

Carbon-covered magnetic nanomaterials and their application for the thermolysis of cancer cells

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Abstract: Three types of graphitic shelled-magnetic core (Fe, Fe/Co, and Co) nanoparticles (named as C-Fe, C-Fe/Co, and C-Co NPs) were synthesized by radio frequency-catalytic chemical vapor deposition (RF-cCVD). X-ray diffraction and X-ray photoelectron spectroscopy analysis revealed that the cores inside the carbon shells of these NPs were preserved in their metallic states. Fluorescence microscopy images indicated effective penetrations of the NPs through the cellular membranes of cultured cancer HeLa cells, both inside the cytoplasm and the nucleus. Low RF radiation of 350 kHz induced localized heating of the magnetic NPs, which triggered cell death. Apoptosis inducement was found to be dependent on the RF irradiation time and NP concentration. It was showed that the Fe-C NPs had a much higher ability of killing the cancer cells (over 99%) compared with the other types of NPs (C-Co or C-Fe/Co), even at a very low concentration of 0.83 $\mu\text{g}/\text{mL}$. The localized heating of NPs inside the cancer cells comes from the hysteresis heating and resistive heating through eddy currents generated under the RF radiation. The RF thermal ablation properties of the magnetic NPs were correlated with the analysis provided by a superconducting quantum interference device (SQUID).

Keywords: graphitic shelled, magnetic nanoparticles, radio frequency, thermal ablation, HeLa cells

Introduction

Magnetic nanoparticles (NPs) present a great interest for a wide range of applications, such as medical diagnostics and therapeutics,^{1,2} magnetic resonance imaging (MRI),^{3,4} catalysis,⁵ data storage,⁶ and environmental remediation.⁷ The low stability of these NPs in severe environments presents a challenge, since it limits their usage without a protective shell. While many methods have been developed for the synthesis of magnetic NPs with different protective coatings, the successful use of such magnetic NPs in biomedical applications is highly dependent on their biostability and surface active properties. Magnetic NPs have a large magnetic moment and behave like giant paramagnetic atoms with fast response to applied magnetic fields with negligible remanence (residual magnetism) and coercivity (the field required to bring the magnetization to zero). Although most studies have focused on the development of polymer or silica protective coatings, carbon-protected magnetic NPs have recently received more attention^{8–10} due to the relatively higher chemical and thermal stability, inertness, as well as enhanced biocompatibility of carbon.^{11,12} Moreover, carbon-coated NPs can be preserved in the metallic state, and thus have a higher magnetic moment than their corresponding oxides.¹³ Core-shell nanostructures often

exhibit improved physical and chemical properties over their single-component counterparts, which leads to a broader range of applications.

In this paper, Fe, Fe/Co, and Co NPs covered with layers of graphitic carbon were respectively synthesized by radio frequency-catalytic chemical vapor deposition (RF-cCVD).¹⁰ C-Fe NPs showed the highest efficiency for inducing cell death compared to their C-Fe/Co and C-Co counterparts. It was determined that the C-Fe NPs had a much higher ability to kill the cancer cells (over 99%) compared with the other types of NPs, such as the C-Co NPs,¹⁰ even at much lower concentrations and for shorter RF radiation times.

The NPs were shown to act as strong RF absorbers and temperature inducers. NPs with various metallic cores induced the death of the HeLa cells during *in vitro* experiments due to their strong magnetic properties. The C-Co NPs exhibited superparamagnetism at room temperature, but at 5 K, C-Fe NPs demonstrated a higher coercivity. Carbon-coated magnetic NPs could be further developed into components of an efficient and reliable tumor therapy for the successful thermal ablation of both individual cancer cells and tumors. To achieve this goal, it is desired to synthesize magnetic NPs that can produce the required heat to kill cancer cells under RF radiation, but do so in low concentrations in order to avoid their possible undesired cytotoxic effects. This may be accomplished by improving the response of the NPs to RF radiation, as well as improving their surface properties of the NPs for specific cellular uptake.

Methods

C-(Co, Fe/Co, Fe) NPs preparation

C-(Co, Fe/Co, Fe) NPs were prepared using the RF-cCVD method as follows: For C-Fe/Co NPs, 1.00 g of nanopowder MgO (Sigma-Aldrich, St. Louis, MO) with 0.145 g of $\text{Fe}(\text{NO}_3)_3 \cdot 9\text{H}_2\text{O}$ and 0.105 g of $\text{Co}(\text{NO}_3)_2 \cdot 6\text{H}_2\text{O}$ were impregnated in 50 mL of ethanol and sonicated for one hour. For C-Fe and C-Co NPs, 0.29 g of $\text{Fe}(\text{NO}_3)_3 \cdot 9\text{H}_2\text{O}$ and 0.21 g of $\text{Co}(\text{NO}_3)_2 \cdot 6\text{H}_2\text{O}$ were used instead, respectively. After drying at 100 °C, the powder was ground finely and typically 0.40 g was used for methane-based cCVD in an RF generator (350 kHz, 5 kW of RF induction). The samples were heated in a H_2 atmosphere at 100 mL/min flow rate up to 800 °C and then subjected to a methane flow of 400 mL/min for five minutes. Finally, the samples were washed with concentrated HCl and collected using a 0.2 μm filter (ISOPORE™; Millipore, Billerica, MA).

