

DIC in Pregnancy – Pathophysiology, Clinical Characteristics, Diagnostic Scores, and Treatments

Offer Erez ^{1,2}
 Maha Othman³
 Anat Rabinovich⁴
 Elad Leron ⁵
 Francesca Gotsch⁶
 Jecko Thachil⁷

¹Maternity Department “D”, Division of Obstetrics and Gynecology, Soroka University Medical Center, School of Medicine, Faculty of Health Sciences Ben Gurion University of the Negev, Beer Sheva, Israel; ²Department of Obstetrics and Gynecology, Hutzel Women’s Hospital, Wayne State University, Detroit, MI, USA; ³Department of Biomedical and Molecular Sciences, School of Medicine, Queen’s University, Kingston, ON, Canada; ⁴Thrombosis and Hemostasis Unit, Hematology Institute, Soroka University Medical Center and Faculty of Health Sciences, Ben-Gurion University of the Negev, Beer Sheva, Israel; ⁵Division of Obstetrics and Gynecology, Soroka University Medical Center, School of Medicine, Faculty of Health Sciences Ben Gurion University of the Negev, Beer Sheva, Israel; ⁶Department of Obstetrics and Gynecology, Azienda Ospedaliera Universitaria Integrata, AOUI Verona, University of Verona, Verona, Italy; ⁷Department of Haematology, Manchester Royal Infirmary, Manchester, UK

Correspondence: Offer Erez
 Maternity Department “D” and
 Obstetrical Day Care Center, Division of
 Obstetrics and Gynecology, Soroka
 University Medical Center and Faculty of
 Health Sciences, Ben Gurion University of
 the Negev, Beer Sheva, 84101, Israel
 Email erezof@bgu.ac.il

Abstract: Obstetrical hemorrhage and especially DIC (disseminated intravascular coagulation) is a leading cause for maternal mortality across the globe, often secondary to underlying maternal and/or fetal complications including placental abruption, amniotic fluid embolism, HELLP syndrome (hemolysis, elevated liver enzymes and low platelets), retained stillbirth and acute fatty liver of pregnancy. Various obstetrical disorders can present with DIC as a complication; thus, increased awareness is key to diagnosing the condition. DIC patients can present to clinicians who may not be experienced in a variety of aspects of thrombosis and hemostasis. Hence, DIC diagnosis is often only entertained when the patient already developed uncontrollable bleeding or multi-organ failure, all of which represent unsalvageable scenarios. Beyond the clinical presentations, the main issue with DIC diagnosis is in relation to coagulation test abnormalities. It is widely believed that in DIC, patients will have prolonged prothrombin time (PT) and partial thromboplastin time (PTT), thrombocytopenia, low fibrinogen, and raised D-dimers. Diagnosis of DIC can be elusive during pregnancy and requires vigilance and knowledge of the physiologic changes during pregnancy. It can be facilitated by using a pregnancy specific DIC score including three components: 1) fibrinogen concentrations; 2) the PT difference – relating to the difference in PT result between the patient’s plasma and the laboratory control; and 3) platelet count. At a cutoff of ≥ 26 points, the pregnancy specific DIC score has 88% sensitivity, 96% specificity, a positive likelihood ratio (LR) of 22, and a negative LR of 0.125. Management of DIC during pregnancy requires a prompt attention to the underlying condition leading to this complication, including the delivery of the patient, and correction of the hemostatic problem that can be guided by point of care testing adjusted for pregnancy.

Keywords: DIC, hyperfibrinolysis, thrombin, pregnancy specific DIC score, maternal mortality, placental abruption

Introduction

The life threatening thrombo-hemorrhagic condition known as disseminated intravascular coagulation (DIC), is generally more common in men than women apart from the reproductive years when pregnancy and its complications increase its prevalence.¹ DIC during pregnancy constitutes one of the leading causes for maternal mortality worldwide,^{2,3} and its rate varies from 0.03%⁴ to 0.35%.⁵ This complication is often secondary to underlying maternal and/or fetal complications including placental abruption,^{6–8} postpartum hemorrhage, amniotic fluid embolism, acute fatty liver of pregnancy, preeclampsia and Hemolysis, Elevated Liver enzymes and Low Platelets (HELLP) syndrome,⁹ and retained stillbirth.¹⁰ DIC is characterized by a concomitant over-activation of the coagulation and fibrinolytic

systems,¹¹ leading to widespread microvascular thrombosis, disruption of blood supply to different organs, ischemia, and multi-organ failure. This extensive activation of the coagulation cascade leads to consumption and depletion of platelets and coagulation proteins, which can provoke concurrent severe bleeding.¹² Of note, obstetric DIC more typically presents with bleeding complications, rather than thrombotic complications. The current review discusses the clinical characteristics, pathophysiology, diagnostic scores, and treatments of DIC in pregnancy.

DIC – Definitions and Types

The Scientific and Standardization Committee (SSC) on DIC of the International Society on Thrombosis and Haemostasis (ISTH) defined DIC as

an acquired syndrome characterized by the intravascular activation of coagulation with a loss of localization arising from different causes. It can originate from and cause damage to the microvasculature, which if sufficiently severe, can produce organ dysfunction.¹³

This definition highlights three important aspects: 1) it is always secondary to other causes, one of them being an obstetric related, such as abruption placentae, pre-eclampsia, or intrauterine fetal death.¹⁴ 2) DIC represents systemic pathological activation of coagulation. Normal haemostasis refers to the formation of a thrombus in response to endothelial damage and, in these cases, the clotting and fibrinolytic processes are limited to the site of endothelial injury.¹⁵ However, in DIC, there is uncontrolled dissemination of this localised thrombotic process which is clearly pathological. In obstetrical DIC, there is extraplacental dissemination of the activated coagulation system which is usually localised to the placenta.¹⁶ 3) The third crucial aspect of the DIC definition is the role of microvasculature. DIC is a process which originates in the microvasculature, or the vascular endothelium, the most ubiquitous organ in the human body. Excessive and dysregulated thrombin generation due to (and causing) marked endothelial dysfunction can result in organ damage from microthrombi.¹⁷ If microvascular dysfunction led to organ damage, it would be considered DIC according to the ISTH definition. If in obstetrical disorders, microthrombi can lead to kidney impairment or cerebrovascular ischaemia, this would be considered as a manifestation of DIC. Microthrombi in the placental vessels can be one of the pathophysiological processes which lead to the complications associated with pre-eclampsia including foetal

growth retardation and organ impairment. The microthrombi can arise from localised coagulation activation with platelet aggregation being a significant contributory factor. Most inflammatory disorders like pre-eclampsia are associated with the release of large amounts of Von Willebrand factor into the circulation. In normal circumstances, the ultra large multimer forms of the Von Willebrand factor is cleaved by a metalloprotease enzyme, ADAMTS-13. But in severe inflammation, the enzyme is quenched by the inordinate amount of ultra-large multimers released and the consequence is binding to platelets causing platelet aggregation and microthrombi. In support of this mechanism,^{18,19} Miodownik et al²⁰ have shown that congenital thrombotic thrombocytopenic purpura can present with clinical complications during pregnancy and maternal vascular lesions of under-perfusion in the placenta.

The clinical presentation of DIC can be either thrombosis and/or bleeding as it is a thrombohemorrhagic disorder.²¹ This may be explained by the extremely rapid pace of thrombin generation in the thrombotic phenotype, while it is relatively slower in the haemorrhagic presentation of this complication (Figure 1). The “hyperfibrinolytic” form of DIC as often observed in postpartum DIC, and abruption²² results from an extremely rapid burst of thrombin generation, whereas a “procoagulant” form of DIC (eg, pre-eclampsia related DIC) may result from an excessive but slower and more gradual form of thrombin generation.²³ Although both procoagulant and hyperfibrinolytic processes may proceed simultaneously in DIC, the clinical presentation of thrombosis or bleeding is determined by the predominant mechanism at a particular time.²⁴

Various obstetrical disorders can present with DIC as a complication; thus, increased awareness is key to diagnosing the condition. DIC patients can present to clinicians who may not be experienced in a variety of aspects of thrombosis and haemostasis. Hence, DIC diagnosis is often only entertained when the patient already developed uncontrollable bleeding or multi-organ failure, all of which represent unsalvageable scenarios.²⁵ Beyond the clinical presentations, the main issue with DIC diagnosis is in relation to coagulation test abnormalities. It is widely believed that in DIC, patients will have thrombocytopenia, prolonged prothrombin time (PT) and partial thromboplastin time (PTT), low fibrinogen and raised D-dimers.²⁶ These lab markers were considered as diagnostic of DIC in an era when the DIC diagnosis was predominantly made

The different types of DIC

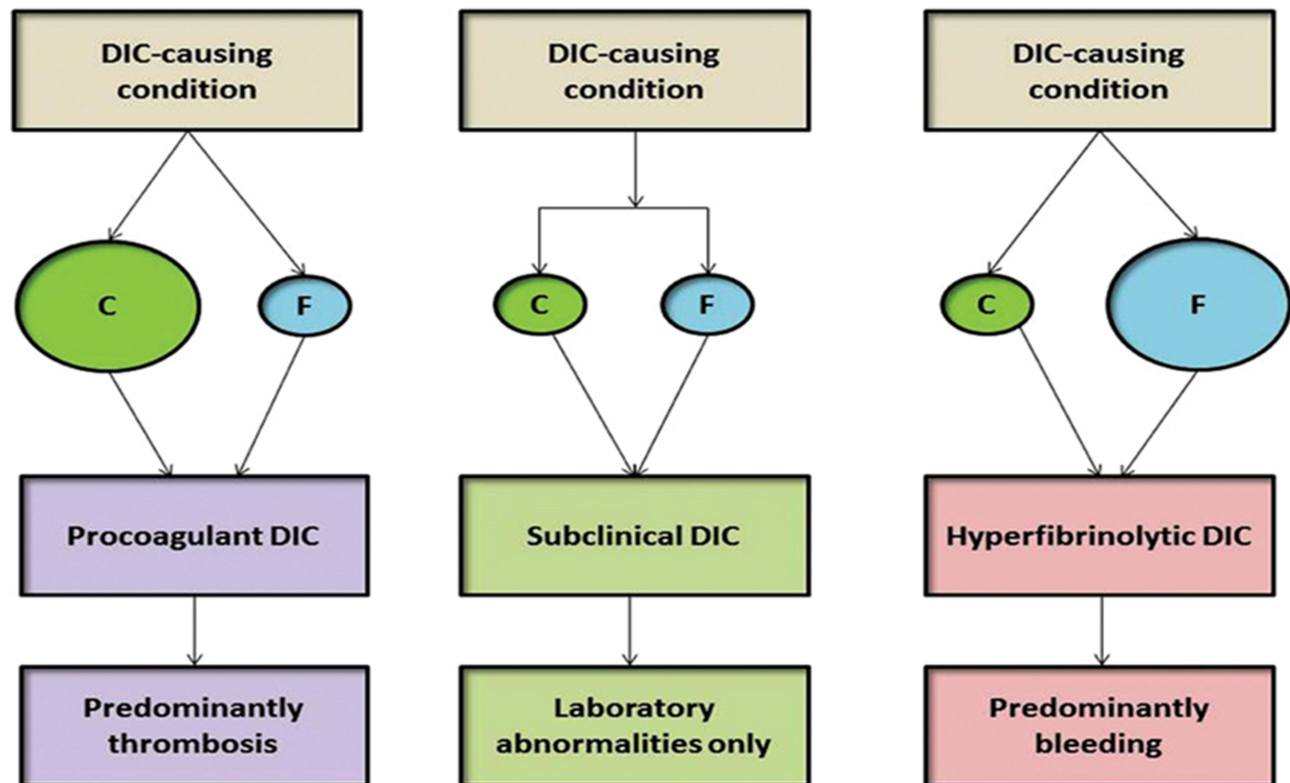


Figure 1 The different types of DIC and their clinical presentation. If there is predominance of coagulation pathway activation (denoted as C), in comparison with the fibrinolytic pathways (denoted as F), procoagulant DIC is the result. While the reverse leads to hyperfibrinolytic DIC.

Notes: Reprinted from: Thachil J. The Elusive Diagnosis of Disseminated Intravascular Coagulation: does a Diagnosis of DIC Exist Anymore? *Semin Thromb Hemost.* 2019;45:100–107.²⁴ With permission. Copyright © Georg Thieme Verlag KG.

post-mortem or in extremely ill patients.⁹ The introduction of DIC scoring discussed below generates cut off values of laboratory tests that enable the clinicians to diagnose DIC at early stages of the disease with a more subtle abnormalities of the laboratory tests.

Epidemiology of DIC in Pregnancy

DIC has a higher lifetime prevalence in men than in women, the only period where the incidence of this disorders is similar between the genders is during women's reproductive years.¹ The prevalence of obstetrical DIC differs worldwide ranging from as low as 0.03% in North America to 0.35% in developing countries.^{2–5,27–29} Differences in quality and accessibility to advance maternity care, as well as inconsistent definitions and diagnostic criteria led to variations in the reported incidence of DIC. Indeed, a cohort from the Nationwide Inpatient Sample from USA,³⁰ reported that the prevalence of DIC during

pregnancy from 1998 to 2009, increased by 35% from 0.092% to 0.125%³⁰ and linked with nearly a quarter of maternal deaths. Furthermore, DIC was the second most common severe maternal morbidity indicator in the USA between 2010 to 2011, affecting 32 per 10,000 delivery hospitalizations.³¹

DIC is always secondary to an underlying disorder. Indeed, it is associated with pregnancy complications such as placental abruption, HELLP syndrome, preeclampsia, retained stillbirth, sepsis, post-partum hemorrhage (PPH), acute fatty liver, and amniotic fluid embolism (Figure 2). The proportions of the contribution of each risk factor varies among countries. Indeed, the most frequent pregnancy complication associated with DIC according to Rattray et al⁴ was placental abruption followed by PPH and preeclampsia. Similarly, Erez et al⁵ reported that placental abruption was the leading underlying obstetrical disorder leading to DIC especially when it

Obstetrical complications associated with DIC

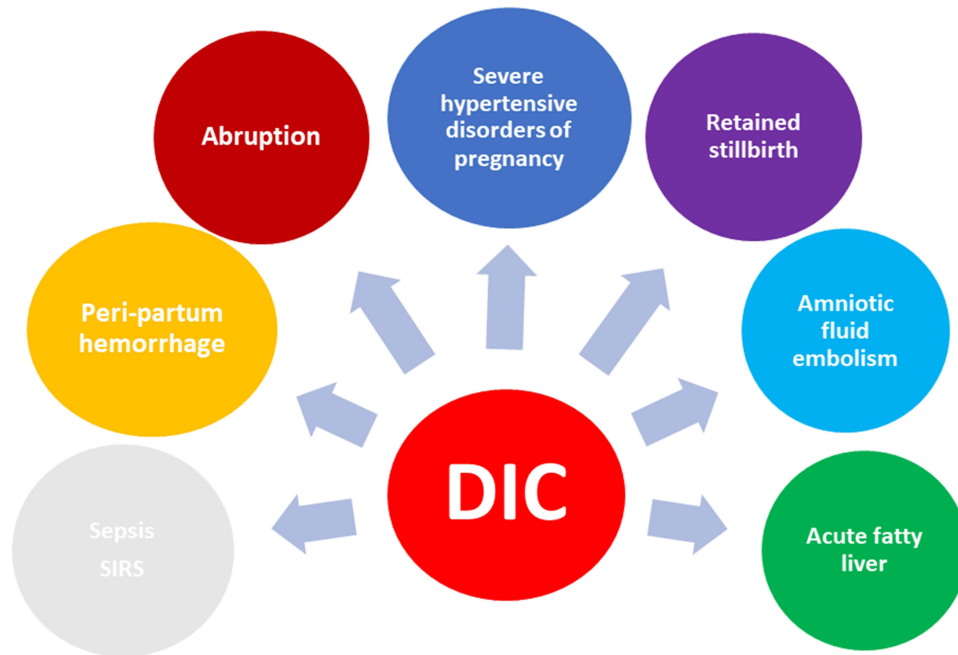


Figure 2 Obstetrical complications associated with DIC in pregnancy.

was accompanied by fetal death. Other groups reported preeclampsia³² and fetal death²⁹ to be the leading obstetrical complications associated with DIC (Figure 3). This

suggests that early recognition and treatment of complications such as preeclampsia or stillbirth may prevent the deterioration to DIC in these patients. Moreover, in countries

Variation in pregnancy complications resulting in DIC in different countries

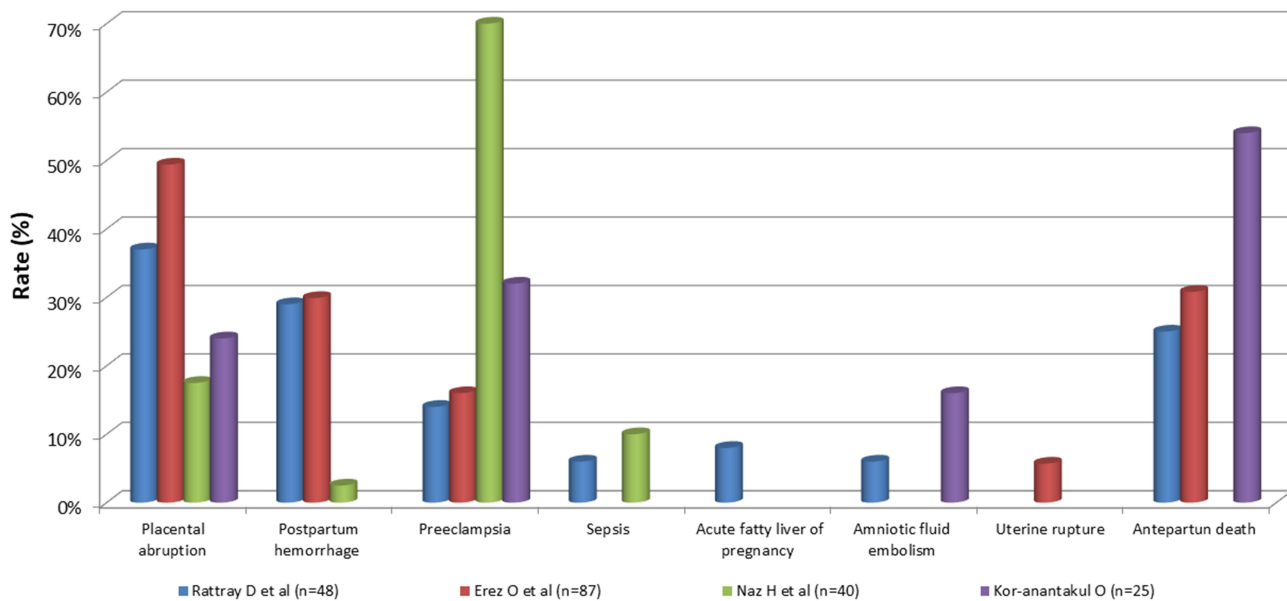


Figure 3 Global distribution of obstetrical complications associated with DIC in pregnancy.

