


Simultaneous Optimization of Oral and Transdermal Nanovesicles for Bioavailability Enhancement of Ivabradine Hydrochloride

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Purpose: Ivabradine hydrochloride is selective pacemaker current (I_f) ion channel inhibitor used in case of chronic heart failure (CHF) with superior efficacy and lower side effects than most β -blockers. However, the drug suffers from low bioavailability ($\approx 40\%$) due to extensive first-pass metabolism. Hence, this work aims to formulate nanovesicular platforms to enhance their bioavailability both orally and transdermally.

Materials and Methods: A central composite face-centered design was employed to formulate the nanovesicles, both phosphatidylcholine: drug ratio and percentage of pluronic F68 were used as independent variables. The nine developed formulae were characterized in terms of vesicle size (nm), polydispersity index, zeta potential (mV), entrapment efficiency (%). Decreasing vesicle size, increasing negative value of the zeta potential, and increasing entrapment efficiency were the chosen constraints to optimize the engineered nanovesicles. The candidate formula was subjected to further investigation including lyophilization, loading into carbopol gel, in vitro release, imaging with a transmission electron microscope, histopathological examination, in vitro cytotoxicity study and in vivo pharmacokinetics.

Results: The optimized nanovesicular formula was composed of lipid: drug ratio of 3.91:1 and 100% pluronic as a stabilizer. It has particle size, zeta potential and entrapment efficiency of 337.6 nm, -40.5 mV and 30.5, respectively. It was then lyophilized in the presence of 5% trehalose as a cryoprotectant, dispersed in 0.5% carbopol to develop the transdermal gel. The two different forms of the candidate formula (lyophilized and gel form) displayed sustained drug release in comparison to drug solution. The histopathological and cytotoxicity studies showed that the optimized formula was safe and highly biocompatible. The pharmacokinetics parameters measured declared a higher C_{max} and half-life of both formulae in comparison to market product (Procoralan[®]) with a 2.54- and 1.85-folds increase in bioavailability, respectively.

Conclusion: Hence, the developed nanovesicles can be reported as the first nanoplatforms to be used for simultaneous ivabradine delivery by both oral and topical routes with enhanced oral and transdermal drug delivery. The developed nanoplatforms hence can be further used to formulate other drugs that suffer from low bioavailability due to extensive first-pass metabolism.

Keywords: ivabradine, oral, transdermal, central composite, lyophilized, extensive first pass

Introduction

Chronic heart failure (CHF) is a critical and serious disease affecting up to 3% of the peoples in advanced countries.¹ Reducing the heart rate (< 70 beats per min) is highly desired to reduce mortality in CHF patients. Ivabradine was first authorized by the European Medicines Agency in 2005 and then approved by the United

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States Food and Drug Administration in 2015.^{2,3} It acts as a selective inhibitor to the I_f ion channels of the sinoatrial node which leads to lowering the heart rate without affecting the cardiac conductivity and repolarization.⁴ Moreover, it is superior over β -blockers in being more effective with fewer side effects, in addition to being capable of improving exercise tolerance.⁵ The drug has excellent physicochemical characteristics including its high solubility (50 mg/mL), suitable partition coefficient (3.17) and small molecular weight (468.6 g/mol).^{6,7} Unfortunately, ivabradine has significantly low oral bioavailability (\approx 40%) due to its extensive first-pass metabolism that yields inactive metabolites.⁸ Ivabradine is a substrate for the CYP3A4 enzyme system which is mainly present in the gut and the liver. The only active metabolite is N-demethylated derivative which is formed in an amount equivalent to 40% of the parent compound.⁷ Moreover, it suffers from a rapid elimination rate with a significantly short elimination half-life (2 h) but still its dose (2.5–7.5 mg) and dosing frequency (two times daily) remain acceptable and does not affect patient compliance.⁹

To avoid the extensive first-pass effect, several trials were performed to deliver ivabradine with different routes that could avoid passing through the small intestine including transdermal and buccal applications.^{10,11} Ivabradine was loaded into transdermal patches composed of hydroxypropyl methylcellulose K15 and ethyl cellulose (2:1 w/w). These patches achieved a controlled release and sustained pharmacokinetic profile after application on Wistar rat skin.¹² Propylene glycol, isopropyl myristate and oleic acid were incorporated by Banu et al into the transdermal patches to further enhance the drug permeation.¹³ Recently, Balata et al developed transfersomes loaded with ivabradine utilizing either tween 80, sodium lauryl sulfate or cetrимide as stabilizers and permeation enhancers mixed in different ratios with the drug and phospholipid.⁶ The optimized transfersomal formula was incorporated into a film composed of hydroxypropyl methylcellulose and starch in ratio 1:1 w/w. The developed nanostructured transdermal film showed improved permeability and retention compared to the unformulated ivabradine. On the other hand, Lodhi et al tried to develop mucoadhesive buccal films loaded with ivabradine.¹⁴ The developed buccal films showed a good *ex vivo* permeation through the porcine buccal membrane, but the study lacked the comparison with the pure drug as a reference.

This study aimed to develop and optimize lipid-based nanovesicles loaded with ivabradine and stabilized by either Pluronic F68, span 60 or a mixture of both with different ratios. These nanovesicles could be able to increase the extent of drug absorption after oral administration through bypassing the hepatic uptake and metabolism due to their built-in lipophilicity and small particle size.^{15,16} Moreover, the developed nanovesicles were embedded in a gel and then topically applied to ensure their abilities to enhance the transdermal permeation of ivabradine depending on the utilized surfactants that could act as potential permeation enhancers.^{17,18} The excipients used to develop the nanovesicles are significantly cheaper than polylactic co-glycolic acid and D- α -tocopherol polyethylene glycol 1000 succinate used in a previous study. Moreover, developing a single formulation for both oral and transdermal delivery of drugs could save time and effort, and facilitate the way for developing a universal formulation for a wide range of applications.

Materials

Ivabradine hydrochloride was supplied by Eva Pharma, Cairo, Egypt. Cholesterol was purchased from Acros Organics, NJ, USA. Phosphatidylcholine (PC), Pluronic F68 (PL) and span 60 (SP) were purchased from Sigma-Aldrich, St. Louis, MO, USA. The remaining chemicals and solvents were of analytical grade used without further processing or purification.