Transmission electron microscopy analysis

Transmission electron microscopy (TEM) images were collected on a JEM2100F TEM (JEOL Inc., Pleasanton, CA) with an acceleration voltage of 200 kV. For this analysis, NP powder was dispersed in 2-propanol and sonicated for 10 minutes. A few drops of the suspension were deposited on a TEM grid. This preparation was then dried and evacuated before analysis. Size distributions for each NP preparation were obtained from TEM analysis based on the measurement of 100 NPs.

Atomic force microscopy analysis

Atomic force microscopy (AFM) image was acquired with a Dimension 3100 instrument (Veeco, Plainview, NY) using tapping mode measurements. The cantilevers exhibited resonant frequencies around 300 kHz and force constants of about 40 N/m. This instrument has a scan range of 90 μm for the “x/y” direction and 6 μm for the “z” direction. The sample was prepared by dropping the NP isopropanol solution on Si substrates. Next, the substrates were air-dried and placed directly under the AFM tip for morphology analysis.

X-ray photoelectron spectrometer analysis

X-ray photoelectron spectrometer (XPS) measurements were performed using K-Alpha (Thermo Scientific, Waltham, MA) at a background pressure of 1×10^{-9} torr and a monochromated Al $K\alpha$ ($h\nu = 1456.6$ eV) X-ray source and a combined low-energy electron/ion flood gun for charge neutralization. The collected data were referenced to the graphite C1s peak to 284.5 eV.¹⁴ Detection limits for XPS were approximately 0.1 to 1.0 atomic% depending upon the sensitivity of the elements.

Temperature-dependent measurements of magnetization

These measurements were performed by using a MPMS-5S superconducting quantum interference device (SQUID) magnetometer from Quantum Design Inc (San Diego, CA). The maximum allowed magnetic field was 5 Tesla and the magnetic field sweeps could be performed for both polarities in a temperature range of 1.8–300 K. The longitudinal magnetic moment was measured using the DC technique. Each point was taken as an average of four single measurements. Samples were mounted in gelatin capsules having a negligible magnetic response. The weight of synthesized

materials was measured using a weight measurement station (XL-410; Denver Instruments, Denver, CO).

Three types of measurements were performed:

1. Zero-field-cooled (ZFC): samples were cooled down to temperature of 5 K with zero applied magnetic field, then a magnetic field of 30 Oersted (unit of magnetizing field and also known as magnetic field strength or intensity Oe) was applied and temperature dependent measurements were taken during warm up.
2. Field-cooled (FC): samples were cooled down to 5 K at an applied magnetic field of 30 Oe and measurements were taken during warm up with the magnetic field held constant.
3. Hysteresis loops were measured for each of the samples at 300 K and 5 K in the range of -2 to 2 Tesla.

HeLa cell culture and incubation with NPs

For the cell culture, mammalian cervical cancer cells (HeLa cells) were seeded in 10 cm² culture plates (0.5×10^6 cells/plate) with growth medium (minimum essential medium containing 10% fetal bovine serum and 1% penicillin 100 unit/mL, streptomycin 100 μ g/mL) and incubated in a humidified incubator (37 °C, 5% CO₂). For the subculture, cells were dissociated by $1 \times$ trypsin/EDTA in PBS and counted and plated into 35 mm culture plates at a density of 5×10^4 cells/plate and supplemented with growth medium containing various concentrations of NPs (0–3.33 μ g/mL). All the NPs were incubated with HeLa cells for 24 hours before RF treatment.

Cellular death studies performed using ethidium bromide and acridine orange (EB/AO) staining methods

For EB/AO staining, 25 μ L of the cell suspension (5×10^4 cells/ μ L) was mixed with 2 μ L of the dye mixture, containing 0.15 mg mL⁻¹ of acridine orange and 0.50 mg mL⁻¹ of ethidium bromide in $1 \times$ phosphate-buffered saline (PBS). After staining, cells were then visualized by using a fluorescence microscope (Olympus, Center Valley, PA) at 10 \times and 40 \times objectives equipped with a digital camera. Photographs were taken using randomly selected fields of view. To determine the percentage of cells undergoing death, photographs taken were used for counting the number of live (green) and dead (orange) cells. Acridine orange stains live cells green whereas ethidium bromide stains dead cells as red. Approximately 200–300 cells per treatment were counted to obtain statistical cell death information.

RF heating and determination on surface temperature of NP powder

Cells were subjected to RF induction at 350 kHz and 5 kW for time periods ranging from 2 to 45 minutes. The calculation of the cells was done under fluorescence microscopy counting the dead and alive cells. NP powders were placed in the Petri dish individually and subject to RF heating for five minutes. Before, during, and after RF heating, an infrared thermometer (PTM 01, Russia) was used to check the temperature of the powdered samples. All heating studies were repeated at least five times in all conditions.

Results and discussions

The magnetic NPs were synthesized by a RF-cCVD process.¹⁰ AFM and TEM analysis revealed that the average size of the C-Fe, C-Fe/Co and C-Co NPs was 8 ± 1.0 nm, 9 ± 1.5 nm and 7 ± 1.2 nm, respectively, with the mean and standard deviation of sizes measured by TEM for ~ 100 NPs in each samples. The NPs were covered with 5–8 layers, 7–10 layers, and 2–4 layers of graphitic carbon respectively, as shown in Figure 1.