Note: Data was modified from references 4, 5, 26 and 32.

with early recognition and treatment of preeclampsia and stillbirth, placental abruption (usually unexpected and non-preventable), becomes the leading obstetrical complication associated with DIC.

Placental abruption seems to be the underlying mechanism for the development of DIC when it is associated with other obstetrical complications including fetal death and HELLP syndrome.^{7,33–39}

Thus, pregnancy is a time point in women's life in which she is at increased risk to develop DIC as a result from different complications of pregnancy, and especially placental abruption. Prompt attention and treatment of underlying obstetrical disorders and rapid diagnosis and management of placental abruption can reduce the risk for DIC substantially.

Pathophysiology of DIC in Pregnancy

DIC has a complex pathophysiology, the understanding of the normal haemostasis and the highly orchestrated balance involving the various pathways including pro-coagulant/anticoagulant, fibrinolytic/antifibrinolytic and platelet/von Willebrand factor axis during pregnancy, is essential for the characterisation of the haemostatic abnormalities observed during DIC.¹⁷

Excessive and uncontrolled *in vivo* thrombin generation is a key to the development of DIC.²¹ The excess thrombin generated will activate these pathways in differential manners. Hence, if the procoagulant and antifibrinolytic pathways are both activated "in excess", then the result would be thrombosis, especially if the natural anticoagulants have also been consumed.²⁵ If the excess thrombin generated drives the fibrinolytic pathway "in excess" of the other pathways, bleeding will commence.⁹

How does this translate to obstetrical cases? In the placental circulation, maintenance of the blood flow is paramount to ensure adequate oxygenation and nutrition of the foetus.¹⁶ At the same time there is local physiological activation of coagulation in the placental bed, in preparation to quench large amounts of bleeding anticipated during parturition.¹⁶ Despite the heightened coagulation, clots are not formed in the placental bed except in cases of abnormal placental pathologies.⁴⁰ In obstetrical disorders, the much-enhanced coagulation process could lead to microvascular clots which impair the appropriate development of the fetus.⁴⁰ In addition, if the activated procoagulant pathway does not come to a halt, this would spill into the maternal circulation causing systemic coagulopathy, otherwise termed as DIC.²⁴ While, if there is

a brisk fibrinolytic response, then this could lead to haemorrhage wherein formation of firm and dense clots is prevented by fibrinolytic proteins.⁴¹ Furthermore, D-dimers and fibrin-degradation products that results from fibrinolysis interfere with platelet activation and can impair myometrial contractility.⁴²

Pathological Mechanisms of DIC in Pregnancy

Endothelial Dysfunction and Platelet Activation

Normal pregnancy is associated with activation of maternal leukocytes⁴³ into a state akin to sepsis.⁴⁴ Nevertheless, placental trophoblast maintain a balanced systemic maternal inflammation during gestation⁴⁵ by inactivation of maternal leukocytes. However, infectious agents, septic abortion, and amniotic fluid embolism⁴⁶ leading to sepsis perturbed this balance and can lead to the development of maternal DIC (Figure 4).

During sepsis the coagulation system can be activated by Endothelial cells or leucocytes (either intact, dysfunctional, or activated), platelets, and remnants of cell surfaces, inflammatory mediators and coagulation proteins participate in the processes leading to uncontrolled activation of coagulation cascade and DIC⁴⁷ (Figure 5). These cells release pro-inflammatory cytokines such as tumor necrosis factor- α , interleukin-1 (IL-1) and interleukin-6 (IL-6), along with propagation of tissue factor (TF) expression on the surface of endothelial cell and leukocytes^{22,48–54} that can initiate an uncontrolled activation of the coagulation cascade via the TF/factor VIIa pathway leading to thrombin generation if this coagulation response is uncontrolled it will eventually lead to DIC.⁵⁵ The most significant source of TF is not completely clear as this activator of the coagulation cascade is expressed not only in mononuclear cells in response to pro-inflammatory cytokines (mainly IL-6), but also by vascular endothelial (Figure 5), cancer cells,^{53,54,56} decidua and placenta.

Despite the potency of TF as a trigger of coagulation, potent anticoagulation pathways can control and limit its activity. However, all the natural anticoagulant pathways (ie, anti-thrombin III, protein C system, and TF pathway inhibitor [TFPI]) are compromised during sepsis mediated DIC.⁵⁷ Indeed, during DIC antithrombin III concentration are markedly reduced, due to its consumption,⁵⁸ degradation (by elastase from activated neutrophils),⁵⁹ and impaired synthesis.⁴⁹ Moreover, protein C system inhibition due to decreased protein synthesis, cytokine-mediated down-regulation of endothelial thrombomodulin and a drop in the concentration of the free fraction of protein S (protein C co-factor) may further

Schematic representation of pathogenic pathways in DIC

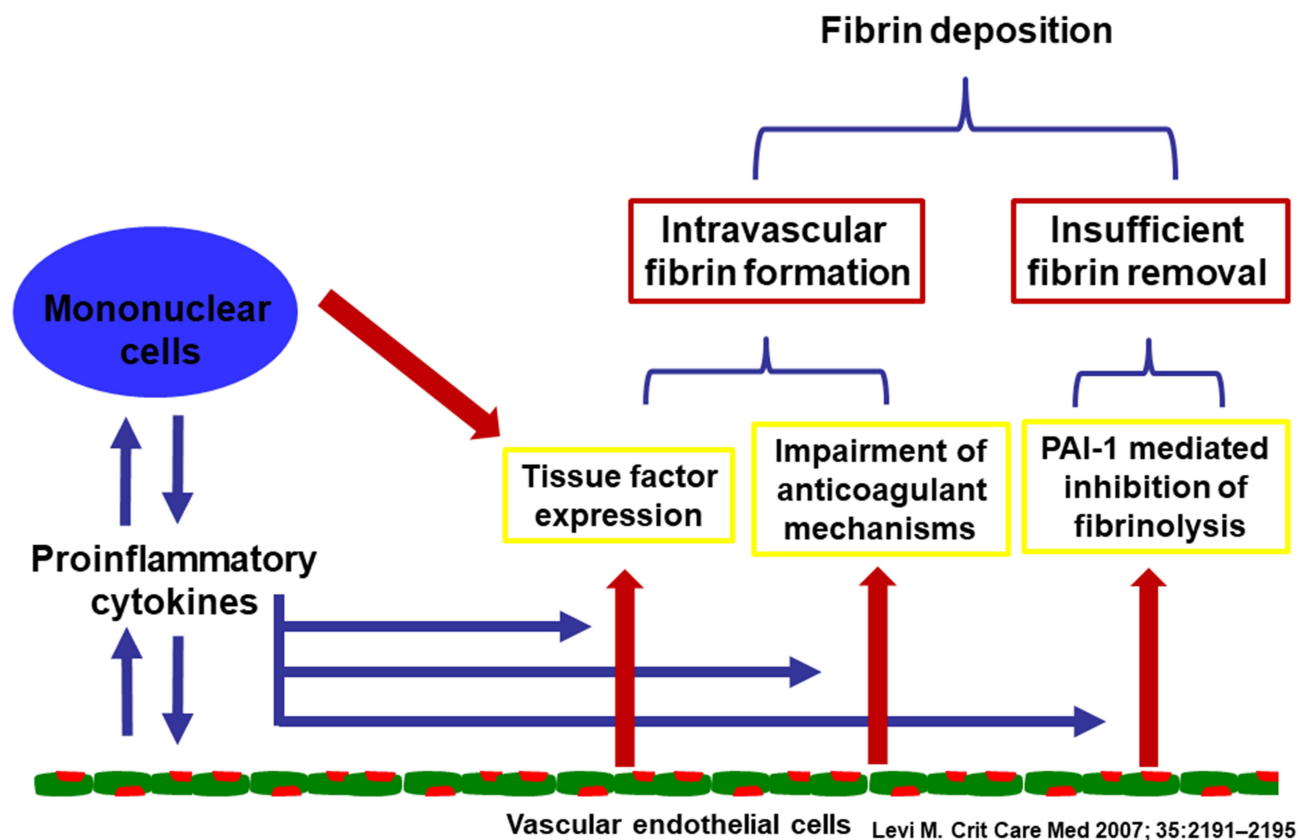


Figure 4 Schematic representation of pathogenic pathways in DIC.

Notes: Adapted from: Levi M. Disseminated intravascular coagulation. *Crit Care Med*. <https://journals.lww.com/ccmjournal/pages/default.aspx>. 2007;35(9):2191-2195.⁴⁹ With permission. © 2007 by the Society of Critical Care Medicine and Lippincott Williams & Wilkins.

compromise an adequate regulation of thrombin generation.⁶⁰ Lastly, reduce TFPI function cannot address the increased activation of the extrinsic coagulation cascade.⁶¹

Fibrin formation is increased during DIC at the time of maximal activation of coagulation due to the following: 1) there is a continuous elevation of plasminogen activator inhibitor-1 (PAI-1) that suppresses fibrinolysis;^{49,62} 2) platelets activation⁶³ increases TF expression on monocytes especially in the presence of granulocytes in a P-selectin dependent reaction.⁵⁶ This effect can be attributed to the induction of nuclear factor kappa B (NF- κ B) activation following the binding of activated platelets to neutrophils and mononuclear cells.⁶⁴ Similar mechanisms are activated during DIC in pregnancy resulting from sepsis or systemic maternal inflammation.

Systemic inflammatory response resulting from maternal infection is not unique only for bacterial infection. Indeed, maternal DIC associated with COVID-19 infection

was previously reported.⁶⁵ Unlike the procoagulant DIC in the non-pregnant state,⁶⁶ during pregnancy, COVID-19 coagulopathy in the third trimester was associated with a clinical manifestation mimicking normotensive HELLP syndrome associated with thrombocytopenia and elevated liver enzymes⁶⁵ and the coagulopathy was of the hyperfibrinolytic type with fibrinogen concentrations and bleeding.⁶⁷ This issue is further discussed in a Communication regarding COVID-19 coagulopathy in pregnancy from the ISTH SSC for Women's Health.

Trophoblast Deportation and Systemic Activation of Maternal Coagulation Cascade

During normal gestation, the trophoblasts face two contradicting hemostatic functions: 1) sustaining of laminar flow of maternal blood in the intervillous space without clotting; and 2) to prevent bleeding at the maternal fetal interface.⁹ This led to the acquisition of endothelial

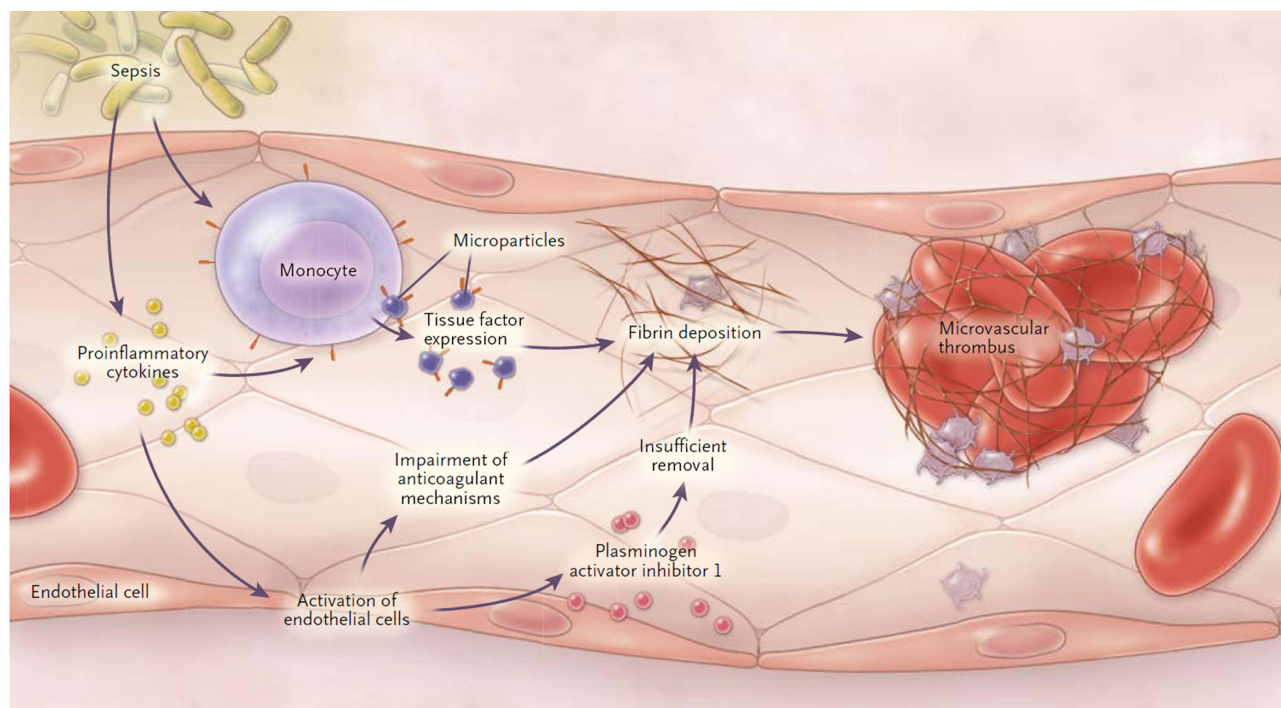


Figure 5 Mechanisms of DIC in sepsis involving endothelial dysfunction and platelet activation.

Notes: Reproduced from: Hunt BJ. Bleeding and Coagulopathies in Critical Care. *N Engl J Med.* 2014;370:847–859.¹⁹⁹ Copyright © 2014 Massachusetts Medical Society. Reprinted with permission from Massachusetts Medical Society.

cell like properties to the syncytiotrophoblast along with the expression of TF to maintain hemostatic function. Placental TF has higher activity in comparison to that expressed on human umbilical vein endothelial cells. The syncytiotrophoblast has also anti coagulation properties as it is able to synthesize Protein C, Protein S and Protein Z as well as pregnancy specific inhibitor of the tissue factor pathway known as TFPI-2 (Placental Protein 5)^{68–70} to prevent uncontrolled activation of the coagulation cascade.^{71–74} The prevention of fibrinolysis by the placenta is achieved by placental production of PAI-2 during normal pregnancy.⁷⁵ These physiological changes during pregnancy are associated with relatively constant tissue plasminogen activator (t-PA) concentrations, leading to a prothrombotic state related in part to reduced clot lysis.⁷⁶

Disruption of trophoblast/decidua integrity would lead to a discharge of potent TF into maternal circulation, and activation of coagulation cascade that in large quantities can lead to uncontrolled systemic thrombin generation and DIC^{22,77} (Figure 6).