Experimental Design

The study was fully designed by Design ExpertTM software (ver. 7; Stat-Ease Inc., MN, USA) with a central-composite response-surface statistical analysis. PC was utilized as a builder for the nanovesicular walls in different ratios to the drug starting from 0.5:1 w/w to 4:1 w/w (A: PC/D). Moreover, SP and PL were used as stabilizers in different ratios with a total amount of 200 mg (B: PL%: 0–100%). On the other hand, the vesicle size (Y_1) of the prepared nanovesicles was measured along with their polydispersity index (Y_2), zeta potential (Y_3) and entrapment efficiency (Y_4). The composition and all measured responses of the prepared nanovesicles are listed in Table 1. The statistical models were validated through the determination of their R^2 , lack of fit and adequate precision. A difference less than 0.2 between the adjusted and the predicted R^2 values, a non-significant lack of fit and an adequate precision of more than 4 could indicate the capability of the model to predict values and navigate the design space with a minimal lack of fit.^{19–22}

Table 1 Experimental Runs, Independent Variables and Measured Responses of Ivabradine Nanovesicles Following a Central Composite Design ^{a,b}

Formulae	PC/D Ratio ^c	PL %	Vesicle Size (nm)	Polydispersity Index	Zeta Potential (mV)	Entrapment Efficiency (% w/w)
N1	0.5	0	1615.5± 8.3	0.801 ± 0.03	-32.85± 1.4	24.82 ± 1.1
N2	0.5	50	1383.0 ± 3.1	0.807 ± 0.01	-34.28± 2.7	16.27 ± 1.8
N3	0.5	100	545.7± 1.8	0.609± 0.05	-35.70 ± 2.1	6.48 ± 0.4
N4	2.25	0	610.0± 2.7	0.676± 0.02	-40.15± 3.9	31.14± 3.9
N5	2.25	50	659.7± 3.0	0.714 ± 0.02	-40.80 ± 0.5	21.61 ± 1.6
			613.0 ± 3.6	0.728± 0.00	-41.85± 0.6	21.79 ± 1.2
			486.2± 0.8	0.561± 0.04	-39.85± 1.0	21.46 ± 2.5
			539.2± 4.2	0.513 ± 0.01	-39.80 ± 0.6	27.33 ± 0.3
			500.0 ± 6.2	0.600 ± 0.07	-40.00 ± 0.8	24.00 ± 0.1
N6	2.25	100	216.0 ± 1.5	0.702± 0.01	-38.85± 1.8	14.57 ± 2.0
N7	4	0	575.5 ± 7.9	0.598± 0.00	-42.20 ± 1.5	31.78 ± 1.1
N8	4	50	513.3 ± 8.8	0.608± 0.05	-43.28± 3.3	30.16± 3.7
N9	4	100	352.9 ± 0.7	0.679 ± 0.03	-40.05± 3.1	33.47± 1.7

Notes: ^aAll the formulae prepared using 100 mg of ivabradine. ^bValues are expressed as mean ± SD; n = 3. ^cCholesterol is added at a constant percent of PC (5%).

Preparation of Ivabradine Nanovesicles

The ivabradine nanovesicles were prepared by the coacervation phase separation method.^{23,24} SP, PL, PC, cholesterol (5%w/w of PC), and ivabradine (100 mg) were accurately weighed and dissolved in 0.25 mL absolute ethanol in a stoppered 10 mL glass vial. Then, the vial was incubated in a thermostatic shaking water bath with the temperature adjusted at 60 °C and a stirring rate of 1000 rpm for at least 5 min to ensure that all ingredients were completely dissolved. Afterward, distilled water (0.1 mL), preheated at 60 °C, was added and mixed with the alcoholic solution on the same water bath until a clear mixture was formed. Finally, the prepared formulae were allowed to cool down to room temperature before being stored in the refrigerator at 4 °C until further characterization.

Evaluation of Ivabradine Nanovesicles

Determination of the Vesicle Size, Polydispersity Index and Zeta Potential

Zetasizer Nano ZS (Malvern Panalytical Ltd, Malvern, UK) adopting the dynamic light scattering technique was employed to measure the vesicle size of the prepared ivabradine nanovesicles. Before analysis, each formula was

hydrated with 10 mL distilled water in a bath sonicator (Clifton™ Heated Timed Ultrasonic Bath, Nickel Electro Ltd, Weston-super-Mare, UK) and then, a 1 mL sample was transferred to a folded capillary zeta cell to measure the vesicle size, polydispersity index and zeta potential. All measurements were performed thrice at 25 °C with a count rate of 200–400 kilo counts per second (Kcps) and the mean values ± standard deviations were recorded in Table 1.

Determination of the Entrapment Efficiency Percentages

The amounts of ivabradine entrapped in the prepared nanovesicles were indirectly determined after diluting each formula with 10 mL distilled water and centrifuging it at 15,000 rpm and a temperature 4 °C for 60 min in a cooling centrifuge (Mikro 200 R, Hettich Lab, Tuttlingen, Germany). The supernatant was then separated, diluted with distilled water, and spectrophotometrically analyzed at ivabradine λ_{max} (284 nm). Finally, the entrapment efficiency percentages were calculated based on the following equation:

$$\text{Entrapment efficiency} = \frac{TA - SA}{TA} \times 100\% \quad (1)$$

where TA is the total amount of ivabradine in each formula while SA is the amount of ivabradine in the supernatant.

Desirability and Optimization

To statistically optimize the composition of the prepared ivabradine nanovesicles, the desirability values were calculated to find the formula with the lowest vesicle size, polydispersity index, and highest zeta potential and entrapment efficiency.²⁵ The desirability value allows merging all the dependent responses in one measurement and concluding the superlative levels of the independent parameters. It is determined by first calculating each desirability function.²⁶

Responses (Y) are maximized as follows

$$d_i = \frac{Y_i - Y_{min}}{Y_{max} - Y_{min}} \quad (2)$$

and minimized as follows

$$D_i = \frac{Y_{max} - Y_i}{Y_{max} - Y_{min}} \quad (3)$$

Where Y_{min} , Y_{max} and Y_i are the minimum, maximum, and individual response values, respectively.

The overall desirability values were calculated from the individual values by using the following equation for n responses:

$$D = (d_1 \times d_2 \dots \dots d_n)^{\frac{1}{n}} \quad (4)$$

The formula with the optimum composition was prepared and subjected to the same characterization tests. The obtained findings were compared to the predicted values and the percentage errors were calculated to ensure the prediction capabilities of the employed statistical models.^{27,28}

Freeze-Drying of the Optimized Ivabradine Nanovesicles

The optimized ivabradine nanovesicles were lyophilized either with and without mannitol or trehalose as cryoprotectants, each in a concentration of 2.5% and 5%. The formulae were stored overnight in the freezer at a temperature -20 °C and then, transferred to Novalyphen NL 500 freeze-dryer (Savant Instruments Corp., NY, USA). The freeze-drying process continued for 24 h at a temperature of -45 °C and a pressure of 7×10^{-2} mbar.^{29,30} The freeze-dried formulae were kept in a desiccator until being reconstituted with distilled water and investigated for their vesicle size, polydispersity

index, zeta potential and entrapment efficiency. The new findings were compared to the respective results obtained before freeze-drying.

Preparation and Characterization of Ivabradine Nanovesicular Gel

The freeze-dried optimized ivabradine formula was reconstituted with 10 mL distilled water and then, carbopol 940 was added to the prepared nanodispersion to give a concentration of 0.5% w/v. The mixture was stirred overnight to ensure hydration of carbopol before being converted to gel using tri-ethanolamine added in a ratio of 1:1 w/w to carbopol.^{31,32} The prepared nanogel was characterized by studying its rheological properties.