XPS analysis was carried out to inspect the oxidation states of the encapsulated NPs. Core level XPS profiles of the C1s, O1s, Fe 2p, and Co2p were taken at a pass energy of 50 eV in 0.01 eV steps for higher resolution. The Co2p and Fe2p peaks were located at 778.6 (± 0.1) eV and 707.1 (± 0.1) eV (Figures 2a and 2b, respectively), indicating the metallic state (Co oxides ~ 780 –781 eV, Fe oxides 709–711 eV).¹⁵ This was also confirmed by the O1s peak at 532.7 eV, since the O1s peak for Co-oxide is 532.35 eV.¹⁶ Therefore, the metallic states of the Fe, Fe/Co, and Co NPs were well protected by the graphite shells.

In order to confirm the biocompatibility of the different types of NPs, cytotoxicity studies involving HeLa Cells were carried out whereby the cells were cultured with the C-Fe, C-Fe/Co, and C-Co NPs and these cultures were analyzed for cell viability. The results indicated that most (99.4 to 99.7%) of the total cultured cells were alive, revealing a low level of toxicity of these magnetic NPs. Furthermore, it was found that RF radiation alone hardly induced any effect to the HeLa cells since around 93.0% of the cells still survived after RF treatment depending on the radiation time.

HeLa Cells cultured with various concentrations of magnetic NPs were introduced inside a water-cooled coil coupled to a radiofrequency generator with the frequency of 350 kHz, which is far lower than the 10 MHz to 300 GHz that is commonly used.¹⁷ The depth of penetration

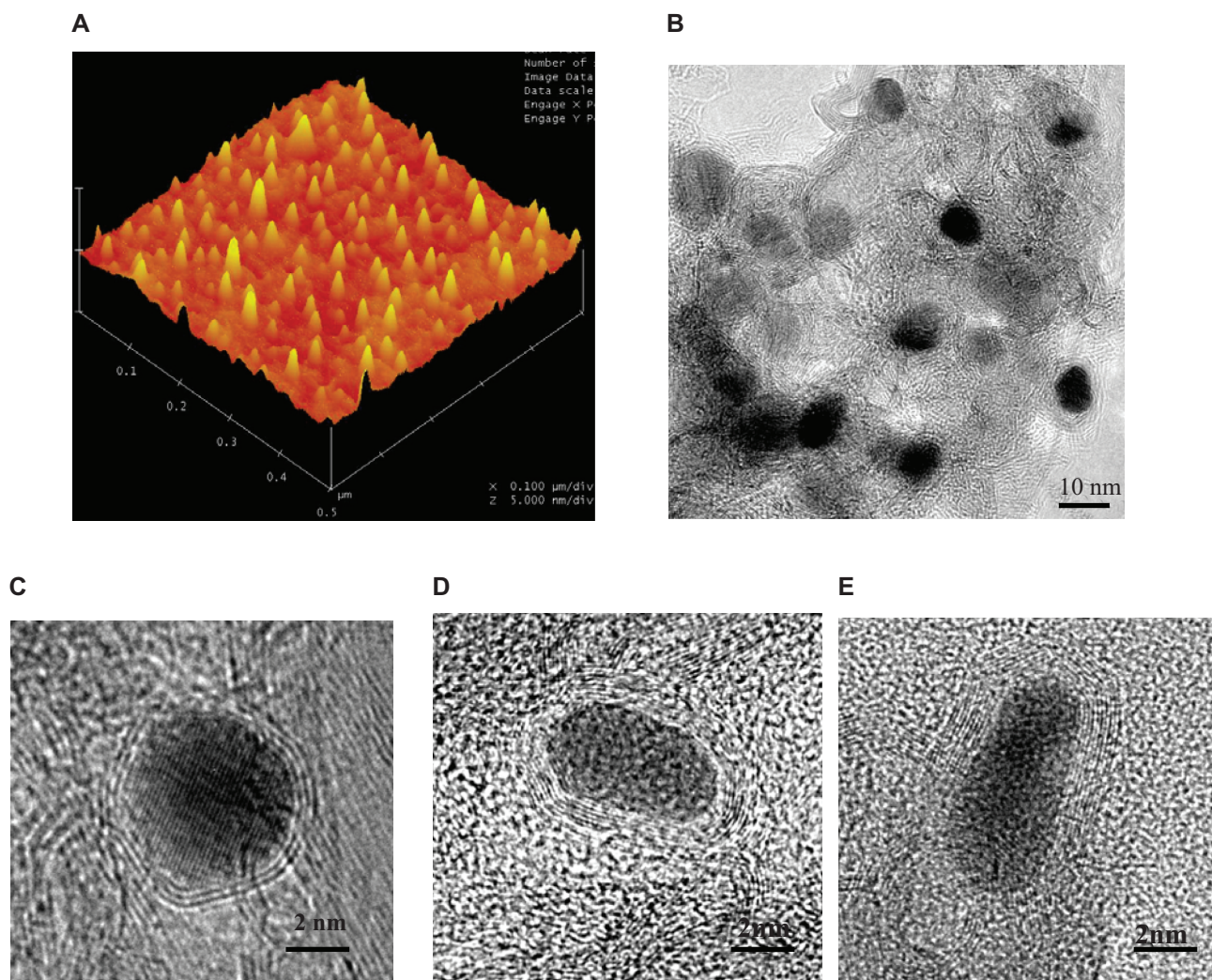


Figure 1 **A)** AFM topographic images showing individual C-Co NPs. **B)** Low and **C, D, E)** high magnification TEM images of graphitic **C)** C-Co, **D)** C-Fe and **E)** C-Fe/Co NPs obtained by RF-cCVD method.