Abruption of the placental is the classica example for this mechanism, especially in cases of concealed abruption or when it is complicated by stillbirth. Even though some

regard that consumption coagulopathy as the underlying mechanism of placental abruption associated DIC, it seems that there is more to it. Indeed, women with a retro placental clot have a faster developing and more severe form of DIC than those with PPH although they have overall lower blood loss. Indeed, patients who develop placental abruption suffer from a combined coagulopathy incorporating consumption coagulopathy and substantial release of thromboplastin (tissue factor) into the maternal circulation.^{9,49} In addition, local hypoxia and hypovolemia trigger endothelial response leading to increased expression of vascular endothelial growth factor, which causes an increased endothelial expression of TF.⁷⁸ This view is supported by the following: 1) Erez et al²² reported that among women with stillbirth, those with abruption, have higher amniotic fluid TAT complexes concentrations and 2) mice who were injected with placental extracts developed DIC and died. The authors also demonstrated that administration of heparin prevented the death of the injected mice.⁷⁹ Further study suggested that thromboplastin triggers this response in mice via its effect on clotting time and chemical properties.⁷⁹ Recent advances in the understanding of the mechanisms of DIC in HELLP syndrome suggest that abruption rather than liver dysfunction

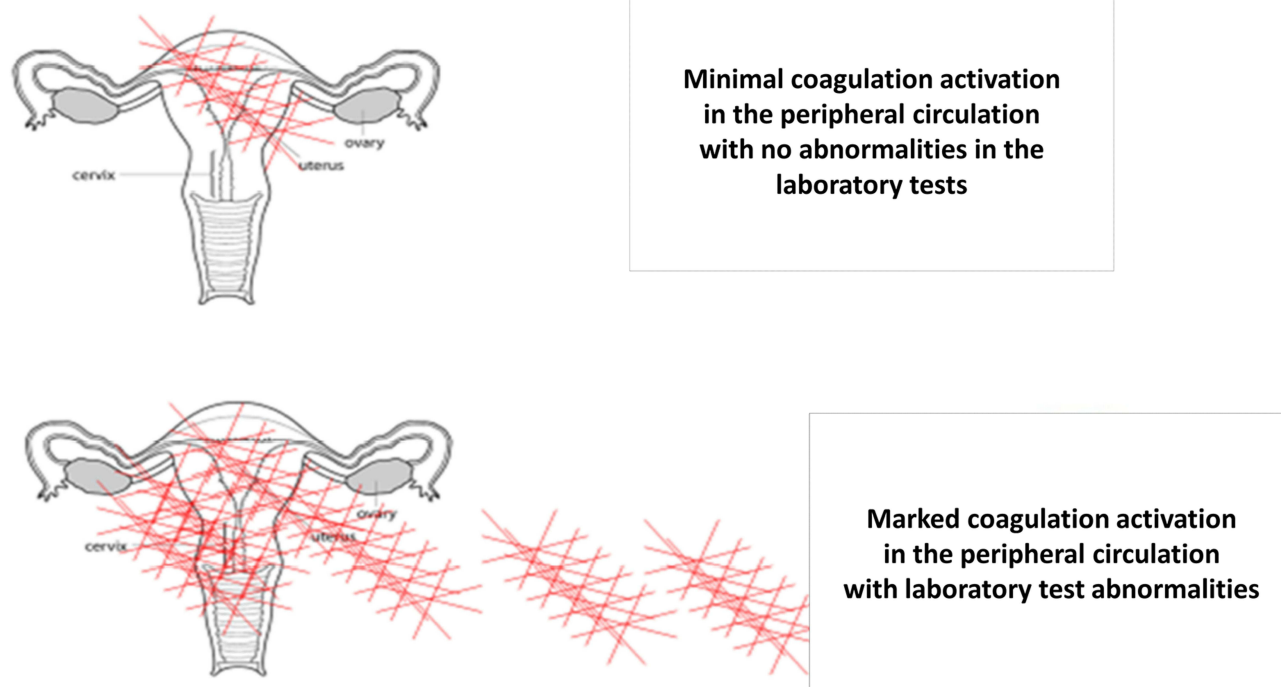


Figure 6 Trophoblast deportation and systemic activation of maternal coagulation cascade. Coagulation system is activated in the placental circulation in the normal pregnant state. This activated coagulation can spill into the peripheral circulation in patients who have obstetrical complications like pre-eclampsia. If coagulation activation in the peripheral circulation becomes uncontrolled, it manifests as DIC.

is the driving force for the development of DIC in these patients.³⁹ Indeed, the rate of placental abruption was 5.6% in women with HELLP syndrome without DIC in comparison to 42.9% among those with DIC,³⁹ suggesting that the main underlying mechanism leading to DIC in HELLP syndrome is related to trophoblast deportation into maternal circulation rather than to an intrinsic liver dysfunction.

Amniotic fluid embolism, although rare, is a major cause for maternal DIC, though a combined mechanism of activation of coagulation by the TF rich amniotic fluid debris that enter the maternal circulation and the acute systemic maternal inflammatory response further propagating the activation of the coagulation cascade. These patients have a fibrinolytic type of DIC⁸⁰ observed as unlike placental abruption autopsies of women who died due to amniotic fluid embolism lack evidence of fibrin deposition.⁸¹ Moreover, Beller et al⁸² studied the clotting and fibrinolytic systems of their patient in three blood samples: the first was drawn on admission, the second at the onset of shock and the third at the start of cardiac massage following cardiac arrest. Clotting parameters on admission were normal. The blood in the third sample did not clot even after adding thrombin in excess. Moreover,

the clotting time remained prolonged after fibrinogen was added in the presence of excess thrombin indicating that anti-thrombin activity was present. However, protamine sulfate did not restore the clotting time to normal, indicating that the antithrombin activity was not attributable to heparin. The contribution of systemic maternal inflammation to the development of DIC in women with amniotic fluid embolism is highlighted in the report of three cases of amniotic fluid embolism in which the patients had elevated maternal circulation TNF α upon admission prior to the clinical presentation of this complication.⁴⁶

Obstetrical Hemorrhage

Hemorrhage, especially post-partum is regarded by many^{4,9,50,83–87} as one of the major causes for DIC in pregnancy. Classically related to PPH, this type of consumption coagulopathy is a complication of uterine atony or rupture, retained placenta or membranes, placenta accreta, or severe lacerations of the birth canal (cervical or vaginal).^{5,88,89} The characteristics of the DIC developing in these cases are the rapid maternal loss of a large blood volume along with its coagulation factors, resulting in patients who are hemodynamically compromised. Currently, there is a debate whether this form of

consumption coagulopathy is truly DIC or just a massive blood loss that depletes the patient's coagulation factors and can lead to death due to exsanguination.⁹⁰ However, maternal coagulopathy associated with PPH is not just the loss of blood volume and coagulation factor but rather a more complex complication that can be considered as DIC. This can be attributed to the procoagulant state and increased thrombin generation associated with parturition attributed to the release of TF to the maternal circulation following the separation of the membranes and the placenta,⁹¹ along with the systemic inflammation accompanies the process of labor and delivery.^{92,93} Indeed, women who had PPH are considered at high risk for development of deep vein thrombosis during the puerperium.⁹⁴ The evidence brought herein, that parturient with PPH have a higher activation of coagulation cascade, beyond physiologic levels for normal pregnancy, suggest that the clinicians who treat these patients must consider them as high risk groups for DIC, even though the fundamental pathology is considered to be the rapid and massive loss of blood and coagulation factors.⁷⁷ Therefore, PPH requires a careful and prompt assessment and to be treated pharmacologically and/or surgically, and by blood products as well as volume expanders to sustain the maternal circulation in an attempt to prevent the subsequent development of DIC.

Disruption of Liver Function

Acute fatty liver of pregnancy is severe, but a rare unique complication of pregnancy mostly observed in nulliparous patients toward the end of the third trimester.^{95–97} Clinicopathological features of this complication include progressive loss of liver function,⁹⁸ abnormal renal function⁹⁹ and DIC.¹⁰⁰ The liver presents accumulation of fatty microvascular infiltration of hepatocytes with, without alteration of the overall structure of the liver.⁹⁸ Nevertheless, the synthetic function of the liver is hampered leading to lower production of fibrinogen and coagulation factors and along with the hemorrhage observed in this syndrome lead to DIC. Indeed, in one of the largest series of acute fatty liver published thus far including 51 patients,¹⁰¹ and 80% of these women according the ISTH DIC score^{102,103} had DIC similar to the report of Vigil-De Gracia et al¹⁰⁰ in which DIC was present in >70% of the patients. The characteristics of the hemostatic parameters of women with DIC in the study of Nelson et al¹⁰¹ demonstrated that patients with DIC had persistent decreased plasma fibrinogen for several days following delivery and mild to moderately elevated fibrin

degradation products.¹⁰¹ Unlike women with abruption that their hemostatic parameters recover within hours after delivery those with acute fatty liver requires several days for this to happen as a function of the recovery of the synthetic function of their liver. Thus, DIC is a central component of acute fatty liver and in a way reflects the severity of the hepatic injury.

Collectively, the alteration in maternal physiology associated with pregnancy is further augmented by different pregnancy complications leading to perturbation of the hemostatic system and the development of DIC. It is important to understand the underlying mechanisms associated with the development of DIC in each complication as it will assist the clinician in the prevention and management of DIC in these cases.

Diagnosis of DIC in Pregnancy: Clinical Definitions, Laboratory Findings and Scoring Systems

The determination whether obstetrics coagulopathy results from consumption of coagulation factors, their loss due to uncontrolled bleeding, or both. Isolated intravascular consumption of coagulation factor is a true DIC, whereas their loss due to perfused obstetrical hemorrhage can be regarded as dilutional/consumption coagulopathy. However, it is very hard to make such distinction.²⁵

The hallmark of successful management of this dire complication depends on prompt and accurate recognition of DIC. Unfortunately, often the diagnosis of DIC by the attending physicians is based mainly on clinical presentation of the patient and often made relatively late in the course of the disease. During pregnancy, there is a physiologic change in the maternal plasma concentrations of many of the coagulation parameters leading to a false perception that the status of the coagulation system is normal in cases when the patient is already developing coagulopathy. Moreover, often there is underestimation of the amount of bleeding and the relevant laboratory test are performed too late when the patient is already compromised. It is important to emphasize that there is no single laboratory or clinical test that is sensitive and specific enough to diagnose DIC and the risk to develop DIC is not evident in all cases.^{4,9,40,46,50,90,104} DIC is a dynamic situation that requires a continuous assessment of the clinical and laboratory parameters including decreasing concentration of fibrinogen and platelet count, prolongation of prothrombin time, and increased concentration of

fibrin split products or d-dimers^{14,105,106} until it resolves. For those reasons, there is a need to: 1) develop means for early diagnosis of DIC by clinicians; and 2) establish a universal definition of DIC in pregnancy. The development of DIC scores was an effort to address both challenges. Such scores need to be simple, easy to perform, and readily available to clinicians worldwide. Indeed, these scores use simple and readily available coagulation tests including platelet count, prothrombin time (PT) prolongation, fibrinogen, and fibrin split products/D-dimer concentrations.

The use of DIC scoring systems was introduced to facilitate the diagnosis of DIC. The ISTH DIC score was proposed in 2001¹⁰² followed by that of the Japanese Association for Acute Medicine (JAAM) in 2005,¹⁰⁷ both scores have good prediction performance for the diagnosis of DIC and the identification of critically ill non-pregnant patients and their prognosis in intensive care units.^{103,108–110} None of these scores is adjusted for the physiologic hemostatic changes occurring in pregnancy, limiting their applicability during gestation.

Pregnancy is associated with adaptive changes in the coagulation system that are aimed to address the challenges posed by the need to have extra circulatory maternal blood flow through the placental bed. Thus, the mother has to protect herself from a life-threatening bleeding especially during labor and delivery on one hand, and secure a continuous blood flow through the placental bed to nourish the developing fetus on the other hand.^{91,111–114} The adaptive changes in the hemostatic system are observed in the maternal circulation, placental bed and amniotic fluid as

summarized in Table 1. Women with a normal pregnancy have: 1) excessive thrombin generation,^{114,115} 2) increased agonist derived platelets aggregation,^{116,117} 3) two to three fold increase in fibrinogen concentrations; and 4) towards term they experience a 20% to 1000% increase in factors VII, VIII, IX, X, and XII,¹¹⁸ as well as up to 400% increase in von Willebrand factor.¹¹⁸ By contrast, factors XIII and XI concentrations decrease during gestation and those of factors II & V unchanged.¹¹⁹ The excessive thrombin generation observed during normal pregnancy,^{114,115} is supported by the observations of elevation in circulating maternal fibrinopeptide A, prothrombin fragments (PF) 1 and 2, and thrombin–antithrombin (TAT) III complexes during pregnancy,^{91,120–122} especially at the time of and after normal labor^{123,124} and delivery,^{121,124} subsequently, in the course of the puerperium their concentrations decrease.¹²⁴ Pregnancy is also associated with changes in the anticoagulation proteins concentrations. Indeed, there is a 55% decline in free protein S plasma concentration reaching a nadir at birth leading to an increase in resistance to activated protein C^{118,125}. This process is exacerbated by cesarean delivery and infection.^{118,126} The concentrations of PAI-1 increase by 3 to 4-fold during pregnancy while plasma PAI-2 values, which are negligible before pregnancy reach concentrations of 160 mg/L at delivery.¹¹⁸ Thus, pregnancy is associated with increased clotting potential, as well as decreased anticoagulant properties, and fibrinolysis.⁹² In addition to the changes in maternal circulation, pregnancy is associated with changes in the local hemostatic mechanisms. Indeed, there is an increase in decidual and myometrial tissue factor.^{93,127–129}

Table 1 Adaptive Changes of the Coagulation System During Pregnancy

Category	Increased	Decreased	No Change
Systemic changes			
Procoagulant factors	I, V, VII, VIII, IX, X	XI	
Anticoagulation proteins	Soluble TM	PS	PC
Adhesive proteins	vWF		
Fibrinolytic proteins	PAI-1, PAI-2	t-PA	TAFI
Microparticles and antiphospholipid antibodies	MP		ALPA
Local placental changes	TF	TFPI	

Notes: Reprinted from: Brenner B. Haemostatic changes in pregnancy. *Thromb Res.* 2004;114(5–6):409–414.¹⁹⁸ With permission from Elsevier. doi: 10.1016/j.thromres.2004.08.004.

