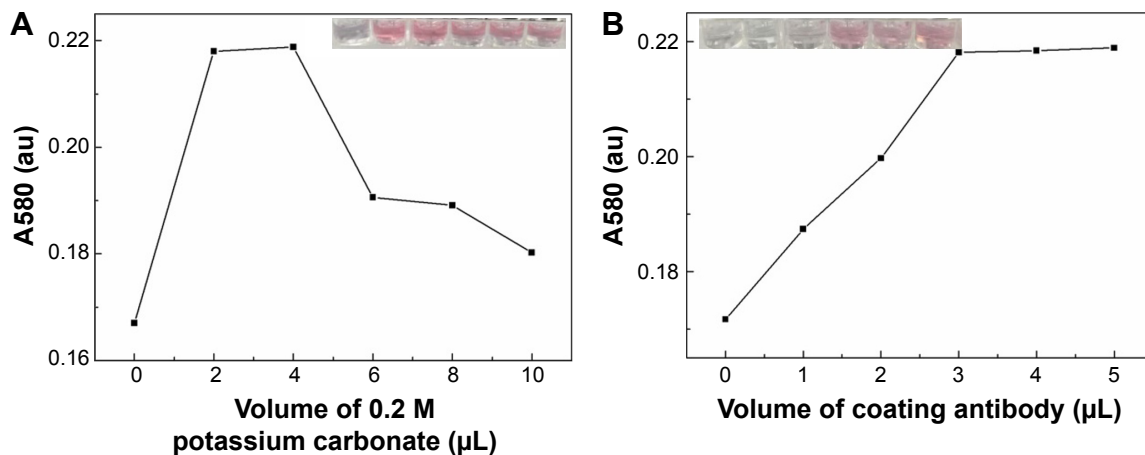


Figure S2 Determination of optimal pH and volume of conjugation.

Notes: (A) Determination of optimal pH for the conjugation of antibody to GNP (The inset shows the images of color change of pH optimization). (B) Determination of optimal volume of coating antibody for conjugation at pH 8 (The inset shows the images of color change of volume optimization).

Abbreviation: GNP, gold nanoparticle.

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