

Characterization and analysis of a novel diguanylate cyclase PA0847 from *Pseudomonas aeruginosa* PAO1

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Background: As a central signaling molecule, cyclic diguanylate (c-di-GMP) is found to regulate various bacterial phenotypes, especially those involved in pathogen infection and drug resistance. Noticeably, many microbes have up to dozens of proteins that are involved in c-di-GMP metabolism. This apparent redundancy and the relevant functional specificity have become the focus of research. While a number of these proteins have been identified and investigated, the functions of PA0847, a PAS and GGDEF domain-containing protein from *Pseudomonas aeruginosa* PAO1, remain unclear.

Materials and methods: In the current study, microbiology, biochemistry and structural biology methods were applied to characterize the gene/protein of PA0847.

Results: We showed that PA0847 affects bacterial motility but not biofilm formation. We recorded the phenotypic influences of amino acids and compounds, and found that PA0847 is involved in response to various environmental nutrients and factors, suggesting its possible role in sensing environmental cues. Both in-vitro and in-vivo studies showed that PA0847 is an active diguanylate cyclase (DGC), whose activity depends on the neighboring PAS domain. Interestingly, PA0847 demonstrates no significant product inhibition, though the key residues of two I-sites for c-di-GMP binding are conserved in its GGDEF domain. A local structural change imposed by an adjacent tyrosine residue was identified, which indicates the structural and functional diversities of the GGDEF family proteins.

Conclusion: Our data provide evidence for understanding the signaling mechanism of the unique c-di-GMP metabolizing protein PA0847.

Keywords: *Pseudomonas aeruginosa*, c-di-GMP, GGDEF domain, diguanylate cyclase, structure

Introduction

Pseudomonas aeruginosa is a widespread Gram-negative bacterium that can cause a variety of acute and chronic infections. It has a complicated regulatory network of signaling molecules, which allow the bacteria to adapt to and thrive in different environments.¹ As a well-established model strain, *P. aeruginosa* PAO1 has been fully sequenced and frequently applied for bacterial phenotypic, metabolic and physiological studies.

In recent years, signaling of the second messenger c-di-GMP (bis-(3,5)-cyclic diguanosine monophosphate) belongs to the most dynamic fields in molecular microbiology.² C-di-GMP is involved in regulation of various bacteria physiological functions, and most noticeably, it modulates the bacteria motility and the switch

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between motile and sessile lifestyles. Typically, elevated intracellular level of c-di-GMP is linked to increased exopolysaccharides and biofilm formation, while decreased c-di-GMP associated with motile planktonic bacteria.³

In bacteria cells, c-di-GMP is synthesized from two molecules of GTP by diguanylate cyclases (DGCs), and degraded to pGpG by phosphodiesterases (PDEs). These enzymes contain the conserved GGDEF and EAL or HD-GYP domains.^{4–6} Strikingly, there is an apparent redundancy of c-di-GMP metabolic proteins encoded in bacteria genome. In *P. aeruginosa* PAO1, these c-di-GMP modulating proteins are highly complex, with 17 GGDEF, 5 EAL, 16 GGDEF/EAL, and 3 HD-GYP domain-containing proteins (http://www.ncbi.nlm.nih.gov/Complete_Genomes/c-di-GMP.html).^{7,8} To date, the biological functions and underlying mechanisms of a quantity of these proteins/signaling systems have been identified and investigated, including the well-studied WspR (PA3702), SadC (PA4332), FimX (PA4959), LapD (from *P. fluorescens* Pf0–1), BdlA (PA1423), and the newly identified FcsR (PA2133), Dcsbis (PA2771) and HsbD (PA3343) et al^{9–16}. The list is still in fast growth.

Noticeably, many of the c-di-GMP modulating proteins contain concatenated distinct N-terminal sensing domains, indicating that they may respond to individual environmental signals.¹⁷ Experimental data in c-di-GMP signaling is emerging dynamically. However, there are still many unresolved issues for understanding the environmental response, functional specificity and physiological importance of these c-di-GMP modulating proteins. In a recent study, a multimodal regulatory strategy, including combinations of ligand-mediated signals, protein–protein interaction and/or transcriptional regulation, was proposed in c-di-GMP networking.¹⁸ In another discussion, a “local” c-di-GMP signaling concept was suggested, where a subset of DGCs and PDEs could operate as “central interaction hubs” in a larger supermodule, with other DGCs and PDEs behaving as “lonely players” without contacting different c-di-GMP-related enzymes any more.¹⁹ These observations signify the networking of c-di-GMP metabolizing molecules far more complicated than previously thought, hence highlighting the need for investigations of individual GGDEF and EAL containing protein in details.

The sequence architecture of PA0847 gene from *P. aeruginosa* PAO1, which encodes a putative GGDEF domain, suggests that it could be involved in c-di-GMP synthesis.²⁰ However, its effects on bacterial behaviors and

its enzymatic mechanisms remain elusive. In this study, we examined and identified the roles of PA0847 in the regulation of bacterial motility and biofilm formation. Further, we found that PA0847 was involved in response to a variety of environmental nutrients and factors, suggesting it could serve as a broad-range environmental sensor on the membrane of *P. aeruginosa*. Domain analysis of PA0847 shows a unique molecular architecture with an N-terminal periplasmic sensory domain and a C-terminal intracellular GGDEF domain. Furthermore, biochemical and structural characterization of the GGDEF domain revealed that PA0847 is an active DGC with weak product feedback inhibition. These new findings provide a basis for studying c-di-GMP signaling through a unique GGDEF domain-containing protein with environmental and clinical relevance.