Similarly, changes are observed in the chorioamniotic membranes (mainly the amnion) and in the amniotic fluid.^{91,130–133}

To address the changes of pregnancy in the definition of DIC, Erez et al,⁵ developed a pregnancy specific DIC score by using platelet count, fibrinogen concentrations and the PT difference – relating to the difference between the patient's PT and the laboratory control. This diagnostic performance of this score for DIC at a cutoff of ≥ 26 points were: 88% sensitivity, 96% specificity, a positive likelihood ratio (LR) of 22, and a negative LR of 0.125 (Table 2). To validate the score vs the ISTH DIC score, Erez et al⁵ compared the diagnostic parameters of both scores in women with placental abruption (n= 684); of them, 21.93% (150/684) needed blood products transfusion and 6.29% (43/684) developed DIC. The pregnancy modified DIC score had 88% sensitivity, 96% specificity, while the modified ISTH score had at a cutoff point of 0.5 had 74% sensitivity, and 95% specificity of for the diagnosis of DIC. An independent validations of

these results was reported in a population of French women with obstetrical hemorrhage who were admitted to intensive care unit.¹³⁴ Among women with liver rupture or hematoma, pregnancy specific DIC score >26 was associated with increased blood product transfusions requirement, longer hospitalizations, and lower neonatal 1 and 5 minutes Apgar scores.¹³⁵ Moreover, a retrospective analysis of blood product transfusion requirements in patients with PPH revealed that according to the pregnancy specific DIC score, blood transfusion was unnecessary in 179 of the 279 postpartum women (64.1%) suggesting that its use may prevent unnecessary transfusions and their related risks and complications.¹³⁶ Recently, Clark et al,¹³⁷ suggested an additional modified version of the ISTH DIC score. Erez et al¹³⁸ tested the performance of this suggested scoring system on their cohort and found that this score had 14.9% sensitivity, 99.9% specificity, and a LR+ score of 14.9. Despite the good likelihood ratio score, the DIC score presented by Clark et al¹³⁷ could identify only 13 out of 87 cases of DIC in

Table 2 An Effect of Components of the New DIC Score – Results of Logistic Regression

	Effect of Individual Analytes		Effect of Individual Analytes Adjusted to Other Tests		Assigned Weight ^a
	Relative Risk	p-value	Relative Risk	p-value	
PT difference (seconds)					
<0.5	1.0		1.0		0
0.5–1	12.7	0.031	29.3	<0.001	5
1.0–1.5	27.7	0.005	68.8	<0.001	12
>1.5	60.3	<0.001	558.1	<0.001	25
Platelets ($10^9/L$)					
<50	3.1	0.06	89.2	<0.001	1
50–100	5.2	<0.001	56.2	<0.001	2
100–185	2.9	0.001	12.8	<0.001	1
>185	1.0		1.0		0
Fibrinogen (g/L)					
<3.0	59.0	<0.001	662.9	<0.001	25
3.0–4.0	13.4	<0.001	59.1	<0.001	6
4.0–4.5	2.4	0.320	6.8	0.03	1
>4.5	1.00		1.0		0

Notes: ^aWeight was calculated as relative risk of each of the adjusted factors to the relative risk of a factor with minimal effect. Adapted from: Erez O, Novack L, Beer-Weisel R, et al. DIC score in pregnant women—a population based modification of the International Society on Thrombosis and Hemostasis score. *PLoS One*. 2014;9:e93240.⁵ Copyright: © 2014 Erez et al. Creative Commons Attribution License (<https://creativecommons.org/licenses/by/4.0/legalcode>)

this cohort, casting a heavy shadow on the clinical utility of this score. Thus, following the study by Erez et al⁵ coagulation parameters standards of non-pregnant patients can no longer use for the diagnosis of DIC in pregnant women, as “Normal” values of coagulation tests are different during pregnancy, parturition, and puerperium than in the non-pregnant state (Figure 7).

Other DIC scores in pregnancy were suggested by: 1) Terao et al¹³⁹ in 1987 developed a DIC score that included three categories, etiology, clinical manifestation, and laboratory tests (PT, fibrinogen, FDP, and platelets). The study included 77 women with DIC recruited from 100 centers in Japan. A DIC score ≥ 7 was considered positive and identified 90% (70/77) of the patients. Of note, this score was not validated in comparison to the normal

obstetric population, and it is currently not in clinical use outside of Japan. 2) the utilization of the fibrinogen/C-reactive protein (CRP) ratio as a tool to diagnose the development of DIC among women with HELLP syndrome by Windsperger et al¹⁴⁰ demonstrated a good sensitivity and perform better than fibrinogen concentrations alone in these patients.

The incorporation of pregnancy specific scoring system to diagnose DIC in pregnant women is an accurate and easy to use and may assist clinicians in real time during at the Labor and Delivery wards. However, there is a need for an international consensus on the diagnostic criteria and scoring for DIC in pregnancy, this will facilitate standardization of the definition and will support international research effort in this dire complication of pregnancy.

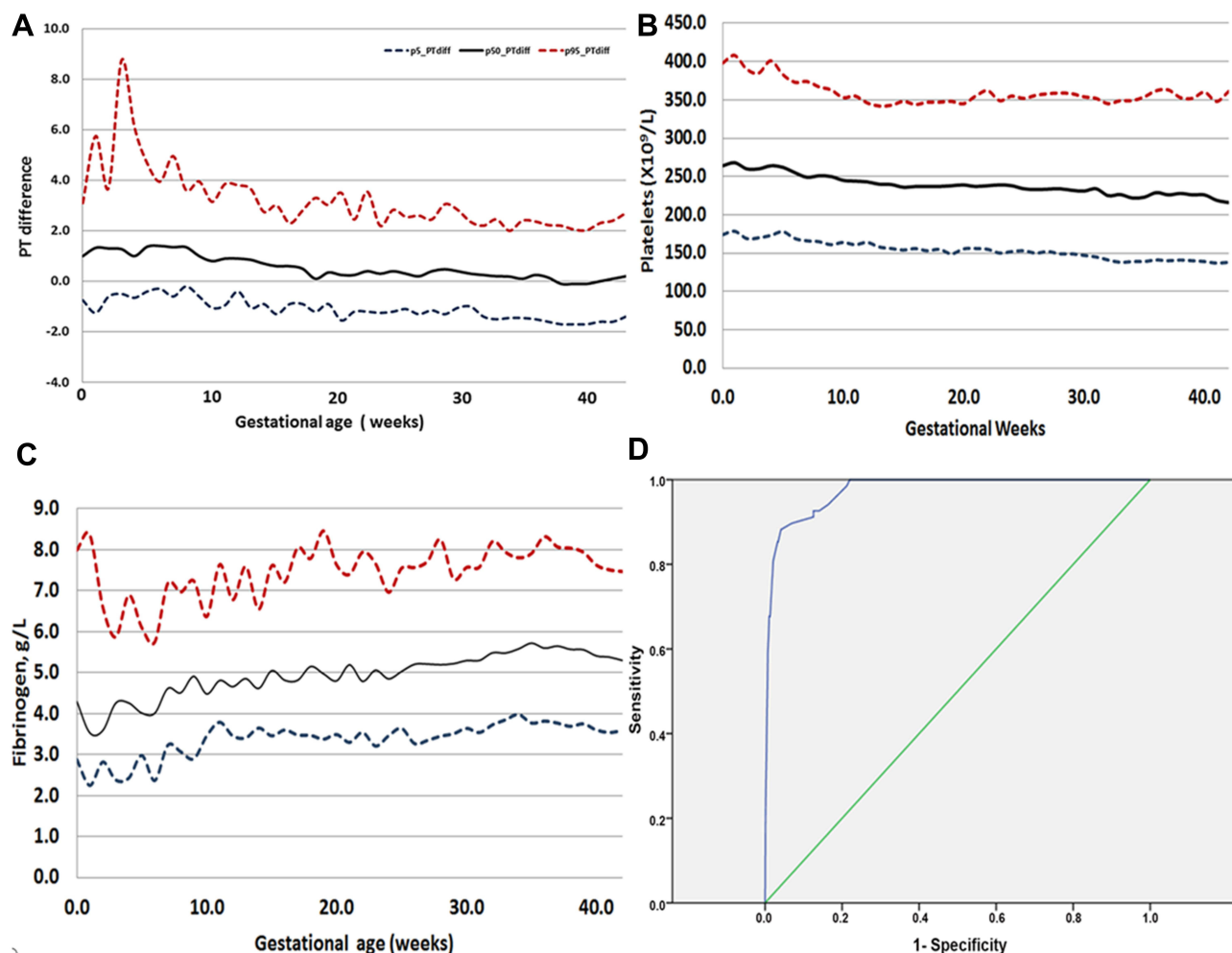


Figure 7 The changes in the major components of the pregnancy modified DIC score: (A) PT difference: (stands for the difference between the patients PT results and the laboratory control); (B) platelets; (C) fibrinogen, with advancing gestations in women with advancing gestation. (D) ROC curve analysis for the association of the pregnancy specific DIC score with the development of DIC.

Notes: Adapted from: Erez O, Novak L, Beer-Weisel et al. DIC score in pregnant women—a population based modification of the International Society on Thrombosis and Hemostasis score. *PLoS One*. 2014;9(4):e93240.⁵ Copyright: © 2014 Erez et al. Creative Commons Attribution License (<https://creativecommons.org/licenses/by/4.0/legalcode>).

Point of Care Testing in the Management of DIC During Pregnancy and Parturition

Standard laboratory tests have limitations, including absence of real-time data and incapacity to determine the functionality of the hemostatic system of whole blood (ie the strength of blood clot, and platelet function). Point of care viscoelastic tests Thromboelastography (TEG); and Rotational Thromboelastometry (ROTEM) are the most widely studied viscoelastic tests which provide a rapid assessment of in vivo coagulation. Specific TEG/ROTEM values were defined for pregnancy^{16,26,84,141,142} and at the time of delivery.¹⁴³ Indeed, in comparison to the non-pregnant state, the characteristics of TEG differ significantly between pregnant and non-pregnant women, displaying a shorter R and K a higher α angle and maximum amplitude suggesting that clot formation is faster and bigger during pregnancy especially during the third trimester.¹⁴⁴ The ROTEM parameters display similar changes in comparison to the non-pregnant state.¹⁴⁵ Moreover, in a study by Rigouzzo et al¹⁴⁶ the TEG parameters (the maximum amplitude of K ≤ 63.5 mm and time to maximum rate of thrombus generation) had excellent diagnostic performance for fibrinogen concentration ≤ 2 g/L and Platelet count $\leq 80,000/\text{mm}^3$ in women with postpartum hemorrhage. Suggesting that TEG can be an excellent point of care testing to identify women with severe PPH at risk to develop DIC. Encouraging data show these tests may enable early detection of dysfunctional coagulation and hyperfibrinolysis,¹⁴⁷ allowing adequate surveillance and prompt intervention.^{14,148} A study of 21 patients classified according to the ISTH DIC score, proposed a thromboelastographic score had 95.2% sensitivity, 81.0% specificity, and an AUC of 0.957 for identifying overt DIC¹⁴⁹ thus allowing, along other diagnostic and prognostic modalities (ie DIC scores), an adequate surveillance and, eventually, a prompt intervention during the early stages of DIC in pregnancy.¹⁵⁰

Treatment of DIC in Pregnancy

Prevention, early diagnosis and treatment of obstetrical hemorrhage are the heart of midwifery.

Prevention of obstetrical hemorrhage is based on identification of patients at risk for peripartum bleeding, active management of the third stage of labor and prompt diagnosis and management of obstetrical complications that may be associated with the development of DIC even in the absence

of labor, the interested reader is referred to the guidelines of the professional committees and selected reviews (ref).

The basic principles for treating obstetrical DIC are presented in Figure 8 and include the following:^{9,14,106,151} 1) treatment and resolution of the underlying condition leading to DIC; 2) fast and prompt delivery or termination of pregnancy (before the threshold of viability). The delivery options should be discussed by a multidisciplinary team and consider the safest mode of delivery to the mother, how fast she is expected to deliver, what are the resources of blood products and other supportive mechanisms available, and can she sustain a surgery; 3) supportive treatment with blood product transfusion, surgical care and related measures; 4) rigorous clinical and laboratory patients surveillance; 5) prompt involvement of needed consultant such as hematologists, gynecological surgeons, anesthesiologists and others; and 6) in small to medium size health care facilities it is important to estimate whether their blood bank can support a massive blood transfusion and, if necessary, contact regional or larger hospitals for assistance or for transferring the patient.

Treatment algorithms for obstetric DIC always involve simultaneous blood product transfusion as a replacement of women's blood loss. Specific transfusion protocols have been previously published and the interested reader is referred to the specific papers on the subject.¹⁵² The harmonized guidance from the ISTH^{151,153} give a fine definition for the thresholds for blood products transfusion, and massive transfusion protocols with fixed RBC: plasma: platelet ratios,^{83,152} are mostly being used, but this recommendation is under debate.¹⁵⁴ The response to blood component therapy should be monitored both clinically and with repeated assessments of the platelet count and coagulation parameters. For further reading, the reader is referred to the published guidelines for blood component replacement from national professional societies^{155–159} and the scientific subcommittees on Women's Health Issues in Thrombosis and Hemostasis and on DIC of the ISTH.^{41,160}

Viscoelastic Hemostatic Assays Guided Transfusion Protocols

During DIC there is a rapid consumption of fibrinogen and coagulation factors, making the monitoring and maintenance of proper blood products administration by conventional laboratory tests challenging, as rapid correction of blood component deficiencies is required at a rate faster than their depletion. ROTEM\TEG allow early detection and dynamic monitoring of clotting abnormalities, and

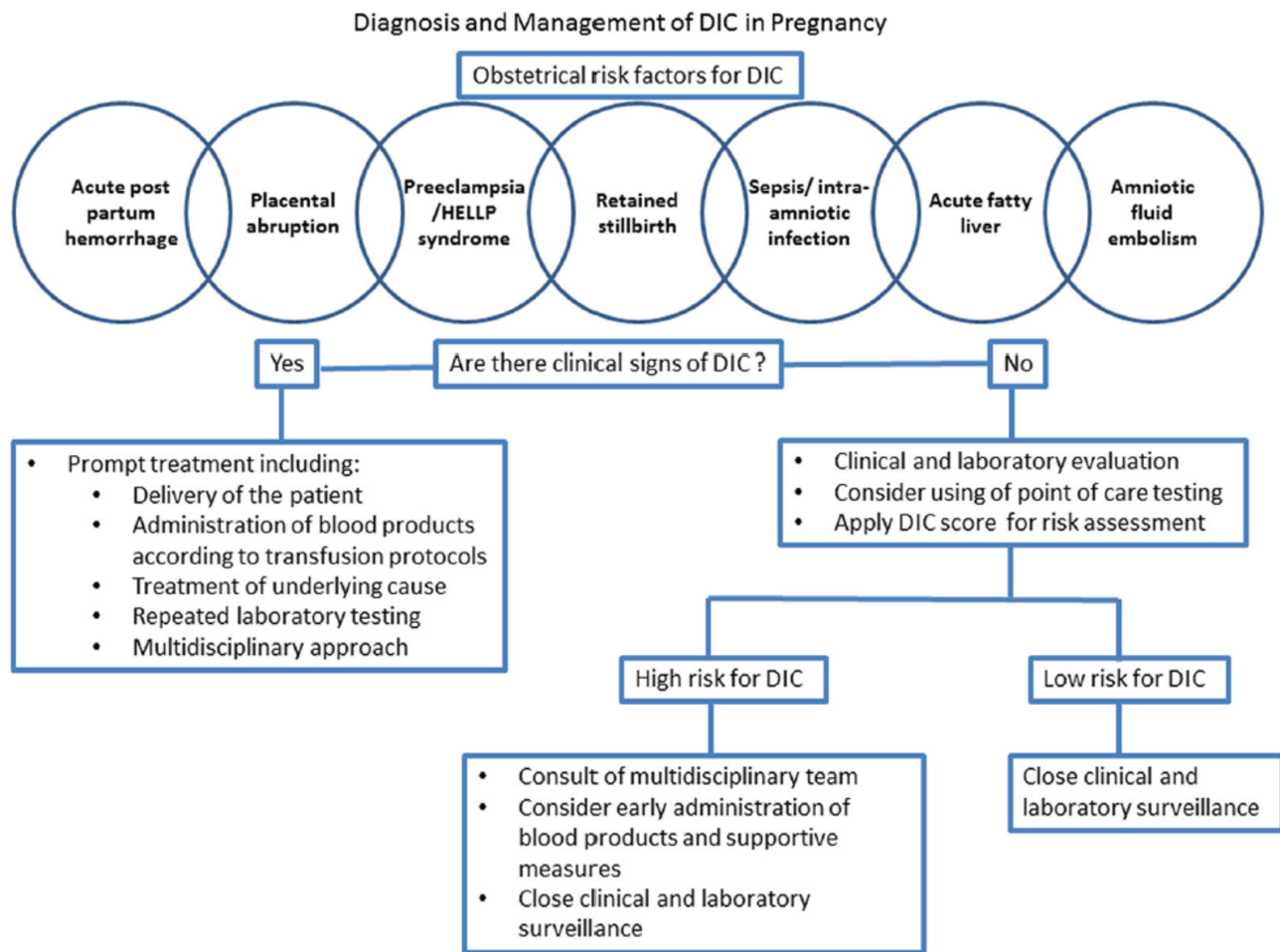


Figure 8 Principles of Diagnosis and management of DIC in pregnancy.

Notes: Reproduced from: Erez O. Disseminated intravascular coagulation in pregnancy - Clinical phenotypes and diagnostic scores. *Thromb Res.* 2017;151203 1:556-560.²⁰⁰ With permission. Copyright © 2017 Elsevier Ltd. All rights reserved.

transfusion needed; hence, they could potentially be used as guidance for administration of blood components. Favorable evidence regarding use of these tests, especially to guide fibrinogen replacement therapy in management of obstetric complications, is accumulating.^{123,147,150,161–166} Further research demonstrated that the use of ROTEM-FIBTEM A5 as a point of care testing for fibrinogen concentration assisted in targeting patients with postpartum hemorrhage who will require blood product transfusion. The rationale for its use was that while Clauss fibrinogen tests results are available within an hour of venipuncture, those of FIBTEM A5 are available within 10 minutes. The authors use a FIBTEM A5 of <15 that can be translated to fibrinogen concentration of 3 grams/Liter or 300mg/Dl as a cutoff point that requires obstetrician attention in women with PPH and developed an algorithm for the management of women with PPH based on the FIBTEM A5 results (Figure 9).^{167–169} These

encouraging results in the pregnant population await further studies before widespread clinical application of VHA scan as point of care testing to guide blood product transfusion in women with PPH can be implemented.^{170–173}

Another possible clinical application of VHAs could be the identification of women who show a severe hypercoagulable state and could benefit from antithrombotic prophylaxis. In a recent case-control study, Spiezia et al evaluated ROTEM profiles in women with preeclampsia in comparison to healthy pregnant women.¹⁷⁴ Characteristic hypercoagulable ROTEM profiles in women with preeclampsia were found.