Abbreviations:

of RF radiation in human tissue decreases as the frequency increases. Current clinical microwave radiometers operate in the GHz frequency range. The effective depth of the RF penetration drops from 17 cm at 85 MHz to 7 cm at 220 MHz. Thus, RF below 200 MHz are more desirable for the thermal ablation of cells or tissues located 1–20 cm deep into the body. Such low frequency radiation has the ability to penetrate the biological tissues efficiently and present a path for cancer treatment deep inside the body (for example, at 400 kHz, field penetration into 15 cm of tissue is >99%).¹⁸ At frequencies below 100 MHz, the RF power deposition in patients is more evenly distributed over the exposed body volume than currently assumed.¹⁹ The frequency of the AC electromagnetic field should be chosen based on a compromise; namely, it must be higher than the frequency capable of provoking

neuromuscular response and lower than the frequencies causing overheating of healthy tissues. It is believed that the frequency should be in the range 100–1000 kHz; in this case, correctly chosen frequencies and electromagnetic field strengths produce no notable side effects of the AC magnetic field *in vivo*.²⁰ The schematic diagram of this setup was shown in our previous work.¹⁰ After the RF irradiation, the total number of live and dead cells was immediately counted through visualization by fluorescence microscopy. Figure 3a shows the results of the influence of the RF exposure for time periods of 2 to 30 minutes on the induction of apoptosis in the cells. If the concentration of the NPs was kept constant, the number of dead cells gradually increased after extending the RF-heating time. The three types of NPs were kept at the same low concentration (about 0.83 μg/mL) and

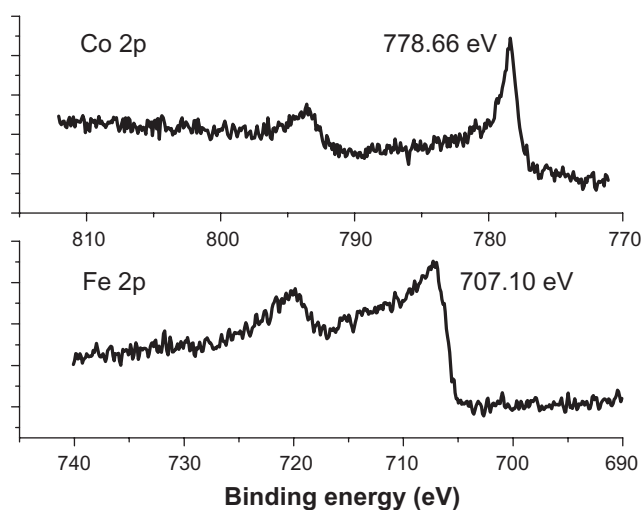


Figure 2 XPS spectra of the Co2p and Fe2p peaks represent the patterns of C-Fe and C-Co magnetic NPs.
Abbreviations:

were incubated with the HeLa Cells. After the RF heating, C-Fe NPs exhibited the highest killing rate of the HeLa Cells, as presented in Figure 3a. During only two minutes of RF exposure, 67.0% of cells were observed to have died. Increasing the heating time to 30 minutes caused the percentage of dead cells to reach about 74.5%, which is 2.14 times and 2.69 times higher than that induced by the C-Fe/Co NPs and C-Fe NPs, respectively. The results indicated that the C-Fe NPs were the best RF absorbers and induced cellular death in the shortest time and at the lowest concentrations. Moreover, for RF exposure times longer than 10 minutes, the percentage of dead cells increased rather slowly (around 10% or less) depending on the type of NPs used in the experiment. As a result, if the exposure time was kept constant, the percentage of dead cells was found to be highly dependent on the NP concentration. As shown in Figure 3b, when the NP concentration was increased to 3.33 $\mu\text{g}/\text{mL}$, C-Fe NPs killed almost 98.88%–100% of the exposed cells with only two minutes of RF exposure. However, for the C-Co NPs, a significantly higher concentration ($\sim 20 \mu\text{g}/\text{mL}$) was required to cause the same effect.¹⁰

It is believed that there will be variations in the amount and the numbers of NPs that will penetrate each individual cell. Although these variations exist, over a certain period of time, all cells will be heated up even at lower concentrations of NPs absorbed. The heat from one cell will propagate to the adjacent ones and therefore create a uniform temperature distribution over a certain area. This process of heat transfer is believed to enhance the overall thermodynamic process used to destroy the cells and the tumors. It is thought that

cell death occurs not only from the NPs that are absorbed into the cells but also from the NPs attached to the cells and the extracellular medium.