Materials and methods

Bacteria phenotype analysis

The *P. aeruginosa* PA0847 mutants (Δ PA0847-1/2) and wild type PAO1 strains were requested from Manoil lab. For biofilm analysis, overnight cultures were adjusted to O.D. (600 nm) 0.5, 1:100 diluted with fresh LB broth (1 ml, in Falcon 352057 tube), and incubated at 37°C, 100 rpm for the indicated periods of culture. Cells bound to the walls of the tubes were stained with 0.2% crystal violet (Sigma) for 20 min at room temperature. The tubes were then rinsed with water, air dried and photographed. For quantification, biofilms were suspended in 5 ml of 95% ethanol, and measured with a spectrophotometer at 595 nm. Each experiment was repeated at least three times.

For motility analysis, overnight cultures were adjusted to O.D. (600 nm) 0.5, 1:100 diluted with fresh LB broth, and inoculated into 100 mm petri plates. Swimming motility medium: LB medium containing 0.3% Bacto agar (BD). Swarming motility medium: 0.8% nutrient broth (OXOID) with 0.45% Bacto agar (BD), add 0.5% Glucose before use.

Complementation of PA0847 was applied by amplification of the full-length gene with its native promoter. The PCR products were cloned into the pACYC184 vector and verified by sequencing (Figure S6). Plasmids were then introduced into *P. aeruginosa* via electroporation.

Amino acids and environmental factors

A number of amino acids and compounds were selected to be applied in the M63 medium, to study their effects on

bacteria biofilm formation and motility. The medium applied is M63 supplemented with individual amino acids at the following concentrations: isoleucine (17.6 mM), leucine (25.6 mM), valine (17.6 mM), arginine (4.8 mM), and phenylalanine (8.0 mM).²¹ These concentrations are the same as those applied for biofilm formation analysis. The chemicals and their working concentrations are included in the [Table S2](#).

Recombinant protein expression and purification

Individual domains of PA0847 was subcloned between the BamH I and Hind III sites of a modified pET-32a (Novagen) vector with the S-tag and thioredoxin gene removed. The His-tagged proteins were induced for expression with optimal IPTG and purified with the Ni-NTA resin (Qiagen). The imidazole in protein solution was removed by dialysis against a Tris buffer (pH 7.6).

Enzymatic activity analysis

The recombinant proteins were adjusted to 10 μ M in 100 mM Tris-Cl buffer, pH 7.6, with 10 mM MgCl₂, 30 nM BSA. Substrate GTP were added to the concentration of 200 μ M, incubate at 37°C for 1 hr. Once the reactions ended, reaction mix were incubate in 95°C for 20 mins, centrifuged and filtered to collect the supernatants. High-performance liquid chromatography analysis was applied with Hitachi L2000 HPLC system (Japan) using Hibar 250-4 LiChrospher 100 column (Merck) at the flow rate of 0.8 ml per minute, with mobile phase containing 95:5 methanol and 10 mM triethylammonium bicarbonate (TEAB).

Congo red staining

Congo red plates preparation: 100ml 10 \times stock medium containing K₂HPO₄ 10g, (NH₄)₂SO₄ 1.75 g, Citric acid 2 g; 0.22 μ m filter filtered. Dilute to 1x, adjust to pH 7.0, add MgCl₂ to 1 mM, agarose 10 g/L, with Congo red 40 μ g/ml.

Planktonic culture

Inoculate *E. coli* BL21 containing individual plasmids (GGDEF-PAS, R595A, R641A, D626A, D609A, D652A, K722A, K614A, R672A) of PA0847 into M63 medium, add 0.02% glucose, 37°C, 200 rpm overnight culture. Adjust to O.D. 600 0.5, 1:100 dilute and culture for around

3 h, add 0.05 mM IPTG and culture for 12 hrs, drop one tip and take photos to record.

Homology modeling

A three-dimensional model of PA0847 GGDEF domain (residues 576–736) was constructed in MODELLER-9 package using DGC DgcZ (PDB ID 4H54) as a structure template.²² All the structural figures were generated using UCSF Chimera.²³

Results

PA0847 regulates bacterial motility without significantly affecting the biofilm formation in PAO1

To analyze the biological functions of gene of PA0847 in PAO1, we applied the PA0847 transposon mutants (Δ PA0847-1 and Δ PA0847-2, [Table S1](#)) of PAO1 and compared the behavior with that of the wild type strain (WT).²⁴ We tested the ability of biofilm formation and motility, two phenotypes most commonly influenced by c-di-GMP metabolizing proteins, in both WT PAO1 and Δ PA0847 strains. Compared with wild type strain, PA0847 mutant bacteria displayed significantly increased swimming and swarming motilities (Figure 1A and B). Additionally, the ability of the PA0847 mutants to form a biofilm was not changed drastically (Figure 1C). Complementation of the mutant strains with full length PA0847 cloned in pACYC184 vector resulted in restoration of both swimming and swarming motility phenotypes (Figure 1A and B), while biofilm formation did not display significant changes (Figure 1C). It is therefore demonstrated that disruption of PA0847 would cause an increase in motility while there seemed no significant influence on biofilm formation. These observations for Δ PA0847 biofilm formation were roughly in accordance to that reported by Stephen Lory et al in 2006.⁸

PA0847 is involved in response to various environmental nutrients and factors

P. aeruginosa PAO1 PA0847 contains N-terminal sensing domains,²⁰ implying its possible roles in responding to different environmental cues. Moreover, it has been reported previously that amino acids can modulate biofilm formation and motility, at least in part, by controlling the intracellular levels of c-di-GMP.²¹ We therefore investigated the effects of various amino acids and compounds on motility and biofilm formation for both WT PAO1 and PA0847 mutants.