Treatments for Coagulopathy

Based on the pathogenesis of microvascular failure and coagulation activation in DIC, strategies aimed at the inhibition of coagulation activation have been found

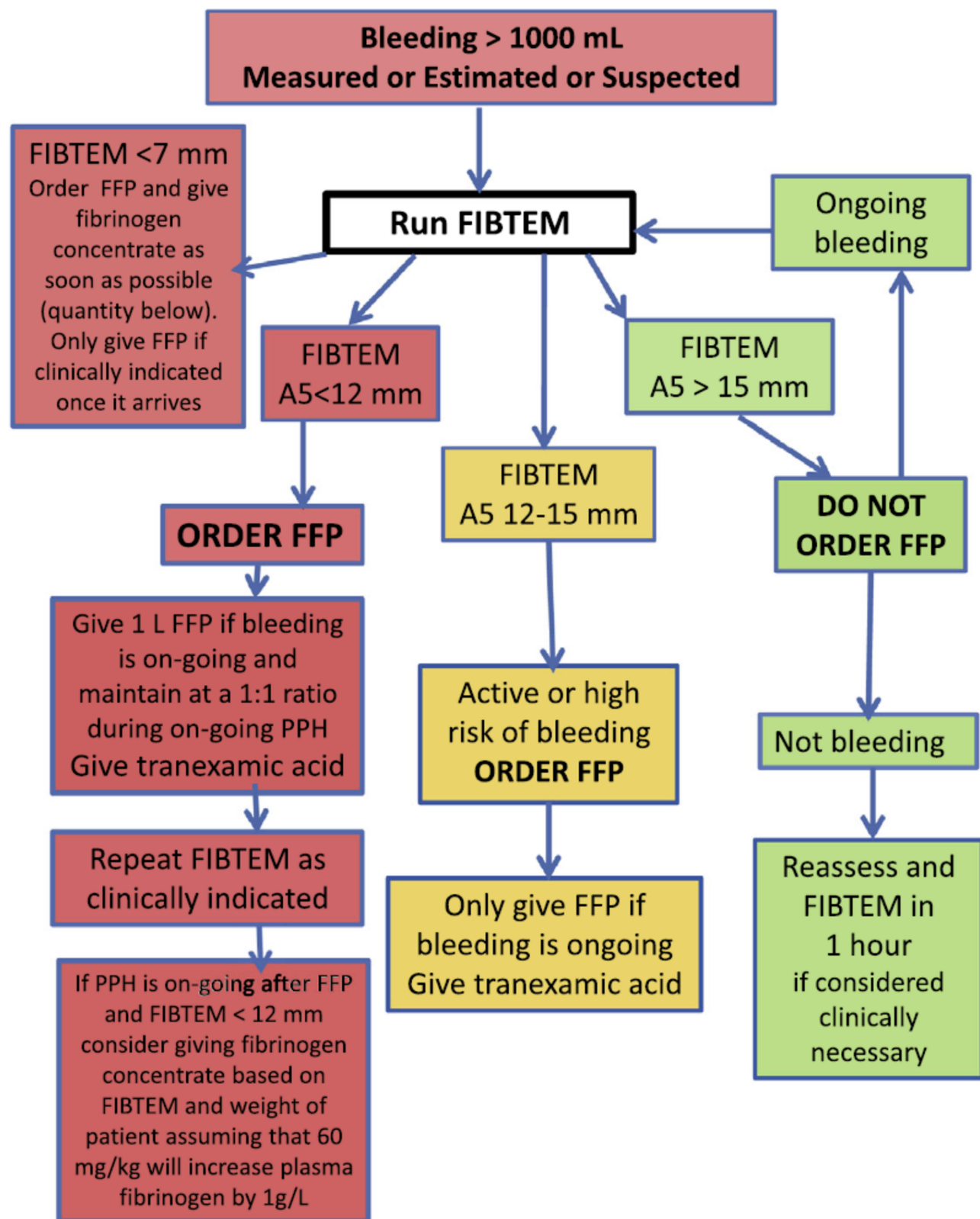


Figure 9 Algorithm for the treatment of obstetrical hemorrhage based on FIBTEM and fibrinogen concentrations.

Notes: Reprinted from: Collins PV, Bell SF, de Lloyd L, Collis RE. Management of postpartum haemorrhage: from research into practice, a narrative review of the literature and the Cardiff experience. *Int J Obstet Anesth.* 2019;37:106–117.¹⁶⁷ With permission Crown Copyright © 2018 Published by Elsevier Ltd. All rights reserved.

favorable in experimental and clinical studies. Most RCTs have been carried out in patients with sepsis, and evidence in obstetric DIC is limited. Tranexamic acid is used as an adjunct to blood product administration and has been studied in PPH. Recombinant activated factor VII (rFVIIa) is used as a second line treatment in women in whom massive transfusion does not halt blood loss. All other treatment modalities are considered experimental at this stage.

Antifibrinolytic Agents

Tranexamic acid (TXA) has been suggested as a treatment for coagulopathy in PPH.¹⁷⁵ TXA prevents the activation of plasminogen by plasmin by blocking its lysine binding sites.¹⁷⁵ Four recent systematic reviews of the use of TXA for reduction of blood loss in PPH came to conflicting results.^{176–179} TXA appears to be a promising drug for the prevention of PPH after cesarean and vaginal

delivery.^{180,181} Patients with the organ failure or non-symptomatic type of DIC may not benefit from antifibrinolytic agents as fibrinolysis is needed for the resolution of widespread fibrin thromboses resulting by DIC.¹⁸² The usual dose is 1gr administered intravenously over 10 minutes up to 4 times daily.¹⁸³

Recombinant Activated Factor VII (rFVIIa)

Guidelines suggest that administration of rFVIIa is warranted in active obstetrical hemorrhage that does not resolve by conventional treatment or to prevent hysterectomy.¹⁶⁰ In such cases, this hemostatic agent decreases maternal mortality due to obstetrical hemorrhage.^{184,185} A review of 99 cases of its use in DIC, of them 32 due to PPH, reported that a median dose of 67.2 mg/kg is successful in controlling the ongoing obstetrical hemorrhage.¹⁸⁶ Optimal use of rFVIIa requires exclusion or correction of metabolic acidosis, hypothermia, hypofibrinogenemia, and thrombocytopenia.¹⁸⁷ A recent RCT, reported

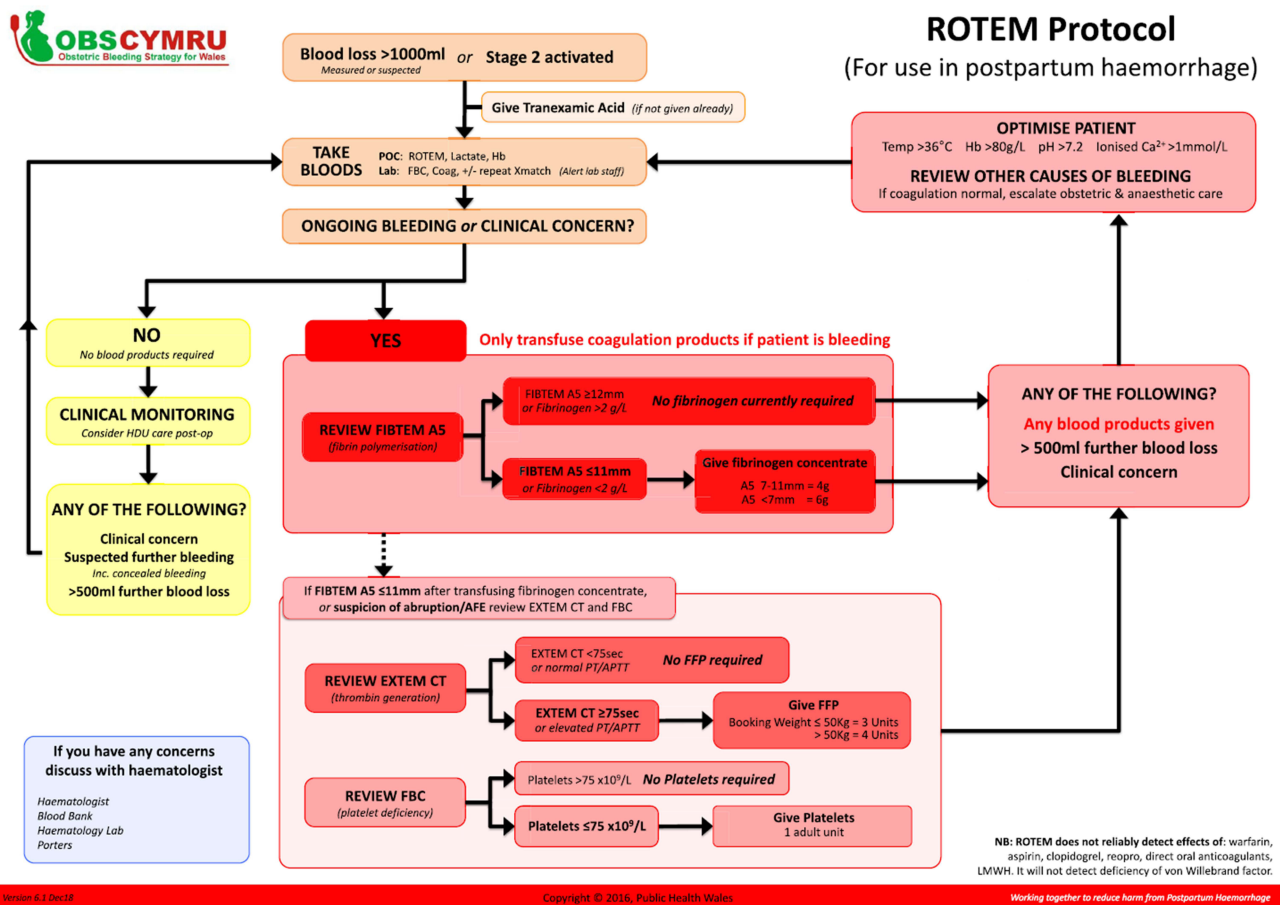


Figure 10 The ROTEM protocol of the OBS Cymru (the Obstetric Bleeding Strategy for Wales) initiative.

Notes: Reproduced with permission from: ROTEM Protocol (For use in postpartum haemorrhage); 2018. Obstetric Bleeding Strategy for Wales. Available from: <https://phw.nhs.wales/services-and-teams/improvement-cymru/improvement-cymru-programmes/maternity-cymru/obs-cymru/obstetric-bleeding-strategy-cymru/rotem-point-of-care-testing>.²⁰¹ Copyright © 2016, Public Health Wales.

that rFVIIa reduces by 30% the need for second-line treatments (interventional hemostatic procedures, blood transfusions).¹⁸⁸ However, as the use of rFVIIa increases by two fold the risk for arterial thrombosis, adequate thromboprophylaxis should be administered to these patients following the acute hemorrhagic event. The optimal dose is unclear, however it is preferable to start with a low dose (40–60mcg/kg) to reduce the risk of thrombotic events. Higher doses (eg 90mcg/kg) can be used if the patient is unstable or if blood loss is brisk and ongoing.

Potential Experimental Treatments

Fibrinogen Concentrate

Human fibrinogen concentrates have been used for substitution therapy in cases of hypofibrinogenemia, dysfibrinogenemia, and afibrinogenemia. This product has the potential to administer relative high quantity of fibrinogen in relatively low volume of transfusion in women with obstetrical hemorrhage. Its benefits are that it is readily available, as it can be kept in room temperature or refrigerators, and it can introduce higher quantity of fibrinogen without the need to transfuse large plasma volume as in FFP transfusion.^{167,189} The obstetric bleeding strategy for Wales (OBSCYMRU) initiative has adopted the use for fibrinogen concentrate at the treatment of choice for women with active obstetrical hemorrhage who has fibrinogen concentrations <2g/L or FIBTEM A5<11mm¹⁶⁷ (Figure 10). However, this Fibrinogen concentrate is not readily available worldwide and the current standard of care for the administration of high concentration of fibrinogen in a low volume transfusion is by cryoprecipitate. Each vial contains approximately 1000mg of fibrinogen, doses are repeated according to TEG/ROTEM guidance.

Desmopressin- 1-Deamino-8-D-Arginine Vasopressin (DDAVP)

The synthetic Anti diuretic hormone analog increases factor VIII concentrations and VWF release from endothelial cells, promoting platelet aggregation and adhesion. DDAVP can reduce bleeding resulting from obstetrical hemorrhage with a good safety profile.¹⁹⁰ The current evidence although limited suggest that DDAVP may be beneficial in DIC resulting from abnormal platelet activity.¹⁹¹ However, it is not part of standard treatment protocols.

Recombinant Human Soluble Thrombomodulin (rhTM)

This agent was reported to potentially lower morbidity and mortality of patients with DIC due to sepsis.^{192–194} This anti-

coagulant reduces excessive thrombin activation and regulates the imbalance of the coagulation system. The administration rhTM to patients with obstetric DIC in a single-center, retrospective cohort study, resulted in improved platelet count, D-dimer and fibrinogen concentrations, and prothrombin time, as well as reduction in the need for platelet transfusions; however, these observations were not associated with a change in the rate of organ failure.¹⁹⁵ There is a need for more substantial evidence prior for the inclusion of rhTM in the treatment protocols for obstetrical DIC.

Anticoagulants

In cases of DIC with predominant hypercoagulation and thrombotic phenotype heparin can partly inhibit coagulation in this setting¹⁹⁶ and low molecular weight heparin (LMWH) is superior to unfractionated heparin (UFH) for treating this type of DIC.¹⁹⁷ However, the majority of cases of DIC in pregnancy are associated with a hemorrhagic phenotype of DIC, and heparin may increase bleeding, especially if adequate replacement therapy for consumed clotting factors has not been achieved, hence it is not recommended.

Conclusions

DIC in obstetrics is a life-threatening complication that is secondary to obstetrical and non-obstetrical related complications of pregnancy. Solid clinical and epidemiologic data is still lacking. The SSC Subcommittees on Disseminated Intravascular Coagulation (DIC) and Women's Issues in Thrombosis and Hemostasis of the International Society for Thrombosis and Hemostasis have launched a global registry of DIC in pregnancy to collect this information. We encourage physicians from across the globe to participate using this link <https://redcap.isth.org/surveys/?s=KFC8RN8XWC>. The diagnosis of DIC can be elusive during pregnancy and requires vigilance and knowledge of the physiologic changes during pregnancy. Pregnancy specific DIC scores and adjustment of the cutoff of point of care testing can facilitate the diagnosis and the management of DIC in pregnancy.

Disclosure

The authors report no conflicts of interest for this work.