Temperature changes of the NP powder upon exposure to RF radiation were studied in order to investigate the heating effects generated by the three different magnetic NPs. Five minutes of RF heating was considered sufficient in order to highlight the heating rates and the temperature differences between the three types of NPs. The high sensitivity thermal analysis indicated that the RF induced temperatures, as well as their heating rates, rose and were found to be dependent mostly upon the mass of the NPs used for the measurements. The C-Fe NPs were shown to be the best hyperthermia agents for killing the cancer cells since they were found to reach temperatures higher than the C-Fe/Co and C-Co NPs under identical RF exposure conditions (Figure 3c). Based on these experimental results and given the significant mass difference between the NPs present inside the cells and the cells themselves, the death of the cells is not expected to occur as a result of the bulk heating of the entire cell structures, but rather due to the localized damages of the cell such as nuclear membrane destruction, DNA fragmentation, and thermal denaturation of proteins (which happen at temperatures higher than 55 °C). As shown in Figure 4, after the NPs were taken up into the HeLa cell cytoplasm they were able to cross the various intercellular membranes and reach the nucleus.¹⁰ Due to the localized RF heating provided by the NPs, the cells were found to go through an apoptotic process, and subsequent cellular decomposition (one of the signs of the apoptotic process: the cell membrane blebbing) was observed as a result (Figure 4c). Compared to different NPs, as shown in Figures 4d–f, the C-Fe NPs revealed the highest efficiency for inducing cell death when compared to the C-Fe/Co NPs and C-Co NPs after two minutes of RF exposure. The disintegration of localized cellular environments such as nucleus, nuclear membranes, and DNA were believed to be the effective response resulting from RF-heat inducement into the NPs.

Magnetism of biological objects is negligibly small, so biologically compatible nontoxic magnetic NPs are used to strengthen the influence of an external magnetic field. Carbon-shelled magnetic NP heating happens under RF radiation and it is possible that the NPs undergo an “auto-heating” event whereby the NPs generate their own heat energy based on molecular rotation. After heating for 60 to 90 seconds, the NP powders were found to reach the heating equilibrium (as shown in the insert in Figure 3c). This relaxation corresponds either to the physical rotation of particles directly inside the

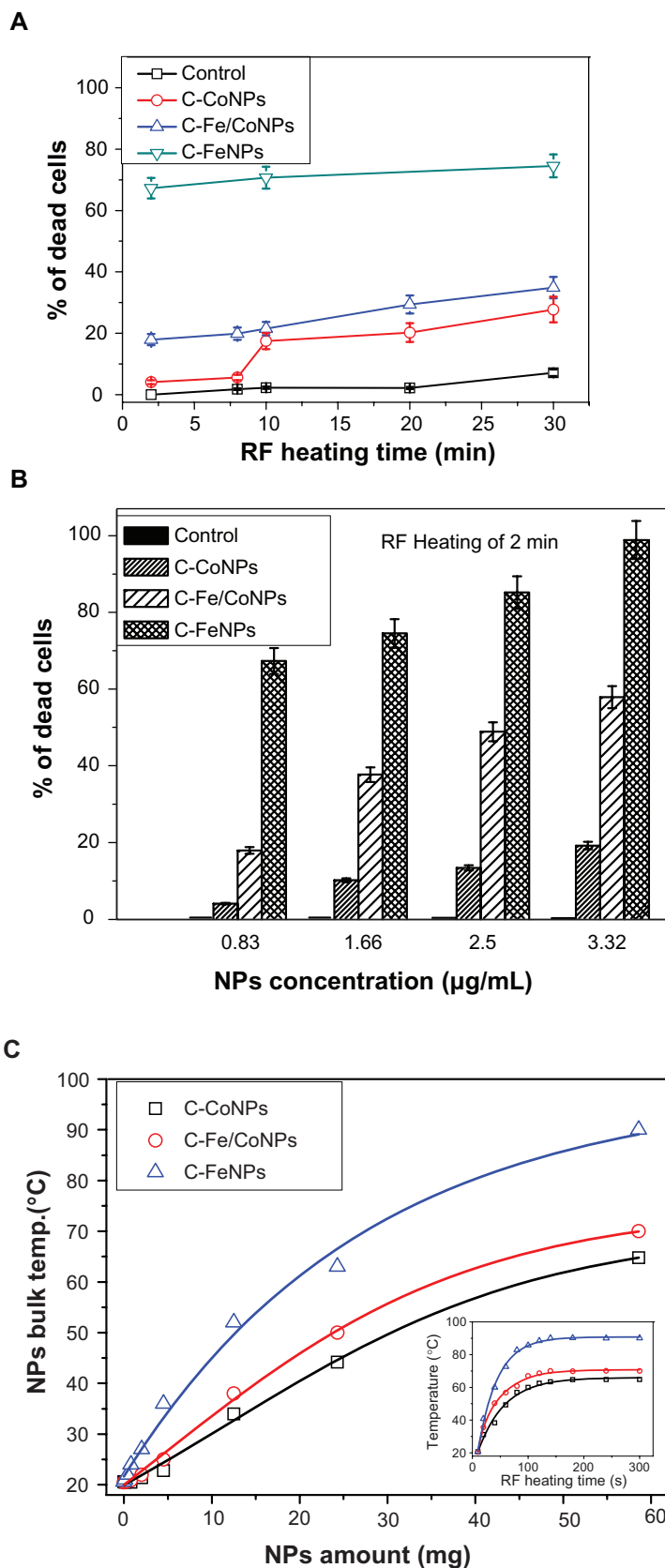


Figure 3 A) Cytotoxicity effects of the low concentration (0.83 µg/mL) C-Co, C-Fe/Co, and C-Fe NPs on the HeLa cancer cells after 2 to 30 minutes of RF exposure. **B)** Effect of different concentrations of the C-Fe, C-Fe/Co, and C-Co NPs on the HeLa cells that died from 350 kHz RF heating after two minutes of exposure time. **C)** Comparative RF-induced temperature variations as the function of different C-Fe, C-Fe/Co, and C-Co NP concentrations. Insert figure shows the temperature-rising characteristics of different magnetic NPs with the same amount under 350 kHz RF exposure.