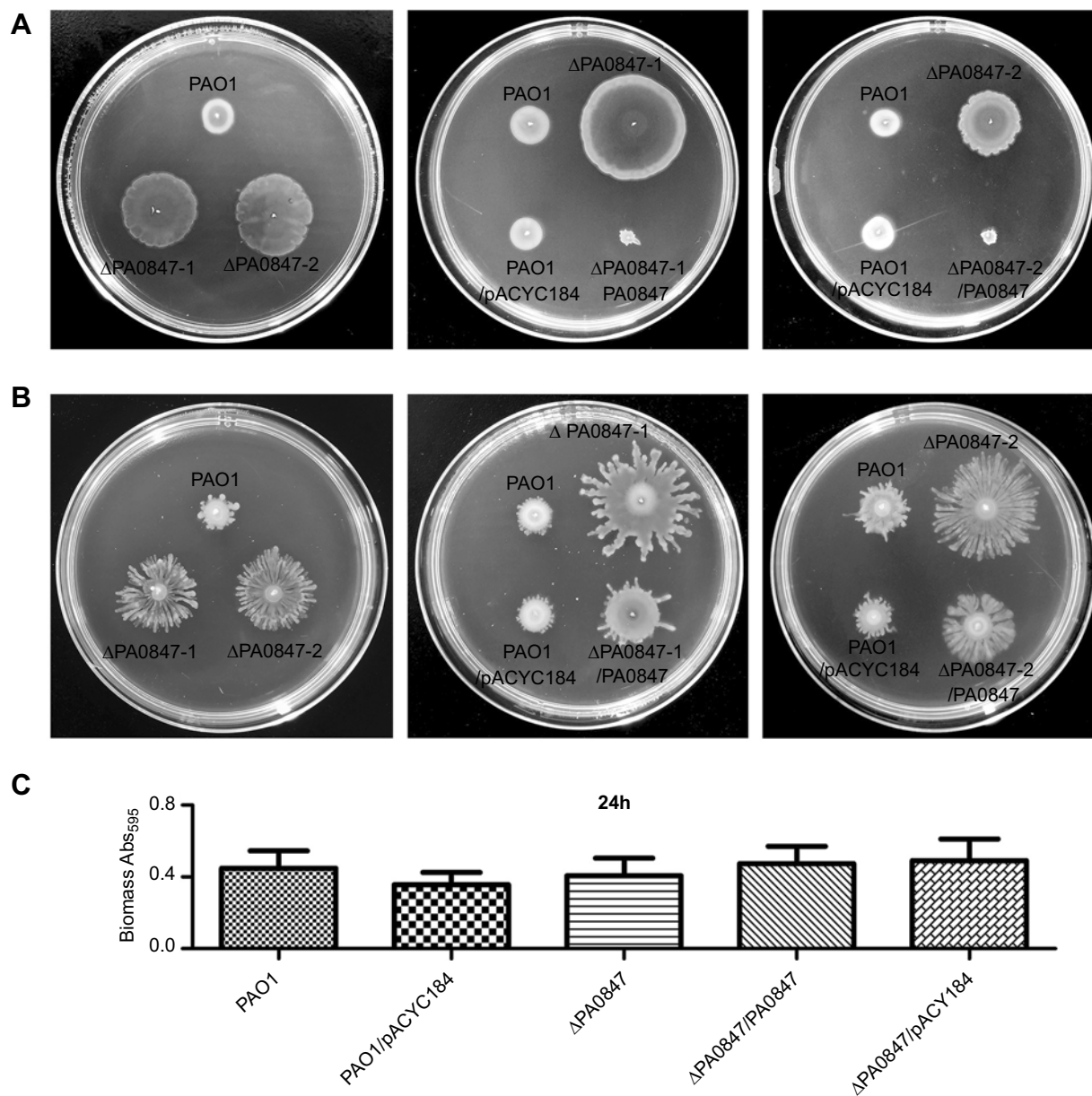


Figure 1 The gene of PA0847 affects biofilm formation and motility phenotypes in *Pseudomonas aeruginosa*.

Notes: (A) Swimming motility of *P. aeruginosa* PAO1 (WT), the PA0847 mutants and the strains complemented with empty vector or PA0847, colony morphologies taken after 20–24 hours of incubation at 37°C, and swimming motility assays performed on medium supplemented with 0.3% agar. (B) Swarming motility of *P. aeruginosa* PAO1 (WT), the PA0847 mutants and the strains complemented with empty vector or PA0847, colony morphologies taken after 36–48 hours of incubation at 37°C, and swarming motility assays performed on medium supplemented with 0.45% agar. (C) Statistical analysis of biofilms of *P. aeruginosa* PAO1, the PA0847 mutant or the strains complemented with empty vector or PA0847. The biofilm data shown are mean \pm SD of at least three replicates. PAO1: *P. aeruginosa* PAO1 WT; ΔPA0847-1 or ΔPA0847-2: two PA0847 transposon mutant strains; PAO1/pACYC184: PAO1 harboring the empty vector pACYC184; ΔPA0847/pACYC184: ΔPA0847-1 or ΔPA0847-2 harboring empty vector pACYC184; ΔPA0847-1/PA0847 or ΔPA0847-2/PA0847: ΔPA0847-1 or ΔPA0847-2 mutant strains harboring full-length PA0847 with its native promoter and RBS cloned into pACYC184.