References

1. Singh B, Hanson AC, Alhurani R, et al. Trends in the incidence and outcomes of disseminated intravascular coagulation in critically ill patients (2004–2010): a population-based study. *Chest*. 2013;143:1235–1242. doi:10.1378/chest.12-2112

2. Cunningham FG, Nelson DB. Disseminated intravascular coagulation syndromes in obstetrics. *Obstet Gynecol.* 2015;126:999–1011. doi:10.1097/AOG.0000000000001110
3. Gulumser C, Engin-Ustun Y, Keskin L, et al. Maternal mortality due to hemorrhage: population-based study in Turkey. *J Mater Fetal Neonatal Med.* 2019;32:3998–4004. doi:10.1080/14767058.2018.1481029
4. Rattray DD, O'Connell CM, Baskett TF. Acute disseminated intravascular coagulation in obstetrics: a tertiary centre population review (1980 to 2009). *J Obstet Gynaecol Can.* 2012;34:341–347. doi:10.1016/S1701-2163(16)35214-8
5. Erez O, Novack L, Beer-Weisel R, et al. DIC score in pregnant women—a population based modification of the International Society on Thrombosis and Hemostasis score. *PLoS One.* 2014;9:e93240. doi:10.1371/journal.pone.0093240
6. Onishi K, Tsuda H, Fuma K, et al. The impact of the abruption severity and the onset-to-delivery time on the maternal and neonatal outcomes of placental abruption. *J Mater Fetal Neonatal Med.* 2019;33:1–9.
7. Qiu Y, Wu L, Xiao Y, Zhang X. Clinical analysis and classification of placental abruption. *J Mater Fetal Neonatal Med.* 2019;2:1–5.
8. Kilicci C, Ozkaya E, Karakus R, et al. Early low molecular weight heparin for postpartum hemorrhage in women with pre-eclampsia. Is it effective to prevent consumptive coagulopathy? *J Mater Fetal Neonatal Med.* 2020;33:410–414. doi:10.1080/14767058.2018.1494708
9. Thachil J, Toh CH. Disseminated intravascular coagulation in obstetric disorders and its acute haematological management. *Blood Rev.* 2009;23:167–176. doi:10.1016/j.blre.2009.04.002
10. Romero R, Copel JA, Hobbins JC. Intrauterine fetal demise and hemostatic failure: the fetal death syndrome. *Clin Obstet Gynecol.* 1985;28:24–31. doi:10.1097/00003081-198528010-00004
11. Levi M, de Jonge E, van der Poll T, Ten Cate H. Advances in the understanding of the pathogenetic pathways of disseminated intravascular coagulation result in more insight in the clinical picture and better management strategies. *Semin Thromb Hemost.* 2001;27:569–575. doi:10.1055/s-2001-18862
12. Franchini M, Lippi G, Manzato F. Recent acquisitions in the pathophysiology, diagnosis and treatment of disseminated intravascular coagulation. *Thromb J.* 2006;4:4. doi:10.1186/1477-9560-4-4
13. Taylor FB Jr, Toh CH, Hoots WK, Wada H, Levi M. Towards definition, clinical and laboratory criteria, and a scoring system for disseminated intravascular coagulation. *Thromb Haemost.* 2001;86:1327–1330. doi:10.1055/s-0037-1616068
14. Levi M, Toh CH, Thachil J, Watson HG. Guidelines for the diagnosis and management of disseminated intravascular coagulation. *Br J Haematol.* 2009;145:24–33. doi:10.1111/j.1365-2141.2009.07600.x
15. Gando S, Levi M. Disseminated intravascular coagulation. *Nat Rev Dis Primers.* 2016;2:16038. doi:10.1038/nrdp.2016.38
16. Erez O, Mastrolia SA, Thachil J. Disseminated intravascular coagulation in pregnancy: insights in pathophysiology, diagnosis and management. *Am J Obstet Gynecol.* 2015;213:452–463. doi:10.1016/j.ajog.2015.03.054
17. Thachil J. Disseminated Intravascular Coagulation: a Practical Approach. *Anesthesiology.* 2016;125:230–236. doi:10.1097/ALN.0000000000001123
18. Matevosyan K, Sarode R. Thrombosis, Microangiopathies, and Inflammation. *Semin Thromb Hemost.* 2015;41:556–562. doi:10.1055/s-0035-1556587
19. Bockmeyer CL, Claus RA, Budde U, et al. Inflammation-associated ADAMTS13 deficiency promotes formation of ultra-large von Willebrand factor. *Haematologica.* 2008;93:137–140. doi:10.3324/haematol.11677
20. Miodownik S, Pikovsky O, Erez O, Kezerle Y, Lavon O, Rabinovich A. Unfolding the pathophysiology of congenital thrombotic thrombocytopenic purpura in pregnancy: lessons from a cluster of familial cases. *Am J Obstet Gynecol.* 2021;225:177.e1–77.e15. doi:10.1016/j.ajog.2021.02.018
21. Thachil J, Toh CH. Current concepts in the management of disseminated intravascular coagulation. *Thromb Res.* 2012;129 (Suppl 1):S54–9. doi:10.1016/S0049-3848(12)70017-8
22. Erez O, Gotsch F, Mazaki-Tovi S, et al. Evidence of maternal platelet activation, excessive thrombin generation, and high amniotic fluid tissue factor immunoreactivity and functional activity in patients with fetal death. *J Mater Fetal Neonatal Med.* 2009;22:672–687. doi:10.1080/14767050902853117
23. Wada H, Matsumoto T, Yamashita Y, Hatada T. Disseminated intravascular coagulation: testing and diagnosis. *Int J Clin Chem.* 2014;436:130–134.
24. Thachil J. The Elusive Diagnosis of Disseminated Intravascular Coagulation: does a Diagnosis of DIC Exist Anymore? *Semin Thromb Hemost.* 2019;45:100–107. doi:10.1055/s-0038-1677042
25. Thachil J. Disseminated intravascular coagulation - new pathophysiological concepts and impact on management. *Expert Rev Hematol.* 2016;9:803–814. doi:10.1080/17474086.2016.1203250
26. Levi M, Meijers JC. DIC: which laboratory tests are most useful. *Blood Rev.* 2011;25:33–37. doi:10.1016/j.blre.2010.09.002
27. Hossain N, Paidas MJ. Disseminated intravascular coagulation. *Semin Perinatol.* 2013;37:257–266. doi:10.1053/j.semperi.2013.04.008
28. Kobayashi T. Obstetrical disseminated intravascular coagulation score. *J Obstet Gynaecol Res.* 2014;40:1500–1506. doi:10.1111/jog.12426
29. Kor-anantakul O, Lekhakula A. Overt disseminated intravascular coagulation in obstetric patients. *J Med Assoc Thai.* 2007;90:857–864.
30. Creanga AA, Berg CJ, Syverson C, Seed K, Bruce FC, Callaghan WM. Pregnancy-related mortality in the United States, 2006–2010. *Obstet Gynecol.* 2015;125:5–12. doi:10.1097/AOG.0000000000000564
31. Callaghan WM, Creanga AA, Kuklina EV. Severe maternal morbidity among delivery and postpartum hospitalizations in the United States. *Obstet Gynecol.* 2012;120:1029–1036. doi:10.1097/AOG.0b013e31826d60c5
32. Naz H, Fawad A, Islam A, Shahid H, Abbasi AU. Disseminated intravascular coagulation. *J Ayub Med Coll Abbottabad.* 2011;23:111–113.
33. Haram K, Mortensen JH, Mastrolia SA, Erez O. Disseminated intravascular coagulation in the HELLP syndrome: how much do we really know? *J Mater Fetal Neonatal Med.* 2017;30:779–788. doi:10.1080/14767058.2016.1189897
34. Latif E, Adam S, Rungruang B, et al. Use of uterine artery embolization to prevent peripartum hemorrhage of placental abruption with fetal demise & severe DIC. *J Neonatal Perinatal Med.* 2016;9:325–331. doi:10.3233/NPM-16915108
35. Myers J, Wu G, Shapiro RE, Vallejo MC. Preeclampsia Induced Liver Dysfunction Complicated by Disseminated Intravascular Coagulopathy and Placental Abruption: a Case Report and Review of the Literature. *Case Rep Anesthesiol.* 2019;2019:4305849.
36. Ni S, Wang X, Cheng X. The comparison of placental abruption coupled with and without preeclampsia and/or intrauterine growth restriction in singleton pregnancies. *J Mater Fetal Neonatal Med.* 2019;54:1–6.
37. Onishi K, Tsuda H, Fuma K, et al. The impact of the abruption severity and the onset-to-delivery time on the maternal and neonatal outcomes of placental abruption. *J Mater Fetal Neonatal Med.* 2020;33:3775–3783. doi:10.1080/14767058.2019.1585424

38. Takeda J, Takeda S. Management of disseminated intravascular coagulation associated with placental abruption and measures to improve outcomes. *Obstet Gynecol Sci.* 2019;62:299–306. doi:10.5468/ogs.2019.62.5.299
39. Gomez-Tolub R, Rabinovich A, Kachko E. et al. Placental abruption as a trigger of DIC in women with HELLP syndrome: a population-based study. *J Mater Fetal Neonatal Med.* 2020;2:1–11. doi:10.1080/14767058.2020.1818200
40. Montagnana M, Franchi M, Danese E, Gotsch F, Guidi GC. Disseminated intravascular coagulation in obstetric and gynecologic disorders. *Semin Thromb Hemost.* 2010;36:404–418. doi:10.1055/s-0030-1254049
41. Rabinovich A, Abdul-Kadir R, Thachil J, Iba T, Othman M, Erez O. DIC in obstetrics: diagnostic score, highlights in management, and international registry-communication from the DIC and Women's Health SSCs of the International Society of Thrombosis and Haemostasis. *J Thromb Haemost.* 2019;17:1562–1566. doi:10.1111/jth.14523
42. Sher G. Pathogenesis and management of uterine inertia complicating abruptio placentae with consumption coagulopathy. *Am J Obstet Gynecol.* 1977;129:164–170. doi:10.1016/0002-9378(77)90739-6
43. Naccasha N, Gervasi MT, Chaiworapongsa T, et al. Phenotypic and metabolic characteristics of monocytes and granulocytes in normal pregnancy and maternal infection. *Am J Obstet Gynecol.* 2001;185:1118–1123. doi:10.1067/mob.2001.117682
44. Sacks GP, Studena K, Sargent K, Redman CW. Normal pregnancy and preeclampsia both produce inflammatory changes in peripheral blood leukocytes akin to those of sepsis. *Am J Obstet Gynecol.* 1998;179:80–86. doi:10.1016/S0002-9378(98)70254-6
45. Zhu A, Romero R, Huang JB, Clark A, Petty HR. Maltoooligosaccharides from JEG-3 trophoblast-like cells exhibit immunoregulatory properties. *Am J Reprod Immunol.* 2011;65:54–64. doi:10.1111/j.1600-0897.2010.00851.x
46. Romero R, Kadar N, Vaisbuch E, Hassan SS. Maternal death following cardiopulmonary collapse after delivery: amniotic fluid embolism or septic shock due to intrauterine infection? *Am J Reprod Immunol.* 2010;64:113–125.
47. Anas AA, Wiersinga WJ, de Vos AF, van der Poll T. Recent insights into the pathogenesis of bacterial sepsis. *Neth J Med.* 2010;68:147–152.
48. Takai H, Kondoh E, Sato Y, Kakui K, Tatsumi K, Konishi I. Disseminated intravascular coagulation as the presenting sign of gastric cancer during pregnancy. *J Obstet Gynaecol Res.* 2011;37:1717–1719. doi:10.1111/j.1447-0756.2011.01561.x
49. Levi M. Disseminated intravascular coagulation. *Crit Care Med.* 2007;35:2191–2195. doi:10.1097/01.CCM.0000281468.94108.4B
50. Levi M. Pathogenesis and management of peripartum coagulopathic calamities (disseminated intravascular coagulation and amniotic fluid embolism). *Thromb Res.* 2013;131(Suppl 1):S32–4. doi:10.1016/S0049-3848(13)70017-3
51. Levi M, van der Poll T. Inflammation and coagulation. *Crit Care Med.* 2010;38:S26–34. doi:10.1097/CCM.0b013e3181c98d21
52. Seki Y, Wada H, Kawasugi K, et al. A prospective analysis of disseminated intravascular coagulation in patients with infections. *Intern Med.* 2013;52:1893–1898. doi:10.2169/internalmedicine.52.0571
53. Sung JY, Kim JE, Kim KS, Han KS, Kim HK. Differential expression of leukocyte receptors in disseminated intravascular coagulation: prognostic value of low protein C receptor expression. *Thromb Res.* 2014;134:1130–1134. doi:10.1016/j.thromres.2014.08.026
54. Kvolik S, Jukic M, Matijevic M, Marjanovic K, Glavas-Obrovac L. An overview of coagulation disorders in cancer patients. *Surg Oncol.* 2010;19:e33–46. doi:10.1016/j.suronc.2009.03.008
55. Levi M, van der Poll T. Disseminated intravascular coagulation: a review for the internist. *Intern Emerg Med.* 2013;8:23–32. doi:10.1007/s11739-012-0859-9
56. Osterud B. Tissue factor expression by monocytes: regulation and pathophysiological roles. *Blood Coagul Fibrinolysis.* 1998;9 (Suppl 1):S9–14.
57. Levi M, de Jonge E, van der Poll T. Rationale for restoration of physiological anticoagulant pathways in patients with sepsis and disseminated intravascular coagulation. *Crit Care Med.* 2001;29: S90–4. doi:10.1097/00003246-200107001-00028
58. Choi Q, Hong KH, Kim JE, Kim HK. Changes in plasma levels of natural anticoagulants in disseminated intravascular coagulation: high prognostic value of antithrombin and protein C in patients with underlying sepsis or severe infection. *Ann Lab Med.* 2014;34:85–91. doi:10.3343/alm.2014.34.2.85
59. Himmelreich G, Jochum M, Bechstein WO, et al. Mediators of leukocyte activation play a role in disseminated intravascular coagulation during orthotopic liver transplantation. *Transplantation.* 1994;57:354–358.
60. Esmon CT. Role of coagulation inhibitors in inflammation. *Thromb Haemost.* 2001;86:51–56. doi:10.1055/s-0037-1616200
61. Levi M. The imbalance between tissue factor and tissue factor pathway inhibitor in sepsis. *Crit Care Med.* 2002;30:1914–1915. doi:10.1097/00003246-200208000-00046
62. Slofstra SH, Spek CA, Ten Cate H. Disseminated intravascular coagulation. *Hematol J.* 2003;4:295–302. doi:10.1038/sj.thj.6200263
63. Löwenberg EC, Meijers JC, Levi M. Platelet-vessel wall interaction in health and disease. *Neth J Med.* 2010;68:242–251.
64. Furie B, Furie BC. Role of platelet P-selectin and microparticle PSGL-1 in thrombus formation. *Trends Mol Med.* 2004;10:171–178.
65. Vlachodimitropoulou Koumoutsea E, Vivanti AJ, Shehata N, et al. COVID-19 and acute coagulopathy in pregnancy. *J Thromb Haemost.* 2020;18:1648–1652. doi:10.1111/jth.14856
66. Tang N, Li D, Wang X, Sun Z. Abnormal coagulation parameters are associated with poor prognosis in patients with novel coronavirus pneumonia. *J Thrombosis Haemostasis.* 2020;18:844–847. doi:10.1111/jth.14768
67. Kadir RA, Kobayashi T, Iba T, et al. COVID-19 coagulopathy in pregnancy: critical review, preliminary recommendations, and ISTH registry-Communication from the ISTH SSC for Women's Health. *J Thromb Haemost.* 2020;18:3086–3098. doi:10.1111/jth.15072
68. Seppälä M, Wahlström T, Bohn H. Circulating levels and tissue localization of placental protein five (PP5) in pregnancy and trophoblastic disease: absence of PP5 expression in the malignant trophoblast. *Int J Cancer.* 1979;24:6–10. doi:10.1002/ijc.2910240103
69. Obiekwe BC, Chard T. Placental protein 5: circulating levels in twin pregnancy and some observations on the analysis of biochemical data from multiple pregnancy. *Eur J Obstet Gynecol Reprod Biol.* 1981;12:135–141. doi:10.1016/0028-2243(81)90068-X
70. Than G, Bohn H, Szabo D. *Advances in Pregnancy- Related Protein Research.* Boca Raton: CRC Press; 2020.
71. Reverdiau P, Jarousseau AC, Thibault G, et al. Tissue factor activity of syncytiotrophoblast plasma membranes and tumoral trophoblast cells in culture. *Thromb Haemost.* 1995;73:49–54. doi:10.1055/s-0038-1653724
72. Aharon A, Brenner B, Katz T, Miyagi Y, Lanir N. Tissue factor and tissue factor pathway inhibitor levels in trophoblast cells: implications for placental hemostasis. *Thromb Haemost.* 2004;92:776–786. doi:10.1160/TH04-01-0033