Brookfield viscometer (Model No. HBDV-I, MA, USA) adopting a CPE-41 spindle was utilized to measure the viscosity and the shear stress (τ , dyne/cm²) of the prepared ivabradine gel at a different rate of shear ($\dot{\gamma}$, s⁻¹), ranged from 2 to 100 rpm within the acceptable ranges of torque percentages. The flow index (n) was calculated based on the power model (Eq. 5) and used to determine the flow behavior of the prepared nanogel if it is Newtonian ($n \approx 1$), shear-thinning ($n < 1$) or shear thickening ($n > 1$).³³

$$\tau = K\dot{\gamma}^n \quad (5)$$

where K is the consistency index (dyne/cm².sⁿ).

In vitro Ivabradine Release

The release profiles of ivabradine from the optimized nanovesicles, before and after freeze-drying and after geling, were investigated and compared to the dissolution profile of the plain drug solution. A sample equivalent to 6 mg ivabradine was withdrawn from each formula and placed in a dialysis bag with 12–14 kDa cut-off (Sigma-Aldrich, St. Louis, MO, USA) immersed in 25 mL phosphate buffer (pH 7.4).³⁴ The dissolution medium was stirred at 100 rpm and heated at 37 °C for the drug solution, the optimized formula and the lyophilized one and at 35 °C for the nanovesicular gel using a thermostatic shaking water bath (Model No. SBS40, Stuart, Staffordshire, UK). Samples of 1 mL each were withdrawn at the pre-planned time intervals; 0.25, 0.5, 1, 2, 3, 4, 6 and 8 h. The ivabradine concentrations in the withdrawn samples were measured using a UV spectrophotometer at λ_{max} of 284 nm. The whole dissolution experiment was performed three times for each formula and the mean percentage dissolved \pm standard deviations were calculated and plotted against its respective time interval. The similarity

factor (f_2) was utilized to compare the ivabradine dissolution profiles after being calculated using the following equation:^{35,36}

$$f_2 = 50 \times \log \left\{ \left[1 + \left(\frac{1}{n} \right) \sum_{j=1}^n |R_j - T_j|^2 \right]^{-0.5} \times 100 \right\} \quad (6)$$

where the percentages of ivabradine released at each time interval (j) were R and T for the reference and the test formulae, respectively, and the number of samples employed in the calculations was n. The dissolution half-life ($T_{50\%}$) values were calculated and utilized to compare the drug release profiles.

Imaging of the Optimized Ivabradine Nanovesicles

The optimized nanovesicles were visualized under JEM-1230 transmission electron microscope (Jeol, Tokyo, Japan), operated at a voltage of 100 kV. Before imaging, sample was withdrawn, sprayed over a copper grid coated with carbon, stained with 2% phosphotungstic acid and finally, dried at room temperature.

Effect of Storage on Ivabradine Lyophilized Nanovesicles

The lyophilized formula (L5) was assessed following storage in a cold place (refrigerator 2–8 °C) for six months.³⁷ At the end of the storage period, the lyophilized nanovesicles were evaluated for their appearance, vesicle size, polydispersity index, zeta potential and entrapment efficiency. Statistical analysis of the obtained results was performed by Student's *t*-test using SPSS 17.0 software (SPSS Inc., Chicago, IL). Difference at *p*-value < 0.05 was considered significant.

Histopathological Examination

To assess the possible irritation effect of the prepared lyophilized nanovesicular gel, the nanovesicular gel formula was applied to shaved back skin of a rabbit in an area of 5 cm² diameter for three consecutive days. Another area was shaved and left untreated as a control. Afterward, the rabbit was euthanized, skin samples from treated and untreated areas were collected, and stored in 10% formalin. The samples were then dehydrated using ethanol, fixed in paraffin and stained with eosin and hematoxylin. Sections of 5 μm were cut using a microtome and examined under a light microscope.²⁵

In vitro Cytotoxicity Study

In flat bottom 96 well-microplates, HepG2 cells (Human liver adenocarcinoma cells) (0.5×10^5) were cultured in 180 μL/well RPMI media supplemented with 10% fetal bovine serum, 2 μmol/mL L-glutamine, 250 ng/mL fungizone, 100 units/mL penicillin-streptomycin solutions at 37°C in a CO₂ incubator.^{38,39} The plates were incubated for 24 h at 37°C in a humidified 5% CO₂ atmosphere to allow cells to settle down. After incubation, the media were removed, and 180 μL/well fresh serum-free medium was added to each well. Cells are then treated with 20 μL of different concentrations of either the drug solution or the lyophilized nanovesicular formula (500–15.62 μM). The plates were then incubated for 24 h at 37°C in a humidified 5% CO₂ atmosphere. After incubation, the media were removed and MTT solution 40 μL/well was added and incubated for an additional 4 h. MTT crystals were solubilized by adding acidified isopropanol (160 μL/well) and the plate was shaken at room temperature. This was followed by photometric determination of the absorbance at 570 nm using the microplate ELISA reader (FLUOstar Omega, BMG, Labtech, Germany). The absorbance of the resulting color is directly proportional to the number of viable cells in each sample. The percentage of relative viability was calculated using the following equation:

$$\text{Percentage of relative viability} = \frac{\text{Absorbance of treated cells}}{\text{Absorbance of control cells}} \times 100 \quad (7)$$

Statistical analysis was done for both drug solution and lyophilized nanovesicles in comparison to normal untreated cells using Student's *t*-test employing SPSS 17.0 software (SPSS Inc., Chicago, IL). The difference at *p*-value < 0.05 was considered significant.

Oral and Transdermal Bioavailability of Ivabradine

Animals

Nine New Zealand male rabbits (2–3 kg each) were involved in the study. Each rabbit was hosted in a separate cage where the humidity and temperature were maintained at $55 \pm 5\%$ and 22 ± 2 °C, respectively. Equal daily cycles of light and dark circumstances were maintained according to the Guide for the Care and Use of Laboratory Animals.⁴⁰

Study Design

The procedure was reviewed and approved by the ethical committee of Faculty of Pharmacy, Cairo University with an application number of PI 2653. The rabbits were randomly assigned to three groups where the first group administered ivabradine market product (Procoralan[®]) and the second group administered the freeze-dried optimized nanovesicles orally while the optimized nanovesicular gel was applied to the shaved back skin of the rabbits assigned to the third group. A randomized three-period crossover design was applied to eliminate subject-to-subject variation with a washout period of a week.⁴¹ The administered dose (0.26 mg/kg) was calculated from the human dose after an adjustment based on the difference in the body surface area according to the following equation.⁴²

$$\text{Animal dose(mg/kg)} = \text{Human dose(mg/kg)} \times \frac{\text{Human } k_m}{\text{Animal } k_m} \quad (8)$$

where

$$k_m = \frac{\text{Average body weight (kg)}}{\text{Body surface area (m}^2\text{)}} \quad (9)$$

Blood samples, 2 mL each, were withdrawn from the tail vein at 0.25, 0.5, 0.75, 1, 1.5, 2, 4, 6, 8 and 24 h following oral administration or topical application. Plasma was separated by centrifugation at 5000 rpm at 5 °C for 20 min and stored in a freezer at – 20 °C until the time of analysis.