Abbreviation: WT, wild type.

As shown in Figure 2, ΔPA0847 displayed higher motility with additional amino acids of valine, arginine, phenylalanine, and isoleucine. Leucine at the concentration of 25.6 mM (Figure 2A) caused almost no difference of motility between PAO1 and ΔPA0847. Biofilm formation for ΔPA0847, compared to PAO1, generally

decreased with the amino acids tested (Figure 2B). In the presence of valine, arginine, phenylalanine, isoleucine, and leucine, the differences were statistically significant. The bacterial growth was not strongly affected by these amino acids at an applied concentration (Figures S1 and S2).

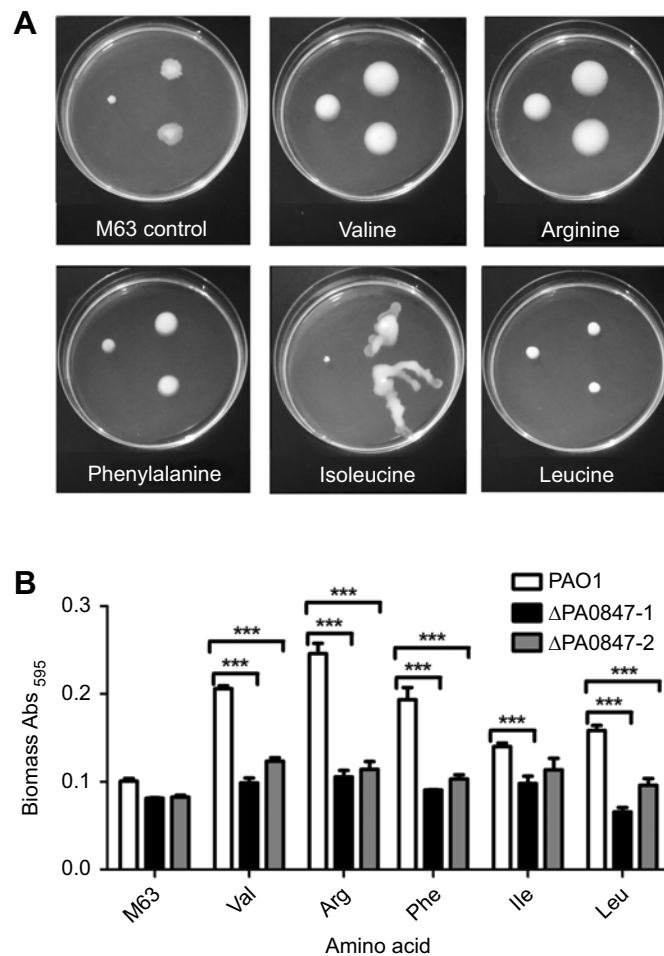


Figure 2 *Pseudomonas aeruginosa* PAO1 and PA0847 mutants motility and biofilm formation in response to individual amino acids differently.

Notes: (A) Differential impacts of individual amino acids on bacterial motility. Swimming motility of both *P. aeruginosa* PAO1 and PA0847 mutants in response to five amino acids. Shown are pictures of plates incubated for 20–24 hours at 30°C. Left: wild type, upper right: ΔPA0847-1, lower right: ΔPA0847-2. (B) *P. aeruginosa* biofilm formation in response to individual amino acids. Shown is the quantification of 24 hour biofilms formed in the tubes and stained with Crystal Violet, solubilized in 95% ethanol and measured at 595 nm. Asterisks indicate values significantly different from the wild type. *** $p < 0.005$.

We also examined some compounds and chemicals. As shown in [Figures S3 and S4 \(Supplemental materials\)](#), with adding of sucrose, raffinose, CaCl₂, MgCl₂, CuSO₄, motility of PA0847 mutants was higher than that of WT strain. When supplemented with spermidine, DTT, SDS, FeCl₂, and NaNO₃, however, there seemed no apparent difference of motility between the two strains. Noticeably, with 10 mM of H₂O₂, WT PAO1 display increased motility. MnCl₂ at the concentration of 2 mM inhibited motility of both wild type and PA0847mutant strains; WT PAO1 growth seemed to be decreased in 4% NaCl, while ΔPA0847 maintained to grow under this condition ([Figure S4](#)).

For biofilm formation, in most of the conditions tested, there were no significant differences between PAO1 and ΔPA0847. With extra sucrose, ΔPA0847 showed an increased biofilm formation. With CaCl₂ and SDS added,

however, the level of biofilm in ΔPA0847 was decreased significantly ([Figure S3](#)). The different tendency indicated that further analysis will be needed, to illustrate the exact function of PA0847 N-terminal domains and their roles in responding to environmental cues.