73. Hubé F, Reverdiau P, Iochmann S, Trassard S, Thibault G, Gruel Y. Demonstration of a tissue factor pathway inhibitor 2 messenger RNA synthesis by pure villous cytotrophoblast cells isolated from term human placentas. *Biol Reprod.* 2003;68:1888–1894. doi:10.1095/biolreprod.102.011858
74. Udagawa K, Miyagi Y, Hirahara F, et al. Specific expression of PP5/TFPI2 mRNA by syncytiotrophoblasts in human placenta as revealed by in situ hybridization. *Placenta.* 1998;19:217–223. doi:10.1016/S0143-4004(98)90011-X
75. van Wersch JW, Ubachs JM. Blood coagulation and fibrinolysis during normal pregnancy. *Eur J Clin Chem Clin Biochem.* 1991;29:45–50.
76. Robb AO, Mills NL, Din JN, et al. Acute endothelial tissue plasminogen activator release in pregnancy. *J Thromb Haemost.* 2009;7:138–142. doi:10.1111/j.1538-7836.2008.03207.x
77. Oyelese Y, Ananth CV. Placental abruption. *Obstet Gynecol.* 2006;108:1005–1016. doi:10.1097/01.AOG.0000239439.04364.9a
78. Krikun G, Huang ST, Schatz F, Salafia C, Stocco C, Lockwood CJ. Thrombin activation of endometrial endothelial cells: a possible role in intrauterine growth restriction. *Thromb Haemost.* 2007;97:245–253. doi:10.1160/TH06-07-0387
79. Schneider CL. The active principle of placental toxin; thromboplastin; its inactivator in blood; antithromboplastin. *Am J Physiol.* 1947;149:123–129. doi:10.1152/ajplegacy.1947.149.1.123
80. Collins NF, Bloor M, McDonnell NJ. Hyperfibrinolysis diagnosed by rotational thromboelastometry in a case of suspected amniotic fluid embolism. *Int J Obstet Anesth.* 2013;22:71–76. doi:10.1016/j.ijoa.2012.09.008
81. Schneider CL. Fibrin embolism (disseminated intravascular coagulation) with defibrination as one of the end results during placenta abruptio. *Surg Gynecol Obstet.* 1951;92:27–34.
82. Beller FK, Douglas GW, Debrovner CH, Robinson R. The fibrinolytic system in amniotic fluid embolism. *Am J Obstet Gynecol.* 1963;87:48–55. doi:10.1016/S0002-9378(16)35142-0
83. McLintock C, James AH. Obstetric hemorrhage. *J Thromb Haemost.* 2011;9:1441–1451. doi:10.1111/j.1538-7836.2011.04398.x
84. Levi M, Ten Cate H. Disseminated intravascular coagulation. *N Engl J Med.* 1999;341:586–592. doi:10.1056/NEJM199908193410807
85. Weiner CP. The obstetric patient and disseminated intravascular coagulation. *Clin Perinatol.* 1986;13:705–717. doi:10.1016/S0095-5108(18)30794-2
86. Grönvall M, Tikkanen M, Metsätähti M, Loukovaara M, Paavonen J, Stefanovic V. Pelvic arterial embolization in severe obstetric hemorrhage. *Acta Obstet Gynecol Scand.* 2014;93:716–719. doi:10.1111/aogs.12376
87. Rath WH. Postpartum hemorrhage—update on problems of definitions and diagnosis. *Acta Obstet Gynecol Scand.* 2011;90:421–428. doi:10.1111/j.1600-0412.2011.01107.x
88. Knight M. The findings of the MBRRACE-UK confidential enquiry into Maternal Deaths and Morbidity. *Obstet Gynaecol Reproduct Med.* 2019;29:21–23. doi:10.1016/j.ogrm.2018.12.003
89. Green L, Knight M, Seeney FM, et al. The epidemiology and outcomes of women with postpartum haemorrhage requiring massive transfusion with eight or more units of red cells: a national cross-sectional study. *Bjog.* 2016;123:2164–2170. doi:10.1111/1471-0528.13831
90. Levi M. Disseminated intravascular coagulation (DIC) in pregnancy and the peri-partum period. *Thromb Res.* 2009;123(Suppl 2):S63–4. doi:10.1016/S0049-3848(09)70013-1
91. de Boer K, Ten Cate JW, Sturk A, Borm JJ, Treffers PE. Enhanced thrombin generation in normal and hypertensive pregnancy. *Am J Obstet Gynecol.* 1989;160:95–100. doi:10.1016/0002-9378(89)90096-3
92. Lockwood CJ. Pregnancy-associated changes in the hemostatic system. *Clin Obstet Gynecol.* 2006;49:836–843. doi:10.1097/01.grf.0000211952.82206.16
93. Kuczyński J, Uszyński W, Zekanowska E, Soszka T, Uszyński M. Tissue factor (TF) and tissue factor pathway inhibitor (TFPI) in the placenta and myometrium. *Eur J Obstet Gynecol Reprod Biol.* 2002;105:15–19. doi:10.1016/S0301-2115(02)00113-6
94. Beller FK, Ebert C. The coagulation and fibrinolytic enzyme system in pregnancy and in the puerperium. *Eur J Obstet Gynecol Reprod Biol.* 1982;13:177–197. doi:10.1016/0028-2243(82)90028-4
95. Castro MA, Goodwin TM, Shaw KJ, Ouzounian JG, McGehee WG. Disseminated intravascular coagulation and antithrombin III depression in acute fatty liver of pregnancy. *Am J Obstet Gynecol.* 1996;174:211–216.
96. Reyes H, Sandoval L, Wainstein A, et al. Acute fatty liver of pregnancy: a clinical study of 12 episodes in 11 patients. *Gut.* 1994;35:101–106. doi:10.1136/gut.35.1.101
97. Pockros PJ, Peters RL, Reynolds TB. Idiopathic fatty liver of pregnancy: findings in ten cases. *Medicine.* 1984;63:1–11. doi:10.1097/00005792-198401000-00001
98. Riely CA, Latham PS, Romero R, Duffy TP. Acute fatty liver of pregnancy. A reassessment based on observations in nine patients. *Ann Intern Med.* 1987;106:703–706. doi:10.7326/0003-4819-106-5-703
99. Slater DN, Hague WM. Renal morphological changes in idiopathic acute fatty liver of pregnancy. *Histopathology.* 1984;8:567–581. doi:10.1111/j.1365-2559.1984.tb02369.x
100. Vigil-de Gracia P. Acute fatty liver and HELLP syndrome: two distinct pregnancy disorders. *Int J Gynaecol Obstet.* 2001;73:215–220. doi:10.1016/S0020-7292(01)00364-2
101. Nelson DB, Yost NP, Cunningham FG. Hemostatic dysfunction with acute fatty liver of pregnancy. *Obstet Gynecol.* 2014;124:40–46. doi:10.1097/AOG.0000000000000296
102. Taylor FB, Toh CH, Hoots WK, Wada H, Levi M. (ISTH) SSoDICDotISoTaH. Towards definition, clinical and laboratory criteria, and a scoring system for disseminated intravascular coagulation. *Thromb Haemost.* 2001;86:1327–1330.
103. Bakhtiari K, Meijers JC, de Jonge E, Levi M. Prospective validation of the International Society of Thrombosis and Haemostasis scoring system for disseminated intravascular coagulation. *Crit Care Med.* 2004;32:2416–2421. doi:10.1097/01.CCM.0000147769.07699.E3
104. Gasem T, Al Jama FE, Burshaid S, Rahman J, Al Suleiman SA, Rahman MS. Maternal and fetal outcome of pregnancy complicated by HELLP syndrome. *J Matern Fetal Neonatal Med.* 2009;22:1140–1143. doi:10.3109/14767050903019627
105. Gando S, Wada H, Thachil J; (ISTH) SaSCoDotSoTaH. Differentiating disseminated intravascular coagulation (DIC) with the fibrinolytic phenotype from coagulopathy of trauma and acute coagulopathy of trauma-shock (COT/ACOTS). *J Thromb Haemost.* 2013;11:826–835. doi:10.1111/jth.12190
106. Wada H, Thachil J, Di Nisio M, et al. Harmonized guidance for disseminated intravascular coagulation from the International Society on Thrombosis and Haemostasis and the current status of anticoagulant therapy in Japan: a rebuttal. *J Thromb Haemost.* 2013;11(11):2078–2079. doi:10.1111/jth.12366
107. Gando S, Iba T, Eguchi Y, et al. A multicenter, prospective validation of disseminated intravascular coagulation diagnostic criteria for critically ill patients: comparing current criteria. *Crit Care Med.* 2006;34:625–631. doi:10.1097/01.CCM.0000202209.42491.38
108. Angstwurm MW, Dempfle CE, Spannagl M. New disseminated intravascular coagulation score: a useful tool to predict mortality in comparison with Acute Physiology and Chronic Health Evaluation II and Logistic Organ Dysfunction scores. *Crit Care Med.* 2006;34:314–20; quiz 28. doi:10.1097/01.CCM.0000196832.27501.B2

109. Asakura H, Wada H, Okamoto K, et al. Evaluation of haemostatic molecular markers for diagnosis of disseminated intravascular coagulation in patients with infections. *Thromb Haemost.* 2006;95:282–287. doi:10.1160/TH05-04-0286
110. Cavkaytar S, Ugurlu EN, Karaer A, Tapisiz OL, Danisman N. Are clinical symptoms more predictive than laboratory parameters for adverse maternal outcome in HELLP syndrome? *Acta Obstet Gynecol Scand.* 2007;86:648–651. doi:10.1080/00016340601185384
111. Yuen PM, Yin JA, Lao TT. Fibrinopeptide A levels in maternal and newborn plasma. *Eur J Obstet Gynecol Reprod Biol.* 1989;30:239–244. doi:10.1016/0028-2243(89)90007-5
112. Sørensen JD, Secher NJ, Jespersen J. Perturbed (procoagulant) endothelium and deviations within the fibrinolytic system during the third trimester of normal pregnancy. A possible link to placental function. *Acta Obstet Gynecol Scand.* 1995;74:257–261. doi:10.3109/00016349509024445
113. Walker MC, Garner PR, Keely EJ, Rock GA, Reis MD. Changes in activated protein C resistance during normal pregnancy. *Am J Obstet Gynecol.* 1997;177:162–169. doi:10.1016/S0002-9378(97)70456-3
114. Bellart J, Gilibert R, Miralles RM, Monasterio J, Cabero L. Endothelial cell markers and fibrinopeptide A to D-dimer ratio as a measure of coagulation and fibrinolysis balance in normal pregnancy. *Gynecol Obstet Invest.* 1998;46:17–21. doi:10.1159/000009989
115. Chaiworapongsa T, Espinoza J, Yoshimatsu J, et al. Activation of coagulation system in preterm labor and preterm premature rupture of membranes. *J Mater Fetal Neonatal Med.* 2002;11:368–373. doi:10.1080/jmf.11.6.368.373
116. Yoneyama Y, Suzuki S, Sawa R, Otsubo Y, Power GG, Araki T. Plasma adenosine levels increase in women with normal pregnancies. *Am J Obstet Gynecol.* 2000;182:1200–1203. doi:10.1067/mob.2000.104832
117. Sheu JR, Hsiao G, Luk HN, et al. Mechanisms involved in the antiplatelet activity of midazolam in human platelets. *Anesthesiology.* 2002;96:651–658. doi:10.1097/00000542-200203000-00022
118. Bremme KA. Haemostatic changes in pregnancy. *Best Pract Res Clin Haematol.* 2003;16:153–168. doi:10.1016/S1521-6926(03)00021-5
119. Eichinger S, Weltermann A, Philipp K, et al. Prospective evaluation of hemostatic system activation and thrombin potential in healthy pregnant women with and without factor V Leiden. *Thromb Haemost.* 1999;82:1232–1236. doi:10.1055/s-0037-1614366
120. Reinhaller A, Mursch-Edlmayr G, Tatra G. Thrombin-antithrombin III complex levels in normal pregnancy with hyper-tensive disorders and after delivery. *Br J Obstet Gynaecol.* 1990;97:506–510. doi:10.1111/j.1471-0528.1990.tb02520.x
121. Uszyński M. Generation of thrombin in blood plasma of non-pregnant and pregnant women studied through concentration of thrombin-antithrombin III complexes. *Eur J Obstet Gynecol Reprod Biol.* 1997;75:127–131. doi:10.1016/S0301-2115(97)00101-2
122. Reber G, Amiral J, de Moerloose P. Modified antithrombin III levels during normal pregnancy and relationship with prothrombin fragment F1 + 2 and thrombin-antithrombin complexes. *Thromb Res.* 1998;91:45–47. doi:10.1016/S0049-3848(98)00043-7
123. Mallaiah S, Barclay P, Harrod I, Chevannes C, Bhalla A. Introduction of an algorithm for ROTEM-guided fibrinogen concentrate administration in major obstetric haemorrhage. *Anaesthesia.* 2015;70:166–175. doi:10.1111/anae.12859
124. Andersson T, Lorentzen B, Hogdahl H, Clausen T, Mowinckel MC, Abildgaard U. Thrombin-inhibitor complexes in the blood during and after delivery. *Thromb Res.* 1996;82:109–117. doi:10.1016/0049-3848(96)00057-6
125. Ku DH, Arkel YS, Paidas MP, Lockwood CJ. Circulating levels of inflammatory cytokines (IL-1 beta and TNF-alpha), resistance to activated protein C, thrombin and fibrin generation in uncomplicated pregnancies. *Thromb Haemost.* 2003;90:1074–1079. doi:10.1160/TH03-02-0119
126. Faught W, Garner P, Jones G, Ivey B. Changes in protein C and protein S levels in normal pregnancy. *Am J Obstet Gynecol.* 1995;172:147–150. doi:10.1016/0002-9378(95)90104-3
127. Lockwood CJ, Krikun G, Schatz F. The decidua regulates hemostasis in human endometrium. *Semin Reprod Endocrinol.* 1999;17:45–51. doi:10.1055/s-2007-1016211
128. Lockwood CJ, Krikun G, Schatz F. Decidual cell-expressed tissue factor maintains hemostasis in human endometrium. *Ann N Y Acad Sci.* 2001;943:77–88. doi:10.1111/j.1749-6632.2001.tb03793.x
129. Erlich J, Parry GC, Fearn C, et al. Tissue factor is required for uterine hemostasis and maintenance of the placental labyrinth during gestation. *Proc Natl Acad Sci U S A.* 1999;96:8138–8143. doi:10.1073/pnas.96.14.8138
130. Creter D. Amnioplastin: new reagent for coagulation tests. *Lancet.* 1977;2:251. doi:10.1016/S0140-6736(77)92871-9
131. Omsjø IH, Oian P, Maltau JM, Osterud B. Thromboplastin activity in amniotic fluid. *Gynecol Obstet Invest.* 1985;19:1–5. doi:10.1159/000299000
132. Lockwood CJ, Bach R, Guha A, Zhou XD, Miller WA, Nemerson Y. Amniotic fluid contains tissue factor, a potent initiator of coagulation. *Am J Obstet Gynecol.* 1991;165:1335–1341. doi:10.1016/S0002-9378(12)90756-5
133. Uszyński M, Zekanowska E, Uszyński W, Kuczyński J. Tissue factor (TF) and tissue factor pathway inhibitor (TFPI) in amniotic fluid and blood plasma: implications for the mechanism of amniotic fluid embolism. *Eur J Obstet Gynecol Reprod Biol.* 2001;95:163–166. doi:10.1016/S0301-2115(00)00448-6
134. Jonard M, Ducloy-Bouthors AS, Fourrier F. Comparison of two diagnostic scores of disseminated intravascular coagulation in pregnant women admitted to the ICU. *PLoS One.* 2016;11:e0166471. doi:10.1371/journal.pone.0166471
135. Hizkiyahu R, Rabinovich A, Thachil J, et al. Modified ISTH pregnancy-specific DIC score in parturients with liver rupture: population-based case series. *J Mater Fetal Neonatal Med.* 2019;32:2517–2523. doi:10.1080/14767058.2018.1439469
136. Goksever Celik H, Celik E, Ozdemir I, Ozge Savkli A, Sanli K, Gorgen H. Is blood transfusion necessary in all patients with disseminated intravascular coagulation associated postpartum hemorrhage? *J Mater Fetal Neonatal Med.* 2019;32:1004–1008. doi:10.1080/14767058.2017.1397125
137. Clark SL, Romero R, Dildy GA, et al. Proposed diagnostic criteria for the case definition of amniotic fluid embolism in research studies. *Am J Obstet Gynecol.* 2016;215:408–412. doi:10.1016/j.ajog.2016.06.037
138. Erez O. Proposed diagnostic criteria for the case definition of amniotic fluid embolism in research studies. *Am J Obstet Gynecol.* 2017;217:228–229. doi:10.1016/j.ajog.2017.04.009
139. Terao T, Maki M, Ikenoue T. A prospective study in 38 patients with abruptio placentae of 70 cases complicated by DIC. *Asia Oceania J Obstet Gynaecol.* 1987;13:1–13. doi:10.1111/j.1447-0756.1987.tb00001.x
140. Windsperger K, Lehner R. The fibrinogen/CRP ratio as a new parameter for the diagnosis of disseminated intravascular coagulation in patients with HELLP syndrome and as a predictive factor for neonatal outcome. *Am J Obstet Gynecol.* 2013;208:118.e1–7. doi:10.1016/j.ajog.2012.11.025