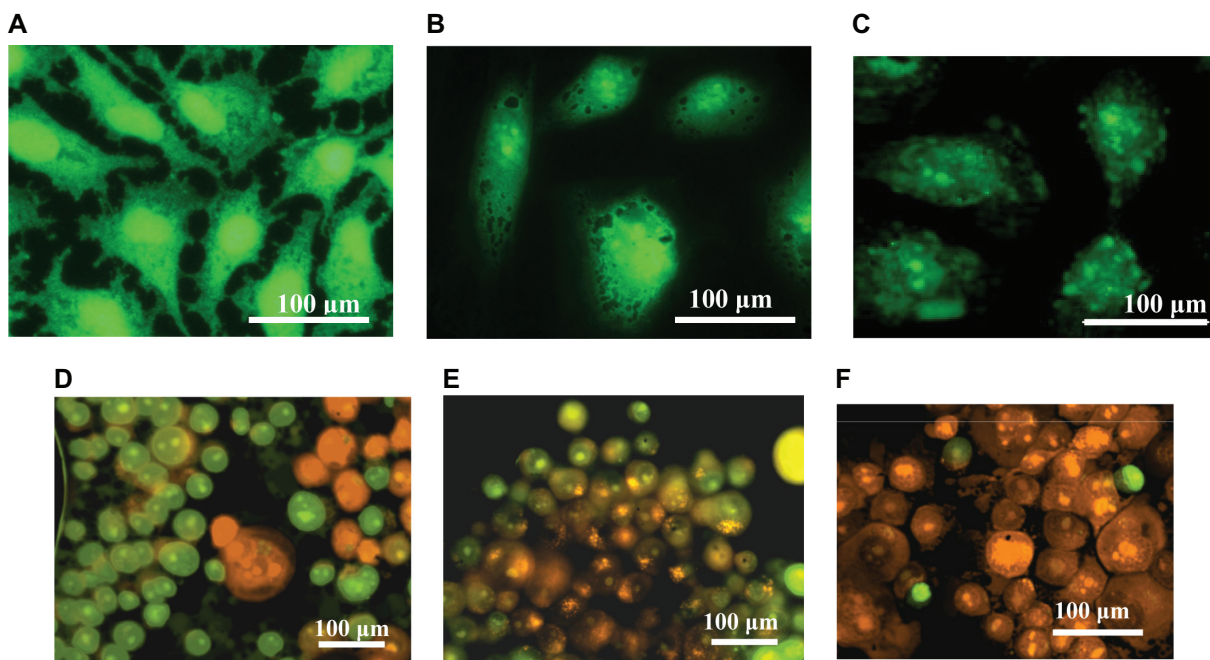


Figure 4 **A)** Normal HeLa cells without incubation of magnetic NPs; **B)** After 24 hours incubation time, the magnetic C-Co NPs were found to aggregate around and further penetrate into the nucleus of HeLa cells. **C)** The images took from the Olympus confocal microscopy indicating the membrane blebbing of HeLa cells incubated with magnetic C-Fe NPs after 350 kHz RF heating for two minutes. Confocal microscopy images indicating the extensive live and dead cells after being incubated with the C-Co NPs **D)** C-Fe/Co NPs **E)**, and C-Fe NPs **F)** after exposure to RF radiation for two minutes. The cells were stained in order to distinguish between the live (green for acridine orange) and the dead cells (orange for ethidium bromide).

liquid or to the rotation of nuclear magnetic moments inside each particle. Rotation of particles is usually referred to as Brownian rotation, and rotation of the magnetic moment inside each particle is called Néel relaxation. This “auto-heating” was attributed to the Néel relaxation²¹ of the spin of the NPs under low frequency (350 kHz) RF radiation. The Néel relaxation time, T_N , of the magnetic NPs under an external magnetic field is expressed by following formula²²:

$$T_N = \frac{\sqrt{\pi}}{2} T_0 \frac{\text{Exp}(KV_m / k_B T)}{(KV_m / k_B T)^{1/2}}$$

where T_0 is the relaxation time constant and has the order of 10^{-9} s, K is the anisotropy constant, and V_M is the magnetic volume of particles. The Néel relaxation time, T_N , was determined as the ratio of the energy of magnetic anisotropy of superparamagnetic particles to the thermal energy. The temperature of the three NPs increased rapidly in the initial stage (from 0–60 s) which can be seen from the curves in Figure 3c inset. After about 200 seconds, the final temperatures of 65 °C, 70 °C, and 90 °C were reached for Co, Fe/Co, and Co NPs, respectively. This is due to the thermally stable spin rotation of the ferromagnetic NPs caused by the RF induced hysteresis loss. These spin properties are attributed to the particle size for the C-Fe, C-Fe/Co, or C-Co NPs with diameters <10 nm as shown in Figure 5a.