PA0847 is an active diguanylate cyclase

Domain analysis of PA0847 shows a unique molecular architecture among known GGDEF-containing proteins in *P. aeruginosa* PAO1.²⁰ PA0847 consists of an uncharacterized N-terminal domain (NTD) of ~90 amino acid residues, a predicted periplasmic sensory domain CHASE (cyclase/histidine kinase-associated sensing extracellular) flanked by two single-pass transmembrane domains (TM), an HAMP (Histidine kinase, Adenylyl cyclase, Methyl-accepting protein, and Phosphatase) domain,

a PAS (Per-Arnt-Sim) domain and a C-terminal canonical GGDEF domain (Figure 3A). To experimentally evaluate its diguanylate cyclase activity in-vitro, two recombinant proteins, GGDEF, and PAS-GGDEF dual domains, were expressed, purified and confirmed by SDS-PAGE analysis (Figure S5). The ability of both PA0847 GGDEF and PAS-GGDEF fusion proteins to synthesize c-di-GMP from GTP was assessed by HPLC (Figure 3B). Although isolated GGDEF domain showed only residual DGC activity (data not shown), PAS-GGDEF dual domain efficiently catalyzed the formation of c-di-GMP in vitro. These results indicate that the PAS domain of PA0847 is required for high levels of DGC activity, presumably by bringing

two GGDEF domains together to form a stable homodimer to initiate c-di-GMP synthesis. Mg^{2+} is an essential cofactor for PA0847 cyclase activity, which can be completely disrupted in the presence of the metal-chelating reagent, EDTA (Figure 3B). Titration of GTP γ S to PAS-GGDEF indicated an apparent dissociation constant (K_d) of $15.3 \pm 1.3 \mu M$ (Figure 3C), slightly higher than other GGDEF domain.²⁵ We further investigate the oligomeric state of PAS-GGDEF in solution. Size-exclusion chromatography (SEC) of the recombinant PAS-GGDEF showed co-existence of monomer (~34 kDa), dimer (~68 kDa), and aggregated species (tetramer or larger oligomer) in solution at a micromolar concentration (Figure 3D). The

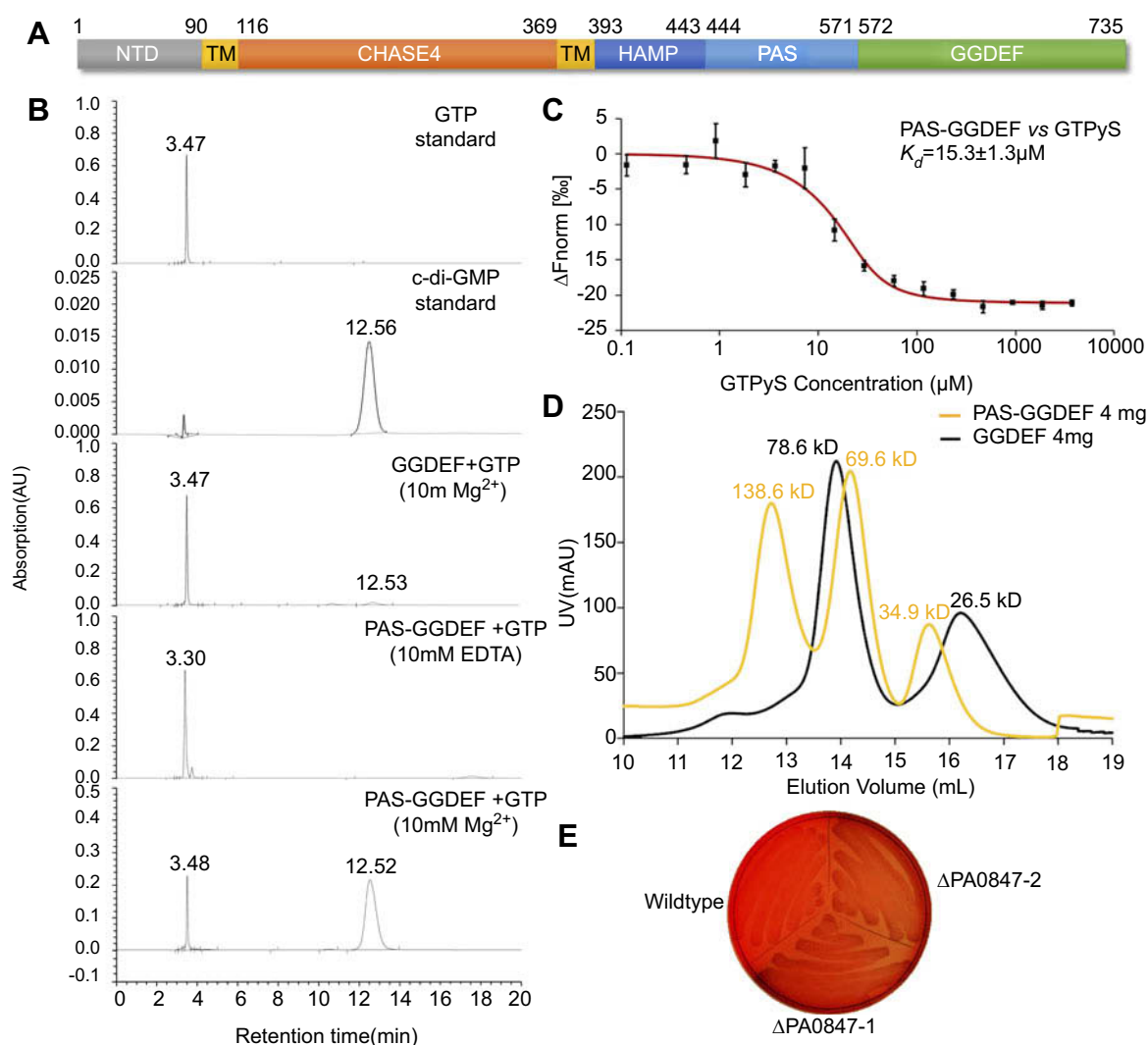


Figure 3 Diguanylate cyclase activity of PA0847 GGDEF domain.