Determination of Ivabradine in Plasma

Plasma samples were thawed at room temperature and then mixed in a ratio of 10:1 v/v with a 200 ng/mL alcoholic solution of the internal standard (clopidogrel). Ivabradine was extracted using methanol added in a ratio of 2:1 v/v to the plasma samples. The extracted drug was dried under vacuum and reconstituted with a mobile phase composed of methanol and 0.2% w/v aqueous solution of formic acid mixed in a ratio of 7:3 v/v, respectively. The samples were injected into an LC/MS/MS (Shimadzu, Tokyo, Japan) equipped with a Poroshell column (120EC-C18, 4.6 x 50 mm, 2.7 μm, Agilent Technologies, DE, USA) and a mobile phase running at a flow rate of 0.6 mL/min. The analysis method was validated with linearity ($R^2 = 0.9998$) and a lower limit of detection of 0.2 ng/mL. The equipment was operated with a positive ion mode, an accelerator cell voltage of 7 V and a dwell of 200 s. Ivabradine and clopidogrel were quantified at

a precursor-to-product ion ratios of 469/177 and 322/212, respectively.

Pharmacokinetic and Statistical Analysis

The in vivo results were processed using noncompartmental pharmacokinetic analysis utilizing Kinetica[®] software (Version 5, Thermo Fisher Scientific Inc., MA, USA) to determine the key pharmacokinetic parameters including the maximum concentration of ivabradine in plasma (C_{max}), its corresponding time (T_{max}), the area under the curve from zero to the last time AUC_0^{24} , the area under the curve from zero to infinity (AUC_0^∞) and the relative bioavailability compared to the oral market product. The results were expressed as the mean of 9 rabbits ±SD. The pharmacokinetic parameters were compared using the crossover ANOVA test. The nonparametric-signed ranks test (Wilcoxon) was used to compare the median values of T_{max} . A p -value < 0.05 was considered statistically significant. Statistical analysis was performed using SPSS 19.0[®].

Results and Discussion

Evaluation of Ivabradine Nanovesicles Determination of the Vesicle Size, Polydispersity Index and Zeta Potential

The vesicle size of the prepared ivabradine nanovesicles ranged from 216.0 (N6) to 1615.5 (N1) nm, as shown in Table 1 and Figure 1A. The results were subjected to statistical analysis using a quadratic model after being converted to their square root values to increase the validity of the statistical analysis.^{43,44} The model was assessed for its capability to cover the design space with an adequate precision of 23.32 and a limited gap between the adjusted (0.9502) and predicted (0.8596) R^2 values, ie less than 0.2.⁴⁵ Moreover, the model was statistically fitting the vesicle size values with a non-significant lack of fit (p -value = 0.5309).⁴⁶ The following quadratic equation correlates the vesicle size findings with the studied independent parameters:

$$\sqrt{Y_1} = 23.54 - 5.89A - 5.34B + 2.91AB + 6.59A^2 - 3.64B^2 \quad (10)$$

Significant inverse relationships were observed between the two independent factors, ie PC/D ratio and PL%, and the vesicle size with p -values < 0.001. The effect of PC was clear at the low level of PL where SP was the predominant component inside the prepared nanovesicles. SP nanovesicles have more tendency to aggregate than the corresponding liposomal structures that are mainly built

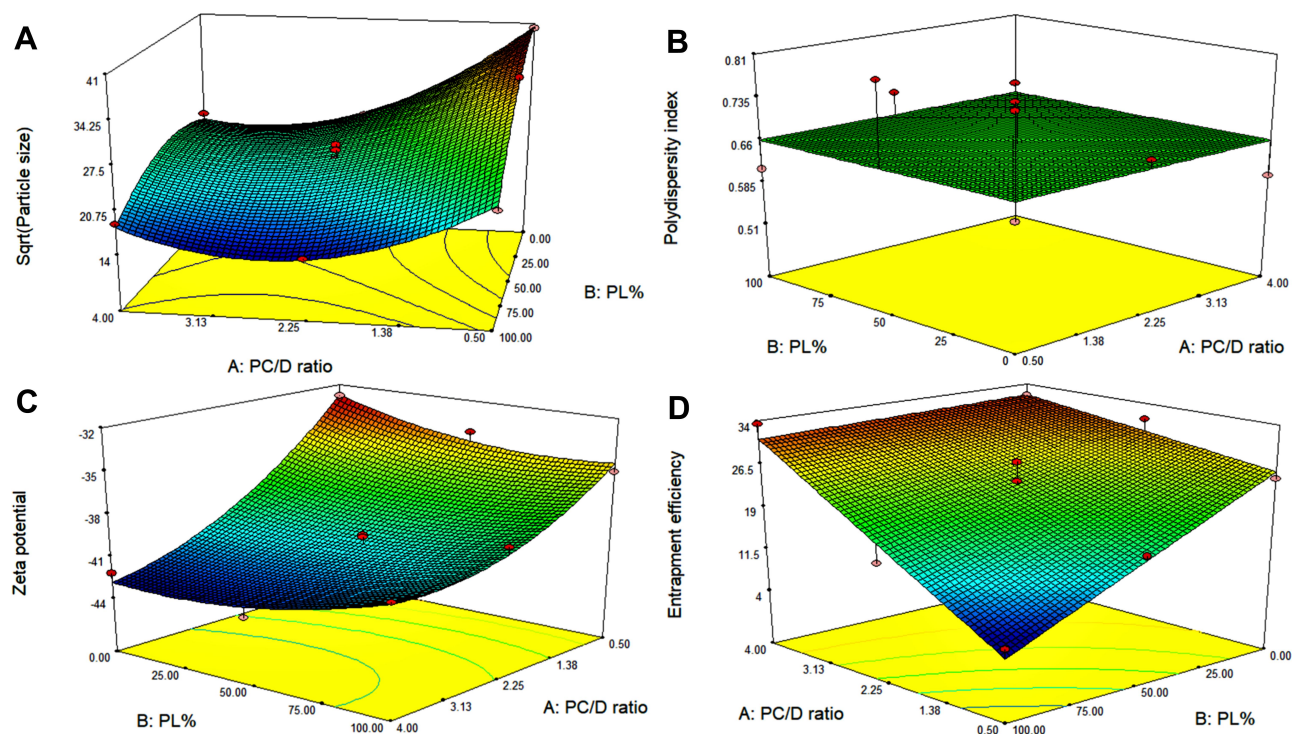


Figure 1 Three-dimensional response surface plots for the effects of PC/D (X₁) and PL% (X₂) ratios on the vesicle size (A), polydispersity index (B), zeta potential (C) and entrapment efficiency (D) of the prepared ivabradine nanovesicles.

out of PC and cholesterol.⁴⁷ Consequently, increasing PC and cholesterol could significantly reduce the size of the prepared nanovesicles. On the other hand, increasing PL% significantly reduced the vesicle size due to its ability to reduce the surface tension between the lipophilic components, ie PC, SP and cholesterol, and the surrounding aqueous phase.^{48,49}

The polydispersity index of the prepared nanovesicles ranged from 0.512 to 0.806, as shown in Table 1 and Figure 1B, indicating their polydisperse nature. This could be referred to the presence of a mixture of polymers that might have a tendency to separate and form two PC-based nanovesicles and PL-based micelles. A similar co-existence of micelles and liposomes was previously reported and imaged by Ogunsola et al.⁵⁰ The studied factors had no statistically significant effect on the polydispersity values as confirmed by the high *p*-value (0.7786).