In the RF-induced localized heating, there are two major heating mechanisms in the magnetic NPs. (1) Eddy currents: ferromagnetic nanomaterials are good conductors, and a solid core made from such a material also constitutes a single short-circuited turn throughout its entire length. Eddy currents therefore circulate within the core in a plane normal to the flux, and are responsible for resistive heating of the core material. (2) Hysteresis losses: Each time the magnetic field is reversed, a small amount of energy is lost due to hysteresis within the core. For a given core material, the loss is proportional to the frequency, and is a function of the peak flux density to which it is subjected.

Ferromagnetic NPs possess hysteretic properties when exposed to alternating magnetic fields, which gives rise to magnetically induced heating. The amount of heat (power dissipation) generated per unit volume, W , is given by the frequency multiplied by the area of the hysteresis loop in the M–H coordinates caused by hysteresis loss²²:

$$W = -\mu_0 f_{\text{appl}} \oint H \cdot dM$$

For ferromagnetic particles with sizes greater than those of superparamagnetic NPs, there is no explicit frequency dependence of the hysteresis loop area, so that the rate

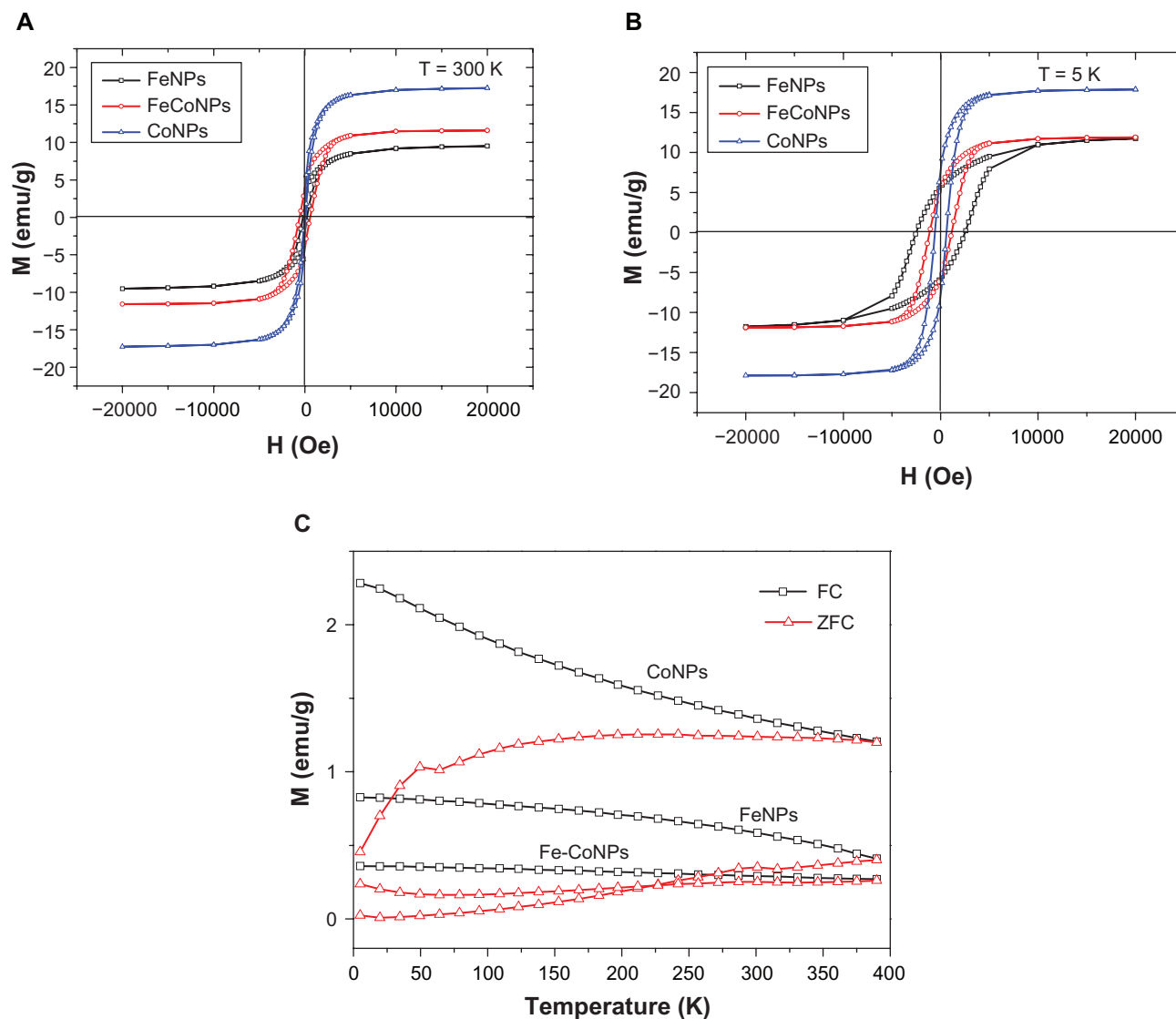


Figure 5 Magnetization curves of different magnetic NPs at 300 K **A**) and 5 K **B**); **C**) Temperature dependence of ZFC and FC measurements recorded at a magnetic field of 30 Oe for these magnetic NPs.