Notes: (A) Molecular architecture of PA0847. Domains are predicted by using the Conserved-Domain Database search from NCBI (<https://www.ncbi.nlm.nih.gov/Structure/cdd/wrpsb.cgi>) and transmembrane helices are predicted in the TMHMM Server (<http://www.cbs.dtu.dk/services/TMHMM/>). The amino acid number of each domain is indicated above the corresponding bar. (B) DGC activity of PA0847 GGDEF domain is detected by HPLC assays. GTP and c-di-GMP standard HPLC profiles are indicated above the assays. (C) The binding of GTP γ S to PAS-GGDEF domain showed a binding affinity (K_d) of $15.3 \pm 1.3 \mu M$. (D) Gel filtration profiles of PAS-GGDEF and GGDEF. The calculated MW of each peak based on protein standards is indicated above the peak. (E) Congo Red staining of wild-type PAO1 and Δ PA0847.

isolated GGDEF domain (~19 kDa) mainly exists in two oligomeric states (Figure 3D). The apparent molecular weight (MW) of the smaller oligomer was estimated to ~26.5 kDa based on size exclusion chromatography, ~40% larger than the theoretical MW of the GGDEF domain. It is most likely that the isolated GGDEF domain rapidly equilibrated between monomeric and dimeric states and form a much less stable dimer in the absence of the PAS domain, which could account for its deficient DGC activity in vitro. It appears that the larger GGDEF oligomer could be a dimer of the smaller one.

In addition to the in-vitro analysis, we stained both WT PAO1 and Δ PA0847 strains using Congo Red to investigate the level of c-di-GMP in vivo, as elevated c-di-GMP results in increased polysaccharides synthesis and stronger Congo Red binding. As exhibited in Figure 3E, wildtype PAO1 is more reddish than Δ PA0847. Combining both in-vitro and in-vivo results, we demonstrated that PA0847 is an active DGC.

Structural model of GGDEF domain and its implications in catalytic function

To understand further the catalytic mechanisms for PA0847, we analyzed the sequence and structural model of its GGDEF domain. Multiple sequence alignment showed that critical residues for DGC activity are all conserved in PA0847 GGDEF domain (Figure 4A). The overall sequence identities between PA0847 GGDEF and other GGDEF domains with known structures are close to 40%, suggesting similarities in global folding. A structural model of the dimeric PA0847 GGDEF domain in complex with two GTPs is presented in Figure 4B. PA0847 GGDEF domain displayed a conserved topology of α - β plait in GGDEF family. It is composed of a five-stranded β -sheet core surrounded by five α -helices. A close examination of the structure reveals seven critical residues (D⁶⁰⁹, K⁶¹⁴, N⁶¹⁷, D⁶²⁶, D⁶⁵², E⁶⁵³, and K⁷²²) involved in GTP binding (Figure 4C). Single or double mutation of these residues completely disrupted GGDEF DGC activity (Figure 4D). PA0847 GGDEF also displays predicted inhibition sites, including a primary I-site (R⁶⁴¹xxD⁶⁴⁴ motif) and a secondary I-site (R⁵⁹⁵ and R⁶⁷²) as shown in Figure 4B. These sites are all conserved in GGDEF-containing proteins with strong product inhibition of some DGCs (ie, PA3702 and PleD in Figure 4A).^{26,27} Nevertheless, the PAS-GGDEF of PA0847 only displays low binding affinity to c-di-GMP with an apparent K_d of ~59 μ M (Figure 4E), indicating weak product inhibition.

Mutation of the key residues in putative I-sites did not significantly increase the catalytic production of c-di-GMP (Figure 4D). Structural analysis of PA0847 GGDEF domain revealed that the first α -helix harboring the secondary I-site is shifted by approximate 15° due to bulky side-chain of Tyr⁷⁰⁰ when compared to the corresponding helix in PleD (Figure 4F). This shift in helix position may sterically hinder potential binding of c-di-GMP. A similar local structural perturbation was also observed in GGDEF domains of PA0575 and PA1120 (YfiN) which does not display product feedback inhibition.²⁵ It is clear that both I-sites and local conformation are necessary for non-competitive inhibition.

To further verify the catalytic function of the GGDEF domain in a cellular context, we expressed the fusion proteins in *Escherichia coli* BL21. After 12-hr induction by IPTG in M63 medium, only the cultures of *E. coli* over-expressing wildtype PAS-GGDEF and mutants of R595A, R641A/R644A, and R672A maintained a similar phenotype of the clear solution with aggregated cell pellets (Figure 4G). These cells possessed lower motility and were prone to aggregate because of their higher cellular level of c-di-GMP, which were in agreement with the HPLC results. Taking all these data together, we concluded that DGC activity of PA0847 is attributed to its C-terminal GGDEF domain which did not undergo strong allosteric feedback inhibition.

Discussion

In current study, mainly through phenotypic analysis, recombinant proteins techniques, structural investigation, and both in-vitro and in-vivo biochemical studies, we demonstrated that gene PA0847 from *P. aeruginosa* PAO1 encodes an active DGC that was involved in c-di-GMP metabolism. PA0847 can also respond to various environmental nutrients and factors, presumably through its putative N-terminal sensory domain.