The prepared nanovesicles had highly repulsive surfaces due to their zeta potential values which ranged from -32.85 and -43.28 mV, as shown in Table 1 and Figure 1C. The obtained values were found to be fitting the quadratic model with a *p*-value of 0.002. The model had been assessed for its capability to predict the zeta

potential upon modulating the studied factors via calculating its adequate precision (16.68), adjusted (0.9178) and predicted (0.7441) R² values. The following equation correlates the zeta potential values with the two investigated factors:

$$Y_3 = -40.48 - 3.78A + 0.10B + 1.25AB + 1.74A^2 + 1.02B^2 \quad (11)$$

Based on the factorial ANOVA, the PC/D ratio had a statistically significant effect on the zeta potential values (*p*-value < 0.0001) where the latter decreased by increasing PC. This could be logically attributed to the negative charge on the PC surfaces that could increase the negative value of the zeta potential which is mathematically considered a decrease.⁵¹ On the other hand, PL% had no statistically significant effect on the zeta potential values and this might be referred to the non-ionic nature of the two incorporated surfactants.⁵²

Determination of the Entrapment Efficiency Percentages

The drug entrapment percentages within the prepared nanovesicles ranged from 6.48 to 33.47%, as shown in Table 1 and Figure 1D. This range was considered relatively low compared to the values obtained by Sharma

et al (55.3–74.9%) who prepared ivabradine polymeric nanoparticles. These nanoparticles were prepared using the w/o/w emulsification technique which could prevent the drug migration to the external aqueous phase during preparation and consequently yielded high entrapment efficiency.⁵³ A two-factor interaction was adopted to statistically analyze the entrapment efficiency values with a p -value < 0.0001. The model had a significantly high adequate precision (19.88) and a difference of less than 0.2 between the adjusted and predicted R^2 values. The effects of the studied factors on the entrapment efficiency were established based on the following equation:

$$Y_4 = 23.45 + 7.79A - 5.54B + 5.01AB \quad (12)$$

Opposite impacts on the entrapment efficiency values were observed upon increasing PC and PL with p -values of < 0.0001 and 0.0004, respectively. The entrapment efficiency was directly proportional with the PC ratio while it was inversely proportional with the PL%. Increasing PC could enhance the formation of multilamellar nanovesicles which might be able to entrap more ivabradine within their layered membranes.⁵⁴ On the other hand, increasing PL% could increase the possibility of micellar formation to which a part of the drug might be partitioned from the original nanovesicles.⁵⁰

Desirability and Optimization

The relation between the desirability values and the studied independent factors is demonstrated in Figure 2. The optimized nanovesicular formula was composed of PC in a ratio of 3.91:1 w/w to the drug, cholesterol (20 mg), in addition to 200 mg PL as the only stabilizer without SP. This composition had the highest desirability value (0.869)

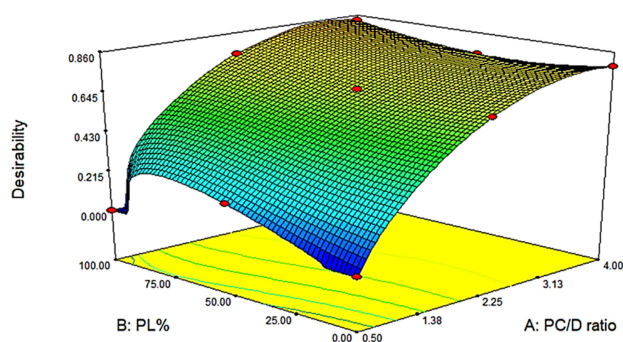


Figure 2 Three-dimensional response surface plot for the effects of PC/D (X_1) and PL% (X_2) ratios on the desirability of the prepared ivabradine nanovesicles.

calculated based on the optimization criteria fed into Design Expert™ software, ie the lowest vesicle size, the highest negative value of the zeta potential and the highest entrapment efficiency. The statistical model predicted that the optimized nanovesicular formula would have vesicle size, zeta potential and entrapment efficiency of 326.53 nm, -40.19 mV and 30.21%. After preparing the formula and measuring its physical characteristics (vesicle size, zeta potential and entrapment efficiency), the actual findings (337.6 nm, -40.5 mV and 30.5%) were found to be in a harmony with the predicted values with deviation percentages of less than $\pm 10\%$.

Characterization of the Freeze-Dried Optimized Ivabradine Nanovesicles

After lyophilization, all formulae were reconstituted to their original volume with sonication for 2 min. Samples were taken from each formula to identify the effects of different cryoprotectants on retaining the original characteristics of the optimized nanovesicular dispersion. Maintaining vesicle size was the main role of the added cryoprotectants through avoidance of the nanovesicular aggregation during lyophilization.^{55,56} As shown in Table 2, Formula L5 containing 5% trehalose showed lower vesicle size to the optimized formula before lyophilization, where the entrapment efficiency was not statistically significant in comparison to the optimized formula without cryoprotectant (L1) and higher than other cryoprotectants used (L2, L3, L4). Trehalose was commonly considered as one of the best cryoprotectants used to preserve the size of different nanoparticles during lyophilization.⁵⁷ Similar results were reported by Campos et al during the lyophilization of nimesulide solid lipid nanoparticles and by Siri et al during the lyophilization of albumin nanoparticles.^{58,59} Finally, the formula L5 was selected to be orally administered by the rabbits during the bioavailability study. It is worth mentioning that Bosch et al proved that trehalose prevents the aggregation of exosomes, where vesicle size showed a significant decrease after lyophilization in the presence of trehalose.⁶⁰