141. Bick RL. Disseminated intravascular coagulation: objective clinical and laboratory diagnosis, treatment, and assessment of therapeutic response. *Semin Thromb Hemost.* 1996;22:69–88. doi:10.1055/s-2007-998993
142. Toh CH, Dennis M. Disseminated intravascular coagulation: old disease, new hope. *BMJ.* 2003;327:974–977. doi:10.1136/bmj.327.7421.974
143. Mallaiah S, Barclay P, Harrod I, Chevannes C, Bhalla A. Introduction of an algorithm for ROTEM-guided fibrinogen concentrate administration in major obstetric haemorrhage. *Anaesthesia.* 2014;1:5874.
144. Yang J, Yang H, Tang A, et al. Trimester-specific reference intervals for kaolin-activated thromboelastography (TEG[®]) in healthy Chinese pregnant women. *Thromb Res.* 2019;184:81–85. doi:10.1016/j.thromres.2019.09.030
145. Shamshirsaz AA, Fox KA, Erfani H, et al. Trimester-specific thromboelastic values and coagulation activation markers in pregnancy compared across trimesters and compared to the nonpregnant state. *Int J Lab Hematol.* 2021;1:5487. doi:10.1111/ijlh.13472
146. Rigouzzo A, Louvet N, Favier R, et al. Assessment of coagulation by thromboelastography during ongoing postpartum hemorrhage: a retrospective cohort analysis. *Anesth Analg.* 2020;130:416–425. doi:10.1213/ANE.0000000000004422
147. Karlsson O, Jeppsson A, Hellgren M. Major obstetric haemorrhage: monitoring with thromboelastography, laboratory analyses or both? *Int J Obstet Anesth.* 2014;23:10–17. doi:10.1016/j.ijoa.2013.07.003
148. Downey C, Kazmi R, Toh CH. Early identification and prognostic implications in disseminated intravascular coagulation through transmittance waveform analysis. *Thromb Haemost.* 1998;80:65–69. doi:10.1055/s-0037-1615140
149. Sharma P, Saxena R. A novel thromboelastographic score to identify overt disseminated intravascular coagulation resulting in a hypocoagulable state. *Am J Clin Pathol.* 2010;134:97–102. doi:10.1309/AJCPPZ4J6CAFYDVM
150. de Lange NM, Lance MD, de Groot R, Beckers EA, Henskens YM, Scheepers HC. Obstetric hemorrhage and coagulation: an update. Thromboelastography, thromboelastometry, and conventional coagulation tests in the diagnosis and prediction of postpartum hemorrhage. *Obstet Gynecol Surv.* 2012;67:426–435. doi:10.1097/OGX.0b013e3182605861
151. Collis RE, Collins PW. Haemostatic management of obstetric haemorrhage. *Anaesthesia.* 2015;70 Suppl 1:78–86, e27–e28. doi:10.1111/anae.12913
152. Abdul-Kadir R, McLintock C, Ducloy AS, et al. Evaluation and management of postpartum hemorrhage: consensus from an international expert panel. *Transfusion.* 2014;54:1756–1768. doi:10.1111/trf.12550
153. Wada H, Thachil J, Di Nisio M, Kurosawa S, Gando S, Toh CH. Harmonized guidance for disseminated intravascular coagulation from the International Society on Thrombosis and Haemostasis and the current status of anticoagulant therapy in Japan: a rebuttal. *J Thromb Haemost.* 2013;11:2078–2079.
154. Dzik WH, Blajchman MA, Fergusson D, et al. Clinical review: Canadian national advisory committee on blood and blood products—massive transfusion consensus conference 2011: report of the panel. *Crit Care.* 2011;15:242. doi:10.1186/cc10498
155. American College of Obstetricians and Gynecologists. ACOG Practice Bulletin: clinical Management Guidelines for Obstetrician-Gynecologists Number 76, October 2006: postpartum hemorrhage. *Obstet Gynecol.* 2006;108:1039–1047.
156. WHO. *WHO Recommendations for the Prevention and Treatment of Postpartum Haemorrhage.* WHO edGeneva: WHO; 2012.
157. Gynaecologists RAaNZCoOa. *Management of Postpartum Haemorrhage (PPH).* 2017:1–17
158. Marshall AL, Durani U, Bartley A, et al. The impact of postpartum hemorrhage on hospital length of stay and inpatient mortality: a National Inpatient Sample-based analysis. *Am J Obstet Gynecol.* 2017;217:344.e1–44.e6. doi:10.1016/j.ajog.2017.05.004
159. Leduc D, Senikas V, Lalonde AB. No. 235-Active Management of the Third Stage of Labour: prevention and Treatment of Postpartum Hemorrhage. *J Obstet Gynaecol Can.* 2018;40: e841–e855. doi:10.1016/j.jogc.2018.09.024
160. Collins P, Abdul-Kadir R, Thachil J. Management of coagulopathy associated with postpartum hemorrhage: guidance from the SSC of the ISTH. *J Thromb Haemost.* 2016;14:205–210. doi:10.1111/jth.13174
161. Solomon C, Collis RE, Collins PW. Haemostatic monitoring during postpartum haemorrhage and implications for management. *Br J Anaesth.* 2012;109:851–863. doi:10.1093/bja/aes361
162. Huissoud C, Carrabin N, Audibert F, et al. Bedside assessment of fibrinogen level in postpartum haemorrhage by thrombelastometry. *BJOG.* 2009;116:1097–1102. doi:10.1111/j.1471-0528.2009.02187.x
163. Hill JS, Devenie G, Powell M. Point-of-care testing of coagulation and fibrinolytic status during postpartum haemorrhage: developing a thrombelastographyI-guided transfusion algorithm. *Anaesth Intensive Care.* 2012;40:1007–1015. doi:10.1177/0310057X1204000612
164. Chitlur M, Lusher J. Standardization of thromboelastography: values and challenges. *Semin Thromb Hemost.* 2010;36:707–711. doi:10.1055/s-0030-1265287
165. Collins PW, Lilley G, Bruynseels D, et al. Fibrin-based clot formation as an early and rapid biomarker for progression of postpartum hemorrhage: a prospective study. *Blood.* 2014;124:1727–1736. doi:10.1182/blood-2014-04-567891
166. McNamara H, Mallaiah S, Barclay P, Chevannes C, Bhalla A. Coagulopathy and placental abruption: changing management with ROTEM-guided fibrinogen concentrate therapy. *Int J Obstet Anesth.* 2015;24:174–179. doi:10.1016/j.ijoa.2014.12.005
167. Collins PW, Bell SF, de Lloyd L, Collis RE. Management of postpartum haemorrhage: from research into practice, a narrative review of the literature and the Cardiff experience. *Int J Obstet Anesth.* 2019;37:106–117. doi:10.1016/j.ijoa.2018.08.008
168. Collins PW, Cannings-John R, Bruynseels D, et al. Viscoelastometry guided fresh frozen plasma infusion for postpartum haemorrhage: OBS2, an observational study. *Br J Anaesth.* 2017;119:422–434. doi:10.1093/bja/aex245
169. Collins PW, Cannings-John R, Bruynseels D, et al. Viscoelastometric-guided early fibrinogen concentrate replacement during postpartum haemorrhage: OBS2, a double-blind randomized controlled trial. *Br J Anaesth.* 2017;119:411–421. doi:10.1093/bja/aex181
170. Amgalan A, Allen T, Othman M, Ahmadzia HK. Systematic review of viscoelastic testing (TEG/ROTEM) in obstetrics and recommendations from the women's SSC of the ISTH. *J Thromb Haemost.* 2020;18:1813–1838. doi:10.1111/jth.14882
171. Amgalan A, Othman M. TEG in pregnancy: current challenges and future potential applications. *Thromb Res.* 2020;191:140–141. doi:10.1016/j.thromres.2020.03.017
172. Favalaro EJ, Amgalan A, Allen T, Othman M, Ahmadzia HK. “Systematic review of viscoelastic testing (TEG/ROTEM) in obstetrics and recommendation from the women's SSC of the ISTH”: response to comment from Kitchen et al. *J Thromb Haemost.* 2020;18:2420–2422. doi:10.1111/jth.15007
173. Othman M, Han K, Elbatarny M, Abdul-Kadir R. The use of viscoelastic hemostatic tests in pregnancy and puerperium: review of the current evidence - communication from the Women's Health SSC of the ISTH. *J Thromb Haemost.* 2019;17:1184–1189. doi:10.1111/jth.14461

174. Spiezia L, Bogana G, Campello E, et al. Whole blood thromboelastometry profiles in women with preeclampsia. *Clin Chem Lab Med.* 2015;53:1793–1798. doi:10.1515/ccm-2014-1128
175. Annecke T, Geisenberger T, Kürzl R, Penning R, Heindl B. Algorithm-based coagulation management of catastrophic amniotic fluid embolism. *Blood Coagul Fibrinolysis.* 2010;21:95–100. doi:10.1097/MBC.0b013e328332cfe2
176. Alam A, Choi S. Prophylactic Use of Tranexamic Acid for Postpartum Bleeding Outcomes: a Systematic Review and Meta-Analysis of Randomized Controlled Trials. *Transfus Med Rev.* 2015;29:231–241. doi:10.1016/j.tmr.2015.07.002
177. Novikova N, Hofmeyr GJ, Cluver C. Tranexamic acid for preventing postpartum haemorrhage. *Cochrane Database Syst Rev.* 2015;2:CD007872.
178. Sentilhes L, Lasocki S, Ducloy-Bouthors AS, et al. Tranexamic acid for the prevention and treatment of postpartum haemorrhage. *Br J Anaesth.* 2015;114:576–587. doi:10.1093/bja/aeu448
179. Simonazzi G, Bisulli M, Saccone G, Moro E, Marshall A, Berghella V. Tranexamic acid for preventing postpartum blood loss after cesarean delivery: a systematic review and meta-analysis of randomized controlled trials. *Acta Obstet Gynecol Scand.* 2016;95:28–37. doi:10.1111/aogs.12798
180. Ahmadzia HK, Phillips JM, Katler QS, James AH. Tranexamic acid for prevention and treatment of postpartum hemorrhage: an update on management and clinical outcomes. *Obstet Gynecol Surv.* 2018;73:587–594. doi:10.1097/OGX.0000000000000597
181. Ducloy-Bouthors AS, Duhamel A, Kipnis E, et al. Postpartum haemorrhage related early increase in D-dimers is inhibited by tranexamic acid: haemostasis parameters of a randomized controlled open labelled trial. *Br J Anaesth.* 2016;116:641–648. doi:10.1093/bja/aew021
182. Mannucci PM, Levi M. Prevention and treatment of major blood loss. *N Engl J Med.* 2007;356:2301–2311. doi:10.1056/NEJMra067742
183. Shakur H, Roberts I, Fawole B, et al. Effect of early tranexamic acid administration on mortality, hysterectomy, and other morbidities in women with post-partum haemorrhage (WOMAN): an international, randomised, double-blind, placebo-controlled trial. *Lancet.* 2017;389:2105–2116. doi:10.1016/S0140-6736(17)30638-4
184. Franchini M, Lippi G, Franchi M. The use of recombinant activated factor VII in obstetric and gynaecological haemorrhage. *BJOG.* 2007;114:8–15. doi:10.1111/j.1471-0528.2006.01156.x
185. Alfirevic Z, Elbourne D, Pavord S, et al. Use of recombinant activated factor VII in primary postpartum hemorrhage: the Northern European registry 2000–2004. *Obstet Gynecol.* 2007;110:1270–1278. doi:10.1097/01.AOG.0000288515.48066.99
186. Franchini M, Manzato F, Salvagno GL, Lippi G. Potential role of recombinant activated factor VII for the treatment of severe bleeding associated with disseminated intravascular coagulation: a systematic review. *Blood Coagul Fibrinolysis.* 2007;18:589–593. doi:10.1097/MBC.0b013e32822d2a3c
187. Franchini M, Franchi M, Bergamini V, Salvagno GL, Montagnana M, Lippi G. A critical review on the use of recombinant factor VIIa in life-threatening obstetric postpartum hemorrhage. *Semin Thromb Hemost.* 2008;34:104–112. doi:10.1055/s-2008-1066022
188. Lavigne-Lissalde G, Aya AG, Mercier FJ, et al. Recombinant human FVIIa for reducing the need for invasive second-line therapies in severe refractory postpartum hemorrhage: a multicenter, randomized, open controlled trial. *J Thromb Haemost.* 2015;13:520–529. doi:10.1111/jth.12844
189. Wikkelsø AJ, Edwards HM, Afshari A, et al. Pre-emptive treatment with fibrinogen concentrate for postpartum haemorrhage: randomized controlled trial. *Br J Anaesth.* 2015;114:623–633. doi:10.1093/bja/aeu444
190. Trigg DE, Stergiotou I, Peitsidis P, Kadir RA. A systematic review: the use of desmopressin for treatment and prophylaxis of bleeding disorders in pregnancy. *Haemophilia.* 2012;18:25–33. doi:10.1111/j.1365-2516.2011.02573.x
191. Ekelund K, Hanke G, Stensballe J, Wikkelsøe A, Albrechtsen CK, Afshari A. Hemostatic resuscitation in postpartum hemorrhage - a supplement to surgery. *Acta Obstet Gynecol Scand.* 2015;94:680–692. doi:10.1111/aogs.12607
192. Vincent JL, Ramesh MK, Ernest D, et al. A randomized, double-blind, placebo-controlled, Phase 2b study to evaluate the safety and efficacy of recombinant human soluble thrombomodulin, ART-123, in patients with sepsis and suspected disseminated intravascular coagulation. *Crit Care Med.* 2013;41:2069–2079. doi:10.1097/CCM.0b013e31828e9b03
193. Saito H, Maruyama I, Shimazaki S, et al. Efficacy and safety of recombinant human soluble thrombomodulin (ART-123) in disseminated intravascular coagulation: results of a Phase III, randomized, double-blind clinical trial. *J Thromb Haemost.* 2007;5:31–41. doi:10.1111/j.1538-7836.2006.02267.x
194. Yamakawa K, Fujimi S, Mohri T, et al. Treatment effects of recombinant human soluble thrombomodulin in patients with severe sepsis: a historical control study. *Crit Care.* 2011;15:R123. doi:10.1186/cc10228
195. Yoshihara M, Uno K, Tano S, et al. The efficacy of recombinant human soluble thrombomodulin for obstetric disseminated intravascular coagulation: a retrospective study. *Crit Care.* 2015;19:369. doi:10.1186/s13054-015-1086-3
196. Pernerstorfer T, Hollenstein U, Hansen J, et al. Heparin blunts endotoxin-induced coagulation activation. *Circulation.* 1999;100:2485–2490. doi:10.1161/01.CIR.100.25.2485
197. Sakuragawa N, Hasegawa H, Maki M, Nakagawa M, Nakashima M. Clinical evaluation of low-molecular-weight heparin (FR-860) on disseminated intravascular coagulation (DIC)—a multicenter cooperative double-blind trial in comparison with heparin. *Thromb Res.* 1993;72:475–500. doi:10.1016/0049-3848(93)90109-2
198. Brenner B. Haemostatic changes in pregnancy. *Thromb Res.* 2004;114(5–6):409–414. doi:10.1016/j.thromres.2004.08.004
199. Hunt BJ. Bleeding and Coagulopathies in Critical Care. *N Engl J Med.* 2014;370:847–859. doi:10.1056/NEJMra1208626
200. Erez O. Disseminated intravascular coagulation in pregnancy - Clinical phenotypes and diagnostic scores. *Thromb Res.* 2017;151(Suppl 1):S56–S60. doi:10.1016/S0049-3848(17)30069-5
201. ROTEM Protocol (For use in postpartum haemorrhage); 2018. Obstetric Bleeding Strategy for Wales. Available from: <https://phw.nhs.wales/services-and-teams/improvement-cymru/improvement-cymru-programmes/maternity-cymru/obs-cymru/obstetric-bleeding-strategy-cymru/rotem-point-of-care-testing>. Accessed September 25, 2021.

Journal of Blood Medicine

Dovepress

Publish your work in this journal

The Journal of Blood Medicine is an international, peer-reviewed, open access, online journal publishing laboratory, experimental and clinical aspects of all aspect pertaining to blood based medicine including but not limited to: Transfusion Medicine; Blood collection, Donor issues, Transmittable diseases, and Blood banking logistics; Immunohematology; Artificial and alternative blood based

therapeutics; Hematology; Biotechnology/nanotechnology of blood related medicine; Legal aspects of blood medicine; Historical perspectives. The manuscript management system is completely online and includes a very quick and fair peer-review system. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Submit your manuscript here: <http://www.dovepress.com/journal-of-blood-medicine-journal>