In phenotypic assays, PA0847 mutant strains exhibited increased swimming and swarming motility phenotypes and no apparent differences in biofilm formation (Figure 1). The obvious redundancy of c-di-GMP metabolizing genes in bacteria regium has long been enigmatic. Recently, the “local signaling” theories have been proposed, indicating that these genes might form interconnected signaling networks.^{18,19} In agreement with this, Overhage J and colleagues had reported that expression of PA0847 could be upregulated drastically while the DGC of PA3177 was knocked out.²⁸ Moreover, when *suhB*, a virulence factors regulator from PAO1, was deleted, the expression of PA0847 decreased slightly.²⁹ Our

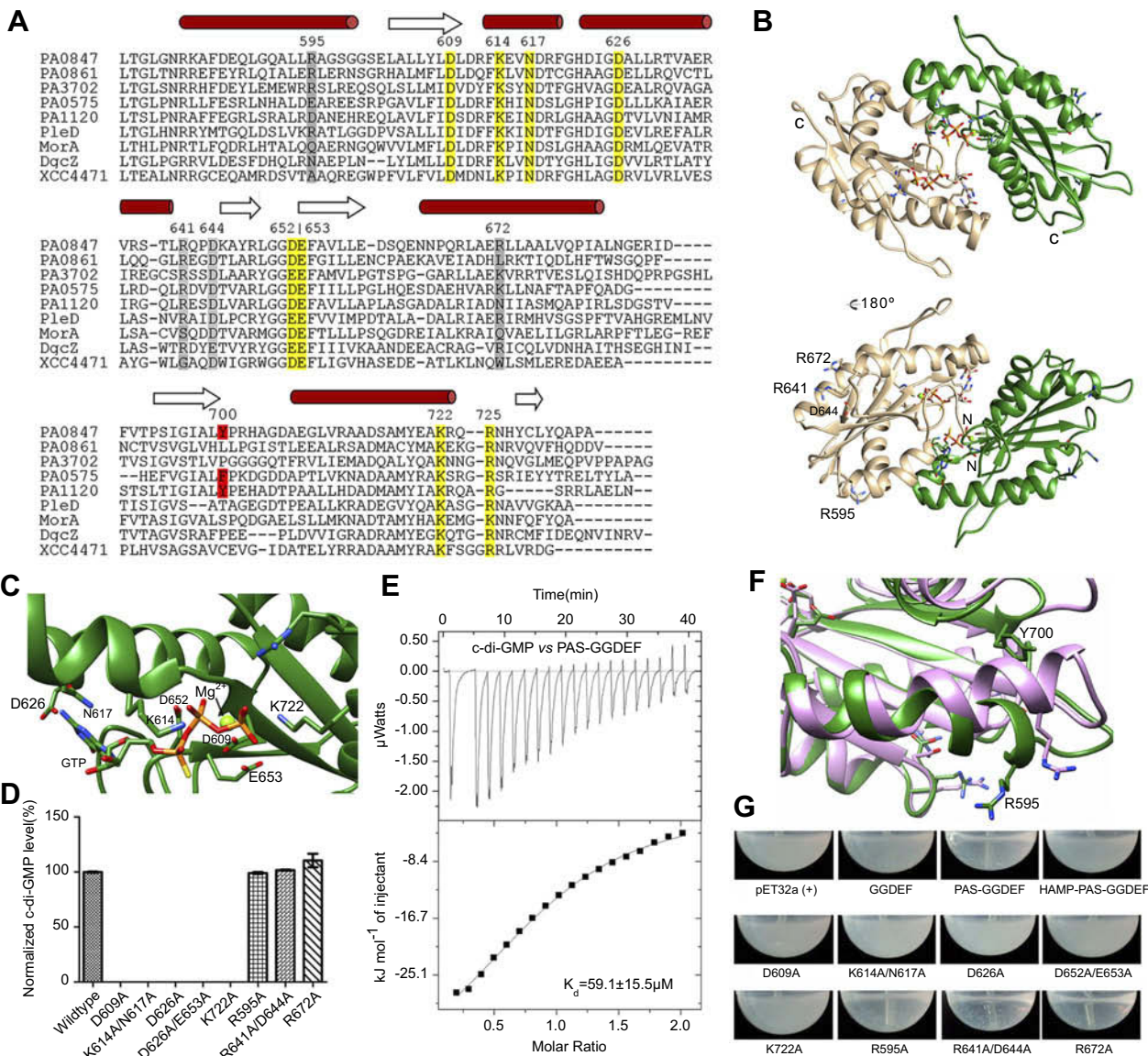


Figure 4 Structural model of PA0847 GGDEF dimer.

Notes: (A) Sequence alignment between PA0847 GGDEF domain and other DGCs of known structure. Secondary structure elements are indicated above the sequences. Key residues involved in binding GTP are highlighted in yellow and putative residues in inhibition sites are highlighted in grey. (B) Structural model of PA0847 GGDEF dimer in complex with GTPs and Mg²⁺. (C) GTP binding residues labeled and depicted as stick models. (D) c-di-GMP synthesis by wild type and GGDEF point mutants of PA0847. (E) Binding of c-di-GMP to PAS-GGDEF dual domain. (F) Structural comparison of GGDEF domains between PA0847 (green) and PleD (pink). Bulky residue Y⁷⁰⁰ was showed in stick. (G) Different phenotype of *Escherichia coli* with overexpressed wild type and mutants of PAS-GGDEF domains.

observations combining these analyses imply that PA0847 might be involved in a complicated c-di-GMP modulating system, where more investigations for their interplays and interactions are needed.

To further analyze the catalytic activities of PA0847, we carried out both in-vitro and in-vivo assays and demonstrated that PA0847 is an active DGC (Figure 3B–E). What worth to mention is that, compared to the dual PAS-GGDEF domain, the recombinant single GGDEF domain of PA0847 exhibited only a trace amount of

DGC activity. This observation implies that the DGC activity of PA0847 might be modulated and affected by the neighboring PAS domain in-vivo.