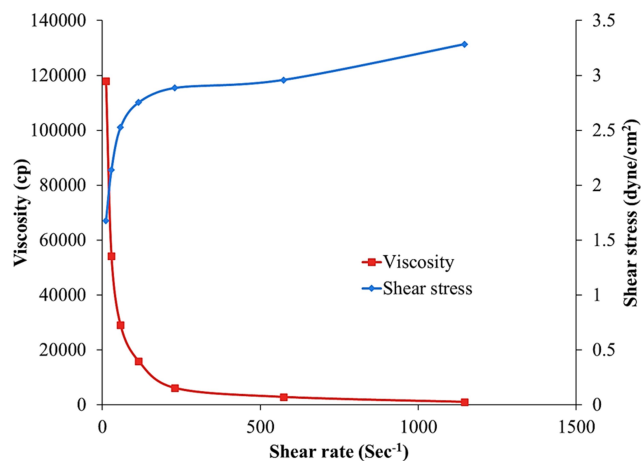
Characterization of Ivabradine Nanovesicular Gel

The nanovesicular gel had a viscosity of 120,000 cp which abruptly reduced by increasing the rate of shear, as shown in Figure 3. This indicated a non-Newtonian pseudoplastic flow behavior confirmed by the power law model that had a flow index (n value) of 0.1321. Carbopol is a common gelling

Table 2 Effect of Lyophilization on Vesicle Size, Zeta Potential, Polydispersity Index and Entrapment Efficiency of Ivabradine Nanovesicles^a

Formulae	% Cryoprotectant	Cryoprotectant Type	Vesicle Size (nm)	Zeta Potential (mV)	Polydispersity Index	Entrapment Efficiency (%)
L1	0	–	387.0 ± 32.43	-35.80 ± 0.42	0.466 ± 0.09	28.27 ± 3.36
L2	2.5	Mannitol	537.4 ± 12.50	-36.35 ± 2.05	0.634 ± 0.11	22.12 ± 2.12
L3	5	Mannitol	426.9 ± 47.80	-33.50 ± 1.84	0.511 ± 0.10	21.54 ± 1.82
L4	2.5	Trehalose	426.3 ± 18.67	-33.75 ± 0.35	0.623 ± 0.20	21.08 ± 2.25
L5	5	Trehalose	195.7 ± 8.49	-36.70 ± 0.71	0.466 ± 0.02	24.46 ± 1.72

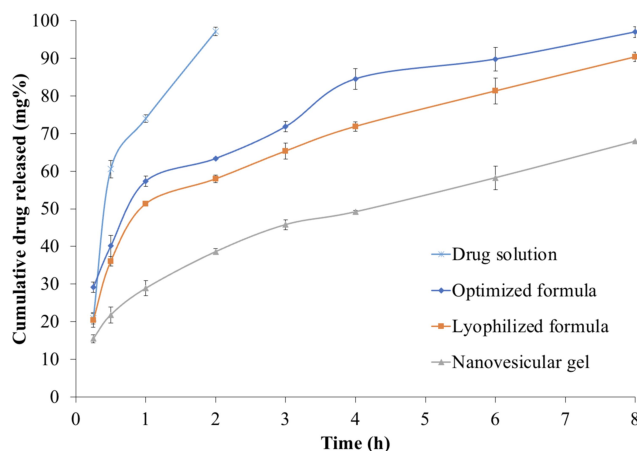
Note: ^aValues are expressed as mean ± SD; n = 3.

**Figure 3** Rheological characteristics of the prepared nanovesicular gel.

agent and known to form a highly viscous shear-thinning gel.⁶¹ The high viscosity during storage is extremely useful to maintain the physical stability of the dispersed nanovesicles by hindering their aggregation.⁶² On the other hand, the gel viscosity will significantly drop-down during application and rubbing which will facilitate spreadability on the skin.^{63,64}

In vitro Ivabradine Release

The study was conducted to determine the formulation effects on ivabradine release from the optimized nanovesicular formula as a dispersion, after lyophilization and after gelling. The drug release profiles are demonstrated in Figure 4. The drug release reached 100% in 2 h, this suggested that the drug could freely diffuse through the dialysis bag. The percentages of ivabradine released from the optimized formula, lyophilized one and nanovesicular gel in 2 h were, 63.34% ± 0.25, 57.93% ± 0.98 and 38.59% ± 0.84, respectively. Those release profiles could indicate a fast initial drug release. A slower release was observed with the gel due to the high viscosity of the gel carrier that could slow down drug release. At the end of 8

**Figure 4** Ivabradine release profiles from the optimized formula before and after lyophilization and gelling, in comparison with the drug solution.

h, percentages of ivabradine released from the optimized formula, lyophilized one and nanovesicular gel were 96.96% ± 1.45, 90.36% ± 1.27 and 67.92% ± 0.12.

On the other hand and in terms of dissolution half-life ($T_{50\%}$), the drug diffused from its solution through the dialysis membrane to the dissolution medium within 2 h ($T_{50\%} = 0.43$ h) while it takes 8 h in the case of the optimized formula ($T_{50\%} = 3.13$ h) with f_2 value of 34 indicating the significant difference between the two release profiles. This might be due to the presence of several lipophilic components in the formula, ie PC, SP and cholesterol, which could be able to control the drug partitioning to the surrounding aqueous medium.^{65,66} Moreover, there was no difference between drug release profiles before and after lyophilization where the f_2 value was equal to 55. On the other hand, converting the optimized formula to gel with carbopol led to a slower release profile ($f_2 = 29$) and higher release $T_{50\%}$ (5.29 h), compared to the optimized formula before gelling. Similar results were reported by Abdelnabi et al who prepared and optimized carbopol in situ nanovesicular gel loaded

with buspirone hydrochloride.⁶⁷ The slow drug release from the prepared nanovesicular gel was expected to give a sustained pharmacokinetic profile upon application to the skin.⁶⁸

Imaging of the Optimized Ivabradine Nanovesicles

The optimized nanovesicles have been imaged under TEM, as shown in Figure 5. The nanovesicles appeared well dispersed with no clear aggregation (Figure 5A). Moreover, the nanovesicles had a smooth surface with an irregularly spherical outline (Figure 5B). The size of captured nanovesicles was in good correlation with the size determined using the light scattering dynamic technique (Figure 5B).

Effect of Storage on Ivabradine Lyophilized Nanovesicles

There was no change in the appearance of the stored lyophilized nanovesicles after six months. The measured vesicle size, polydispersity index, zeta potential and entrapment efficiency were $231.3 \text{ nm} \pm 12.7$, 0.495 ± 0.01 , $39.05 \text{ mV} \pm 2.75$, $25.84\% \pm 1.03$. The statistical analysis using Student's *t*-test showed an absence of significance in the measured variables ($p < 0.05$) compared to the original findings before storage. Similar results were declared by Soliman et al who formulated proniosomes

containing the drug lacidipine and no significant difference was monitored after storage for 90 days.⁶⁹

Histopathological Examination

The histopathological test was done on rabbit shaved back skin to assess the possible irritation effect of the applied nanovesicular gel. Microscopical examination for normal rabbit skin used as a negative control revealed normal epidermis, squamous epithelium, underlying dermis containing normal adnexa, hair follicles and sweat glands (Figure 6A). Upon application of the nanovesicular gel, no significant changes were monitored with epidermis, dermis or adnexa (Figure 6B). Also, no signs of inflammatory cell infiltration or any skin irritation were observed. These results declared the safety of the nanovesicular Carbopol gel for the transdermal delivery of ivabradine.⁷⁰