Abbreviations:

of heat emission can be determined from quasistatistical measurements of the hysteresis loop, for example, by means of SQUID magnetometry. Figure 5 compares magnetization loops measured at 5 K and 300 K for the C-Fe, C-Fe/Co, and C-Co NPs. The C-Co nanostructures revealed a larger saturation magnetic induction (M_s), larger residual magnetization (M_r), and smaller coercivity (H_c) relative to the C-Fe/Co and C-Fe nanostructures. One reason is the NP size distributions. The specific energy absorption multiplied by the particle density yields the thermal emission power in unit volume, W , which represents the efficiency of heating of the magnetic particles. This calculation allows NPs of various sizes to be compared as heat generators. The Fe and Co NPs in this work have sizes ranging from 7–15 nanometers.

For this size, the Fe NPs are about the size of the Fe domain (14 nm); however the Co NPs are much smaller than the Co domain size (70 nm).²³ Since the Fe NPs are composed most probably of single domains they exhibit the maximum possible coercivity for Fe, since the Co NPs which are much smaller compared to their domain size show a consequent diminished coercivity. This phenomenon is highlighted by the almost nonexistent hysteresis loss for the Co NPs. In the last decade, magnetic hyperthermia has been given a new impetus due to superparamagnetic liquids used as working agents.^{24,25} These are ferroliquids with one-domain magnetic NPs that are suspended in water and show superparamagnetic properties. When a ferroliquid is removed from the magnetic field, its magnetization vanishes because of the thermal energy of

the environment. The other reason is that the greater pairing of electrons in Co and the stronger nuclear charge of the Co atoms lead to less surface delta multibonding of Co surface atoms in the Co NPs relative the Fe NPs. Such Co-C bonding lowers the coercivity of Co which results in far softer magnetic properties relative to Fe NPs. It is noteworthy that the magnetic hysteresis of Co NPs is almost zero at 300 K, which indicates that the Co NPs are not agglomerated and are superparamagnetic. However, the Fe/Co and Fe NPs have more agglomerations, which were indicated in the larger hysteresis loops for these particles relative to Co NPs. As a result, Fe and Fe/Co NPs can generate more thermal energy when they were exposure to the external AC field compared to the Co NPs.

A ferromagnetic particle becomes “superparamagnetic” below a critical size on the order of 100 nm, depending on the material. Such a particle cannot develop internal magnetic domains and, therefore, acts as a paramagnetic particle with a magnetic moment of up to 10^7 Bohr magnetons. The blocking phenomenon is a signature of the superparamagnetic regime that depends on the particle size, degree of crystallinity, and interparticle interactions.²⁶ Each NP then becomes a single magnetic domain and shows superparamagnetic behavior when the temperature is above the so-called blocking temperature, T_B .²⁷ Below T_B , NPs are “blocked”, which means that initially random magnetic moments of individual NPs cannot readily align with the applied field, because magnetic Zeeman (H) and thermal fluctuation ($k_B T$) energies are insufficient to overcome the energy barrier set by the magnetic anisotropy and interparticle dipolar interactions. Experimentally, T_B is marked by the peak in the $M_{(T)}$ curve measured upon warming after a magnetic field is applied at a low temperature to a ZFC sample. It is well established that T_B is reduced for smaller particles and for particles with reduced crystallinity; both properties lead to the reduction of the magnetic moment per particle.²⁸ Another way to probe the magnetic response of a superparamagnetic assembly is to measure its magnetic moment at a fixed temperature as a function of the magnetic field. It is important to compare $M_{(H)}$ curves below and above T_B to show the absence of the magnetic hysteresis above T_B . Figure 5c shows the results of ZFC measurements performed upon warming after a magnetic field of 30 Oe was applied at 5 K after cooling in zero applied fields. Clearly, Co NPs exhibit the lowest blocking temperature about 50 K. Fe and Fe/Co NPs show an elevated T_B of 300 K. These results were fully consistent with the results of TEM; that is, Co NPs shown a relative smaller size and lower crystallinity than Fe and Fe/Co NPs.

Conclusions

This report demonstrated that Fe, Fe/Co, and Co magnetic NPs with several graphitic layers were successfully synthesized. They were introduced to HeLa cervical cancer cell cultures and were found to be taken up by the cells and acted as high-efficiency RF absorbers. Under low frequency (350 kHz) RF radiation for a short exposure time of two minutes, the lowest concentration (0.83 $\mu\text{g/mL}$) of C-Fe NPs was found to be the most suitable for inducing death in over 99% of the cancer cells. The RF-driven heating of the NPs is believed to be responsible for the generation of extremely localized damage inside and around the cells. The process described in this research has significant promise for various medical therapies and efficient ablation of tumors. A further continuation of this work involving the binding of these magnetic NPs to antibodies, proteins, and growth factors will lead to their specific delivery to the individual cancer cells, causing comparable thermal ablation of cancer cells while diminishing the potential toxicity of these entities to normal cells. This research should also advance towards finding more RF-sensitive nanomaterials materials that are able to provide thermal ablation of cancer cells, but in very low concentrations that avoid the possible cytotoxic and undesirable effects that these materials could induce in healthy tissues.

Acknowledgments

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