PA0847 contains the canonical product inhibition site (primary I site) of the RxxD motif, which is located upstream of the conserved GGDEF motif. It also has a secondary inhibition site which is conserved in GGDEF domains that can undergo feedback inhibition (Figure 4A and B). These two sites may form a pocket for c-di-GMP binding.³⁰ To analyze the mechanisms involved in its enzymatic activity

and the regulation, we carried out site-directed mutagenesis using alanine substitutions to generate a series of PA0847 PAS-GGDEF mutants. Unexpectedly, the DGC activity of PA0847 was not significantly affected by product inhibition (Figure 4D and G), in line with its weak binding affinity to c-di-GMP. Nevertheless, different from degenerated I-sites in PA0575 and PA1120 GGDEF domains, all the key residues for c-di-GMP binding are conserved in PA0847 GGDEF domain. Further analysis of the c-di-GMP-binding site indicated a local structural change imposed by a tyrosine residue from an adjacent β -strand when compared to other GGDEF-containing proteins with significant feedback inhibition. Therefore, the primary and secondary I-sites together with I-site local conformation are necessary conditions for product feedback inhibition. The weak product feedback inhibition found in PA0847 GGDEF domain suggests that allosteric regulation from the sensory domain could play a dominant role in the control of intracellular c-di-GMP synthesis.

Recently, there emerged a number of studies reporting that amino acids affect bacteria motility and biofilm formation, mainly through specific and functional c-di-GMP signaling networks. In *B. cenocepacia*, a PDE of BCAL1069 (CdpA) has been reported to regulate swimming motility by sensing the amino acids arginine and glutamate.³¹ For *S. Typhimurium*, it specifically responded to L-arginine with an increase in c-di-GMP, which required the periplasmic regulatory domain of the diguanylate cyclase (DGC) STM1987.³² In *P. aeruginosa* PA14, it has been reported that the two DGCs, SadC (PA4332) and RoeA (PA1107), participate in repression of swarming motility as well as boosting biofilm formation when the amino acid arginine exists.²¹ In a deeper analysis, it was demonstrated that sensing of the amino acid glutamate by NicD (PA4929) in *P. aeruginosa* results in dephosphorylation and increased cyclase activity of this DGC, glutamate thus becomes the cue to induce *P. aeruginosa* dispersion.³³ In the current study, we probed the effects of several amino acids on PA0847. As shown in Figure 2A, with the addition of L-valine, L-arginine, L-phenylalanine and L-isoleucine, *P. aeruginosa* PA0847 mutant cells displayed the trend of increased motility, while their biofilm formation was also affected by L-valine, L-arginine, L-phenylalanine and L-isoleucine, and L-leucine (Figure 2B).

Amino acids have long been noticed to have profound effects on infection. For example, Cystic fibrosis (CF) patients are among the top victims of *P. aeruginosa*. Strikingly, there are CF pseudomonas auxotrophy strains emerged, which are frequently associated with a few

specific amino acids. These CF-adapted *P. aeruginosa* strains reflected bacterium-host symbiosis and coevolution during long-term chronic infection, but not nutrient availability.³⁴ The linking of amino acids with bacteria c-di-GMP modulating genes in previous research and our current study might therefore provide an angle to deepen our understandings, mainly in bacteria physiological regulation.

Likewise, c-di-GMP signaling pathways are believed to be affected by various ions and compounds. A number of environmental signals, including CaCl₂, MgCl₂, CuSO₄, FeCl₂ or NaCl, sucrose, sodium dodecyl sulfate, and dithiothreitol, were shown to differentially regulate the expression of DGCs and PDE in *Y. pestis*, resulting in changed intracellular levels of c-di-GMP and biofilm formation.³⁵ In *P. aeruginosa* PAO1, hypochlorite (HClO), a phagocyte-derived host defense compound, was shown to act as an activator of the DGC PA3177, whose expression leads to biofilm formation and inhibited motility.²⁸ Very recently, in a detailed dissection in *P. fluorescens* Pf0-1, the authors displayed that the citrate and isocitrate are ligands for the N-terminal CACHE domain of GcbC, a DGC which is involved in the complicated LapD GcbC signaling pathway. This ligand sensing may enhance the interaction between GcbC and LapD, promote higher levels of c-di-GMP production and biofilm formation, and finally, contribute to the signaling specificity of the c-di-GMP network.³⁶ Therefore, sensing of environmental cues not only linked to phenotypic changes but also to gene expression regulations, via the c-di-GMP signaling pathway. We also tested ions and some compounds' effects on both wildtype PAO1 and PA0847 mutants (Figures S2–S4). Basing on present results, it is difficult to set direct connections between PA0847 and specific environmental cues due to complicated signaling networks.

It has long been noticed that c-di-GMP metabolizing proteins contain concatenated N-terminus sensory domains, such as CHASE, PAS, GAF, and HAMP.¹⁷ The molecular and structural mechanisms of individual sensory domains and their corresponding environment cues will be the focus of future research to decipher PA0847-mediated signaling in *P. aeruginosa* PAO1.

Conclusion

In current study, we demonstrated that as an active diguanylate cyclase from *P. aeruginosa* PAO1, PA0847 is involved in response to a variety of environmental nutrients and factors, suggesting its role as a broad-range

environmental sensor on the membrane. We also identified a local structural change imposed by an adjacent tyrosine residue, which indicates structural and functional diversities of the GGDEF family proteins. Our work thus contributes to the understandings of c-di-GMP metabolism and signaling in bacteria.

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Disclosure

The authors report no conflicts of interest in this work.

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