In vitro Cytotoxicity Study

The results of the cytotoxicity are shown in Figure 7. Analysis of results revealed the absence of significance in cytotoxicity for both drug solution and lyophilized nanovesicular formula in comparison to normal untreated cells for concentrations equal to or less than $125 \mu\text{g/mL}$. Further increase in drug concentration revealed significant cytotoxicity. However, no significant differences were observed between the drug solution and the lyophilized nanovesicular formula. These results are in accordance

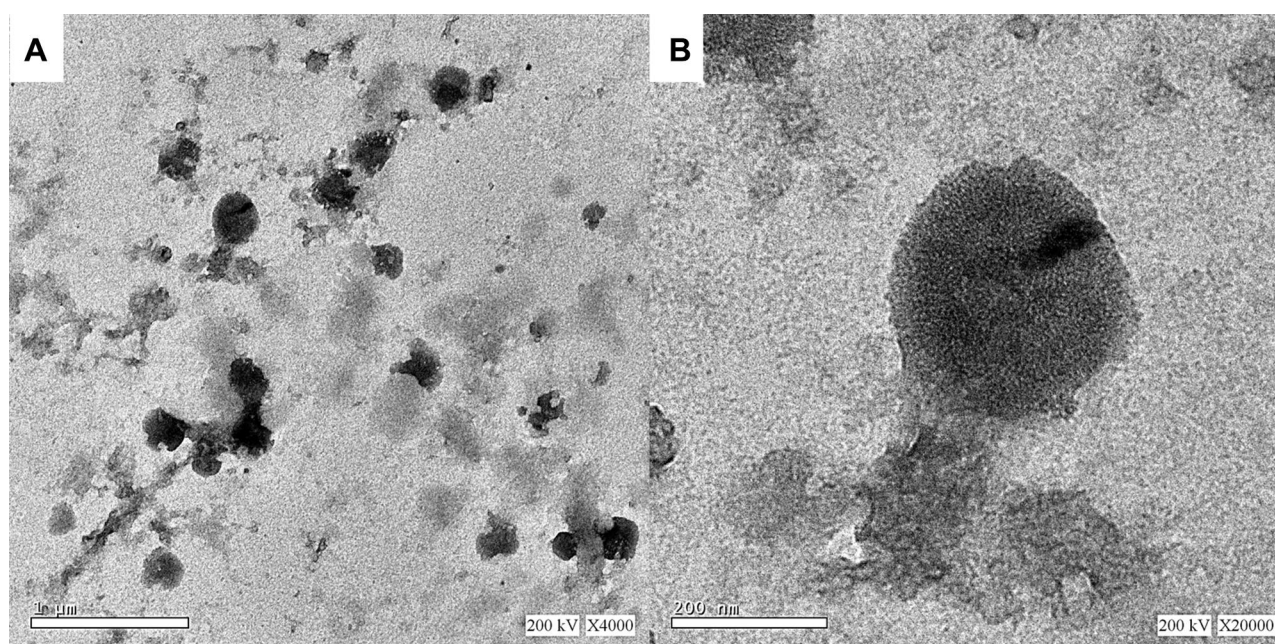


Figure 5 Transmission electron micrographs of the optimized ivabradine nanovesicles (A) X4000, (B) X20000.

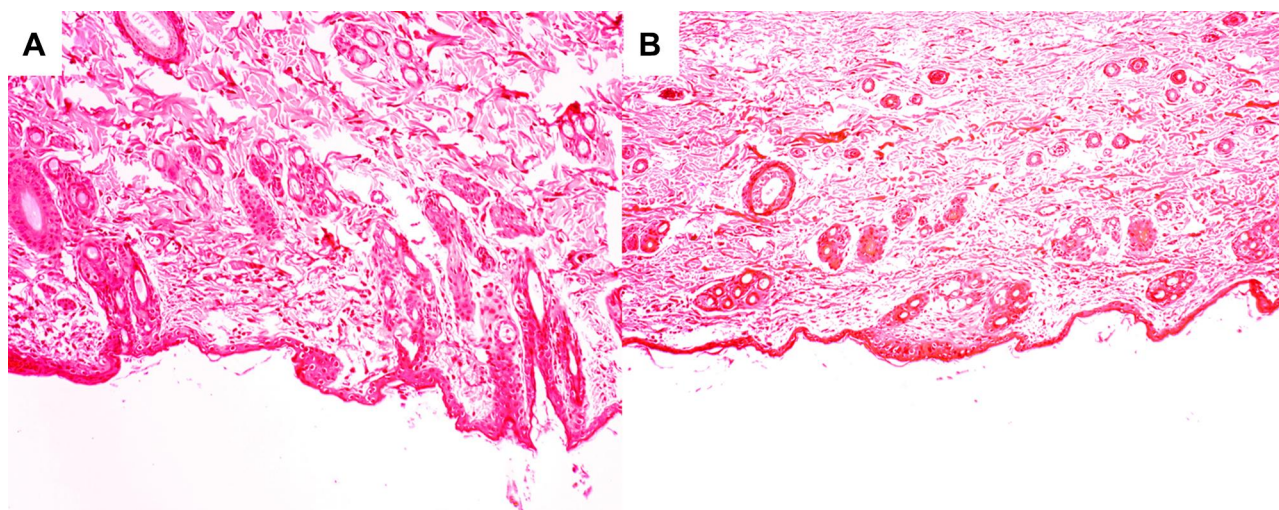


Figure 6 Histopathological examination of rabbit back skin untreated (control) (A) and treated with lyophilized nanovesicular gel (B) after treatment for three days (X100).

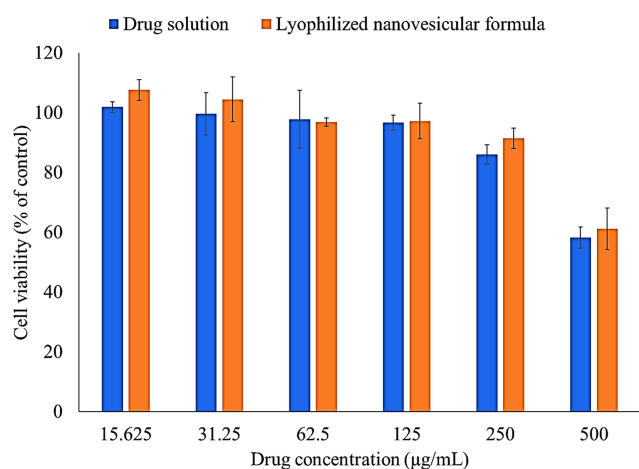


Figure 7 Cytotoxicity assay of ivabradine (drug solution and lyophilized nanovesicular gel) on HepG2 cells at different concentrations (15.625 to 500 µg/mL at 24 h), compared to respective untreated cells used as control.

with Ahmed et al who studied the cytotoxic effect of halloysite nanotubes against HepG2, their results also declared the safety of the used nanotubes up to 125 µg/mL with a further decrease in safety upon the increase of concentration to 250 and 500 µg/mL.³⁹

Oral and Transdermal Bioavailability of Ivabradine

The in vivo profiles of ivabradine after oral administration of the optimized formula and transdermal application of the nanovesicular gel to rabbits, compared to the market product (Procoralan®) are presented in Figure 8. The pharmacokinetic parameters are also shown in Table 3. The orally administered optimized

formula showed a significant increase in the drug bioavailability (254.61%) when compared to the market product. These results are in line with Varshozaz et al who tried to encapsulate the water-soluble drug buspirone HCl in solid lipid nanoparticles.⁷¹ Their optimized formula entrapped \approx 32.8% of the drug with an increase in bioavailability that reached 2.53 folds in comparison to buspirone solution. This could be attributed to the avoidance of the hepatic 1st pass effect through enhancing the lymphatic uptake as previously reported by Elsayed et al and Baek et al.^{16,72} A similar bioavailability enhancement (185.22%) was observed upon transdermal application of the nanovesicular gel. This might be referred to the small nanovesicular size (< 300 nm) and the presence of PL and SP as surfactants that could solubilize intercellular lipids and increase the fluidity of the stratum corneum layer, in addition to disrupting corneocytes because of the possible interaction with keratin filaments.^{73,74} On the other hand, the pharmacokinetic data depicted in Table 3 indicated a significant increase in C_{max} of ivabradine nanovesicles either as an oral lyophilized product or as a transdermal gel ($p < 0.05$) in comparison to market product. These results are in accordance with the results declared by Yaghoobian et al who fabricated repaglinide niosomes using different surfactants.⁷⁵ All the engineered niosomes showed higher C_{max} in comparison to the drug suspension.⁷⁶ Also, it is worth noticing that the developed nanovesicles could successfully increase ivabradine half-life from 2.88 ± 0.73 h for the market product to $6.6 \pm$

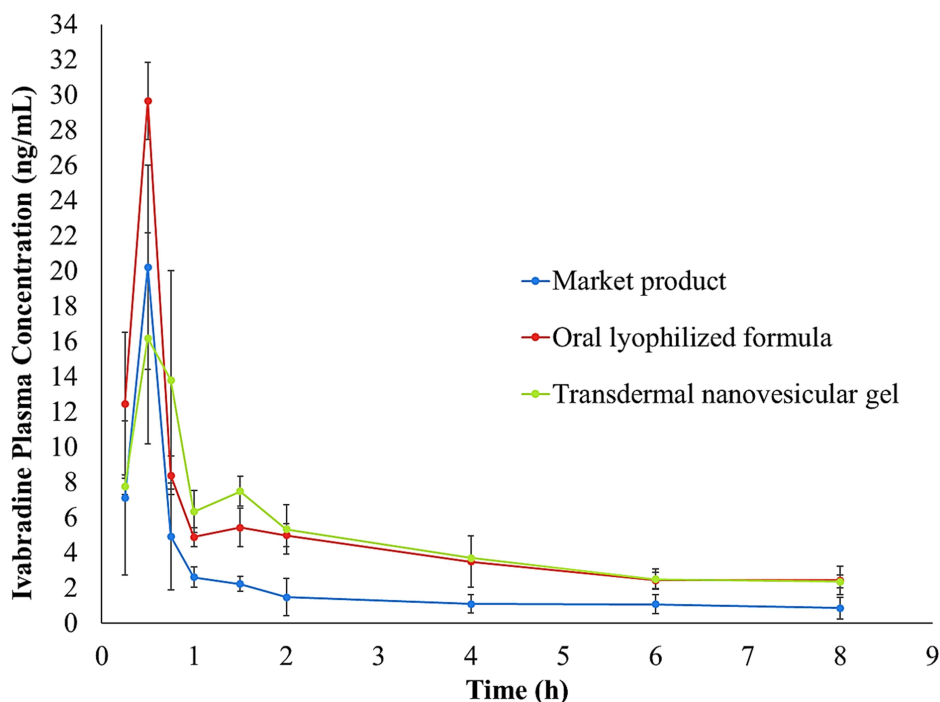


Figure 8 Mean plasma concentrations (ng/mL) of ivabradine after oral administration of the lyophilized formula and transdermal application of the nanovesicular gel, compared to the marketed product (Procoralan®).

Table 3 Pharmacokinetic Parameters of Ivabradine After Oral and Transdermal Administration of Lyophilized Nanovesicles and Nanovesicular Gel to Rabbits, Compared to the Market Oral Tablets (Procoralan®) ^{a,b}

Pharmacokinetic Parameters	The Market Product	The Oral Optimized Formula	The Transdermal Nanovesicular Gel
K (h ⁻¹)	0.36 ± 0.01	0.10 ± 0.01	0.08 ± 0.02
T _{1/2} (h ⁻¹)	2.88 ± 0.73	6.60 ± 0.11	8.02 ± 0.34
T _{max} (h) ^c	0.75	0.5	0.5
C _{max} (ng/mL)	16.11 ± 1.57	29.66 ± 1.90	20.43 ± 1.28
AUC ₀₋₈ (ng.h/mL)	22.00 ± 1.84	38.60 ± 2.77	39.22 ± 2.01
AUC _{0-∞} (ng.h/mL)	23.29 ± 1.59	59.30 ± 3.25	43.14 ± 1.94
Relative bioavailability (%)	-	254.61	185.22

Notes: ^aThe administered drug dose was 0.26 mg/kg. ^bValues are expressed as mean ± SD; n = 9. ^cMedian of T_{max} is displayed instead of the mean values.

0.11 h in the case of oral optimized formula and 8.02 ± 0.34 h for the transdermal vesicles (p <0.05). These results coincide with the in vitro release profile of the drug and also with the in vivo results declared by

Ramkanth et al who tried to formulate atenolol niosomes for transdermal administration, their results showed a higher drug bioavailability with prolonged T_{1/2}.^{77,78}

Conclusions

The optimized nanovesicles were capable of doubling ivabradine bioavailability either after oral administration of its lyophilizate or transdermal application of its carbopol gel. The orally administered optimized formula was able to avoid the hepatic 1st pass effect while the transdermally applied nanovesicular gel was capable to penetrate deeply through the skin layer and reaching the bloodstream. Having the same formula tailored for both oral and transdermal administration could be considered a promising cost-effective approach for industrial applications. Future work may consider trying the same formula as a platform for bioavailability enhancement of other active ingredients highly susceptible to hepatic 1st pass metabolism.

Acknowledgment

The authors would like to thank Cell Culture unit Laboratory, Faculty of Pharmacy, October 6 University, Giza, Egypt for their contribution in the in vitro cytotoxicity study.

Disclosure

The authors report no conflicts of interest in this work.

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